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- (54) Title: COMPOSITIONS AND METHODS FOR USE IN RECOMBINATIONAL CLONING OF NUCLEIC ACIDS
(54) Titre: COMPOSITIONS ET METHODES DE CLONAGE RECOMBINATOIRE D'ACIDES NUCLEIQUES ACIDS

(57) Abstract

The present invention relates generally to compositions and methods for use in recombinational cloning of nucleic acid molecules. In particular, the invention relates to nucleic acid molecules encoding one or more recombination sites or portions thereof, to nucleic acid molecules comprising one or more of these recombination site nucleotide sequences and optionally comprising one or more additional physical or functional nucleotide sequences. The invention also relates to vectors comprising the nucleic acid molecules of the invention, to host cells comprising the vectors or nucleic acid molecules of the invention, to methods of producing polypeptides using the nucleic acid molecules of the invention, and to polypeptides encoded by these nucleic acid molecules or produced by the methods of the invention. The invention also relates to antibodies that bind to one or more polypeptides of the invention or epitopes thereof. The invention also relates to the use of these compositions in methods for recombinational cloning of nucleic acids, in vitro and in vivo, to provide chimeric DNA molecules that have particular characteristics and/or DNA segments.

(57) Abrégé

La présente invention concerne de façon générale des compositions et des méthodes de clonage recombinatoire de molécules d'acide nucléique. Elle concerne en particulier des molécules d'acide nucléique codant tout ou partie d'un ou de plusieurs sites de recombinaison, et concerne également des molécules d'acide nucléique comprenant une ou plusieurs de ces séquences nucléotidiques de sites de recombinaison, et comprenant éventuellement une ou plusieurs séquences nucléotidiques supplémentaires fonctionnelles ou physiques. En outre, cette invention concerne des vecteurs comprenant ces molécules d'acide nucléique, des cellules hôtes comprenant ces vecteurs ou ces molécules d'acide nucléique, des méthodes de production de polypeptides au moyen de ces molécules d'acide nucléique, ainsi que des polypeptides codés par ces molécules d'acide nucléique ou produits à l'aide des méthodes de l'invention. Par ailleurs, cette invention concerne des anticorps se liant à un ou plusieurs polypeptides de l'invention ou à des épitopes de ceux-ci. L'invention concerne également l'utilisation de ces compositions dans des méthodes de clonage recombinatoire d'acides nucléiques, in vitro et in vivo, afin d'obtenir des molécules d'ADN chimères présentant des caractéristiques et/ou segments d'ADN particuliers.

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**Compositions and Methods for Use in
Recombinational Cloning of Nucleic Acids**

BACKGROUND OF THE INVENTION

Field of the Invention

The present invention relates generally to recombinant DNA technology. More particularly, the present invention relates to compositions and methods for use in recombinational cloning of nucleic acid molecules. The invention relates specifically to nucleic acid molecules encoding one or more recombination sites of one or more partial recombination sites, particularly *attB*, *attP*, *attL*, and *attR*, and fragments, mutants, variants and derivatives thereof. The invention also relates to such nucleic acid molecules wherein the one or more recombination site nucleotide sequences is operably linked to the one or more additional physical or functional nucleotide sequences. The invention also relates to vectors comprising the nucleic acid molecules of the invention, to host cells comprising the vectors or nucleic acid molecules of the invention, to methods of producing polypeptides and RNAs encoded by the nucleic acid molecules of the invention, and to polypeptides encoded by these nucleic acid molecules or produced by the methods of the invention, which may be fusion proteins. The invention also relates to antibodies that bind to one or more polypeptides of the invention or epitopes thereof, which may be monoclonal or polyclonal antibodies. The invention also relates to the use of these nucleic acid molecules, vectors, polypeptides and antibodies in methods for recombinational cloning of nucleic acids, *in vitro* and *in vivo*, to provide chimeric DNA molecules that have particular characteristics and/or DNA segments. More particularly, the antibodies of the invention may be used to identify and/or purify proteins or fusion proteins encoded by the nucleic acid molecules or vectors of the invention, or to identify and/or purify the nucleic acid molecules of the invention.

Related Art

Site-specific recombinases. Site-specific recombinases are proteins that are present in many organisms (e.g. viruses and bacteria) and have been characterized to have both endonuclease and ligase properties. These recombinases (along with associated proteins in some cases) recognize specific sequences of bases in DNA and exchange the DNA segments flanking those segments. The recombinases and associated proteins are collectively referred to as "recombination proteins" (see, e.g., Landy, A., *Current Opinion in Biotechnology* 3:699-707 (1993)).

Numerous recombination systems from various organisms have been described. See, e.g., Hoess *et al.*, *Nucleic Acids Research* 14(6):2287 (1986); Abremski *et al.*, *J. Biol. Chem.* 261(1):391 (1986); Campbell, *J. Bacteriol.* 174(23):7495 (1992); Qian *et al.*, *J. Biol. Chem.* 267(11):7794 (1992); Araki *et al.*, *J. Mol. Biol.* 225(1):25 (1992); Maeser and Kahnmann *Mol. Gen. Genet.* 230:170-176 (1991); Esposito *et al.*, *Nucl. Acids Res.* 25(18):3605 (1997).

Many of these belong to the integrase family of recombinases (Argos *et al.* *EMBO J.* 5:433-440 (1986); Voznyanov *et al.*, *Nucl. Acids Res.* 27:930 (1999)). Perhaps the best studied of these are the Integrase/*att* system from bacteriophage λ (Landy, A. *Current Opinions in Genetics and Devel.* 3:699-707 (1993)), the Cre/*loxP* system from bacteriophage P1 (Hoess and Abremski (1990) In *Nucleic Acids and Molecular Biology*, vol. 4. Eds.: Eckstein and Lilley, Berlin-Heidelberg: Springer-Verlag; pp. 90-109), and the FLP/FRT system from the *Saccharomyces cerevisiae* 2 μ circle plasmid (Broach *et al.* *Cell* 29:227-234 (1982)).

Backman (U.S. Patent No. 4,673,640) discloses the *in vivo* use of λ recombinase to recombine a protein producing DNA segment by enzymatic site-specific recombination using wild-type recombination sites *attB* and *attP*.

Hasan and Szybalski (*Gene* 56:145-151 (1987)) discloses the use of λ Int recombinase *in vivo* for intramolecular recombination between wild type *attP* and *attB* sites which flank a promoter. Because the orientations of these sites are

5 inverted relative to each other, this causes an irreversible flipping of the promoter region relative to the gene of interest.

10 Palazzolo *et al.* *Gene* 88:25-36 (1990), discloses phage lambda vectors having bacteriophage λ arms that contain restriction sites positioned outside a
5 cloned DNA sequence and between wild-type *loxP* sites. Infection of *E. coli* cells that express the Cre recombinase with these phage vectors results in
15 recombination between the *loxP* sites and the *in vivo* excision of the plasmid replicon, including the cloned cDNA.

10 Pósfai *et al.* (*Nucl. Acids Res.* 22:2392-2398 (1994)) discloses a method for inserting into genomic DNA partial expression vectors having a selectable
20 marker, flanked by two wild-type FRT recognition sequences. FLP site-specific recombinase as present in the cells is used to integrate the vectors into the genome at predetermined sites. Under conditions where the replicon is functional, this
25 cloned genomic DNA can be amplified.

15 Bebee *et al.* (U.S. Patent No. 5,434,066) discloses the use of site-specific recombinases such as Cre for DNA containing two *loxP* sites for *in vivo* recombination between the sites.

30 Boyd (*Nucl. Acids Res.* 21:817-821 (1993)) discloses a method to facilitate the cloning of blunt-ended DNA using conditions that encourage
20 intermolecular ligation to a dephosphorylated vector that contains a wild-type *loxP* site acted upon by a Cre site-specific recombinase present in *E. coli* host cells.

35 Waterhouse *et al.* (WO 93/19172 and *Nucleic Acids Res.* 21 (9):2265 (1993)) disclose an *in vivo* method where light and heavy chains of a particular
40 antibody were cloned in different phage vectors between *loxP* and *loxP 511* sites and used to transfect new *E. coli* cells. Cre, acting in the host cells on the two
25 parental molecules (one plasmid, one phage), produced four products in equilibrium: two different cointegrates (produced by recombination at either *loxP*
45 or *loxP 511* sites), and two daughter molecules, one of which was the desired product.

30 Schlake & Bode (*Biochemistry* 33:12746-12751 (1994)) discloses an *in vivo* method to exchange expression cassettes at defined chromosomal locations,
50 each flanked by a wild type and a spacer-mutated FRT recombination site. A

double-reciprocal crossover was mediated in cultured mammalian cells by using this FLP/FRT system for site-specific recombination.

Hartley *et al.* (U.S. Patent No. 5,888,732) disclose compositions and methods for recombinational exchange of nucleic acid segments and molecules, including for use in recombinational cloning of a variety of nucleic acid molecules *in vitro* and *in vivo*, using a combination of wildtype and mutated recombination sites and recombination proteins.

Transposases. The family of enzymes, the transposases, has also been used to transfer genetic information between replicons. Transposons are structurally variable, being described as simple or compound, but typically encode the recombinase gene flanked by DNA sequences organized in inverted orientations. Integration of transposons can be random or highly specific. Representatives such as Tn7, which are highly site-specific, have been applied to the *in vivo* movement of DNA segments between replicons (Lucklow *et al.*, *J. Virol.* 67:4566-4579 (1993)).

Devine and Boeke *Nucl. Acids Res.* 22:3765-3772 (1994), discloses the construction of artificial transposons for the insertion of DNA segments, *in vitro*, into recipient DNA molecules. The system makes use of the integrase of yeast TY1 virus-like particles. The DNA segment of interest is cloned, using standard methods, between the ends of the transposon-like element TY1. In the presence of the TY1 integrase, the resulting element integrates randomly into a second target DNA molecule.

Recombination Sites. Also key to the integration/recombination reactions mediated by the above-noted recombination proteins and/or transposases are recognition sequences, often termed "recombination sites," on the DNA molecules participating in the integration/recombination reactions. These recombination sites are discrete sections or segments of DNA on the participating nucleic acid molecules that are recognized and bound by the recombination proteins during the initial stages of integration or recombination. For example, the recombination site for Cre recombinase is *loxP* which is a 34 base pair sequence comprised of two 13 base pair inverted repeats (serving as the recombinase binding sites) flanking an 8 base pair core sequence. See Figure 1 of Sauer, B., *Curr. Opin. Biotech.*

5:521-527 (1994). Other examples of recognition sequences include the *attB*, *attP*, *attL*, and *attR* sequences which are recognized by the recombination protein λ Int. *attB* is an approximately 25 base pair sequence containing two 9 base pair core-type Int binding sites and a 7 base pair overlap region, while *attP* is an approximately 240 base pair sequence containing core-type Int binding sites and arm-type Int binding sites as well as sites for auxiliary proteins integration host factor (IHF), FIS and excisionase (Xis). See Landy, *Curr. Opin. Biotech.* 3:699-707 (1993); see also U.S. Patent No. 5,888,732, which is incorporated by reference herein.

DNA cloning. The cloning of DNA segments currently occurs as a daily routine in many research labs and as a prerequisite step in many genetic analyses. The purpose of these clonings is various, however, two general purposes can be considered: (1) the initial cloning of DNA from large DNA or RNA segments (chromosomes, YACs, PCR fragments, mRNA, etc.), done in a relative handful of known vectors such as pUC, pGem, pBlueScript, and (2) the subcloning of these DNA segments into specialized vectors for functional analysis. A great deal of time and effort is expended both in the transfer of DNA segments from the initial cloning vectors to the more specialized vectors. This transfer is called subcloning.

The basic methods for cloning have been known for many years and have changed little during that time. A typical cloning protocol is as follows:

- (1) digest the DNA of interest with one or two restriction enzymes;
- (2) gel purify the DNA segment of interest when known;
- (3) prepare the vector by cutting with appropriate restriction enzymes, treating with alkaline phosphatase, gel purify etc., as appropriate;
- (4) ligate the DNA segment to the vector, with appropriate controls to eliminate background of uncut and self-ligated vector;
- (5) introduce the resulting vector into an *E. coli* host cell;
- (6) pick selected colonies and grow small cultures overnight;
- (7) make DNA minipreps; and

(8) analyze the isolated plasmid on agarose gels (often after diagnostic restriction enzyme digestions) or by PCR.

The specialized vectors used for subcloning DNA segments are functionally diverse. These include but are not limited to: vectors for expressing nucleic acid molecules in various organisms; for regulating nucleic acid molecule expression; for providing tags to aid in protein purification or to allow tracking of proteins in cells; for modifying the cloned DNA segment (*e.g.*, generating deletions); for the synthesis of probes (*e.g.*, riboprobes); for the preparation of templates for DNA sequencing; for the identification of protein coding regions; for the fusion of various protein-coding regions; to provide large amounts of the DNA of interest, *etc.* It is common that a particular investigation will involve subcloning the DNA segment of interest into several different specialized vectors.

As known in the art, simple subclonings can be done in one day (*e.g.*, the DNA segment is not large and the restriction sites are compatible with those of the subcloning vector). However, many other subclonings can take several weeks, especially those involving unknown sequences, long fragments, toxic genes, unsuitable placement of restriction sites, high backgrounds, impure enzymes, *etc.* Subcloning DNA fragments is thus often viewed as a chore to be done as few times as possible.

Several methods for facilitating the cloning of DNA segments have been described, *e.g.*, as in the following references.

Ferguson, J., *et al. Gene* 16:191 (1981), discloses a family of vectors for subcloning fragments of yeast DNA. The vectors encode kanamycin resistance. Clones of longer yeast DNA segments can be partially digested and ligated into the subcloning vectors. If the original cloning vector conveys resistance to ampicillin, no purification is necessary prior to transformation, since the selection will be for kanamycin.

Hashimoto-Gotoh, T., *et al. Gene* 41:125 (1986), discloses a subcloning vector with unique cloning sites within a streptomycin sensitivity gene; in a streptomycin-resistant host, only plasmids with inserts or deletions in the dominant sensitivity gene will survive streptomycin selection.

Accordingly, traditional subcloning methods, using restriction enzymes and ligase, are time consuming and relatively unreliable. Considerable labor is expended, and if two or more days later the desired subclone can not be found among the candidate plasmids, the entire process must then be repeated with alternative conditions attempted. Although site specific recombinases have been used to recombine DNA *in vivo*, the successful use of such enzymes *in vitro* was expected to suffer from several problems. For example, the site specificities and efficiencies were expected to differ *in vitro*; topologically linked products were expected; and the topology of the DNA substrates and recombination proteins was expected to differ significantly *in vitro* (see, e.g., Adams *et al*, *J. Mol. Biol.* 226:661-73 (1992)). Reactions that could go on for many hours *in vivo* were expected to occur in significantly less time *in vitro* before the enzymes became inactive. In addition, the stabilities of the recombination enzymes after incubation for extended periods of time in *in vitro* reactions was unknown, as were the effects of the topologies (*i.e.*, linear, coiled, supercoiled, etc.) of the nucleic acid molecules involved in the reaction. Multiple DNA recombination products were expected in the biological host used, resulting in unsatisfactory reliability, specificity or efficiency of subcloning. Thus, *in vitro* recombination reactions were not expected to be sufficiently efficient to yield the desired levels of product.

Accordingly, there is a long felt need to provide an alternative subcloning system that provides advantages over the known use of restriction enzymes and ligases.

SUMMARY OF THE INVENTION

The present invention relates to nucleic acid molecules encoding one or more recombination sites or one or more partial recombination sites, particularly *attB*, *attP*, *attL*, and *attR*, and fragments, mutants, variants and derivatives thereof. The invention also relates to such nucleic acid molecules comprising one or more of the recombination site nucleotide sequences or portions thereof and one or more additional physical or functional nucleotide sequences, such as those

5 encoding one or more multiple cloning sites, one or more transcription termination
sites, one or more transcriptional regulatory sequences (*e.g.*, one or more
10 promoters, enhancers, or repressors), one or more translational signal sequences,
one or more nucleotide sequences encoding a fusion partner protein or peptide
5 (*e.g.*, GST, His₆ or thioredoxin), one or more selection markers or modules, one
or more nucleotide sequences encoding localization signals such as nuclear
15 localization signals or secretion signals, one or more origins of replication, one or
more protease cleavage sites, one or more desired proteins or peptides encoded
by a gene or a portion of a gene, and one or more 5' or 3' polynucleotide tails
20 (particularly a poly-G tail). The invention also relates to such nucleic acid
molecules wherein the one or more recombination site nucleotide sequences is
operably linked to the one or more additional physical or functional nucleotide
sequences.

25 The invention also relates to primer nucleic acid molecules comprising the
15 recombination site nucleotide sequences of the invention (or portions thereof), and
to such primer nucleic acid molecules linked to one or more target-specific (*e.g.*,
one or more gene-specific) primer nucleic acid sequences. Such primers may also
30 comprise sequences complementary or homologous to DNA or RNA sequences
to be amplified, *e.g.*, by PCR, RT-PCR, etc. Such primers may also comprise
20 sequences or portions of sequences useful in the expression of protein genes
(ribosome binding sites, localization signals, protease cleavage sites, repressor
35 binding sites, promoters, transcription stops, stop codons, etc.). Said primers may
also comprise sequences or portions of sequences useful in the manipulation of
DNA molecules (restriction sites, transposition sites, sequencing primers, etc.).
40 25 The primers of the invention may be used in nucleic acid synthesis and preferably
are used for amplification (*e.g.*, PCR) of nucleic acid molecules. When the
primers of the invention include target- or gene-specific sequences (any sequence
45 contained within the target to be synthesized or amplified including translation
signals, gene sequences, stop codons, transcriptional signals (*e.g.*, promoters) and
30 the like), amplification or synthesis of target sequences or genes may be
accomplished. Thus, the invention relates to synthesis of a nucleic acid molecules
50 comprising mixing one or more primers of the invention with a nucleic acid

5 template, and incubating said mixture under conditions sufficient to make a first
nucleic acid molecule complementary to all or a portion of said template. Thus,
10 the invention relates specifically to a method of synthesizing a nucleic acid
molecule comprising:

- 5 (a) mixing a nucleic acid template with a polypeptide having polymerase
activity and one or more primers comprising one or more
15 recombination sites or portions thereof; and
(b) incubating said mixture under conditions sufficient to synthesize a first
nucleic acid molecule complementary to all or a portion of said
20 10 template and which preferably comprises one or more recombination
sites or portions thereof.

Such method of the invention may further comprise incubating said first
synthesized nucleic acid molecule under conditions sufficient to synthesize a
25 second nucleic acid molecule complementary to all or a portion of said first nucleic
acid molecule. Such synthesis may provide for a first nucleic acid molecule having
15 a recombination site or portion thereof at one or both of its termini.

30 In a preferred aspect, for the synthesis of the nucleic acid molecules, at
least two primers are used wherein each primer comprises a homologous sequence
at its terminus and/or within internal sequences of each primer (which may have
20 a homology length of about 2 to about 500 bases, preferably about 3 to about 100
bases, about 4 to about 50 bases, about 5 to about 25 bases and most preferably
35 about 6 to about 18 base overlap). In a preferred aspect, the first such primer
comprises at least one target-specific sequence and at least one recombination site
or portion thereof while the second primer comprises at least one recombination
40 25 site or portion thereof. Preferably, the homologous regions between the first and
second primers comprise at least a portion of the recombination site. In another
aspect, the homologous regions between the first and second primers may
45 comprise one or more additional sequences, *e.g.*, expression signals, translational
start motifs, or other sequences adding functionality to the desired nucleic acid
sequence upon amplification. In practice, two pairs of primers prime synthesis or
30 amplification of a nucleic acid molecule. In a preferred aspect, all or at least a
portion of the synthesized or amplified nucleic acid molecule will be homologous
50

5 to all or a portion of the template and further comprises a recombination site or
a portion thereof at at least one terminus and preferably both termini of the
10 synthesized or amplified molecule. Such synthesized or amplified nucleic acid
molecule may be double stranded or single stranded and may be used in the
5 recombinational cloning methods of the invention. The homologous primers of
the invention provide a substantial advantage in that one set of the primers may
15 be standardized for any synthesis or amplification reaction. That is, the primers
providing the recombination site sequences (without the target specific sequences)
can be pre-made and readily available for use. This in practice allows the use of
20 shorter custom made primers that contain the target specific sequence needed to
synthesize or amplify the desired nucleic acid molecule. Thus, this provides
reduced time and cost in preparing target specific primers (e.g., shorter primers
containing the target specific sequences can be prepared and used in synthesis
25 reactions). The standardized primers, on the other hand, may be produced in mass
to reduce cost and can be readily provided (e.g., in kits or as a product) to
15 facilitate synthesis of the desired nucleic acid molecules.

30 Thus, in one preferred aspect, the invention relates to a method of
synthesizing or amplifying one or more nucleic acid molecules comprising:

- 20 (a) mixing one or more nucleic acid templates with at least one
polypeptide having polymerase or reverse transcriptase activity
35 and at least a first primer comprising a template specific sequence
(complementary to or capable of hybridizing to said templates) and
at least a second primer comprising all or a portion of a
40 recombination site wherein said at least a portion of said second
25 primer is homologous to or complementary to at least a portion of
said first primer; and
- 45 (b) incubating said mixture under conditions sufficient to synthesize or
amplify one or more nucleic acid molecules complementary to all
or a portion of said templates and comprising one or more
30 recombination sites or portions thereof at one and preferably both
50 termini of said molecules.

More specifically, the invention relates to a method of synthesizing or amplifying one or more nucleic acid molecules comprising:

- (a) mixing one or more nucleic acid templates with at least one polypeptide having polymerase or reverse transcriptase activity and at least a first primer comprising a template specific sequence (complementary to or capable of hybridizing to said templates) and at least a portion of a recombination site, and at least a second primer comprising all or a portion of a recombination site wherein said at least a portion of said recombination site on said second primer is complementary to or homologous to at least a portion of said recombination site on said first primer; and
- (b) incubating said mixture under conditions sufficient to synthesize or amplify one or more nucleic acid molecules complementary to all or a portion of said templates and comprising one or more recombination sites or portions thereof at one and preferably both termini of said molecules.

In a more preferred aspect, the invention relates to a method of amplifying or synthesizing one or more nucleic acid molecules comprising:

- (a) mixing one or more nucleic acid templates with at least one polypeptide having polymerase or reverse transcriptase activity and one or more first primers comprising at least a portion of a recombination site and a template specific sequence (complementary to or capable of hybridizing to said template);
- (b) incubating said mixture under conditions sufficient to synthesize or amplify one or more first nucleic acid molecules complementary to all or a portion of said templates wherein said molecules comprise at least a portion of a recombination site at one and preferably both termini of said molecules;
- (c) mixing said molecules with one or more second primers comprising one or more recombination sites, wherein said recombination sites of said second primers are homologous to or

5 complementary to at least a portion of said recombination sites on
said first nucleic acid molecules; and

- 10 (d) incubating said mixture under conditions sufficient to synthesize or
amplify one or more second nucleic acid molecules complementary
5 to all or a portion of said first nucleic acid molecules and which
comprise one or more recombination sites at one and preferably
15 both termini of said molecules.

The invention also relates to vectors comprising the nucleic acid molecules
of the invention, to host cells comprising the vectors or nucleic acid molecules of
10 the invention, to methods of producing polypeptides encoded by the nucleic acid
20 molecules of the invention, and to polypeptides encoded by these nucleic acid
molecules or produced by the methods of the invention, which may be fusion
25 proteins. The invention also relates to antibodies that bind to one or more
polypeptides of the invention or epitopes thereof, which may be monoclonal or
15 polyclonal antibodies. The invention also relates to the use of these nucleic acid
molecules, primers, vectors, polypeptides and antibodies in methods for
recombinational cloning of nucleic acids, *in vitro* and *in vivo*, to provide chimeric
30 DNA molecules that have particular characteristics and/or DNA segments.

The antibodies of the invention may have particular use to identify and/or
20 purify peptides or proteins (including fusion proteins produced by the invention),
35 and to identify and/or purify the nucleic acid molecules of the invention or
portions thereof.

The methods for *in vitro* or *in vivo* recombinational cloning of nucleic acid
40 molecule generally relate to recombination between at least a first nucleic acid
25 molecule having at least one recombination site and a second nucleic acid
molecule having at least one recombination site to provide a chimeric nucleic acid
molecule. In one aspect, the methods relate to recombination between and first
45 vector having at least one recombination site and a second vector having at least
one recombination site to provide a chimeric vector. In another aspect, a nucleic
30 acid molecule having at least one recombination site is combined with a vector
having at least one recombination site to provide a chimeric vector. In a most
50 preferred aspect, the nucleic acid molecules or vectors used in recombination

comprise two or more recombination sites. In a more specific embodiment of the invention, the recombination methods relate to a Destination Reaction (also referred to herein as an "LR reaction") in which recombination occurs between an Entry clone and a Destination Vector. Such a reaction transfers the nucleic acid molecule of interest from the Entry Clone into the Destination Vector to create an Expression Clone. The methods of the invention also specifically relate to an Entry or Gateway reaction (also referred to herein as a "BP reaction") in which an Expression Clone is recombined with a Donor vector to produce an Entry clone. In other aspects, the invention relates to methods to prepare Entry clones by combining an Entry vector with at least one nucleic acid molecule (e.g., gene or portion of a gene). The invention also relates to conversion of a desired vector into a Destination Vector by including one or more (preferably at least two) recombination sites in the vector of interest. In a more preferred aspect, a nucleic acid molecule (e.g., a cassette) having at least two recombination sites flanking a selectable marker (e.g., a toxic gene or a genetic element preventing the survival of a host cell containing that gene or element, and/or preventing replication, partition or heritability of a nucleic acid molecule (e.g., a vector or plasmid) comprising that gene or element) is added to the vector to make a Destination Vector of the invention.

Preferred vectors for use in the invention include prokaryotic vectors, eukaryotic vectors, or vectors which may shuttle between various prokaryotic and/or eukaryotic systems (e.g. shuttle vectors). Preferred prokaryotic vectors for use in the invention include but are not limited to vectors which may propagate and/or replicate in gram negative and/or gram positive bacteria, including bacteria of the genera *Escherichia*, *Salmonella*, *Proteus*, *Clostridium*, *Klebsiella*, *Bacillus*, *Streptomyces*, and *Pseudomonas* and preferably in the species *E. coli*. Eukaryotic vectors for use in the invention include vectors which propagate and/or replicate in yeast cells, plant cells, mammalian cells, (particularly human and mouse), fungal cells, insect cells, nematode cells, fish cells and the like. Particular vectors of interest include but are not limited to cloning vectors, sequencing vectors, expression vectors, fusion vectors, two-hybrid vectors, gene therapy vectors, phage display vectors, gene-targeting vectors, PACs, BACs, YACs, MACs, and

reverse two-hybrid vectors. Such vectors may be used in prokaryotic and/or eukaryotic systems depending on the particular vector.

In another aspect, the invention relates to kits which may be used in carrying out the methods of the invention, and more specifically relates to cloning or subcloning kits and kits for carrying out the LR Reaction (*e.g.*, making an Expression Clone), for carrying out the BP Reaction (*e.g.*, making an Entry Clone), and for making Entry Clone and Destination Vector molecules of the invention. Such kits may comprise a carrier or receptacle being compartmentalized to receive and hold therein any number of containers. Such containers may contain any number of components for carrying out the methods of the invention or combinations of such components. In particular, a kit of the invention may comprise one or more components (or combinations thereof) selected from the group consisting of one or more recombination proteins or auxiliary factors or combinations thereof, one or more compositions comprising one or more recombination proteins or auxiliary factors or combinations thereof (for example, GATEWAY™ LR Clonase™ Enzyme Mix or GATEWAY™ BP Clonase™ Enzyme Mix), one or more reaction buffers, one or more nucleotides, one or more primers of the invention, one or more restriction enzymes, one or more ligases, one or more polypeptides having polymerase activity (*e.g.*, one or more reverse transcriptases or DNA polymerases), one or more proteinases (*e.g.*, proteinase K or other proteinases), one or more Destination Vector molecules, one or more Entry Clone molecules, one or more host cells (*e.g.* competent cells, such as *E. coli* cells, yeast cells, animal cells (including mammalian cells, insect cells, nematode cells, avian cells, fish cells, etc.), plant cells, and most particularly *E. coli* DB3.1 host cells, such as *E. coli* LIBRARY EFFICIENCY® DB3.1™ Competent Cells), instructions for using the kits of the invention (*e.g.*, to carry out the methods of the invention), and the like. In related aspects, the kits of the invention may comprise one or more nucleic acid molecules encoding one or more recombination sites or portions thereof, particularly one or more nucleic acid molecules comprising a nucleotide sequence encoding the one or more recombination sites or portions thereof of the invention. Preferably, such nucleic acid molecules comprise at least two recombination sites which flank a selectable

marker (*e.g.*, a toxic gene and/or antibiotic resistance gene). In a preferred aspect, such nucleic acid molecules are in the form of a cassette (*e.g.*, a linear nucleic acid molecule comprising one or more and preferably two or more recombination sites or portions thereof).

Kits for inserting or adding recombination sites to nucleic acid molecules of interest may comprise one or more nucleases (preferably restriction endonucleases), one or more ligases, one or more topoisomerases, one or more polymerases, and one or more nucleic acid molecules or adapters comprising one or more recombination sites. Kits for integrating recombination sites into one or more nucleic acid molecules of interest may comprise one or more components (or combinations thereof) selected from the group consisting of one or more integration sequences comprising one or more recombination sites. Such integration sequences may comprise one or more transposons, integrating viruses, homologous recombination sequences, RNA molecules, one or more host cells and the like.

Kits for making the Entry Clone molecules of the invention may comprise any or a number of components and the composition of such kits may vary depending on the specific method involved. Such methods may involve inserting the nucleic acid molecules of interest into an Entry or Donor Vector by the recombinational cloning methods of the invention, or using conventional molecular biology techniques (*e.g.*, restriction enzyme digestion and ligation). In a preferred aspect, the Entry Clone is made using nucleic acid amplification or synthesis products. Kits for synthesizing Entry Clone molecules from amplification or synthesis products may comprise one or more components (or combinations thereof) selected from the group consisting of one or more Donor Vectors (*e.g.*, one or more attP vectors including, but not limited to, pDONR201 (Figure 49), pDONR202 (Figure 50), pDONR203 (Figure 51), pDONR204 (Figure 52), pDONR205 (Figure 53), pDONR206 (Figure 53), and the like), one or more polypeptides having polymerase activity (preferably DNA polymerases and most preferably thermostable DNA polymerases), one or more proteinases, one or more reaction buffers, one or more nucleotides, one or more primers comprising one or

5 more recombination sites or portions thereof, and instructions for making one or more Entry Clones.

10 Kits for making the Destination vectors of the invention may comprise any number of components and the compositions of such kits may vary depending on
5 the specific method involved. Such methods may include the recombination methods of the invention or conventional molecular biology techniques (e.g.,
15 restriction endonuclease digestion and ligation). In a preferred aspect, the Destination vector is made by inserting a nucleic acid molecule comprising at least one recombination site (or portion thereof) of the invention (preferably a nucleic
10 acid molecule comprising at least two recombination sites or portions thereof flanking a selectable marker) into a desired vector to convert the desired vector into a Destination vector of the invention. Such kits may comprise at least one
20 component (or combinations thereof) selected from the group consisting of one or more restriction endonucleases, one or more ligases, one or more polymerases,
25 one or more nucleotides, reaction buffers, one or more nucleic acid molecules comprising at least one recombination site or portion thereof (preferably at least one nucleic acid molecule comprising at least two recombination sites flanking at
30 least one selectable marker, such as a cassette comprising at least one selectable marker such as antibiotic resistance genes and/or toxic genes), and instructions for making such Destination vectors.
20

35 The invention also relates to kits for using the antibodies of the invention in identification and/or isolation of peptides and proteins (which may be fusion proteins) produced by the nucleic acid molecules of the invention, and for
40 identification and/or isolation of the nucleic acid molecules of the invention or portions thereof. Such kits may comprise one or more components (or
25 combination thereof) selected from the group consisting of one or more antibodies of the invention, one or more detectable labels, one or more solid supports and the like.
45

30 Other preferred embodiments of the present invention will be apparent to one of ordinary skill in light of what is known in the art, in light of the following drawings and description of the invention, and in light of the claims.
50

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 depicts one general method of the present invention, wherein the starting (parent) DNA molecules can be circular or linear. The goal is to exchange the new subcloning vector D for the original cloning vector B. It is desirable in one embodiment to select for AD and against all the other molecules, including the Cointegrate. The square and circle are sites of recombination: *e.g.*, *lox* (such as *loxP*) sites, *att* sites, *etc.* For example, segment D can contain expression signals, protein fusion domains, new drug markers, new origins of replication, or specialized functions for mapping or sequencing DNA. It should be noted that the cointegrate molecule contains Segment D (Destination vector) adjacent to segment A (Insert), thereby juxtaposing functional elements in D with the insert in A. Such molecules can be used directly in vitro (*e.g.*, if a promoter is positioned adjacent to a gene-for in vitro transcription/translation) or in vivo (following isolation in a cell capable of propagating *ccdB*-containing vectors) by selecting for the selection markers in Segments B+D. As one skilled in the art will recognize, this single step method has utility in certain envisioned applications of the invention.

Figure 2 is a more detailed depiction of the recombinational cloning system of the invention, referred to herein as the "GATEWAY™ Cloning System." This figure depicts the production of Expression Clones via a "Destination Reaction," which may also be referred to herein as an "LR Reaction." A *kan^r* vector (referred to herein as an "Entry clone") containing a DNA molecule of interest (*e.g.*, a gene) localized between an *attL1* site and an *attL2* site is reacted with an *amp^r* vector (referred to herein as a "Destination Vector") containing a toxic or "death" gene localized between an *attR1* site and an *attR2* site, in the presence of GATEWAY™ LR Clonase™ Enzyme Mix (a mixture of Int, IHF and Xis). After incubation at 25°C for about 60 minutes, the reaction yields an *amp^r* Expression Clone containing the DNA molecule of interest localized between an *attB1* site and an *attB2* site, and a *kan^r* byproduct molecule, as well as intermediates. The reaction mixture may then be transformed into host cells (*e.g.*, *E. coli*) and clones containing the nucleic acid molecule of interest may

5 be selected by plating the cells onto ampicillin-containing media and picking amp^r colonies.

10 **Figure 3** is a schematic depiction of the cloning of a nucleic acid molecule from an Entry clone into multiple types of Destination vectors, to produce a
5 variety of Expression Clones. Recombination between a given Entry clone and different types of Destination vectors (not shown), via the LR Reaction depicted in Figure 2, produces multiple different Expression Clones for use in a variety of
15 applications and host cell types.

10 **Figure 4** is a detailed depiction of the production of Entry Clones via a "BP reaction," also referred to herein as an "Entry Reaction" or a "Gateway Reaction." In the example shown in this figure, an amp^r expression vector containing a DNA molecule of interest (*e.g.*, a gene) localized between an *attB1* site and an *attB2* site is reacted with a kan^r Donor vector (*e.g.*, an *attP* vector; here, GATEWAY™ pDONR201 (see Figure 49A-C)) containing a toxic or
25 "death" gene localized between an *attP1* site and an *attP2* site, in the presence of GATEWAY™ BP Clonase™ Enzyme Mix (a mixture of Int and IHF). After incubation at 25°C for about 60 minutes, the reaction yields a kan^r Entry clone containing the DNA molecule of interest localized between an *attL1* site and an
30 *attL2* site, and an amp^r by-product molecule. The Entry clone may then be transformed into host cells (*e.g.*, *E. coli*) and clones containing the Entry clone (and therefore the nucleic acid molecule of interest) may be selected by plating the
35 cells onto kanamycin-containing media and picking kan^r colonies. Although this figure shows an example of use of a kan^r Donor vector, it is also possible to use Donor vectors containing other selection markers, such as the gentamycin resistance or tetracycline resistance markers, as discussed herein.
40 25

45 **Figure 5** is a more detailed schematic depiction of the LR ("Destination") reaction (Figure 5A) and the BP ("Entry" or "Gateway") reaction (Figure 5B) of the GATEWAY™ Cloning System, showing the reactants, products and byproducts of each reaction.

5 **Figure 6** shows the sequences of the attB1 and attB2 sites flanking a gene of interest after subcloning into a Destination Vector to create an Expression Clone.

10 **Figure 7** is a schematic depiction of four ways to make Entry Clones using the compositions and methods of the invention: 1. using restriction enzymes and ligase; 2. starting with a cDNA library prepared in an attL Entry Vector; 3. using
15 an Expression Clone from a library prepared in an attB Expression Vector via the BxP reaction; and 4. recombinational cloning of PCR fragments with terminal attB sites, via the BxP reaction. Approaches 3 and 4 rely on recombination with a
10 Donor vector (here, an attP vector such as pDONR201 (see Figure 49A-C), pDONR202 (see Figure 50A-C), pDONR203 (see Figure 51A-C), pDONR204
20 (see Figure 52A-C), pDONR205 (see Figure 53A-C), or pDONR206 (see Figure 54A-C), for example) that provides an Entry Clone carrying a selection marker such as kan^r, gen^r, tet^r, or the like.

25 **Figure 8** is a schematic depiction of cloning of a PCR product by a BxP (Entry or Gateward) reaction. A PCR product with 25 bp terminal attB sites (plus four Gs) is shown as a substrate for the BxP reaction. Recombination
30 between the attB-PCR product of a gene and a Donor vector (which donates an Entry Vector that carries kan^r) results in an Entry Clone of the PCR product.

15 **Figure 9** is a listing of the nucleotide sequences of the recombination sites designated herein as attB1, attB2, attP1, attP2, attL1, attL2, attR1 and attR2. Sequences are written conventionally, from 5' to 3'.

20 **Figures 10-20:** The plasmid backbone for all the Entry Vectors depicted herein is the same, and is shown in Figure 10A for the Entry Vector pENTR1A.
35 For other Entry Vectors shown in Figures 11-20, only the sequences shown in Figure "A" for each figure set (*i.e.*, Figure 11A, Figure 12A, etc.) are different (within the attL1-attL2 cassettes) from those shown in Figure 10 – the plasmid backbone is identical.

40 **Figure 10** is a schematic depiction of the physical map and cloning sites (Figure 10A), and the nucleotide sequence (Figure 10B), of the Entry Vector pENTR1A.
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5 **Figure 11** is a schematic depiction of the cloning sites (Figure 11A) and the nucleotide sequence (Figure 11B) of the Entry Vector pENTR2B.

10 **Figure 12** is a schematic depiction of the cloning sites (Figure 12A) and the nucleotide sequence (Figure 12B) of the Entry Vector pENTR3C.

5 **Figure 13** is a schematic depiction of the cloning sites (Figure 13A) and the nucleotide sequence (Figure 13B) of the Entry Vector pENTR4.

15 **Figure 14** is a schematic depiction of the cloning sites (Figure 14A) and the nucleotide sequence (Figure 14B) of the Entry Vector pENTR5.

10 **Figure 15** is a schematic depiction of the cloning sites (Figure 15A) and the nucleotide sequence (Figure 15B) of the Entry Vector pENTR6.

20 **Figure 16** is a schematic depiction of the cloning sites (Figure 16A) and the nucleotide sequence (Figure 16B) of the Entry Vector pENTR7.

25 **Figure 17** is a schematic depiction of the cloning sites (Figure 17A) and the nucleotide sequence (Figure 17B) of the Entry Vector pENTR8.

15 **Figure 18** is a schematic depiction of the cloning sites (Figure 18A) and the nucleotide sequence (Figure 18B) of the Entry Vector pENTR9.

30 **Figure 19** is a schematic depiction of the cloning sites (Figure 19A) and the nucleotide sequence (Figure 19B) of the Entry Vector pENTR10.

20 **Figure 20** is a schematic depiction of the cloning sites (Figure 20A) and the nucleotide sequence (Figure 20B) of the Entry Vector pENTR11.

35 **Figure 21** is a schematic depiction of the physical map and the Trc expression cassette (Figure 21A) showing the promoter sequences at -35 and at -10 from the initiation codon, and the nucleotide sequence (Figure 21B-D), of Destination Vector pDEST1. This vector may also be referred to as pTrc-DEST1.

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45 **Figure 22** is a schematic depiction of the physical map and the His6 expression cassette (Figure 22A) showing the promoter sequences at -35 and at -10 from the initiation codon, and the nucleotide sequence (Figure 22B-D), of Destination Vector pDEST2. This vector may also be referred to as pHis6-DEST2.
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Figure 23 is a schematic depiction of the physical map and the GST expression cassette (Figure 23A) showing the promoter sequences at -35 and at -10 from the initiation codon, and the nucleotide sequence (Figure 23B-D), of Destination Vector pDEST3. This vector may also be referred to as pGST-DEST3.

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Figure 24 is a schematic depiction of the physical map and the His6-Trx expression cassette (Figure 24A) showing the promoter sequences at -35 and at -10 from the initiation codon and a TEV protease cleavage site, and the nucleotide sequence (Figure 24B-D), of Destination Vector pDEST4. This vector may also be referred to as pTrx-DEST4.

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Figure 25 is a schematic depiction of the attR1 and attR2 sites (Figure 25A), the physical map (Figure 25B), and the nucleotide sequence (Figure 25C-D), of Destination Vector pDEST5. This vector may also be referred to as pSPORT(+)-DEST5.

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Figure 26 is a schematic depiction of the attR1 and attR2 sites (Figure 26A), the physical map (Figure 26B), and the nucleotide sequence (Figure 26C-D), of Destination Vector pDEST6. This vector may also be referred to as pSPORT(-)-DEST6.

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Figure 27 is a schematic depiction of the attR1 site, CMV promoter, and the physical map (Figure 27A), and the nucleotide sequence (Figure 27B-C), of Destination Vector pDEST7. This vector may also be referred to as pCMV-DEST7.

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Figure 28 is a schematic depiction of the attR1 site, baculovirus polyhedrin promoter, and the physical map (Figure 28A), and the nucleotide sequence (Figure 28B-D), of Destination Vector pDEST8. This vector may also be referred to as pFastBac-DEST8.

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Figure 29 is a schematic depiction of the attR1 site, Semliki Forest Virus promoter, and the physical map (Figure 29A), and the nucleotide sequence (Figure 29B-E), of Destination Vector pDEST9. This vector may also be referred to as pSFV-DEST9.

Figure 30 is a schematic depiction of the attR1 site, baculovirus polyhedrin promoter, His6 fusion domain, and the physical map (Figure 30A), and the nucleotide sequence (Figure 30B-D), of Destination Vector pDEST10. This vector may also be referred to as pFastBacHT-DEST10.

Figure 31 is a schematic depiction of the attR1 cassette containing a tetracycline-regulated CMV promoter and the physical map (Figure 31A), and the nucleotide sequence (Figure 31B-D), of Destination Vector pDEST11. This vector may also be referred to as pTet-DEST11.

Figure 32 is a schematic depiction of the attR1 site, the start of the mRNA of the CMV promoter, and the physical map (Figure 32A), and the nucleotide sequence (Figure 32B-D), of Destination Vector pDEST12.2. This vector may also be referred to as pCMVneo-DEST12, as pCMV-DEST12, or as pDEST12.

Figure 33 is a schematic depiction of the attR1 site, the λP_L promoter, and the physical map (Figure 33A), and the nucleotide sequence (Figure 33B-C), of Destination Vector pDEST13. This vector may also be referred to as p λP_L -DEST13.

Figure 34 is a schematic depiction of the attR1 site, the T7 promoter, and the physical map (Figure 34A), and the nucleotide sequence (Figure 34B-D), of Destination Vector pDEST14. This vector may also be referred to as pT7-DEST14.

Figure 35 is a schematic depiction of the attR1 site, the T7 promoter, and the N-terminal GST fusion sequence, and the physical map (Figure 35A), and the nucleotide sequence (Figure 35B-D), of Destination Vector pDEST15. This vector may also be referred to as pT7 GST-DEST15.

Figure 36 is a schematic depiction of the attR1 site, the T7 promoter, and the N-terminal thioredoxin fusion sequence, and the physical map (Figure 36A), and the nucleotide sequence (Figure 36B-D), of Destination Vector pDEST16. This vector may also be referred to as pT7 Trx-DEST16.

Figure 37 is a schematic depiction of the attR1 site, the T7 promoter, and the N-terminal His6 fusion sequence, and the physical map (Figure 37A), and the

nucleotide sequence (Figure 37B-D), of Destination Vector pDEST17. This vector may also be referred to as pT7 His-DEST17.

Figure 38 is a schematic depiction of the attR1 site and the p10 baculovirus promoter, and the physical map (Figure 38A), and the nucleotide sequence (Figure 38B-D), of Destination Vector pDEST18. This vector may also be referred to as pFBp10-DEST18.

Figure 39 is a schematic depiction of the attR1 site, and the 39k baculovirus promoter, and the physical map (Figure 39A), and the nucleotide sequence (Figure 39B-D), of Destination Vector pDEST19. This vector may also be referred to as pFB39k-DEST19.

Figure 40 is a schematic depiction of the attR1 site, the *polh* baculovirus promoter, and the N-terminal GST fusion sequence, and the physical map (Figure 40A), and the nucleotide sequence (Figure 40B-D), of Destination Vector pDEST20. This vector may also be referred to as pFB GST-DEST20.

Figure 41 is a schematic depiction of a 2-hybrid vector with a DNA-binding domain, the attR1 site, and the ADH promoter, and the physical map (Figure 41A), and the nucleotide sequence (Figure 41B-E), of Destination Vector pDEST21. This vector may also be referred to as pDB Leu-DEST21.

Figure 42 is a schematic depiction of a 2-hybrid vector with an activation domain, the attR1 site, and the ADH promoter, and the physical map (Figure 42A), and the nucleotide sequence (Figure 42B-D), of Destination Vector pDEST22. This vector may also be referred to as pPC86-DEST22.

Figure 43 is a schematic depiction of the attR1 and attR2 sites, the T7 promoter, and the C-terminal His6 fusion sequence, and the physical map (Figure 43A), and the nucleotide sequence (Figure 43B-D), of Destination Vector pDEST23. This vector may also be referred to as pC-term-His6-DEST23.

Figure 44 is a schematic depiction of the attR1 and attR2 sites, the T7 promoter, and the C-terminal GST fusion sequence, and the physical map (Figure 44A), and the nucleotide sequence (Figure 44B-D), of Destination Vector pDEST24. This vector may also be referred to as pC-term-GST-DEST24.

Figure 45 is a schematic depiction of the attR1 and attR2 sites, the T7 promoter, and the C-terminal thioredoxin fusion sequence, and the physical map (Figure 45A), and the nucleotide sequence (Figure 45B-D), of Destination Vector pDEST25. This vector may also be referred to as pC-term-Trx-DEST25.

Figure 46 is a schematic depiction of the attR1 site, the CMV promoter, and an N-terminal His6 fusion sequence, and the physical map (Figure 46A), and the nucleotide sequence (Figure 46B-D), of Destination Vector pDEST26. This vector may also be referred to as pCMV-SPneo-His-DEST26.

Figure 47 is a schematic depiction of the attR1 site, the CMV promoter, and an N-terminal GST fusion sequence, and the physical map (Figure 47A), and the nucleotide sequence (Figure 47B-D), of Destination Vector pDEST27. This vector may also be referred to as pCMV-Spneo-GST-DEST27.

Figure 48 is a depiction of the physical map (Figure 48A), the cloning sites (Figure 48B), and the nucleotide sequence (Figure 48C-D), for the attB cloning vector plasmid pEXP501. This vector may also be referred to equivalently herein as pCMV•SPORT6, pCMVSPORT6, and pCMVSport6.

Figure 49 is a depiction of the physical map (Figure 49A), and the nucleotide sequence (Figure 49B-C), for the Donor plasmid pDONR201 which donates a kanamycin-resistant vector in the BP Reaction. This vector may also be referred to as pAttPkanr Donor Plasmid, or as pAttPkan Donor Plasmid

Figure 50 is a depiction of the physical map (Figure 50A), and the nucleotide sequence (Figure 50B-C), for the Donor plasmid pDONR202 which donates a kanamycin-resistant vector in the BP Reaction.

Figure 51 is a depiction of the physical map (Figure 51A), and the nucleotide sequence (Figure 51B-C), for the Donor plasmid pDONR203 which donates a kanamycin-resistant vector in the BP Reaction.

Figure 52 is a depiction of the physical map (Figure 52A), and the nucleotide sequence (Figure 52B-C), for the Donor plasmid pDONR204 which donates a kanamycin-resistant vector in the BP Reaction.

Figure 53 is a depiction of the physical map (Figure 53A), and the nucleotide sequence (Figure 53B-C), for the Donor plasmid pDONR205 which donates a tetracycline-resistant vector in the BP Reaction.

Figure 54 is a depiction of the physical map (Figure 54A), and the nucleotide sequence (Figure 54B-C), for the Donor plasmid pDONR206 which donates a gentamycin-resistant vector in the BP Reaction. This vector may also be referred to as pENTR22 attP Donor Plasmid, pAttPGenr Donor Plasmid, or pAttPgnt Donor Plasmid.

Figure 55 depicts the attB1 site, and the physical map, of an Entry Clone (pENTR7) of CAT subcloned into the Destination Vector pDEST2 (Figure 22).

Figure 56 depicts the DNA components of Reaction B of the one-tube BxP reaction described in Example 16, pEZC7102 and attB-tet-PCR.

Figure 57 is a physical map of the desired product of Reaction B of the one-tube BxP reaction described in Example 16, tetx7102.

Figure 58 is a physical map of the Destination Vector pEZC8402.

Figure 59 is a physical map of the expected tet^r subclone product, tetx8402, resulting from the LxR Reaction with tetx7102 (Figure 57) plus pEZC8402 (Figure 58).

Figure 60 is a schematic depiction of the bacteriophage lambda recombination pathways in *E. coli*.

Figure 61 is a schematic depiction of the DNA molecules participating in the LR Reaction. Two different co-integrates form during the LR Reaction (only one of which is shown here), depending on whether attL1 and attR1 or attL2 and attR2 are first to recombine. In one aspect, the invention provides directional cloning of a nucleic acid molecule of interest, since the recombination sites react with specificity (attL1 reacts with attR1; attL2 with attR2; attB1 with attP1; and attB2 with attP2). Thus, positioning of the sites allows construction of desired vectors having recombined fragments in the desired orientation.

Figure 62 is a depiction of native and fusion protein expression using the recombinational cloning methods and compositions of the invention. In the upper figure depicting native protein expression, all of the translational start signals are

5 included between the attB1 and attB2 sites; therefore, these signals must be
present in the starting Entry Clone. The lower figure depicts fusion protein
10 expression (here showing expression with both N-terminal and C-terminal fusion
tags so that ribosomes read through attB1 and attB2 to create the fusion protein).
5 Unlike native protein expression vectors, N-terminal fusion vectors have their
translational start signals upstream of the attB1 site.

15 **Figure 63** is a schematic depiction of three GATEWAY™ Cloning System
cassettes. Three blunt-ended cassettes are depicted which convert standard
expression vectors to Destination Vectors. Each of the depicted cassettes
10 provides amino-terminal fusions in one of three possible reading frames, and each
has a distinctive restriction cleavage site as shown.

20 **Figure 64** shows the physical maps of plasmids containing three attR
reading frame cassettes, pEVC15101 (reading frame A; Figure 64A), pEVC15102
(reading frame B; Figure 64B), and pEVC15103 (reading frame C; Figure 64C).

25 **Figure 65** depicts the attB primers used for amplifying the tet^r and amp^r
genes from pBR322 by the cloning methods of the invention.

30 **Figure 66** is a table listing the results of recombinational cloning of the tet^r
and amp^r PCR products made using the primers shown in Figure 65.

35 **Figure 67** is a graph showing the effect of the number of guanines (G's)
20 contained on the 5' end of the PCR primers on the cloning efficiency of PCR
products. It is noted, however, that other nucleotides besides guanine (including
A, T, C, U or combinations thereof) may be used as 5' extensions on the PCR
primers to enhance cloning efficiency of PCR products.

40 **Figure 68** is a graph showing a titration of various amounts of attP and
25 attB reactants in the BxP reaction, and the effects on cloning efficiency of PCR
products.

45 **Figure 69** is a series of graphs showing the effects of various weights
(Figure 69A) or moles (Figure 69B) of a 256 bp PCR product on formation of
colonies, and on efficiency of cloning of the 256 bp PCR product into a Donor
30 Vector (Figure 69C).

5 **Figure 70** is a series of graphs showing the effects of various weights
(Figure 70A) or moles (Figure 70B) of a 1 kb PCR product on formation of
colonies, and on efficiency of cloning of the 1 kb PCR product into a Donor
10 Vector (Figure 70C).

5 **Figure 71** is a series of graphs showing the effects of various weights
(Figure 71A) or moles (Figure 71B) of a 1.4 kb PCR product on formation of
colonies, and on efficiency of cloning of the 1.4 kb PCR product into a Donor
15 Vector (Figure 71C).

10 **Figure 72** is a series of graphs showing the effects of various weights
(Figure 72A) or moles (Figure 72B) of a 3.4 kb PCR product on formation of
colonies, and on efficiency of cloning of the 3.4 kb PCR product into a Donor
20 Vector (Figure 72C).

25 **Figure 73** is a series of graphs showing the effects of various weights
(Figure 73A) or moles (Figure 73B) of a 4.6 kb PCR product on formation of
colonies, and on efficiency of cloning of the 4.6 kb PCR product into a Donor
15 Vector (Figure 73C).

30 **Figure 74** is photograph of an ethidium bromide-stained gel of a titration
of a 6.9 kb PCR product in a BxP reaction.

20 **Figure 75** is a graph showing the effects of various amounts of a 10.1 kb
PCR product on formation of colonies upon cloning of the 10.1 kb PCR product
35 into a Donor Vector.

Figure 76 is photograph of an ethidium bromide-stained gel of a titration
of a 10.1 kb PCR product in a BxP reaction.

40 **Figure 77** is a table summarizing the results of the PCR product cloning
efficiency experiments depicted in Figures 69-74, for PCR fragments ranging in
25 size from 0.256 kb to 6.9 kb.

45 **Figure 78** is a depiction of the sequences at the ends of attR Cassettes.
Sequences contributed by the *Cm^r-ccdB* cassette are shown, including the outer
ends of the flanking attR sites (boxed). The staggered cleavage sites for Int are
30 indicated in the boxed regions. Following recombination with an Entry Clone,
only the outer sequences in attR sites contribute to the resulting attB sites in the
50

5 Expression Clone. The underlined sequences at both ends dictate the different reading frames (reading frames A, B, or C, with two alternative reading frame C cassettes depicted) for fusion proteins.

10 **Figure 79** is a depiction of several different attR cassettes (in reading frames A, B, or C) which may provide fusion codons at the amino-terminus of the encoded protein.

15 **Figure 80** illustrates the single-cutting restriction sites in an attR reading frame A cassette of the invention.

Figure 81 illustrates the single-cutting restriction sites in an attR reading frame B cassette of the invention.

20 **Figure 82** illustrates the single-cutting restriction sites in two alternative attR reading frame C cassettes of the invention (Figures 82A and 82B) depicted in Figure 78.

25 **Figure 83** shows the physical map (Figure 83A), and the nucleotide sequence (Figure 83B-C), for an attR reading frame C parent plasmid prfC Parent III, which contains an attR reading frame C cassette of the invention (alternative A in Figures 78 and 82).

30 **Figure 84** is a physical map of plasmid pEZC1301.

Figure 85 is a physical map of plasmid pEZC1313.

20 **Figure 86** is a physical map of plasmid pEZ14032.

35 **Figure 87** is a physical map of plasmid pMAB58.

Figure 88 is a physical map of plasmid pMAB62.

40 **Figure 89** is a depiction of a synthesis reaction using two pairs of homologous primers of the invention.

25 **Figure 90** is a schematic depiction of the physical map (Figure 90A), and the nucleotide sequence (Figure 90B-D), of Destination Vector pDEST28.

45 **Figure 91** is a schematic depiction of the physical map (Figure 91A), and the nucleotide sequence (Figure 91B-D), of Destination Vector pDEST29.

30 **Figure 92** is a schematic depiction of the physical map (Figure 92A), and the nucleotide sequence (Figure 92B-D), of Destination Vector pDEST30.

Figure 93 is a schematic depiction of the physical map (Figure 93A), and the nucleotide sequence (Figure 93B-D), of Destination Vector pDEST31.

Figure 94 is a schematic depiction of the physical map (Figure 94A), and the nucleotide sequence (Figure 94B-E), of Destination Vector pDEST32.

Figure 95 is a schematic depiction of the physical map (Figure 95A), and the nucleotide sequence (Figure 95B-D), of Destination Vector pDEST33.

Figure 96 is a schematic depiction of the physical map (Figure 96A), and the nucleotide sequence (Figure 96B-D), of Destination Vector pDEST34.

Figure 97 is a depiction of the physical map (Figure 97A), and the nucleotide sequence (Figure 97B-C), for the Donor plasmid pDONR207 which donates a gentamycin-resistant vector in the BP Reaction.

Figure 98 is a schematic depiction of the physical map (Figure 98A), and the nucleotide sequence (Figure 98B-D), of the 2-hybrid vector pMAB85.

Figure 99 is a schematic depiction of the physical map (Figure 99A), and the nucleotide sequence (Figure 99B-D), of the 2-hybrid vector pMAB86.

DETAILED DESCRIPTION OF THE INVENTION

Definitions

In the description that follows, a number of terms used in recombinant DNA technology are utilized extensively. In order to provide a clear and consistent understanding of the specification and claims, including the scope to be given such terms, the following definitions are provided.

Byproduct: is a daughter molecule (a new clone produced after the second recombination event during the recombinational cloning process) lacking the segment which is desired to be cloned or subcloned.

Cointegrate: is at least one recombination intermediate nucleic acid molecule of the present invention that contains both parental (starting) molecules. It will usually be linear. In some embodiments it can be circular. RNA and polypeptides may be expressed from cointegrates using an appropriate host cell strain, for example *E. coli* DB3.1 (particularly *E. coli* LIBRARY EFFICIENCY®).

5 DB3.1™ Competent Cells), and selecting for both selection markers found on the cointegrate molecule.

10 **Host:** is any prokaryotic or eukaryotic organism that can be a recipient of the recombinational cloning Product, vector, or nucleic acid molecule of the invention. A "host," as the term is used herein, includes prokaryotic or eukaryotic organisms that can be genetically engineered. For examples of such hosts, see 5 *Maniatis et al., Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratory, Cold Spring Harbor, New York (1982).

10 **Insert or Inserts:** include the desired nucleic acid segment or a population of nucleic acid segments (segment *A* of Figure 1) which may be manipulated by the methods of the present invention. Thus, the terms Insert(s) are meant to include a particular nucleic acid (preferably DNA) segment or a population of segments. Such Insert(s) can comprise one or more nucleic acid molecules.

20 **Insert Donor:** is one of the two parental nucleic acid molecules (e.g. RNA or DNA) of the present invention which carries the Insert. The Insert Donor molecule comprises the Insert flanked on both sides with recombination sites. The Insert Donor can be linear or circular. In one embodiment of the invention, 30 the Insert Donor is a circular DNA molecule and further comprises a cloning vector sequence outside of the recombination signals (see Figure 1). When a population of Inserts or population of nucleic acid segments are used to make the Insert Donor, a population of Insert Donors results and may be used in accordance with the invention. Examples of such Insert Donor molecules are GATEWAY™ Entry Vectors, which include but are not limited to those Entry Vectors depicted in Figures 10-20, as well as other vectors comprising a gene of interest flanked by 40 one or more *attL* sites (e.g., *attL1*, *attL2*, etc.), or by one or more *attB* sites (e.g., *attB1*, *attB2*, etc.) for the production of library clones.

45 **Product:** is one of the desired daughter molecules comprising the *A* and *D* sequences which is produced after the second recombination event during the recombinational cloning process (see Figure 1). The Product contains the nucleic acid which was to be cloned or subcloned. In accordance with the invention, 30 when a population of Insert Donors are used, the resulting population of Product

5 molecules will contain all or a portion of the population of Inserts of the Insert Donors and preferably will contain a representative population of the original molecules of the Insert Donors.

10 **Promoter:** is a DNA sequence generally described as the 5'-region of a gene, located proximal to the start codon. The transcription of an adjacent DNA segment is initiated at the promoter region. A repressible promoter's rate of transcription decreases in response to a repressing agent. An inducible promoter's rate of transcription increases in response to an inducing agent. A constitutive promoter's rate of transcription is not specifically regulated, though it can vary under the influence of general metabolic conditions.

20 **Recognition sequence:** Recognition sequences are particular sequences which a protein, chemical compound, DNA, or RNA molecule (e.g., restriction endonuclease, a modification methylase, or a recombinase) recognizes and binds. In the present invention, a recognition sequence will usually refer to a recombination site. For example, the recognition sequence for Cre recombinase is *loxP* which is a 34 base pair sequence comprised of two 13 base pair inverted repeats (serving as the recombinase binding sites) flanking an 8 base pair core sequence. See Figure 1 of Sauer, B., *Current Opinion in Biotechnology* 5:521-527 (1994). Other examples of recognition sequences are the *attB*, *attP*, *attL*, and *attR* sequences which are recognized by the recombinase enzyme λ Integrase. *attB* is an approximately 25 base pair sequence containing two 9 base pair core-type Int binding sites and a 7 base pair overlap region. *attP* is an approximately 240 base pair sequence containing core-type Int binding sites and arm-type Int binding sites as well as sites for auxiliary proteins integration host factor (IHF), FIS and excisionase (Xis). See Landy, *Current Opinion in Biotechnology* 3:699-707 (1993). Such sites may also be engineered according to the present invention to enhance production of products in the methods of the invention. When such engineered sites lack the P1 or H1 domains to make the recombination reactions irreversible (e.g., *attR* or *attP*), such sites may be designated *attR'* or *attP'* to show that the domains of these sites have been modified in some way.

Recombination proteins: include excisive or integrative proteins, enzymes, co-factors or associated proteins that are involved in recombination reactions involving one or more recombination sites, which may be wild-type proteins (See Landy, *Current Opinion in Biotechnology* 3:699-707 (1993)), or mutants, derivatives (e.g., fusion proteins containing the recombination protein sequences or fragments thereof), fragments, and variants thereof.

Recombination site: is a recognition sequence on a DNA molecule participating in an integration/recombination reaction by the recombinational cloning methods of the invention. Recombination sites are discrete sections or segments of DNA on the participating nucleic acid molecules that are recognized and bound by a site-specific recombination protein during the initial stages of integration or recombination. For example, the recombination site for Cre recombinase is *loxP* which is a 34 base pair sequence comprised of two 13 base pair inverted repeats (serving as the recombinase binding sites) flanking an 8 base pair core sequence. See Figure 1 of Sauer, B., *Curr. Opin. Biotech.* 5:521-527 (1994). Other examples of recognition sequences include the *attB*, *attP*, *attL*, and *attR* sequences described herein, and mutants, fragments, variants and derivatives thereof, which are recognized by the recombination protein λ Int and by the auxiliary proteins integration host factor (IHF), FIS and excisionase (Xis). See Landy, *Curr. Opin. Biotech.* 3:699-707 (1993).

Recombinational Cloning: is a method described herein, whereby segments of nucleic acid molecules or populations of such molecules are exchanged, inserted, replaced, substituted or modified, *in vitro* or *in vivo*. By "*in vitro*" and "*in vivo*" herein is meant recombinational cloning that is carried out outside of host cells (e.g., in cell-free systems) or inside of host cells (e.g., using recombination proteins expressed by host cells), respectively.

Repression cassette: is a nucleic acid segment that contains a repressor or a Selectable marker present in the subcloning vector.

Selectable marker: is a DNA segment that allows one to select for or against a molecule (e.g., a replicon) or a cell that contains it, often under particular conditions. These markers can encode an activity, such as, but not limited to,

production of RNA, peptide, or protein, or can provide a binding site for RNA, peptides, proteins, inorganic and organic compounds or compositions and the like. Examples of Selectable markers include but are not limited to: (1) DNA segments that encode products which provide resistance against otherwise toxic compounds (*e.g.*, antibiotics); (2) DNA segments that encode products which are otherwise lacking in the recipient cell (*e.g.*, tRNA genes, auxotrophic markers); (3) DNA segments that encode products which suppress the activity of a gene product; (4) DNA segments that encode products which can be readily identified (*e.g.*, phenotypic markers such as β -galactosidase, green fluorescent protein (GFP), and cell surface proteins); (5) DNA segments that bind products which are otherwise detrimental to cell survival and/or function; (6) DNA segments that otherwise inhibit the activity of any of the DNA segments described in Nos. 1-5 above (*e.g.*, antisense oligonucleotides); (7) DNA segments that bind products that modify a substrate (*e.g.* restriction endonucleases); (8) DNA segments that can be used to isolate or identify a desired molecule (*e.g.* specific protein binding sites); (9) DNA segments that encode a specific nucleotide sequence which can be otherwise non-functional (*e.g.*, for PCR amplification of subpopulations of molecules); (10) DNA segments, which when absent, directly or indirectly confer resistance or sensitivity to particular compounds; (11) DNA segments that encode products which are toxic in recipient cells; (12) DNA segments that inhibit replication, partition or heritability of nucleic acid molecules that contain them; and/or (13) DNA segments that encode conditional replication functions, *e.g.*, replication in certain hosts or host cell strains or under certain environmental conditions (*e.g.*, temperature, nutritional conditions, etc.).

Selection scheme: is any method which allows selection, enrichment, or identification of a desired Product or Product(s) from a mixture containing an Entry Clone or Vector, a Destination Vector, a Donor Vector, an Expression Clone or Vector, any intermediates (*e.g.* a Cointegrate or a replicon), and/or Byproducts. The selection schemes of one preferred embodiment have at least two components that are either linked or unlinked during recombinational cloning. One component is a Selectable marker. The other component controls the expression *in vitro* or *in vivo* of the Selectable marker, or survival of the cell (or

5 the nucleic acid molecule, *e.g.*, a replicon) harboring the plasmid carrying the
Selectable marker. Generally, this controlling element will be a repressor or
10 inducer of the Selectable marker, but other means for controlling expression or
activity of the Selectable marker can be used. Whether a repressor or activator
5 is used will depend on whether the marker is for a positive or negative selection,
and the exact arrangement of the various DNA segments, as will be readily
15 apparent to those skilled in the art. A preferred requirement is that the selection
scheme results in selection of or enrichment for only one or more desired
Products. As defined herein, selecting for a DNA molecule includes (a) selecting
10 or enriching for the presence of the desired DNA molecule, and (b) selecting or
20 enriching against the presence of DNA molecules that are not the desired DNA
molecule.

In one embodiment, the selection schemes (which can be carried out in
25 reverse) will take one of three forms, which will be discussed in terms of Figure 1.
15 The first, exemplified herein with a Selectable marker and a repressor therefore,
selects for molecules having segment *D* and lacking segment *C*. The second
selects against molecules having segment *C* and for molecules having segment *D*.
30 Possible embodiments of the second form would have a DNA segment carrying
a gene toxic to cells into which the *in vitro* reaction products are to be introduced.
20 A toxic gene can be a DNA that is expressed as a toxic gene product (a toxic
protein or RNA), or can be toxic in and of itself. (In the latter case, the toxic gene
35 is understood to carry its classical definition of "heritable trait".)

Examples of such toxic gene products are well known in the art, and
include, but are not limited to, restriction endonucleases (*e.g.*, *DpnI*), apoptosis-
40 25 related genes (*e.g.* ASK1 or members of the *bcl-2/ced-9* family), retroviral genes
including those of the human immunodeficiency virus (HIV), defensins such as
NP-1, inverted repeats or paired palindromic DNA sequences, bacteriophage lytic
45 genes such as those from Φ X174 or bacteriophage T4; antibiotic sensitivity genes
such as *rpsL*, antimicrobial sensitivity genes such as *pheS*, plasmid killer genes,
30 eukaryotic transcriptional vector genes that produce a gene product toxic to
bacteria, such as GATA-1, and genes that kill hosts in the absence of a
50 suppressing function, *e.g.*, *kicB*, *ccdB*, Φ X174 *F* (Liu, Q. *et al.*, *Curr. Biol.*

8:1300-1309 (1998)), and other genes that negatively affect replicon stability and/or replication. A toxic gene can alternatively be selectable *in vitro*, e.g., a restriction site.

Many genes coding for restriction endonucleases operably linked to inducible promoters are known, and may be used in the present invention. See, e.g. U.S. Patent Nos. 4,960,707 (*DpnI* and *DpnII*); 5,000,333, 5,082,784 and 5,192,675 (*KpnI*); 5,147,800 (*NgoAIII* and *NgoAI*); 5,179,015 (*FspI* and *HaeIII*); 5,200,333 (*HaeII* and *TaqI*); 5,248,605 (*HpaII*); 5,312,746 (*ClaI*); 5,231,021 and 5,304,480 (*XhoI* and *XhoII*); 5,334,526 (*AluI*); 5,470,740 (*NsiI*); 5,534,428 (*SstI/SacI*); 5,202,248 (*NcoI*); 5,139,942 (*NdeI*); and 5,098,839 (*PacI*). See also Wilson, G.G., *Nucl. Acids Res.* 19:2539-2566 (1991); and Lunnen, K.D., *et al.*, *Gene* 74:25-32 (1988).

In the second form, segment *D* carries a Selectable marker. The toxic gene would eliminate transformants harboring the Vector Donor, Cointegrate, and Byproduct molecules, while the Selectable marker can be used to select for cells containing the Product and against cells harboring only the Insert Donor.

The third form selects for cells that have both segments *A* and *D* in *cis* on the same molecule, but not for cells that have both segments in *trans* on different molecules. This could be embodied by a Selectable marker that is split into two inactive fragments, one each on segments *A* and *D*.

The fragments are so arranged relative to the recombination sites that when the segments are brought together by the recombination event, they reconstitute a functional Selectable marker. For example, the recombinational event can link a promoter with a structural nucleic acid molecule (e.g., a gene), can link two fragments of a structural nucleic acid molecule, or can link nucleic acid molecules that encode a heterodimeric gene product needed for survival, or can link portions of a replicon.

Site-specific recombinase: is a type of recombinase which typically has at least the following four activities (or combinations thereof): (1) recognition of one or two specific nucleic acid sequences; (2) cleavage of said sequence or sequences; (3) topoisomerase activity involved in strand exchange; and (4) ligase

activity to reseat the cleaved strands of nucleic acid. See Sauer, B., *Current Opinions in Biotechnology* 5:521-527 (1994). Conservative site-specific recombination is distinguished from homologous recombination and transposition by a high degree of sequence specificity for both partners. The strand exchange mechanism involves the cleavage and rejoining of specific DNA sequences in the absence of DNA synthesis (Landy, A. (1989) *Ann. Rev. Biochem.* 58:913-949).

Subcloning vector: is a cloning vector comprising a circular or linear nucleic acid molecule which includes preferably an appropriate replicon. In the present invention, the subcloning vector (segment *D* in Figure 1) can also contain functional and/or regulatory elements that are desired to be incorporated into the final product to act upon or with the cloned DNA Insert (segment *A* in Figure 1). The subcloning vector can also contain a Selectable marker (preferably DNA).

Vector: is a nucleic acid molecule (preferably DNA) that provides a useful biological or biochemical property to an Insert. Examples include plasmids, phages, autonomously replicating sequences (ARS), centromeres, and other sequences which are able to replicate or be replicated *in vitro* or in a host cell, or to convey a desired nucleic acid segment to a desired location within a host cell. A Vector can have one or more restriction endonuclease recognition sites at which the sequences can be cut in a determinable fashion without loss of an essential biological function of the vector, and into which a nucleic acid fragment can be spliced in order to bring about its replication and cloning. Vectors can further provide primer sites, *e.g.*, for PCR, transcriptional and/or translational initiation and/or regulation sites, recombinational signals, replicons, Selectable markers, *etc.* Clearly, methods of inserting a desired nucleic acid fragment which do not require the use of homologous recombination, transpositions or restriction enzymes (such as, but not limited to, UDG cloning of PCR fragments (U.S. Patent No. 5,334,575, entirely incorporated herein by reference), T:A cloning, and the like) can also be applied to clone a fragment into a cloning vector to be used according to the present invention. The cloning vector can further contain one or more selectable markers suitable for use in the identification of cells transformed with the cloning vector.

Vector Donor: is one of the two parental nucleic acid molecules (e.g. RNA or DNA) of the present invention which carries the DNA segments comprising the DNA vector which is to become part of the desired Product. The Vector Donor comprises a subcloning vector *D* (or it can be called the cloning vector if the Insert Donor does not already contain a cloning vector (e.g., for PCR fragments containing *attB* sites; see below)) and a segment *C* flanked by recombination sites (see Figure 1). Segments *C* and/or *D* can contain elements that contribute to selection for the desired Product daughter molecule, as described above for selection schemes. The recombination signals can be the same or different, and can be acted upon by the same or different recombinases. In addition, the Vector Donor can be linear or circular. Examples of such Vector Donor molecules include GATEWAY™ Destination Vectors, which include but are not limited to those Destination Vectors depicted in Figures 21-47 and 90-96.

Primer: refers to a single stranded or double stranded oligonucleotide that is extended by covalent bonding of nucleotide monomers during amplification or polymerization of a nucleic acid molecule (e.g. a DNA molecule). In a preferred aspect, a primer comprises one or more recombination sites or portions of such recombination sites. Portions of recombination sites comprise at least 2 bases (or basepairs, abbreviated herein as "bp"), at least 5-200 bases, at least 10-100 bases, at least 15-75 bases, at least 15-50 bases, at least 15-25 bases, or at least 16-25 bases, of the recombination sites of interest, as described in further detail below and in the Examples. When using portions of recombination sites, the missing portion of the recombination site may be provided as a template by the newly synthesized nucleic acid molecule. Such recombination sites may be located within and/or at one or both termini of the primer. Preferably, additional sequences are added to the primer adjacent to the recombination site(s) to enhance or improve recombination and/or to stabilize the recombination site during recombination. Such stabilization sequences may be any sequences (preferably G/C rich sequences) of any length. Preferably, such sequences range in size from 1 to about 1000 bases, 1 to about 500 bases, and 1 to about 100 bases, 1 to about 60 bases, 1 to about 25, 1 to about 10, 2 to about 10 and preferably about 4 bases.

5 Preferably, such sequences are greater than 1 base in length and preferably greater than 2 bases in length.

10 **Template:** refers to double stranded or single stranded nucleic acid molecules which are to be amplified, synthesized or sequenced. In the case of
5 double stranded molecules, denaturation of its strands to form a first and a second strand is preferably performed before these molecules will be amplified, synthesized or sequenced, or the double stranded molecule may be used directly
15 as a template. For single stranded templates, a primer complementary to a portion of the template is hybridized under appropriate conditions and one or more polypeptides having polymerase activity (e.g. DNA polymerases and/or reverse transcriptases) may then synthesize a nucleic acid molecule complementary to all or a portion of said template. Alternatively, for double stranded templates, one or more promoters may be used in combination with one or more polymerases to make nucleic acid molecules complementary to all or a portion of the template.
20 The newly synthesized molecules, according to the invention, may be equal or shorter in length than the original template. Additionally, a population of nucleic acid templates may be used during synthesis or amplification to produce a population of nucleic acid molecules typically representative of the original template population.

25 **Adapter:** is an oligonucleotide or nucleic acid fragment or segment (preferably DNA) which comprises one or more recombination sites (or portions of such recombination sites) which in accordance with the invention can be added to a circular or linear Insert Donor molecule as well as other nucleic acid molecules described herein. When using portions of recombination sites, the
30 missing portion may be provided by the Insert Donor molecule. Such adapters may be added at any location within a circular or linear molecule, although the adapters are preferably added at or near one or both termini of a linear molecule. Preferably, adapters are positioned to be located on both sides (flanking) a particular nucleic acid molecule of interest. In accordance with the invention,
35 adapters may be added to nucleic acid molecules of interest by standard recombinant techniques (e.g. restriction digest and ligation). For example, adapters may be added to a circular molecule by first digesting the molecule with
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an appropriate restriction enzyme, adding the adapter at the cleavage site and reforming the circular molecule which contains the adapter(s) at the site of cleavage. In other aspects, adapters may be added by homologous recombination, by integration of RNA molecules, and the like. Alternatively, adapters may be ligated directly to one or more and preferably both termini of a linear molecule thereby resulting in linear molecule(s) having adapters at one or both termini. In one aspect of the invention, adapters may be added to a population of linear molecules, (e.g. a cDNA library or genomic DNA which has been cleaved or digested) to form a population of linear molecules containing adapters at one and preferably both termini of all or substantial portion of said population.

Adapter-Primer: is primer molecule which comprises one or more recombination sites (or portions of such recombination sites) which in accordance with the invention can be added to a circular or linear nucleic acid molecule described herein. When using portions of recombination sites, the missing portion may be provided by a nucleic acid molecule (e.g., an adapter) of the invention. Such adapter-primers may be added at any location within a circular or linear molecule, although the adapter-primers are preferably added at or near one or both termini of a linear molecule. Examples of such adapter-primers and the use thereof in accordance with the methods of the invention are shown in Example 25 herein. Such adapter-primers may be used to add one or more recombination sites or portions thereof to circular or linear nucleic acid molecules in a variety of contexts and by a variety of techniques, including but not limited to amplification (e.g., PCR), ligation (e.g., enzymatic or chemical/synthetic ligation), recombination (e.g., homologous or non-homologous (illegitimate) recombination) and the like.

Library: refers to a collection of nucleic acid molecules (circular or linear). In one embodiment, a library may comprise a plurality (i.e., two or more) of DNA molecules, which may or may not be from a common source organism, organ, tissue, or cell. In another embodiment, a library is representative of all or a portion or a significant portion of the DNA content of an organism (a "genomic" library), or a set of nucleic acid molecules representative of all or a portion or a significant portion of the expressed nucleic acid molecules (a cDNA library) in a

cell, tissue, organ or organism. A library may also comprise random sequences made by *de novo* synthesis, mutagenesis of one or more sequences and the like. Such libraries may or may not be contained in one or more vectors.

Amplification: refers to any *in vitro* method for increasing a number of copies of a nucleotide sequence with the use of a polymerase. Nucleic acid amplification results in the incorporation of nucleotides into a DNA and/or RNA molecule or primer thereby forming a new molecule complementary to a template. The formed nucleic acid molecule and its template can be used as templates to synthesize additional nucleic acid molecules. As used herein, one amplification reaction may consist of many rounds of replication. DNA amplification reactions include, for example, polymerase chain reaction (PCR). One PCR reaction may consist of 5-100 "cycles" of denaturation and synthesis of a DNA molecule.

Oligonucleotide: refers to a synthetic or natural molecule comprising a covalently linked sequence of nucleotides which are joined by a phosphodiester bond between the 3' position of the deoxyribose or ribose of one nucleotide and the 5' position of the deoxyribose or ribose of the adjacent nucleotide. This term may be used interchangeably herein with the terms "nucleic acid molecule" and "polynucleotide," without any of these terms necessarily indicating any particular length of the nucleic acid molecule to which the term specifically refers.

Nucleotide: refers to a base-sugar-phosphate combination. Nucleotides are monomeric units of a nucleic acid molecule (DNA and RNA). The term nucleotide includes ribonucleoside triphosphates ATP, UTP, CTG, GTP and deoxyribonucleoside triphosphates such as dATP, dCTP, dITP, dUTP, dGTP, dTTP, or derivatives thereof. Such derivatives include, for example, [α S]dATP, 7-deaza-dGTP and 7-deaza-dATP. The term nucleotide as used herein also refers to dideoxyribonucleoside triphosphates (ddNTPs) and their derivatives. Illustrated examples of dideoxyribonucleoside triphosphates include, but are not limited to, ddATP, ddCTP, ddGTP, ddITP, and ddTTP. According to the present invention, a "nucleotide" may be unlabeled or detectably labeled by well known techniques. Detectable labels include, for example, radioactive isotopes, fluorescent labels, chemiluminescent labels, bioluminescent labels and enzyme labels.

Hybridization: The terms "hybridization" and "hybridizing" refers to base pairing of two complementary single-stranded nucleic acid molecules (RNA and/or DNA) to give a double stranded molecule. As used herein, two nucleic acid molecules may be hybridized, although the base pairing is not completely complementary. Accordingly, mismatched bases do not prevent hybridization of two nucleic acid molecules provided that appropriate conditions, well known in the art, are used. In some aspects, hybridization is said to be under "stringent conditions." By "stringent conditions" as used herein is meant overnight incubation at 42°C in a solution comprising: 50% formamide, 5x SSC (150 mM NaCl, 15mM trisodium citrate), 50 mM sodium phosphate (pH 7.6), 5x Denhardt's solution, 10% dextran sulfate, and 20 g/ml denatured, sheared salmon sperm DNA, followed by washing the filters in 0.1x SSC at about 65°C.

Other terms used in the fields of recombinant DNA technology and molecular and cell biology as used herein will be generally understood by one of ordinary skill in the applicable arts.

Overview

Two reactions constitute the recombinational cloning system of the present invention, referred to herein as the "GATEWAY™ Cloning System," as depicted generally in Figure 1. The first of these reactions, the **LR Reaction** (Figure 2), which may also be referred to interchangeably herein as the **Destination Reaction**, is the main pathway of this system. The LR Reaction is a recombination reaction between an Entry vector or clone and a Destination Vector, mediated by a cocktail of recombination proteins such as the GATEWAY™ LR Clonase™ Enzyme Mix described herein. This reaction transfers nucleic acid molecules of interest (which may be genes, cDNAs, cDNA libraries, or fragments thereof) from the Entry Clone to an Expression Vector, to create an Expression Clone.

The sites labeled L, R, B, and P are respectively the attL, attR, attB, and attP recombination sites for the bacteriophage λ recombination proteins that constitute the Clonase cocktail (referred to herein variously as "Clonase" or

5 “GATEWAY™ LR Clonase™ Enzyme Mix” (for recombination protein mixtures
mediating attL x attR recombination reactions, as described herein) or
10 “GATEWAY™ BP Clonase™ Enzyme Mix” (for recombination protein mixtures
mediating attB x attP recombination reactions, as described herein)). The
5 Recombinational Cloning reactions are equivalent to concerted, highly specific,
cutting and ligation reactions. Viewed in this way, the recombination proteins cut
15 to the left and right of the nucleic acid molecule of interest in the Entry Clone and
ligate it into the Destination vector, creating a new Expression Clone.

The nucleic acid molecule of interest in an Expression Clone is flanked by
10 the small attB1 and attB2 sites. The orientation and reading frame of the nucleic
acid molecule of interest are maintained throughout the subcloning, because attL1
20 reacts only with attR1, and attL2 reacts only with attR2. Likewise, attB1 reacts
only with attP1, and attB2 reacts only with attP2. Thus, the invention also relates
25 to methods of controlled or directional cloning using the recombination sites of the
invention (or portions thereof), including variants, fragments, mutants and
15 derivatives thereof which may have altered or enhanced specificity. The invention
also relates more generally to any number of recombination site partners or pairs
30 (where each recombination site is specific for and interacts with its corresponding
recombination site). Such recombination sites are preferably made by mutating or
20 modifying the recombination site to provide any number of necessary specificities
(e.g., attB1-10, attP1-10, attL1-10, attR1-10, etc.), non-limiting examples of
35 which are described in detail in the Examples herein.

When an aliquot from the recombination reaction is transformed into host
40 cells (e.g., *E. coli*) and spread on plates containing an appropriate selection agent,
25 e.g., an antibiotic such as ampicillin with or without methicillin, cells that take up
the desired clone form colonies. The unreacted Destination Vector does not give
ampicillin-resistant colonies, even though it carries the ampicillin-resistance gene,
45 because it contains a toxic gene, e.g., *ccdB*. Thus selection for ampicillin
resistance selects for *E. coli* cells that carry the desired product, which usually
30 comprise >90% of the colonies on the ampicillin plate.

To participate in the Recombinational (or “GATEWAY™”) Cloning
50 Reaction, a nucleic acid molecule of interest first may be cloned into an Entry

5 Vector, creating an Entry Clone. Multiple options are available for creating Entry
Clones, including: cloning of PCR sequences with terminal attB recombination
10 sites into Entry Vectors; using the GATEWAY™ Cloning System recombination
reaction; transfer of genes from libraries prepared in GATEWAY™ Cloning System
5 vectors by recombination into Entry Vectors; and cloning of restriction enzyme-
generated fragments and PCR fragments into Entry Vectors by standard
15 recombinant DNA methods. These approaches are discussed in further detail
herein.

A key advantage of the GATEWAY™ Cloning System is that a nucleic acid
10 molecule of interest (or even a population of nucleic acid molecules of interest)
present as an Entry Clone can be subcloned in parallel into one or more
20 Destination Vectors in a simple reactions for anywhere from about 30 seconds to
about 60 minutes (preferably about 1-60 minutes, about 1-45 minutes, about 1-30
25 minutes, about 2-60 minutes, about 2-45 minutes, about 2-30 minutes, about 1-2
15 minutes, about 30-60 minutes, about 45-60 minutes, or about 30-45 minutes).
Longer reaction times (e.g., 2-24 hours, or overnight) may increase recombination
efficiency, particularly where larger nucleic acid molecules are used, as described
30 in the Examples herein. Moreover, a high percentage of the colonies obtained
carry the desired Expression Clone. This process is illustrated schematically in
20 Figure 3, which shows an advantage of the invention in which the molecule of
interest can be moved simultaneously or separately into multiple Destination
35 Vectors. In the LR Reaction, one or both of the nucleic acid molecules to be
recombined may have any topology (e.g., linear, relaxed circular, nicked circular,
supercoiled, etc.), although one or both are preferably linear.

40 The second major pathway of the GATEWAY™ Cloning System is the
25 **BP Reaction** (Figure 4), which may also be referred to interchangeably herein as
the **Entry Reaction** or the **Gateward Reaction**. The BP Reaction may
45 recombine an Expression Clone with a Donor Plasmid (the counterpart of the
byproduct in Figure 2). This reaction transfers the nucleic acid molecule of
30 interest (which may have any of a variety of topologies, including linear, coiled,
supercoiled, etc.) in the Expression Clone into an Entry Vector, to produce a new
50 Entry Clone. Once this nucleic acid molecule of interest is cloned into an Entry
55

5 Vector, it can be transferred into new Expression Vectors, through the LR
Reaction as described above. In the BP Reaction, one or both of the nucleic acid
10 molecules to be recombined may have any topology (*e.g.*, linear, relaxed circular,
nicked circular, supercoiled, etc.), although one or both are preferably linear.

5 A useful variation of the BP Reaction permits rapid cloning and expression
of products of amplification (*e.g.*, PCR) or nucleic acid synthesis. Amplification
15 (*e.g.*, PCR) products synthesized with primers containing terminal 25 bp attB sites
serve as efficient substrates for the Gateway Cloning reaction. Such amplification
products may be recombined with a Donor Vector to produce an Entry Clone (see
20 Figure 7). The result is an Entry Clone containing the amplification fragment.
Such Entry Clones can then be recombined with Destination Vectors -- through
the LR Reaction -- to yield Expression Clones of the PCR product.

Additional details of the LR Reaction are shown in Figure 5A. The
25 GATEWAY™ LR Clonase™ Enzyme Mix that mediates this reaction contains
lambda recombination proteins Int (Integrase), Xis (Excisionase), and IHF
15 (Integration Host Factor). In contrast, the GATEWAY™ BP Clonase™ Enzyme
Mix, which mediates the BP Reaction (Figure 5B), comprises Int and IHF alone.

30 The recombination (att) sites of each vector comprise two distinct
segments, donated by the parental vectors. The staggered lines dividing the two
portions of each att site, depicted in Figures 5A and 5B, represent the seven-base
20 staggered cut produced by Int during the recombination reactions. This structure
is seen in greater detail in Figure 6, which displays the attB recombination
35 sequences of an Expression Clone, generated by recombination between the attL1
and attL2 sites of an Entry Clone and the attR1 and attR2 sites of a Destination
40 Vector.

25 The nucleic acid molecule of interest in the Expression Clone is flanked by
attB sites: attB1 to the left (amino terminus) and attB2 to the right (carboxy
45 terminus). The bases in attB1 to the left of the seven-base staggered cut produced
by Int are derived from the Destination vector, and the bases to the right of the
staggered cut are derived from the Entry Vector (see Figure 6). Note that the
30 sequence is displayed in triplets corresponding to an open reading frame. If the
50 reading frame of the nucleic acid molecule of interest cloned in the Entry Vector

is in phase with the reading frame shown for attB1, amino-terminal protein fusions can be made between the nucleic acid molecule of interest and any GATEWAY™ Cloning System Destination Vector encoding an amino-terminal fusion domain. Entry Vectors and Destination Vectors that enable cloning in all three reading frames are described in more detail herein, particularly in the Examples.

The LR Reaction allows the transfer of a desired nucleic acid molecule of interest into new Expression Vectors by recombining a Entry Clone with various Destination Vectors. To participate in the LR or Destination Reaction, however, a nucleic acid molecule of interest preferably is first converted to a Entry Clone. Entry Clones can be made in a number of ways, as shown in Figure 7.

One approach is to clone the nucleic acid molecule of interest into one or more of the Entry Vectors, using standard recombinant DNA methods, with restriction enzymes and ligase. The starting DNA fragment can be generated by restriction enzyme digestion or as a PCR product. The fragment is cloned between the attL1 and attL2 recombination sites in the Entry Vector. Note that a toxic or "death" gene (*e.g.*, *ccdB*), provided to minimize background colonies from incompletely digested Entry Vector, must be excised and replaced by the nucleic acid molecule of interest.

A second approach to making an Entry Clone (Figure 7) is to make a library (genomic or cDNA) in an Entry Vector, as described in detail herein. Such libraries may then be transferred into Destination Vectors for expression screening, for example in appropriate host cells such as yeast cells or mammalian cells.

A third approach to making Entry Clones (Figure 7) is to use Expression Clones obtained from cDNA molecules or libraries prepared in Expression Vectors. Such cDNAs or libraries, flanked by attB sites, can be introduced into a Entry Vector by recombination with a Donor Vector via the BP Reaction. If desired, an entire Expression Clone library can be transferred into the Entry Vector through the BP Reaction. Expression Clone cDNA libraries may also be constructed in a variety of prokaryotic and eukaryotic GATEWAY™-modified vectors (*e.g.*, the pEXP501 Expression Vector (see Figure 48), and 2-hybrid and

attB library vectors), as described in detail herein, particularly in the Examples below.

A fourth, and potentially most versatile, approach to making an Entry Clone (Figure 7) is to introduce a sequence for a nucleic acid molecule of interest into an Entry Vector by amplification (*e.g.*, PCR) fragment cloning. This method is diagramed in Figure 8. The DNA sequence first is amplified (for example, with PCR) as outlined in detail below and in the Examples herein, using primers containing one or more bp, two or more bp, three or more bp, four or more bp, five or more bp, preferably six or more bp, more preferably 6-25 bp (particularly 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24 or 25) bp of the attB nucleotide sequences (such as, but not limited to, those depicted in Figure 9), and optionally one or more, two or more, three or more, four or more, and most preferably four or five or more additional terminal nucleotide bases which preferably are guanines. The PCR product then may be converted to a Entry Clone by performing a BP Reaction, in which the attB-PCR product recombines with a Donor Vector containing one or more attP sites. Details of this approach and protocols for PCR fragment subcloning are provided in Examples 8 and 21-25.

A variety of Entry Clones may be produced by these methods, providing a wide array of cloning options; a number of specific Entry Vectors are also available commercially from Life Technologies, Inc. (Rockville, MD). The Examples herein provide a more in-depth description of selected Entry Vectors and details of their cloning sites. Choosing the optimal Entry Vector for a particular application is discussed in Example 4.

Entry Vectors and Destination Vectors should be constructed so that the amino-terminal region of a nucleic acid molecule of interest (*e.g.*, a gene, cDNA library or insert, or fragment thereof) will be positioned next to the attL1 site. Entry Vectors preferably contain the *rrnB* transcriptional terminator upstream of the attL1 site. This sequence ensures that expression of cloned nucleic acid molecules of interest is reliably "off" in *E. coli*, so that even toxic genes can be successfully cloned. Thus, Entry Clones may be designed to be transcriptionally silent. Note also that Entry Vectors, and hence Entry Clones, may contain the kanamycin antibiotic resistance (*kan'*) gene to facilitate selection of host cells

5 containing Entry Clones after transformation. In certain applications, however,
Entry Clones may contain other selection markers, including but not limited to a
10 gentamycin resistance (*gen^r*) or tetracycline resistance (*tet^r*) gene, to facilitate
selection of host cells containing Entry Clones after transformation.

5 Once a nucleic acid molecule of interest has been cloned into an Entry
Vector, it may be moved into a Destination Vector. The upper right portion of
15 Figure 5A shows a schematic of a Destination Vector. The thick arrow represents
some function (often transcription or translation) that will act on the nucleic acid
molecule of interest in the clone. During the recombination reaction, the region
20 between the attR1 and attR2 sites, including a toxic or "death" gene (*e.g.*, *ccdB*),
is replaced by the DNA segment from the Entry Clone. Selection for
recombinants that have acquired the ampicillin resistance (*amp^r*) gene (carried on
the Destination Vector) and that have also lost the death gene ensures that a high
25 percentage (usually >90%) of the resulting colonies will contain the correct insert.

15 To move a nucleic acid molecule of interest into a Destination Vector, the
Destination Vector is mixed with the Entry Clone comprising the desired nucleic
acid molecule of interest, a cocktail of recombination proteins (*e.g.*,
30 GATEWAY™ LR Clonase™ Enzyme Mix) is added, the mixture is incubated
(preferably at about 25°C for about 60 minutes, or longer under certain
20 circumstances, *e.g.* for transfer of large nucleic acid molecules, as described
below) and any standard host cell (including bacterial cells such as *E. coli*; animal
35 cells such as insect cells, mammalian cells, nematode cells and the like; plant cells;
and yeast cells) strain is transformed with the reaction mixture. The host cell used
will be determined by the desired selection (*e.g.*, *E. coli* DB3.1, available
40 commercially from Life Technologies, Inc., allows survival of clones containing
25 the *ccdB* death gene, and thus can be used to select for cointegrate molecules --
i.e., molecules that are hybrids between the Entry Clone and Destination Vector).
45 The Examples below provide further details and protocols for use of Entry and
Destination Vectors in transferring nucleic acid molecules of interest and
30 expressing RNAs or polypeptides encoded by these nucleic acid molecules in a
variety of host cells.
50
55

The cloning system of the invention therefore offers multiple advantages:

- Once a nucleic acid molecule of interest is cloned into the GATEWAY™ Cloning System, it can be moved into and out of other vectors with complete fidelity of reading frame and orientation. That is, since the reactions proceed whereby attL1 on the Entry Clone recombines with attR1 on the Destination Vector, the directionality of the nucleic acid molecule of interest is maintained or may be controlled upon transfer from the Entry Clone into the Destination Vector. Hence, the GATEWAY™ Cloning System provides a powerful and easy method of directional cloning of nucleic acid molecule of interest.
- One-step cloning or subcloning: Mix the Entry Clone and the Destination Vector with Clonase, incubate, and transform.
- Clone PCR products readily by *in vitro* recombination, by adding attB sites to PCR primers. Then directly transfer these Entry Clones into Destination Vectors. This process may also be carried out in one step (see Examples below).
- Powerful selections give high reliability: >90% (and often >99%) of the colonies contain the desired DNA in its new vector.
- One-step conversion of existing standard vectors into GATEWAY™ Cloning System vectors.
- Ideal for large vectors or those with few cloning sites.
- Recombination sites are short (25 bp), and may be engineered to contain no stop codons or secondary structures.
- Reactions may be automated, for high-throughput applications (*e.g.*, for diagnostic purposes or for therapeutic candidate screening).
- The reactions are economical: 0.3 µg of each DNA; no restriction enzymes, phosphatase, ligase, or gel purification. Reactions work well with miniprep DNA.
- Transfer multiple clones, and even libraries, into one or more Destination Vectors, in a single experiment.
- A variety of Destination Vectors may be produced, for applications including, but not limited to:

- Protein expression in *E. coli*: native proteins; fusion proteins with GST, His6, thioredoxin, etc., for purification, or one or more epitope tags; any promoter useful in expressing proteins in *E. coli* may be used, such as p, λ P_L, and T7 promoters.
- Protein expression in eukaryotic cells: CMV promoter, baculovirus (with or without His6 tag), Semliki Forest virus, Tet regulation.
- DNA sequencing (all *lac* primers), RNA probes, phagemids (both strands)
- A variety of Entry Vectors (for recombinational cloning entry by standard recombinant DNA methods) may be produced:
 - Strong transcription stop just upstream, for genes toxic to *E. coli*.
 - Three reading frames.
 - With or without TEV protease cleavage site.
 - Motifs for prokaryotic and / or eukaryotic translation.
 - Compatible with commercial cDNA libraries.
- Expression Clone cDNA (attB) libraries, for expression screening, including 2-hybrid libraries and phage display libraries, may also be constructed.

Recombination Site Sequences

In one aspect, the invention relates to nucleic acid molecules, which may or may not be isolated nucleic acid molecules, comprising one or more nucleotide sequences encoding one or more recombination sites or portions thereof. In particular, this aspect of the invention relates to such nucleic acid molecules comprising one or more nucleotide sequences encoding *attB*, *attP*, *attL*, or *attR*, or portions of these recombination site sequences. The invention also relates to mutants, derivatives, and fragments of such nucleic acid molecules. Unless otherwise indicated, all nucleotide sequences that may have been determined by sequencing a DNA molecule herein were determined using manual or automated DNA sequencing, such as dideoxy sequencing, according to methods that are routine to one of ordinary skill in the art (Sanger, F., and Coulson, A.R., *J. Mol. Biol.* 94:444-448 (1975); Sanger, F., *et al.*, *Proc. Natl. Acad. Sci. USA* 74:5463-5467 (1977)). All amino acid sequences of polypeptides encoded by DNA

5 molecules determined herein were predicted by conceptual translation of a DNA
sequence determined as above. Therefore, as is known in the art for any DNA
10 sequence determined by these approaches, any nucleotide sequence determined
herein may contain some errors. Nucleotide sequences determined by such
5 methods are typically at least about 90% identical, more typically at least about
95% to at least about 99.9% identical to the actual nucleotide sequence of the
15 sequenced DNA molecule. As is also known in the art, a single insertion or
deletion in a determined nucleotide sequence compared to the actual sequence will
cause a frame shift in translation of the nucleotide sequence such that the predicted
20 amino acid sequence encoded by a determined nucleotide sequence will be
completely different from the amino acid sequence actually encoded by the
sequenced DNA molecule, beginning at the point of such an insertion or deletion.

25 Unless otherwise indicated, each "nucleotide sequence" set forth herein is
presented as a sequence of deoxyribonucleotides (abbreviated A, G, C and T).
15 However, by "nucleotide sequence" of a nucleic acid molecule or polynucleotide
is intended, for a DNA molecule or polynucleotide, a sequence of
deoxyribonucleotides, and for an RNA molecule or polynucleotide, the
30 corresponding sequence of ribonucleotides (A, G, C and U), where each thymidine
deoxyribonucleotide (T) in the specified deoxyribonucleotide sequence is replaced
20 by the ribonucleotide uridine (U). Thus, the invention relates to sequences of the
invention in the form of DNA or RNA molecules, or hybrid DNA/RNA molecules,
35 and their corresponding complementary DNA, RNA, or DNA/RNA strands.

40 In a first such aspect, the invention provides nucleic acid molecules comprising
one or more nucleotide sequences encoding *attB1*, or mutants, fragments, variants
25 or derivatives thereof. Such nucleic acid molecules may comprise an *attB1*
nucleotide sequence having the sequence set forth in Figure 9, such as:
ACAAGTTTGTACAAAAAAGCAGGCT, or a nucleotide sequence
45 complementary to the nucleotide sequence set forth in Figure 9 for *attB1*, or
mutants, fragments, variants or derivatives thereof. As one of ordinary skill will
30 appreciate, however, certain mutations, insertions, or deletions of one or more
bases in the *attB1* sequence contained in the nucleic acid molecules of the
50 invention may be made without compromising the structural and functional

integrity of these molecules; hence, nucleic acid molecules comprising such mutations, insertions, or deletions in the *attB1* sequence are encompassed within the scope of the invention.

In a related aspect, the invention provides nucleic acid molecules comprising one or more nucleotide sequences encoding *attB2*, or mutants, fragments, variants or derivatives thereof. Such nucleic acid molecules may comprise an *attB2* nucleotide sequence having the sequence set forth in Figure 9, such as: ACCCAGCTTTCTTGTAACAAGTGGT, or a nucleotide sequence complementary to the nucleotide sequence set forth in Figure 9 for *attB2*, or mutants, fragments, variants or derivatives thereof. As noted above for *attB1*, certain mutations, insertions, or deletions of one or more bases in the *attB2* sequence contained in the nucleic acid molecules of the invention may be made without compromising the structural and functional integrity of these molecules; hence, nucleic acid molecules comprising such mutations, insertions, or deletions in the *attB2* sequence are encompassed within the scope of the invention.

A recombinant host cell comprising a nucleic acid molecule containing *attB1* and *attB2* sites (the vector pEXP501, also known as pCMVSPORT6; see Figure 48), *E. coli* DB3.1(pCMVSPORT6), was deposited on February 27, 1999, with the Collection, Agricultural Research Culture Collection (NRRL), 1815 North University Street, Peoria, Illinois 61604 USA, as Deposit No. NRRL B-30108. The *attB1* and *attB2* sites within the deposited nucleic acid molecule are contained in nucleic acid cassettes in association with one or more additional functional sequences as described in more detail below.

In another related aspect, the invention provides nucleic acid molecules comprising one or more nucleotide sequences encoding *attP1*, or mutants, fragments, variants or derivatives thereof. Such nucleic acid molecules may comprise an *attP1* nucleotide sequence having the sequence set forth in Figure 9, such as: TACAGGTCATAATACCATCTAAGTAGTTGATTCATAGTGA-CTGGATATGTTGTGTTTACAGTATTATGTAGTCTGTTTTTAT-GCAAAATCTAATTTAATATATTGATATTTATATCATTTCAGTT-TCTCGTTCAGCTTTTTGTACAAAGTTGGCATTATAAAAAAGCATTG-CTCATCAATTTGTTGCAACGAACAGGTCATATCAGTCAAAATAA-

5 AATCATTATTTG, or a nucleotide sequence complementary to the nucleotide
sequence set forth in Figure 9 for *attP1*, or mutants, fragments, variants or
10 derivatives thereof. As noted above for *attB1*, certain mutations, insertions, or
deletions of one or more bases in the *attP1* sequence contained in the nucleic acid
5 molecules of the invention may be made without compromising the structural and
functional integrity of these molecules; hence, nucleic acid molecules comprising
15 such mutations, insertions, or deletions in the *attP1* sequence are encompassed
within the scope of the invention.

In another related aspect, the invention provides nucleic acid molecules
10 comprising one or more nucleotide sequences encoding *attP2*, or mutants,
fragments, variants or derivatives thereof. Such nucleic acid molecules may
comprise an *attP2* nucleotide sequence having the sequence set forth in Figure 9,
such as: CAAATAATGATTTTATTTTGACTGATAGTGACCTGTTTCGTTG-
25 CAACAAATTGATAAGCAATGCTTTCTTATAATGCCAACTTT-
GTACAAGAAAGCTGAACGAGAAACGTAAAATGATA-
15 TAAATATCAATATATTAAATTAGATTTTGCATAAAAAACAG-
ACTACATAATACTGTAAAACACAACATATCCAGTCACTATGAATCAA-
30 CTACTTAGATGGTATTAGTGACCTGTA, or a nucleotide sequence
complementary to the nucleotide sequence set forth in Figure 9 for *attP2*, or
20 mutants, fragments, variants or derivatives thereof. As noted above for *attB1*,
certain mutations, insertions, or deletions of one or more bases in the *attP2*
35 sequence contained in the nucleic acid molecules of the invention may be made
without compromising the structural and functional integrity of these molecules;
hence, nucleic acid molecules comprising such mutations, insertions, or deletions
40 in the *attP2* sequence are encompassed within the scope of the invention.

A recombinant host cell comprising a nucleic acid molecule (the *attP* vector
pDONR201, also known as pENTR21-*attPkan* or p*AttPkan*; see Figure 49)
45 containing *attP1* and *attP2* sites, *E. coli* DB3.1(p*AttPkan*) (also called *E. coli*
DB3.1(pAHK*kan*)), was deposited on February 27, 1999, with the Collection,
30 Agricultural Research Culture Collection (NRRL), 1815 North University Street,
Peoria, Illinois 61604 USA, as Deposit No. NRRL B-30099. The *attP1* and *attP2*
50 sites within the deposited nucleic acid molecule are contained in nucleic acid

cassettes in association with one or more additional functional sequences as described in more detail below.

In another related aspect, the invention provides nucleic acid molecules comprising one or more nucleotide sequences encoding *attR1*, or mutants, fragments, variants or derivatives thereof. Such nucleic acid molecules may comprise an *attR1* nucleotide sequence having the sequence set forth in Figure 9, such as: A C A A G T T T G T A C A A A A A G C T G A A C G A G - A A A C G T A A A A T G A T A T A A A T A T C A A T A T A T T A A A T T A G A T T T T G C A T - A A A A A C A G A C T A C A T A A T A C T G T A A A A C A C A A C A T A T C C A G T C A - C T A T G, or a nucleotide sequence complementary to the nucleotide sequence set forth in Figure 9 for *attR1*, or mutants, fragments, variants or derivatives thereof. As noted above for *attB1*, certain mutations, insertions, or deletions of one or more bases in the *attR1* sequence contained in the nucleic acid molecules of the invention may be made without compromising the structural and functional integrity of these molecules; hence, nucleic acid molecules comprising such mutations, insertions, or deletions in the *attR1* sequence are encompassed within the scope of the invention.

In another related aspect, the invention provides nucleic acid molecules comprising one or more nucleotide sequences encoding *attR2*, or mutants, fragments, variants or derivatives thereof. Such nucleic acid molecules may comprise an *attR2* nucleotide sequence having the sequence set forth in Figure 9, such as: G C A G G T C G A C C A T A G T G A C T G G A T A T - G T T G T G T T T T A C A G T A T T A T G T A G T C T G T T T T T A T G C A A A A T C T A - A T T T A A T A T A T T G A T A T T T A T A T C A T T T T A C G T T T C T C G T T C A G C T T - T C T T G T A C A A G T G G T, or a nucleotide sequence complementary to the nucleotide sequence set forth in Figure 9 for *attR2*, or mutants, fragments, variants or derivatives thereof. As noted above for *attB1*, certain mutations, insertions, or deletions of one or more bases in the *attR2* sequence contained in the nucleic acid molecules of the invention may be made without compromising the structural and functional integrity of these molecules; hence, nucleic acid molecules comprising such mutations, insertions, or deletions in the *attR2* sequence are encompassed within the scope of the invention.

5 Recombinant host cell strains containing attR1 sites apposed to cloning sites
in reading frame A, reading frame B, and reading frame C, *E. coli*
10 DB3.1(pEZC15101) (reading frame A; see Figure 64A), *E. coli*
DB3.1(pEZC15102) (reading frame B; see Figure 64B), and *E. coli*
5 DB3.1(pEZC15103) (reading frame C; see Figure 64C), and containing
corresponding attR2 sites, were deposited on February 27, 1999, with the
15 Collection, Agricultural Research Culture Collection (NRRL), 1815 North
University Street, Peoria, Illinois 61604 USA, as Deposit Nos. NRRL B-30103,
NRRL B-30104, and NRRL B-30105, respectively. The attR1 and attR2 sites
10 within the deposited nucleic acid molecules are contained in nucleic acid cassettes
in association with one or more additional functional sequences as described in
20 more detail below.

In another related aspect, the invention provides nucleic acid molecules
25 comprising one or more nucleotide sequences encoding attL1, or mutants,
15 fragments, variants and derivatives thereof. Such nucleic acid molecules may
comprise an attL1 nucleotide sequence having the sequence set forth in Figure 9,
such as: CAA ATA ATG ATT TTA TTT TGA CTG ATA GTG ACC TGT TCG
30 TTG CAA CAA ATT GAT AAG CAA TGC TTT TTT ATA ATG CCA ACT
TTG TAC AAA AAA GCA GGC T, or a nucleotide sequence complementary to
20 the nucleotide sequence set forth in Figure 9 for attL1, or mutants, fragments,
35 variants or derivatives thereof. As noted above for attB1, certain mutations,
insertions, or deletions of one or more bases in the attL1 sequence contained in
the nucleic acid molecules of the invention may be made without compromising
40 the structural and functional integrity of these molecules; hence, nucleic acid
25 molecules comprising such mutations, insertions, or deletions in the attL1
sequence are encompassed within the scope of the invention.

In another related aspect, the invention provides nucleic acid molecules
45 comprising one or more nucleotide sequences encoding attL2, or mutants,
fragments, variants and derivatives thereof. Such nucleic acid molecules may
30 comprise an attL2 nucleotide sequence having the sequence set forth in Figure 9,
such as: C AAA TAA TGA TTT TAT TTT GAC TGA TAG TGA CCT GTT
50 CGT TGC AAC AAA TTG ATA AGC AAT GCT TTC TTA TAA TGC CAA

CTT TGT ACA AGA AAG CTG GGT, or a nucleotide sequence complementary to the nucleotide sequence set forth in Figure 9 for *attL2*, or mutants, fragments, variants or derivatives thereof. As noted above for *attB1*, certain mutations, insertions, or deletions of one or more bases in the *attL2* sequence contained in the nucleic acid molecules of the invention may be made without compromising the structural and functional integrity of these molecules; hence, nucleic acid molecules comprising such mutations, insertions, or deletions in the *attL2* sequence are encompassed within the scope of the invention.

Recombinant host cell strains containing *attL1* sites apposed to cloning sites in reading frame A, reading frame B, and reading frame C, *E. coli* DB3.1(pENTR1A) (reading frame A; see Figure 10), *E. coli* DB3.1(pENTR2B) (reading frame B; see Figure 11), and *E. coli* DB3.1(pENTR3C) (reading frame C; see Figure 12), and containing corresponding *attL2* sites, were deposited on February 27, 1999, with the Collection, Agricultural Research Culture Collection (NRRL), 1815 North University Street, Peoria, Illinois 61604 USA, as Deposit Nos. NRRL B-30100, NRRL B-30101, and NRRL B-30102, respectively. The *attL1* and *attL2* sites within the deposited nucleic acid molecules are contained in nucleic acid cassettes in association with one or more additional functional sequences as described in more detail below.

Each of the recombination site sequences described herein or portions thereof, or the nucleotide sequence cassettes contained in the deposited clones, may be cloned or inserted into a vector of interest (for example, using the recombinational cloning methods described herein and/or standard restriction cloning techniques that are routine in the art) to generate, for example, Entry Vectors or Destination Vectors which may be used to transfer a desired segment of a nucleic acid molecule of interest (*e.g.*, a gene, cDNA molecule, or cDNA library) into a desired vector or into a host cell.

Using the information provided herein, such as the nucleotide sequences for the recombination site sequences described herein, an isolated nucleic acid molecule of the present invention encoding one or more recombination sites or portions thereof may be obtained using standard cloning and screening procedures, such as those for cloning cDNAs using mRNA as starting material. Preferred such

5 methods include PCR-based cloning methods, such as reverse transcriptase-PCR
(RT-PCR) using primers such as those described herein and in the Examples
10 below. Alternatively, vectors comprising the cassettes containing the
recombination site sequences described herein are available commercially from
5 Life Technologies, Inc. (Rockville, MD).

15 The invention is also directed to nucleic acid molecules comprising one or
more of the recombination site sequences or portions thereof and one or more
additional nucleotide sequences, which may encode functional or structural sites
20 such as one or more multiple cloning sites, one or more transcription termination
sites, one or more transcriptional regulatory sequences (which may be promoters,
enhancers, repressors, and the like), one or more translational signals (*e.g.*,
secretion signal sequences), one or more origins of replication, one or more fusion
25 partner peptides (particularly glutathione S-transferase (GST), hexahistidine
(His₆), and thioredoxin (Trx)), one or more selection markers or modules, one or
15 more nucleotide sequences encoding localization signals such as nuclear
localization signals or secretion signals, one or more origins of replication, one or
more protease cleavage sites, one or more genes or portions of genes encoding a
30 protein or polypeptide of interest, and one or more 5' polynucleotide extensions
(particularly an extension of guanine residues ranging in length from about 1 to
20 about 20, from about 2 to about 15, from about 3 to about 10, from about 4 to
35 about 10, and most preferably an extension of 4 or 5 guanine residues at the 5' end
of the recombination site nucleotide sequence. The one or more additional
functional or structural sequences may or may not flank one or more of the
40 recombination site sequences contained on the nucleic acid molecules of the
25 invention.

45 In some nucleic acid molecules of the invention, the one or more nucleotide
sequences encoding one or more additional functional or structural sites may be
operably linked to the nucleotide sequence encoding the recombination site. For
example, certain nucleic acid molecules of the invention may have a promoter
30 sequence operably linked to a nucleotide sequence encoding a recombination site
or portion thereof of the invention, such as a T7 promoter, a phage lambda PL
50

5 promoter, an *E. coli lac*, *trp* or *tac* promoter, and other suitable promoters which will be familiar to the skilled artisan.

10 Nucleic acid molecules of the present invention, which may be isolated nucleic acid molecules, may be in the form of RNA, such as mRNA, or in the form of DNA, including, for instance, cDNA and genomic DNA obtained by cloning or
5 produced synthetically, or in the form of DNA-RNA hybrids. The nucleic acid molecules of the invention may be double-stranded or single-stranded.
15 Single-stranded DNA or RNA may be the coding strand, also known as the sense strand, or it may be the non-coding strand, also referred to as the anti-sense strand. The nucleic acid molecules of the invention may also have a number of
10 topologies, including linear, circular, coiled, or supercoiled.
20

By "isolated" nucleic acid molecule(s) is intended a nucleic acid molecule, DNA or RNA, which has been removed from its native environment. For
25 example, recombinant DNA molecules contained in a vector are considered isolated for the purposes of the present invention. Further examples of isolated
15 DNA molecules include recombinant DNA molecules maintained in heterologous host cells, and those DNA molecules purified (partially or substantially) from a
30 solution whether produced by recombinant DNA or synthetic chemistry techniques. Isolated RNA molecules include *in vivo* or *in vitro* RNA transcripts of the DNA molecules of the present invention.
20

35 The present invention further relates to mutants, fragments, variants and derivatives of the nucleic acid molecules of the present invention, which encode portions, analogs or derivatives of one or more recombination sites. Variants may
40 occur naturally, such as a natural allelic variant. By an "allelic variant" is intended
25 one of several alternate forms of a gene occupying a given locus on a chromosome of an organism (*see* Lewin, B., ed., *Genes II*, , John Wiley & Sons, New York (1985)). Non-naturally occurring variants may be produced using art-known
45 mutagenesis techniques, such as those described hereinbelow.

Such variants include those produced by nucleotide substitutions, deletions or
30 additions or portions thereof, or combinations thereof. The substitutions, deletions or additions may involve one or more nucleotides. The variants may be
50 altered in coding regions, non-coding regions, or both. Alterations in the coding

regions may produce conservative or non-conservative amino acid substitutions, deletions or additions. Especially preferred among these are silent substitutions, additions and deletions, which do not alter the properties and activities of the encoded polypeptide(s) or portions thereof, and which also do not substantially alter the reactivities of the recombination site nucleic acid sequences in recombination reactions. Also especially preferred in this regard are conservative substitutions.

Particularly preferred mutants, fragments, variants, and derivatives of the nucleic acid molecules of the invention include, but are not limited to, insertions, deletions or substitutions of one or more nucleotide bases within the 15 bp core region (GCTTTTTTATACTAA) which is identical in all four wildtype lambda *att* sites, *attB*, *attP*, *attL* and *attR* (see U.S. Application Nos. 08/663,002, filed June 7, 1996 (now U.S. Patent No. 5,888,732), 09/005,476, filed January 12, 1998, and 09/177,387, filed October 23, 1998, which describes the core region in further detail, and the disclosures of which are incorporated herein by reference in their entireties). Analogously, the core regions in *attB1*, *attP1*, *attL1* and *attR1* are identical to one another, as are the core regions in *attB2*, *attP2*, *attL2* and *attR2*. Particularly preferred in this regard are nucleic acid molecules comprising insertions, deletions or substitutions of one or more nucleotides within the seven bp overlap region (TTTATAC, which is defined by the cut sites for the integrase protein and is the region where strand exchange takes place) that occurs within this 15 bp core region (GCTTTTTTATACTAA). Examples of such preferred mutants, fragments, variants and derivatives according to this aspect of the invention include, but are not limited to, nucleic acid molecules in which the thymine at position 1 of the seven bp overlap region has been deleted or substituted with a guanine, cytosine, or adenine; in which the thymine at position 2 of the seven bp overlap region has been deleted or substituted with a guanine, cytosine, or adenine; in which the thymine at position 3 of the seven bp overlap region has been deleted or substituted with a guanine, cytosine, or adenine; in which the adenine at position 4 of the seven bp overlap region has been deleted or substituted with a guanine, cytosine, or thymine; in which the thymine at position 5 of the seven bp overlap region has been deleted or substituted with a

5 guanine, cytosine, or adenine; in which the adenine at position 6 of the seven bp
overlap region has been deleted or substituted with a guanine, cytosine, or
10 thymine; and in which the cytosine at position 7 of the seven bp overlap region has
been deleted or substituted with a guanine, thymine, or adenine; or any
5 combination of one or more such deletions and/or substitutions within this seven
bp overlap region. As described in detail in Example 21 herein, mutants of the
15 nucleic acid molecules of the invention in which substitutions have been made
within the first three positions of the seven bp overlap (TTTATAC) have been
found in the present invention to strongly affect the specificity of recombination,
20 mutant nucleic acid molecules in which substitutions have been made in the last
four positions (TTTATAC) only partially alter recombination specificity, and
mutant nucleic acid molecules comprising nucleotide substitutions outside of the
seven bp overlap, but elsewhere within the 15 bp core region, do not affect
25 specificity of recombination but do influence the efficiency of recombination.

15 Hence, in an additional aspect, the present invention is also directed to nucleic
acid molecules comprising one or more recombination site nucleotide sequences
that affect recombination specificity, particularly one or more nucleotide
30 sequences that may correspond substantially to the seven base pair overlap within
the 15 bp core region, having one or more mutations that affect recombination
20 specificity. Particularly preferred such molecules may comprise a consensus
sequence (described in detail in Example 21 herein) such as NNNATAC, wherein
35 "N" refers to any nucleotide (*i.e.*, may be A, G, T/U or C), with the proviso that
if one of the first three nucleotides in the consensus sequence is a T/U, then at
least one of the other two of the first three nucleotides is not a T/U.

40 25 In a related aspect, the present invention is also directed to nucleic acid
molecules comprising one or more recombination site nucleotide sequences that
enhance recombination efficiency, particularly one or more nucleotide sequences
45 that may correspond substantially to the core region and having one or more
mutations that enhance recombination efficiency. By sequences or mutations that
30 "enhance recombination efficiency" is meant a sequence or mutation in a
recombination site, preferably in the core region (*e.g.*, the 15 bp core region of *att*
50 recombination sites), that results in an increase in cloning efficiency (typically

measured by determining successful cloning of a test sequence, *e.g.*, by determining CFU/ml for a given cloning mixture) when recombining molecules comprising the mutated sequence or core region as compared to molecules that do not comprise the mutated sequence or core region (*e.g.*, those comprising a wildtype recombination site core region sequence). More specifically, whether or not a given sequence or mutation enhances recombination efficiency may be determined using the sequence or mutation in recombinational cloning as described herein, and determining whether the sequence or mutation provides enhanced recombinational cloning efficiency when compared to a non-mutated (*e.g.*, wildtype) sequence. Methods of determining preferred cloning efficiency-enhancing mutations for a number of recombination sites, particularly for *att* recombination sites, are described herein, for example in Examples 22-25. Examples of preferred such mutant recombination sites include but are not limited to the *attL* consensus core sequence of caactntntnnannaagttg (wherein "n" represents any nucleotide), for example the *attL5* sequence agcctgctttattatactaagttggcatta and the *attL6* sequence agcctgctttttatattaagttggcatta; the *attB1.6* sequence ggggacaactttgtacaaaaagttggct; the *attB2.2* sequence ggggacaactttgtacaagaaagctgggt; and the *attB2.10* sequence ggggacaactttgtacaagaaagttgggt. Those of skill in the art will appreciate that, in addition to the core region, other portions of the *att* site may affect the efficiency of recombination. There are five so-called arm binding sites for the integrase protein in the bacteriophage lambda *attP* site, two in *attR* (P1 and P2), and three in *attL* (P'1, P'2 and P'3). Compared to the core binding sites, the integrase protein binds to arm sites with high affinity and interacts with core and arm sites through two different domains of the protein. As with the core binding site a consensus sequence for the arm binding site consisting of C/AAGTCACTAT has been inferred from sequence comparison of the five arm binding sites and seven non-*att* sites (Ross and Landy, *Proc. Natl. Acad. Sci. USA* 79:7724-7728 (1982)). Each arm site has been mutated and tested for its effect in the excision and integration reactions (Numrych *et al.*, *Nucl. Acids Res.* 18:3953 (1990)). Hence, specific sites are utilized in each reaction in different ways, namely, the P1 and P'3

5 sites are essential for the integration reaction whereas the other three sites are
dispensable to the integration reaction to varying degrees. Similarly, the P2, P'1
10 and P'2 sites are most important for the excision reaction, whereas P1 and P'3 are
completely dispensable. Interestingly, when P2 is mutated the integration reaction
5 occurs more efficiently than with the wild type attP site. Similarly, when P1 and
P'3 are mutated the excision reaction occurs more efficiently. The stimulatory
15 effect of mutating integrase arm binding sites can be explained by removing sites
that compete or inhibit a specific recombination pathway or that function in a
reaction that converts products back to starting substrates. In fact there is
10 evidence for an XIS-independent LR reaction (Abremski and Gottesman, *J. Mol.*
Biol. 153:67-78 (1981)). Thus, in addition to modifications in the core region of
the att site, the present invention contemplates the use of att sites containing one
or more modifications in the integrase arm-type binding sites. In some preferred
25 embodiments, one or more mutations may be introduced into one or more of the
P1, P'1, P2, P'2 and P'3 sites. In some preferred embodiments, multiple mutations
15 may be introduced into one or more of these sites. Preferred such mutations
include those which increase the recombination *in vitro*. For example, in some
embodiments mutations may be introduced into the arm-type binding sites such
30 that integrative recombination, corresponding to the BP reaction, is enhanced. In
other embodiments, mutations may be introduced into the arm-type binding sites
20 such that excisive recombination, corresponding to the LR reaction, is enhanced.
Of course, based on the guidance contained herein, particularly in the construction
and evaluation of effects of mutated recombination sites upon recombinational
35 specificity and efficiency, analogous mutated or engineered sequences may be
produced for other recombination sites described herein (including but not limited
40 to *lox*, FRT, and the like) and used in accordance with the invention. For
example, much like the mutagenesis strategy used to select core binding sites that
enhance recombination efficiency, similar strategies can be employed to select
45 changes in the arms of attP, attL and attR, and in analogous sequences in other
recombination sites such as *lox*, FRT and the like, that enhance recombination
30 efficiency. Hence, the construction and evaluation of such mutants is well within
the abilities of those of ordinary skill in the art without undue experimentation.

55

One suitable methodology for preparing and evaluating such mutations is found in Numrych, *et al.*, (1990) *Nucleic Acids Research* 18(13): 3953-3959.

Other mutant sequences and nucleic acid molecules that may be suitable to enhance recombination efficiency will be apparent from the description herein, or may be easily determined by one of ordinary skill using only routine experimentation in molecular biology in view of the description herein and information that is readily available in the art

Since the genetic code is well known in the art, it is also routine for one of ordinary skill in the art to produce degenerate variants of the nucleic acid molecules described herein without undue experimentation. Hence, nucleic acid molecules comprising degenerate variants of nucleic acid sequences encoding the recombination sites described herein are also encompassed within the scope of the invention.

Further embodiments of the invention include isolated nucleic acid molecules comprising a polynucleotide having a nucleotide sequence at least 50% identical, at least 60% identical, at least 70% identical, at least 75% identical, at least 80% identical, at least 85% identical, at least 90% identical, and more preferably at least 95%, 96%, 97%, 98% or 99% identical to the nucleotide sequences of the seven bp overlap region within the 15 bp core region of the recombination sites described herein, or the nucleotide sequences of *attB1*, *attB2*, *attP1*, *attP2*, *attL1*, *attL2*, *attR1* or *attR2* as set forth in Figure 9 (or portions thereof), or a nucleotide sequence complementary to any of these nucleotide sequences, or fragments, variants, mutants, and derivatives thereof.

By a polynucleotide having a nucleotide sequence at least, for example, 95% "identical" to a reference nucleotide sequence encoding a particular recombination site or portion thereof is intended that the nucleotide sequence of the polynucleotide is identical to the reference sequence except that the polynucleotide sequence may include up to five point mutations (*e.g.*, insertions, substitutions, or deletions) per each 100 nucleotides of the reference nucleotide sequence encoding the recombination site. For example, to obtain a polynucleotide having a nucleotide sequence at least 95% identical to a reference *attB1* nucleotide sequence, up to 5% of the nucleotides in the *attB1* reference sequence may be

5 deleted or substituted with another nucleotide, or a number of nucleotides up to
5% of the total nucleotides in the *attB1* reference sequence may be inserted into
10 the *attB1* reference sequence. These mutations of the reference sequence may
occur at the 5' or 3' terminal positions of the reference nucleotide sequence or
5 anywhere between those terminal positions, interspersed either individually among
nucleotides in the reference sequence or in one or more contiguous groups within
15 the reference sequence.

As a practical matter, whether any particular nucleic acid molecule is at least
50%, 60%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98% or 99% identical
10 to, for instance, a given recombination site nucleotide sequence or portion thereof
can be determined conventionally using known computer programs such as
20 DNAsis software (Hitachi Software, San Bruno, California) for initial sequence
alignment followed by ESEE version 3.0 DNA/protein sequence software
(cabot@trog.mbb.sfu.ca) for multiple sequence alignments. Alternatively, such
25 determinations may be accomplished using the BESTFIT program (Wisconsin
Sequence Analysis Package, Genetics Computer Group, University Research Park,
575 Science Drive, Madison, WI 53711), which employs a local homology
30 algorithm (Smith and Waterman, *Advances in Applied Mathematics* 2: 482-489
(1981)) to find the best segment of homology between two sequences. When
20 using DNAsis, ESEE, BESTFIT or any other sequence alignment program to
determine whether a particular sequence is, for instance, 95% identical to a
35 reference sequence according to the present invention, the parameters are set such
that the percentage of identity is calculated over the full length of the reference
nucleotide sequence and that gaps in homology of up to 5% of the total number
40 of nucleotides in the reference sequence are allowed.

The present invention is directed to nucleic acid molecules at least 50%, 60%,
70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98% or 99% identical to the *attB1*,
45 *attB2*, *attP1*, *attP2*, *attL1*, *attL2*, *attR1* or *attR2* nucleotide sequences as set forth
in Figure 9, or to the nucleotide sequence of the deposited clones, irrespective of
30 whether they encode particular functional polypeptides. This is because even
where a particular nucleic acid molecule does not encode a particular functional
50 polypeptide, one of skill in the art would still know how to use the nucleic acid

5 molecule, for instance, as a hybridization probe or a polymerase chain reaction (PCR) primer.

10 Mutations can also be introduced into the recombination site nucleotide sequences for enhancing site specific recombination or altering the specificities of the reactants, etc. Such mutations include, but are not limited to: recombination
5 sites without translation stop codons that allow fusion proteins to be encoded; recombination sites recognized by the same proteins but differing in base sequence
15 such that they react largely or exclusively with their homologous partners allowing multiple reactions to be contemplated; and mutations that prevent hairpin formation of recombination sites. Which particular reactions take place can be
20 specified by which particular partners are present in the reaction mixture.

There are well known procedures for introducing specific mutations into nucleic acid sequences. A number of these are described in Ausubel, F.M. *et al.*,
25 *Current Protocols in Molecular Biology*, Wiley Interscience, New York (1989-1996). Mutations can be designed into oligonucleotides, which can be used to modify existing cloned sequences, or in amplification reactions. Random
15 mutagenesis can also be employed if appropriate selection methods are available to isolate the desired mutant DNA or RNA. The presence of the desired mutations can be confirmed by sequencing the nucleic acid by well known
30 methods.

20 The following non-limiting methods can be used to modify or mutate a given nucleic acid molecule encoding a particular recombination site to provide mutated sites that can be used in the present invention:

- 35 1. By recombination of two parental DNA sequences by site-specific (e.g. attL and attR to give attP) or other (e.g. homologous) recombination mechanisms where the parental DNA segments contain one or more base alterations resulting in the final mutated nucleic acid molecule;
- 40 2. By mutation or mutagenesis (site-specific, PCR, random, spontaneous, etc) directly of the desired nucleic acid molecule;
- 45 3. By mutagenesis (site-specific, PCR, random, spontaneous, etc) of parental DNA sequences, which are recombined to generate a desired nucleic acid molecule;
- 50

4. By reverse transcription of an RNA encoding the desired core sequence;
and

5. By *de novo* synthesis (chemical synthesis) of a sequence having the desired
base changes, or random base changes followed by sequencing or
functional analysis according to methods that are routine in the art.

The functionality of the mutant recombination sites can be demonstrated in
ways that depend on the particular characteristic that is desired. For example, the
lack of translation stop codons in a recombination site can be demonstrated by
expressing the appropriate fusion proteins. Specificity of recombination between
homologous partners can be demonstrated by introducing the appropriate
molecules into *in vitro* reactions, and assaying for recombination products as
described herein or known in the art. Other desired mutations in recombination
sites might include the presence or absence of restriction sites, translation or
transcription start signals, protein binding sites, particular coding sequences, and
other known functionalities of nucleic acid base sequences. Genetic selection
schemes for particular functional attributes in the recombination sites can be used
according to known method steps. For example, the modification of sites to
provide (from a pair of sites that do not interact) partners that do interact could
be achieved by requiring deletion, via recombination between the sites, of a DNA
sequence encoding a toxic substance. Similarly, selection for sites that remove
translation stop sequences, the presence or absence of protein binding sites, etc.,
can be easily devised by those skilled in the art.

Accordingly, the present invention also provides a nucleic acid molecule,
comprising at least one DNA segment having at least one, and preferably at least
two, engineered recombination site nucleotide sequences of the invention flanking
a selectable marker and/or a desired DNA segment, wherein at least one of said
recombination site nucleotide sequences has at least one engineered mutation that
enhances recombination *in vitro* in the formation of a Cointegrate DNA or a
Product DNA. Such engineered mutations may be in the core sequence of the
recombination site nucleotide sequence of the invention; *see* U.S. Application Nos.
08/486,139, filed June 7, 1995, 08/663,002, filed June 7, 1996 (now U.S. Patent
No. 5,888,732), 09/005,476, filed January 12, 1998, and 09/177,387, filed

October 23, 1998, the disclosures of which are all incorporated herein by reference in their entireties.

While in the preferred embodiment the recombination sites differ in sequence and do not interact with each other, it is recognized that sites comprising the same sequence, which may interact with each other, can be manipulated or engineered to inhibit recombination with each other. Such conceptions are considered and incorporated herein. For example, a protein binding site (*e.g.*, an antibody-binding site, a histone-binding site, an enzyme-binding site, or a binding site for any nucleic acid molecule-binding protein) can be engineered adjacent to one of the sites. In the presence of the protein that recognizes the engineered site, the recombinase fails to access the site and another recombination site in the nucleic acid molecule is therefore used preferentially. In the cointegrate this site can no longer react since it has been changed, *e.g.*, from attB to attL. During or upon resolution of the cointegrate, the protein can be inactivated (*e.g.*, by antibody, heat or a change of buffer) and the second site can undergo recombination.

The nucleic acid molecules of the invention can have at least one mutation that confers at least one enhancement of said recombination, said enhancement selected from the group consisting of substantially (i) favoring integration; (ii) favoring recombination; (ii) relieving the requirement for host factors; (iii) increasing the efficiency of said Cointegrate DNA or Product DNA formation; (iv) increasing the specificity of said Cointegrate DNA or Product DNA formation; and (v) adding or deleting protein binding sites.

In other embodiments, the nucleic acid molecules of the invention may be PCR primer molecules, which comprise one or more of the recombination site sequences described herein or portions thereof, particularly those shown in Figure 9 (or sequences complementary to those shown in Figure 9), or mutants, fragments, variants or derivatives thereof, attached at the 3' end to a target-specific template sequence which specifically interacts with a target nucleic acid molecule which is to be amplified. Primer molecules according to this aspect of the invention may further comprise one or more, (*e.g.*, 1, 2, 3, 4, 5, 10, 20, 25, 50, 100, 500, 1000, or more) additional bases at their 5' ends, and preferably comprise one or more (particularly four or five) additional bases, which are preferably

guanines, at their 5' ends, to increase the efficiency of the amplification products incorporating the primer molecules in the recombinational cloning system of the invention. Such nucleic acid molecules and primers are described in detail in the examples herein, particularly in Examples 22-25.

Certain primers of the invention may comprise one or more nucleotide deletions in the *attB1*, *attB2*, *attP1*, *attP2*, *attL1*, *attL2*, *attR1* or *attR2* sequences as set forth in Figure 9. In one such aspect, for example, *attB2* primers may be constructed in which one or more of the first four nucleotides at the 5' end of the *attB2* sequence shown in Figure 9 have been deleted. Primers according to this aspect of the invention may therefore have the sequence:

(*attB2*(-1)): CCCAGCTTCTTGTACAAAGTGGTnnnnnnnnnnnn . . . n

(*attB2*(-2)): CCAGCTTCTTGTACAAAGTGGTnnnnnnnnnnnn . . . n

(*attB2*(-3)): CAGCTTCTTGTACAAAGTGGTnnnnnnnnnnnn . . . n

(*attB2*(-4)): AGCTTCTTGTACAAAGTGGTnnnnnnnnnnnn . . . n,

wherein "nnnnnnnnnnnn . . . n" at the 3' end of the primer represents a target-specific sequence of any length, for example from one base up to all of the bases of a target nucleic acid molecule (e.g., a gene) or a portion thereof, the sequence and length which will depend upon the identity of the target nucleic acid molecule which is to be amplified.

The primer nucleic acid molecules according to this aspect of the invention may be produced synthetically by attaching the recombination site sequences depicted in Figure 9, or portions thereof, to the 5' end of a standard PCR target-specific primer according to methods that are well-known in the art. Alternatively, additional primer nucleic acid molecules of the invention may be produced synthetically by adding one or more nucleotide bases, which preferably correspond to one or more, preferably five or more, and more preferably six or more, contiguous nucleotides of the *att* nucleotide sequences described herein (see, e.g., Example 20 herein; see also U.S. Application Nos. 08/663,002, filed June 7, 1996 (now U.S. Patent No. 5,888,732), 09/005,476, filed January 12, 1998, and 09/177,387, filed October 23, 1998, the disclosures of which are all incorporated herein by reference in their entireties), to the 5' end of a standard PCR target-specific primer according to methods that are well-known in the art, to provide

primers having the specific nucleotide sequences described herein. As noted above, primer nucleic acid molecules according to this aspect of the invention may also optionally comprise one, two, three, four, five, or more additional nucleotide bases at their 5' ends, and preferably will comprise four or five guanines at their 5' ends. In one particularly preferred such aspect, the primer nucleic acid molecules of the invention may comprise one or more, preferably five or more, more preferably six or more, still more preferably 6-18 or 6-25, and most preferably 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24 or 25, contiguous nucleotides or bp of the *attB1* or *attB2* nucleotide sequences depicted in Figure 9 (or nucleotides complementary thereto), linked to the 5' end of a target-specific (e.g., a gene-specific) primer molecule. Primer nucleic acid molecules according to this aspect of the invention include, but are not limited to, *attB1*- and *attB2*-derived primer nucleic acid molecules having the following nucleotide sequences:

ACAAGTTTGTACAAAAAGCAGGCT-nnnnnnnnnnnnnnnnn . . . n
ACCACTTTGTACAAGAAAGCTGGGT-nnnnnnnnnnnnnnnnn . . . n
TGTACAAAAAGCAGGCT-nnnnnnnnnnnnnnnnn . . . n
TGTACAAGAAAGCTGGGT-nnnnnnnnnnnnnnnnn . . . n
ACAAAAAGCAGGCT-nnnnnnnnnnnnnnnnn . . . n
ACAAGAAAGCTGGGT-nnnnnnnnnnnnnnnnn . . . n
AAAAAGCAGGCT-nnnnnnnnnnnnnnnnn . . . n
AGAAAGCTGGGT-nnnnnnnnnnnnnnnnn . . . n
AAAAGCAGGCT-nnnnnnnnnnnnnnnnn . . . n
GAAAGCTGGGT-nnnnnnnnnnnnnnnnn . . . n
AAAGCAGGCT-nnnnnnnnnnnnnnnnn . . . n
AAAGCTGGGT-nnnnnnnnnnnnnnnnn . . . n
AAGCAGGCT-nnnnnnnnnnnnnnnnn . . . n
AAGCTGGGT-nnnnnnnnnnnnnnnnn . . . n
AGCAGGCT-nnnnnnnnnnnnnnnnn . . . n
AGCTGGGT-nnnnnnnnnnnnnnnnn . . . n
GCAGGCT-nnnnnnnnnnnnnnnnn . . . n
GCTGGGT-nnnnnnnnnnnnnnnnn . . . n

CAGGCT-nnnnnnnnnnnnn . . . n

CTGGGT-nnnnnnnnnnnnn . . . n,

wherein "nnnnnnnnnnnn . . . n" at the 3' end of the primer represents a target-specific sequence of any length, for example from one base up to all of the bases of a target nucleic acid molecule (*e.g.*, a gene) or a portion thereof, the sequence and length which will depend upon the identity of the target nucleic acid molecule which is to be amplified.

Of course, it will be apparent to one of ordinary skill from the teachings contained herein that additional primer nucleic acid molecules analogous to those specifically described herein may be produced using one or more, preferably five or more, more preferably six or more, still more preferably ten or more, 15 or more, 20 or more, 25 or more, 30 or more, etc. (through to and including all) of the contiguous nucleotides or bp of the *attP1*, *attP2*, *attL1*, *attL2*, *attR1* or *attR2* nucleotide sequences depicted in Figure 9 (or nucleotides complementary thereto), linked to the 5' end of a target-specific (*e.g.*, a gene-specific) primer molecule. As noted above, such primer nucleic acid molecules may optionally further comprise one, two, three, four, five, or more additional nucleotide bases at their 5' ends, and preferably will comprise four guanines at their 5' ends. Other primer molecules comprising the *attB1*, *attB2*, *attP1*, *attP2*, *attL1*, *attL2*, *attR1* and *attR2* sequences depicted in Figure 9, or portions thereof, may be made by one of ordinary skill without resorting to undue experimentation in accordance with the guidance provided herein.

The primers of the invention described herein are useful in producing PCR fragments having a nucleic acid molecule of interest flanked at each end by a recombination site sequence (as described in detail below in Example 9), for use in cloning of PCR-amplified DNA fragments using the recombination system of the invention (as described in detail below in Examples 8, 19 and 21-25).

Vectors

The invention also relates to vectors comprising one or more of the nucleic acid molecules of the invention, as described herein. In accordance with the invention, any vector may be used to construct the vectors of the invention. In

particular, vectors known in the art and those commercially available (and variants or derivatives thereof) may in accordance with the invention be engineered to include one or more nucleic acid molecules encoding one or more recombination sites (or portions thereof), or mutants, fragments, or derivatives thereof, for use in the methods of the invention. Such vectors may be obtained from, for example, Vector Laboratories Inc., InVitrogen, Promega, Novagen, New England Biolabs, Clontech, Roche, Pharmacia, EpiCenter, OriGenes Technologies Inc., Stratagene, Perkin Elmer, Pharmingen, Life Technologies, Inc., and Research Genetics. Such vectors may then for example be used for cloning or subcloning nucleic acid molecules of interest. General classes of vectors of particular interest include prokaryotic and/or eukaryotic cloning vectors, Expression Vectors, fusion vectors, two-hybrid or reverse two-hybrid vectors, shuttle vectors for use in different hosts, mutagenesis vectors, transcription vectors, vectors for receiving large inserts and the like.

Other vectors of interest include viral origin vectors (M13 vectors, bacterial phage λ vectors, bacteriophage P1 vectors, adenovirus vectors, herpesvirus vectors, retrovirus vectors, phage display vectors, combinatorial library vectors), high, low, and adjustable copy number vectors, vectors which have compatible replicons for use in combination in a single host (pACYC184 and pBR322) and eukaryotic episomal replication vectors (pCDM8).

Particular vectors of interest include prokaryotic Expression Vectors such as pcDNA II, pSL301, pSE280, pSE380, pSE420, pTrcHisA, B, and C, pRSET A, B, and C (Invitrogen, Inc.), pGEMEX-1, and pGEMEX-2 (Promega, Inc.), the pET vectors (Novagen, Inc.), pTrc99A, pKK223-3, the pGEX vectors, pEZZ18, pRIT2T, and pMC1871 (Pharmacia, Inc.), pKK233-2 and pKK388-1 (Clontech, Inc.), and pProEx-HT (Life Technologies, Inc.) and variants and derivatives thereof. Destination Vectors can also be made from eukaryotic Expression Vectors such as pFastBac, pFastBac HT, pFastBac DUAL, pSFV, and pTet-Splice (Life Technologies, Inc.), pEUK-C1, pPUR, pMAM, pMAMneo, pBI101, pBI121, pDR2, pCMVEBNA, and pYACneo (Clontech), pSVK3, pSVL, pMSG, pCH110, and pKK232-8 (Pharmacia, Inc.), p3'SS, pXT1, pSG5, pPbac, pMbac, pMC1neo, and pOG44 (Stratagene, Inc.), and pYES2, pAC360, pBlueBacHis A,

B, and C, pVL1392, pBsueBacIII, pCDM8, pcDNA1, pZeoSV, pcDNA3 pREP4, pCEP4, and pEBVHis (Invitrogen, Inc.) and variants or derivatives thereof.

Other vectors of particular interest include pUC18, pUC19, pBlueScript, pSPORT, cosmids, phagemids, YACs (yeast artificial chromosomes), BACs (bacterial artificial chromosomes), MACs (mammalian artificial chromosomes), pQE70, pQE60, pQE9 (Quiagen), pBS vectors, PhageScript vectors, BlueScript vectors, pNH8A, pNH16A, pNH18A, pNH46A (Stratagene), pcDNA3 (Invitrogen), pGEX, pTrsfus, pTrc99A, pET-5, pET-9, pKK223-3, pKK233-3, pDR540, pRIT5 (Pharmacia), pSPORT1, pSPORT2, pCMVSPORT2.0 and pSV-SPORT1 (Life Technologies, Inc.) and variants or derivatives thereof.

Additional vectors of interest include pTrxFus, pThioHis, pLEX, pTrcHis, pTrcHis2, pRSET, pBlueBacHis2, pcDNA3.1/His, pcDNA3.1(-)/Myc-His, pSecTag, pEBVHis, pPIC9K, pPIC3.5K, pAO815, pPICZ, pPICZ α , pGAPZ, pGAPZ α , pBlueBac4.5, pBlueBacHis2, pMelBac, pSinRep5, pSinHis, pIND, pIND(SP1), pVgRXR, pcDNA2.1, pYES2, pZER01.1, pZER0-2.1, pCR-Blunt, pSE280, pSE380, pSE420, pVL1392, pVL1393, pCDM8, pcDNA1.1, pcDNA1.1/Amp, pcDNA3.1, pcDNA3.1/Zeo, pSe,SV2, pRc/CMV2, pRc/RSV, pREP4, pREP7, pREP8, pREP9, pREP10, pCEP4, pEBVHis, pCR3.1, pCR2.1, pCR3.1-Uni, and pCRBac from Invitrogen; λ ExCell, λ gt11, pTrc99A, pKK223-3, pGEX-1 λ T, pGEX-2T, pGEX-2TK, pGEX-4T-1, pGEX-4T-2, pGEX-4T-3, pGEX-3X, pGEX-5X-1, pGEX-5X-2, pGEX-5X-3, pEZZ18, pRIT2T, pMC1871, pSVK3, pSVL, pMSG, pCH110, pKK232-8, pSL1180, pNEO, and pUC4K from Pharmacia; pSCREEN-1b(+), pT7Blue(R), pT7Blue-2, pCITE-4abc(+), pOCUS-2, pTag, pET-32LIC, pET-30LIC, pBAC-2cpLIC, pBACgus-2cpLIC, pT7Blue-2LIC, pT7Blue-2, λ SCREEN-1, λ BlueSTAR, pET-3abcd, pET-7abc, pET9abcd, pET11abcd, pET12abc, pET-14b, pET-15b, pET-16b, pET-17b-pET-17xb, pET-19b, pET-20b(+), pET-21abcd(+), pET-22b(+), pET-23abcd(+), pET-24abcd(+), pET-25b(+), pET-26b(+), pET-27b(+), pET-28abc(+), pET-29abc(+), pET-30abc(+), pET-31b(+), pET-32abc(+), pET-33b(+), pBAC-1, pBACgus-1, pBAC4x-1, pBACgus4x-1, pBAC-3cp, pBACgus-2cp, pBACsurf-1, pIg, Signal pIg, pYX, Selecta Vecta-Neo, Selecta Vecta - Hyg, and Selecta Vecta - Gpt from Novagen; pLexA, pB42AD, pGBT9, pAS2-1,

pGAD424, pACT2, pGAD GL, pGAD GH, pGAD10, pGilda, pEZM3, pEGFP, pEGFP-1, pEGFP-N, pEGFP-C, pEBFP, pGFPuv, pGFP, p6xHis-GFP, pSEAP2-Basic, pSEAP2-Contral, pSEAP2-Promoter, pSEAP2-Enhancer, p β gal-Basic, p β gal-Control, p β gal-Promoter, p β gal-Enhancer, pCMV β , pTet-Off, pTet-On, pTK-Hyg, pRetro-Off, pRetro-On, pIRES1neo, pIRES1hyg, pLXSN, pLNCX, pLAPSN, pMAMneo, pMAMneo-CAT, pMAMneo-LUC, pPUR, pSV2neo, pYEX4T-1/2/3, pYEX-S1, pBacPAK-His, pBacPAK8/9, pAcUW31, BacPAK6, pTriplEx, λ gt10, λ gt11, pWE15, and λ TriplEx from Clontech; Lambda ZAP II, pBK-CMV, pBK-RSV, pBluescript II KS +/-, pBluescript II SK +/-, pAD-GAL4, pBD-GAL4 Cam, pSurfscrip, Lambda FIX II, Lambda DASH, Lambda EMBL3, Lambda EMBL4, SuperCos, pCR-Script Amp, pCR-Script Cam, pCR-Script Direct, pBS +/-, pBC KS +/-, pBC SK +/-, Phagescript, pCAL-n-EK, pCAL-n, pCAL-c, pCAL-kc, pET-3abcd, pET-11abcd, pSPUTK, pESP-1, pCMVLacI, pOPRSVI/MCS, pOPI3 CAT, pXT1, pSG5, pPbac, pMbac, pMC1neo, pMC1neo Poly A, pOG44, pOG45, pFRT β GAL, pNEO β GAL, pRS403, pRS404, pRS405, pRS406, pRS413, pRS414, pRS415, and pRS416 from Stratagene.

Two-hybrid and reverse two-hybrid vectors of particular interest include pPC86, pDBLeu, pDBTrp, pPC97, p2.5, pGAD1-3, pGAD10, pAct, pACT2, pGADGL, pGADGH, pAS2-1, pGAD424, pGBT8, pGBT9, pGAD-GAL4, pLexA, pBD-GAL4, pHISi, pHISi-1, placZi, pB42AD, pDG202, pJK202, pJG4-5, pNLexA, pYESTrp and variants or derivatives thereof.

Yeast Expression Vectors of particular interest include pESP-1, pESP-2, pESC-His, pESC-Trp, pESC-URA, pESC-Leu (Stratagene), pRS401, pRS402, pRS411, pRS412, pRS421, pRS422, and variants or derivatives thereof.

According to the invention, the vectors comprising one or more nucleic acid molecules encoding one or more recombination sites, or mutants, variants, fragments, or derivatives thereof, may be produced by one of ordinary skill in the art without resorting to undue experimentation using standard molecular biology methods. For example, the vectors of the invention may be produced by introducing one or more of the nucleic acid molecules encoding one or more recombination sites (or mutants, fragments, variants or derivatives thereof) into one or more of the vectors described herein, according to the methods described,

for example, in Maniatis *et al.*, *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratory, Cold Spring Harbor, New York (1982). In a related aspect of the invention, the vectors may be engineered to contain, in addition to one or more nucleic acid molecules encoding one or more recombination sites (or portions thereof), one or more additional physical or functional nucleotide sequences, such as those encoding one or more multiple cloning sites, one or more transcription termination sites, one or more transcriptional regulatory sequences (e.g., one or more promoters, enhancers, or repressors), one or more selection markers or modules, one or more genes or portions of genes encoding a protein or polypeptide of interest, one or more translational signal sequences, one or more nucleotide sequences encoding a fusion partner protein or peptide (e.g., GST, His₆ or thioredoxin), one or more origins of replication, and one or more 5' or 3' polynucleotide tails (particularly a poly-G tail). According to this aspect of the invention, the one or more recombination site nucleotide sequences (or portions thereof) may optionally be operably linked to the one or more additional physical or functional nucleotide sequences described herein.

Preferred vectors according to this aspect of the invention include, but are not limited to: pENTR1A (Figures 10A and 10B), pENTR2B (Figures 11A and 11B), pENTR3C (Figures 12A and 12B), pENTR4 (Figures 13A and 13B), pENTR5 (Figures 14A and 14B), pENTR6 (Figures 15A and 15B), pENTR7 (Figures 16A and 16B), pENTR8 (Figures 17A and 17B), pENTR9 (Figures 18A and 18B), pENTR10 (Figures 19A and 19B), pENTR11 (Figures 20A and 20B), pDEST1 (Figures 21A-D), pDEST2 (Figure 22A-D), pDEST3 (Figure 23A-D), pDEST4 (Figure 24A-D), pDEST5 (Figure 25A-D), pDEST6 (Figure 26A-D), pDEST7 (Figure 27A-C), pDEST8 (Figure 28A-D), pDEST9 (Figure 29A-E), pDEST10 (Figure 30A-D), pDEST11 (Figure 31A-D), pDEST12.2 (also known as pDEST12) (Figure 32A-D), pDEST13 (Figure 33A-C), pDEST14 (Figure 34A-D), pDEST15 (Figure 35A-D), pDEST16 (Figure 36A-D), pDEST17 (Figure 37A-D), pDEST18 (Figure 38A-D), pDEST19 (Figure 39A-D), pDEST20 (Figure 40A-D), pDEST21 (Figure 41A-E), pDEST22 (Figure 42A-D), pDEST23 (Figure 43A-D), pDEST24 (Figure 44A-D), pDEST25 (Figure 45A-D), pDEST26 (Figure 46A-D), pDEST27 (Figure 47A-D), pEXP501 (also known

as pCMVSPORT6) (Figure 48A-B), pDONR201 (also known as pENTR21 attP vector or pAttPkan Donor Vector) (Figure 49), pDONR202 (Figure 50), pDONR203 (also known as pEZ15812) (Figure 51), pDONR204 (Figure 52), pDONR205 (Figure 53), pDONR206 (also known as pENTR22 attP vector or pAttPgen Donor Vector) (Figure 54), pMAB58 (Figure 87), pMAB62 (Figure 88), pDEST28 (Figure 90), pDEST29 (Figure 91), pDEST30 (Figure 92), pDEST31 (Figure 93), pDEST32 (Figure 94), pDEST33 (Figure 95), pDEST34 (Figure 96), pDONR207 (Figure 97), pMAB85 (Figure 98), pMAB86 (Figure 99), and fragments, mutants, variants, and derivatives thereof. However, it will be understood by one of ordinary skill that the present invention also encompasses other vectors not specifically designated herein, which comprise one or more of the isolated nucleic acid molecules of the invention encoding one or more recombination sites or portions thereof (or mutants, fragments, variants or derivatives thereof), and which may further comprise one or more additional physical or functional nucleotide sequences described herein which may optionally be operably linked to the one or more nucleic acid molecules encoding one or more recombination sites or portions thereof. Such additional vectors may be produced by one of ordinary skill according to the guidance provided in the present specification.

Polymerases

Preferred polypeptides having reverse transcriptase activity (*i.e.*, those polypeptides able to catalyze the synthesis of a DNA molecule from an RNA template) for use in accordance with the present invention include, but are not limited to Moloney Murine Leukemia Virus (M-MLV) reverse transcriptase, Rous Sarcoma Virus (RSV) reverse transcriptase, Avian Myeloblastosis Virus (AMV) reverse transcriptase, Rous Associated Virus (RAV) reverse transcriptase, Myeloblastosis Associated Virus (MAV) reverse transcriptase, Human Immunodeficiency Virus (HIV) reverse transcriptase, retroviral reverse transcriptase, retrotransposon reverse transcriptase, hepatitis B reverse transcriptase, cauliflower mosaic virus reverse transcriptase and bacterial reverse transcriptase. Particularly preferred are those polypeptides having reverse

transcriptase activity that are also substantially reduced in RNase H activity (*i.e.*, “RNase H” polypeptides). By a polypeptide that is “substantially reduced in RNase H activity” is meant that the polypeptide has less than about 20%, more preferably less than about 15%, 10% or 5%, and most preferably less than about 2%, of the RNase H activity of a wildtype or RNase H⁺ enzyme such as wildtype M-MLV reverse transcriptase. The RNase H activity may be determined by a variety of assays, such as those described, for example, in U.S. Patent No. 5,244,797, in Kotewicz, M.L. *et al.*, *Nucl. Acids Res.* 16:265 (1988) and in Gerard, G.F., *et al.*, *FOCUS* 14(5):91 (1992), the disclosures of all of which are fully incorporated herein by reference. Suitable RNase H⁺ polypeptides for use in the present invention include, but are not limited to, M-MLV H⁺ reverse transcriptase, RSV H⁺ reverse transcriptase, AMV H⁺ reverse transcriptase, RAV H⁺ reverse transcriptase, MAV H⁺ reverse transcriptase, HIV H⁺ reverse transcriptase, THERMOSCRIPT™ reverse transcriptase and THERMOSCRIPT™ II reverse transcriptase, and SUPERScript™ I reverse transcriptase and SUPERScript™ II reverse transcriptase, which are obtainable, for example, from Life Technologies, Inc. (Rockville, Maryland). See generally published PCT application WO 98/47912.

Other polypeptides having nucleic acid polymerase activity suitable for use in the present methods include thermophilic DNA polymerases such as DNA polymerase I, DNA polymerase III, Klenow fragment, T7 polymerase, and T5 polymerase, and thermostable DNA polymerases including, but not limited to, *Thermus thermophilus* (*Tth*) DNA polymerase, *Thermus aquaticus* (*Taq*) DNA polymerase, *Thermotoga neopolitana* (*Tne*) DNA polymerase, *Thermotoga maritima* (*Tma*) DNA polymerase, *Thermococcus litoralis* (*Tli* or VENT®) DNA polymerase, *Pyrococcus furiosus* (*Pfu*) DNA polymerase, *Pyrococcus* species GB-D (or DEEPVENT®) DNA polymerase, *Pyrococcus woosii* (*Pwo*) DNA polymerase, *Bacillus stearothermophilus* (*Bst*) DNA polymerase, *Sulfolobus acidocaldarius* (*Sac*) DNA polymerase, *Thermoplasma acidophilum* (*Tac*) DNA polymerase, *Thermus flavus* (*Tfl/Tub*) DNA polymerase, *Thermus ruber* (*Tru*) DNA polymerase, *Thermus brockianus* (DYNAZYME®) DNA polymerase, *Methanobacterium thermoautotrophicum* (*Mth*) DNA polymerase, and mutants,

variants and derivatives thereof. Such polypeptides are available commercially, for example from Life Technologies, Inc. (Rockville, MD), New England BioLabs (Beverly, MA), and Sigma/Aldrich (St. Louis, MO).

Host Cells

The invention also relates to host cells comprising one or more of the nucleic acid molecules or vectors of the invention, particularly those nucleic acid molecules and vectors described in detail herein. Representative host cells that may be used according to this aspect of the invention include, but are not limited to, bacterial cells, yeast cells, plant cells and animal cells. Preferred bacterial host cells include *Escherichia* spp. cells (particularly *E. coli* cells and most particularly *E. coli* strains DH10B, Stbl2, DH5 α , DB3, DB3.1 (preferably *E. coli* LIBRARY EFFICIENCY \otimes DB3.1TM Competent Cells; Life Technologies, Inc., Rockville, MD), DB4 and DB5; see U.S. Provisional Application No. 60/122,392, filed on March 2, 1999, the disclosure of which is incorporated by reference herein in its entirety), *Bacillus* spp. cells (particularly *B. subtilis* and *B. megaterium* cells), *Streptomyces* spp. cells, *Erwinia* spp. cells, *Klebsiella* spp. cells, *Serratia* spp. cells (particularly *S. marcessans* cells), *Pseudomonas* spp. cells (particularly *P. aeruginosa* cells), and *Salmonella* spp. cells (particularly *S. typhimurium* and *S. typhi* cells). Preferred animal host cells include insect cells (most particularly *Drosophila melanogaster* cells, *Spodoptera frugiperda* Sf9 and Sf21 cells and *Trichoplusia* High-Five cells), nematode cells (particularly *C. elegans* cells), avian cells, amphibian cells (particularly *Xenopus laevis* cells), reptilian cells, and mammalian cells (most particularly CHO, COS, VERO, BHK and human cells). Preferred yeast host cells include *Saccharomyces cerevisiae* cells and *Pichia pastoris* cells. These and other suitable host cells are available commercially, for example from Life Technologies, Inc. (Rockville, Maryland), American Type Culture Collection (Manassas, Virginia), and Agricultural Research Culture Collection (NRRL; Peoria, Illinois).

Methods for introducing the nucleic acid molecules and/or vectors of the invention into the host cells described herein, to produce host cells comprising one or more of the nucleic acid molecules and/or vectors of the invention, will be

5 familiar to those of ordinary skill in the art. For instance, the nucleic acid
molecules and/or vectors of the invention may be introduced into host cells using
well known techniques of infection, transduction, transfection, and transformation.
10 The nucleic acid molecules and/or vectors of the invention may be introduced
5 alone or in conjunction with other the nucleic acid molecules and/or vectors.
Alternatively, the nucleic acid molecules and/or vectors of the invention may be
15 introduced into host cells as a precipitate, such as a calcium phosphate precipitate,
or in a complex with a lipid. Electroporation also may be used to introduce the
nucleic acid molecules and/or vectors of the invention into a host. Likewise, such
20 molecules may be introduced into chemically competent cells such as *E. coli*. If
the vector is a virus, it may be packaged *in vitro* or introduced into a packaging
cell and the packaged virus may be transduced into cells. Hence, a wide variety
of techniques suitable for introducing the nucleic acid molecules and/or vectors of
25 the invention into cells in accordance with this aspect of the invention are well
known and routine to those of skill in the art. Such techniques are reviewed at
length, for example, in Sambrook, J., *et al.*, *Molecular Cloning, a Laboratory*
30 *Manual*, 2nd Ed., Cold Spring Harbor, NY: Cold Spring Harbor Laboratory
Press, pp. 16.30-16.55 (1989), Watson, J.D., *et al.*, *Recombinant DNA*, 2nd Ed.,
New York: W.H. Freeman and Co., pp. 213-234 (1992), and Winnacker, E.-L.,
20 *From Genes to Clones*, New York: VCH Publishers (1987), which are illustrative
of the many laboratory manuals that detail these techniques and which are
35 incorporated by reference herein in their entireties for their relevant disclosures.

40 *Polypeptides*

25 In another aspect, the invention relates to polypeptides encoded by the nucleic
acid molecules of the invention (including polypeptides and amino acid sequences
encoded by all possible reading frames of the nucleic acid molecules of the
45 invention), and to methods of producing such polypeptides. Polypeptides of the
present invention include purified or isolated natural products, products of
chemical synthetic procedures, and products produced by recombinant techniques
30 from a prokaryotic or eukaryotic host, including, for example, bacterial, yeast,
50 insect, mammalian, avian and higher plant cells.

5 The polypeptides of the invention may be produced by synthetic organic chemistry, and are preferably produced by standard recombinant methods, employing one or more of the host cells of the invention comprising the vectors or isolated nucleic acid molecules of the invention. According to the invention, 10 polypeptides are produced by cultivating the host cells of the invention (which comprise one or more of the nucleic acid molecules of the invention, preferably contained within an Expression Vector) under conditions favoring the expression of the nucleotide sequence contained on the nucleic acid molecule of the invention, such that the polypeptide encoded by the nucleic acid molecule of the invention is produced by the host cell. As used herein, "conditions favoring the expression 15 of the nucleotide sequence" or "conditions favoring the production of a polypeptide" include optimal physical (e.g., temperature, humidity, etc.) and nutritional (e.g., culture medium, ionic) conditions required for production of a recombinant polypeptide by a given host cell. Such optimal conditions for a variety of host cells, including prokaryotic (bacterial), mammalian, insect, yeast, and plant cells will be familiar to one of ordinary skill in the art, and may be found, for example, in Sambrook, J., *et al.*, *Molecular Cloning, A Laboratory Manual, 2nd Ed.*, Cold Spring Harbor, NY: Cold Spring Harbor Laboratory Press, (1989), Watson, J.D., *et al.*, *Recombinant DNA, 2nd Ed.*, New York: W.H. Freeman and Co., and Winnacker, E.-L., *From Genes to Clones*, New York: VCH Publishers 20 (1987).

35 In some aspects, it may be desirable to isolate or purify the polypeptides of the invention (e.g., for production of antibodies as described below), resulting in the production of the polypeptides of the invention in isolated form. The polypeptides 40 of the invention can be recovered and purified from recombinant cell cultures by well-known methods of protein purification that are routine in the art, including ammonium sulfate or ethanol precipitation, acid extraction, anion or cation exchange chromatography, phosphocellulose chromatography, hydrophobic interaction chromatography, affinity chromatography, hydroxylapatite 45 chromatography and lectin chromatography. For example, His6 or GST fusion tags on polypeptides made by the methods of the invention may be isolated using appropriate affinity chromatography matrices which bind polypeptides bearing 50

His6 or GST tags, as will be familiar to one of ordinary skill in the art. Polypeptides of the present invention include naturally purified products, products of chemical synthetic procedures, and products produced by recombinant techniques from a prokaryotic or eukaryotic host, including, for example, bacterial, yeast, higher plant, insect and mammalian cells. Depending upon the host employed in a recombinant production procedure, the polypeptides of the present invention may be glycosylated or may be non-glycosylated. In addition, polypeptides of the invention may also include an initial modified methionine residue, in some cases as a result of host-mediated processes.

Isolated polypeptides of the invention include those comprising the amino acid sequences encoded by one or more of the reading frames of the polynucleotides comprising one or more of the recombination site-encoding nucleic acid molecules of the invention, including those encoding *attB1*, *attB2*, *attP1*, *attP2*, *attL1*, *attL2*, *attR1* and *attR2* having the nucleotide sequences set forth in Figure 9 (or nucleotide sequences complementary thereto), or fragments, variants, mutants and derivatives thereof; the complete amino acid sequences encoded by the polynucleotides contained in the deposited clones described herein; the amino acid sequences encoded by polynucleotides which hybridize under stringent hybridization conditions to polynucleotides having the nucleotide sequences encoding the recombination site sequences of the invention as set forth in Figure 9 (or a nucleotide sequence complementary thereto); or a peptide or polypeptide comprising a portion or a fragment of the above polypeptides. The invention also relates to additional polypeptides having one or more additional amino acids linked (typically by peptidyl bonds to form a nascent polypeptide) to the polypeptides encoded by the recombination site nucleotide sequences or the deposited clones. Such additional amino acid residues may comprise one or more functional peptide sequences, for example one or more fusion partner peptides (e.g., GST, His₆, Trx, etc.) and the like.

As used herein, the terms "protein," "peptide," "oligopeptide" and "polypeptide" are considered synonymous (as is commonly recognized) and each term can be used interchangeably as the context requires to indicate a chain of two or more amino acids, preferably five or more amino acids, or more preferably ten

5 or more amino acids, coupled by (a) peptidyl linkage(s), unless otherwise defined
in the specific contexts below. As is commonly recognized in the art, all
10 polypeptide formulas or sequences herein are written from left to right and in the
direction from amino terminus to carboxy terminus.

5 It will be recognized by those of ordinary skill in the art that some amino acid
sequences of the polypeptides of the invention can be varied without significant
15 effect on the structure or function of the polypeptides. If such differences in
sequence are contemplated, it should be remembered that there will be critical
areas on the protein which determine structure and activity. In general, it is
20 possible to replace residues which form the tertiary structure, provided that
residues performing a similar function are used. In other instances, the type of
residue may be completely unimportant if the alteration occurs at a non-critical
region of the polypeptide.

25 Thus, the invention further includes variants of the polypeptides of the
invention, including allelic variants, which show substantial structural homology
15 to the polypeptides described herein, or which include specific regions of these
polypeptides such as the portions discussed below. Such mutants may include
deletions, insertions, inversions, repeats, and type substitutions (for example,
30 substituting one hydrophilic residue for another, but not strongly hydrophilic for
strongly hydrophobic as a rule). Small changes or such "neutral" or "conservative"
20 amino acid substitutions will generally have little effect on activity.

35 Typical conservative substitutions are the replacements, one for another,
among the aliphatic amino acids Ala, Val, Leu and Ile; interchange of the
hydroxylated residues Ser and Thr; exchange of the acidic residues Asp and Glu;
40 substitution between the amidated residues Asn and Gln; exchange of the basic
25 residues Lys and Arg; and replacements among the aromatic residues Phe and Tyr.

45 Thus, the fragment, derivative or analog of the polypeptides of the invention,
such as those comprising peptides encoded by the recombination site nucleotide
sequences described herein, may be (i) one in which one or more of the amino acid
30 residues are substituted with a conservative or non-conservative amino acid
residue (preferably a conservative amino acid residue), and such substituted amino
50 acid residue may be encoded by the genetic code or may be an amino acid (*e.g.*,

desmosine, citrulline, ornithine, etc.) that is not encoded by the genetic code; (ii) one in which one or more of the amino acid residues includes a substituent group (e.g., a phosphate, hydroxyl, sulfate or other group) in addition to the normal "R" group of the amino acid; (iii) one in which the mature polypeptide is fused with another compound, such as a compound to increase the half-life of the polypeptide (for example, polyethylene glycol), or (iv) one in which additional amino acids are fused to the mature polypeptide, such as an immunoglobulin Fc region peptide, a leader or secretory sequence, a sequence which is employed for purification of the mature polypeptide (such as GST) or a proprotein sequence. Such fragments, derivatives and analogs are intended to be encompassed by the present invention, and are within the scope of those skilled in the art from the teachings herein and the state of the art at the time of invention.

The polypeptides of the present invention are preferably provided in an isolated form, and preferably are substantially purified. Recombinantly produced versions of the polypeptides of the invention can be substantially purified by the one-step method described in Smith and Johnson, *Gene* 67:31-40 (1988). As used herein, the term "substantially purified" means a preparation of an individual polypeptide of the invention wherein at least 50%, preferably at least 60%, 70%, or 75% and more preferably at least 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98% or 99% (by mass) of contaminating proteins (*i.e.*, those that are not the individual polypeptides described herein or fragments, variants, mutants or derivatives thereof) have been removed from the preparation.

The polypeptides of the present invention include those which are at least about 50% identical, at least 60% identical, at least 65% identical, more preferably at least about 70%, at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 96%, at least about 97%, at least about 98% or at least about 99% identical, to the polypeptides described herein. For example, preferred *attB1*-containing polypeptides of the invention include those that are at least about 50% identical, at least 60% identical, at least 65% identical, more preferably at least about 70%, at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 96%, at least about 97%, at least about 98% or at least about 99% identical,

5 to the polypeptide(s) encoded by the three reading frames of a polynucleotide
comprising a nucleotide sequence of *attB1* having a nucleic acid sequence as set
10 forth in Figure 9 (or a nucleic acid sequence complementary thereto), to a
polypeptide encoded by a polynucleotide contained in the deposited cDNA clones
5 described herein, or to a polypeptide encoded by a polynucleotide hybridizing
under stringent conditions to a polynucleotide comprising a nucleotide sequence
15 of *attB1* having a nucleic acid sequence as set forth in Figure 9 (or a nucleic acid
sequence complementary thereto). Analogous polypeptides may be prepared that
are at least about 65% identical, more preferably at least about 70%, at least about
20 75%, at least about 80%, at least about 85%, at least about 90%, at least about
95%, at least about 96%, at least about 97%, at least about 98% or at least about
99% identical, to the *attB2*, *attP1*, *attP2*, *attL1*, *attL2*, *attR1* and *attR2*
polypeptides of the invention as depicted in Figure 9. The present polypeptides
25 also include portions or fragments of the above-described polypeptides with at
least 5, 10, 15, 20, or 25 amino acids.

By a polypeptide having an amino acid sequence at least, for example, 65%
30 "identical" to a reference amino acid sequence of a given polypeptide of the
invention is intended that the amino acid sequence of the polypeptide is identical
to the reference sequence except that the polypeptide sequence may include up to
20 35 amino acid alterations per each 100 amino acids of the reference amino acid
sequence of a given polypeptide of the invention. In other words, to obtain a
35 polypeptide having an amino acid sequence at least 65% identical to a reference
amino acid sequence, up to 35% of the amino acid residues in the reference
sequence may be deleted or substituted with another amino acid, or a number of
40 amino acids up to 35% of the total amino acid residues in the reference sequence
25 may be inserted into the reference sequence. These alterations of the reference
sequence may occur at the amino (N-) or carboxy (C-) terminal positions of the
reference amino acid sequence or anywhere between those terminal positions,
45 interspersed either individually among residues in the reference sequence or in one
or more contiguous groups within the reference sequence. As a practical matter,
30 whether a given amino acid sequence is, for example, at least 65% identical to the
amino acid sequence of a given polypeptide of the invention can be determined
50

5 conventionally using known computer programs such as those described above for
nucleic acid sequence identity determinations, or more preferably using the
10 CLUSTAL W program (Thompson, J.D., *et al.*, *Nucleic Acids Res.* 22:4673-4680
(1994)).

5 The polypeptides of the present invention can be used as molecular weight
markers on SDS-PAGE gels or on molecular sieve gel filtration columns using
15 methods well known to those of skill in the art. In addition, as described in detail
below, the polypeptides of the present invention can be used to raise polyclonal
and monoclonal antibodies which are useful in a variety of assays for detecting
20 protein expression, localization, detection of interactions with other molecules, or
for the isolation of a polypeptide (including a fusion polypeptide) of the invention.

In another aspect, the present invention provides a peptide or polypeptide
25 comprising an epitope-bearing portion of a polypeptide of the invention, which
may be used to raise antibodies, particularly monoclonal antibodies, that bind
15 specifically to a one or more of the polypeptides of the invention. The epitope of
this polypeptide portion is an immunogenic or antigenic epitope of a polypeptide
of the invention. An "immunogenic epitope" is defined as a part of a protein that
30 elicits an antibody response when the whole protein is the immunogen. These
immunogenic epitopes are believed to be confined to a few loci on the molecule.
On the other hand, a region of a protein molecule to which an antibody can bind
20 is defined as an "antigenic epitope." The number of immunogenic epitopes of a
protein generally is less than the number of antigenic epitopes (*see, e.g.*, Geysen
35 *et al.*, *Proc. Natl. Acad. Sci. USA* 81:3998-4002 (1983)).

40 As to the selection of peptides or polypeptides bearing an antigenic epitope
25 (*i.e.*, that contain a region of a protein molecule to which an antibody can bind),
it is well-known in the art that relatively short synthetic peptides that mimic part
of a protein sequence are routinely capable of eliciting an antiserum that reacts
45 with the partially mimicked protein (*see, e.g.*, Sutcliffe, J.G., *et al.*, *Science*
219:660-666 (1983)). Peptides capable of eliciting protein-reactive sera are
30 frequently represented in the primary sequence of a protein, can be characterized
by a set of simple chemical rules, and are not confined to the immunodominant
50 regions of intact proteins (*i.e.*, immunogenic epitopes) or to the amino or carboxy

termini. Peptides that are extremely hydrophobic and those of six or fewer residues generally are ineffective at inducing antibodies that bind to the mimicked protein; longer peptides, especially those containing proline residues, usually are effective (Sutcliffe, J.G., *et al.*, *Science* 219:660-666 (1983)).

Epitope-bearing peptides and polypeptides of the invention designed according to the above guidelines preferably contain a sequence of at least five, more preferably at least seven or more amino acids contained within the amino acid sequence of a polypeptide of the invention. However, peptides or polypeptides comprising a larger portion of an amino acid sequence of a polypeptide of the invention, containing about 30 to about 50 amino acids, or any length up to and including the entire amino acid sequence of a given polypeptide of the invention, also are considered epitope-bearing peptides or polypeptides of the invention and also are useful for inducing antibodies that react with the mimicked protein. Preferably, the amino acid sequence of the epitope-bearing peptide is selected to provide substantial solubility in aqueous solvents (*i.e.*, the sequence includes relatively hydrophilic residues and highly hydrophobic sequences are preferably avoided); sequences containing proline residues are particularly preferred.

Non-limiting examples of epitope-bearing polypeptides or peptides that can be used to generate antibodies specific for the polypeptides of the invention include certain epitope-bearing regions of the polypeptides comprising amino acid sequences encoded by polynucleotides comprising one or more of the recombination site-encoding nucleic acid molecules of the invention, including those encoding *attB1*, *attB2*, *attP1*, *attP2*, *attL1*, *attL2*, *attR1* and *attR2* having the nucleotide sequences set forth in Figure 9 (or a nucleotide sequence complementary thereto); the complete amino acid sequences encoded by the three reading frames of the polynucleotides contained in the deposited clones described herein; and the amino acid sequences encoded by all reading frames of polynucleotides which hybridize under stringent hybridization conditions to polynucleotides having the nucleotide sequences encoding the recombination site sequences (or portions thereof) of the invention as set forth in Figure 9 (or a nucleic acid sequence complementary thereto). Other epitope-bearing polypeptides or peptides that may be used to generate antibodies specific for the polypeptides

of the invention will be apparent to one of ordinary skill in the art based on the primary amino acid sequences of the polypeptides of the invention described herein, via the construction of Kyte-Doolittle hydrophilicity and Jameson-Wolf antigenic index plots of the polypeptides of the invention using, for example, PROTEAN computer software (DNASTAR, Inc.; Madison, Wisconsin).

The epitope-bearing peptides and polypeptides of the invention may be produced by any conventional means for making peptides or polypeptides including recombinant means using nucleic acid molecules of the invention. For instance, a short epitope-bearing amino acid sequence may be fused to a larger polypeptide which acts as a carrier during recombinant production and purification, as well as during immunization to produce anti-peptide antibodies. Epitope-bearing peptides also may be synthesized using known methods of chemical synthesis (*see, e.g.*, U.S. Patent No. 4,631,211 and Houghten, R. A., *Proc. Natl. Acad. Sci. USA* 82:5131-5135 (1985), both of which are incorporated by reference herein in their entireties).

As one of skill in the art will appreciate, the polypeptides of the present invention and epitope-bearing fragments thereof may be immobilized onto a solid support, by techniques that are well-known and routine in the art. By "solid support" is intended any solid support to which a peptide can be immobilized. Such solid supports include, but are not limited to nitrocellulose, diazocellulose, glass, polystyrene, polyvinylchloride, polypropylene, polyethylene, dextran, Sepharose, agar, starch, nylon, beads and microtitre plates. Linkage of the peptide of the invention to a solid support can be accomplished by attaching one or both ends of the peptide to the support. Attachment may also be made at one or more internal sites in the peptide. Multiple attachments (both internal and at the ends of the peptide) may also be used according to the invention. Attachment can be via an amino acid linkage group such as a primary amino group, a carboxyl group, or a sulfhydryl (SH) group or by chemical linkage groups such as with cyanogen bromide (CNBr) linkage through a spacer. For non-covalent attachments to the support, addition of an affinity tag sequence to the peptide can be used such as GST (Smith, D.B., and Johnson, K.S., *Gene* 67:31 (1988)), polyhistidines (Hochuli, E., *et al.*, *J. Chromatog.* 411:77 (1987)), or biotin. Such affinity tags

5 may be used for the reversible attachment of the peptide to the support. Such
immobilized polypeptides or fragments may be useful, for example, in isolating
10 antibodies directed against one or more of the polypeptides of the invention, or
other proteins or peptides that recognize other proteins or peptides that bind to
5 one or more of the polypeptides of the invention, as described below.

As one of skill in the art will also appreciate, the polypeptides of the present
15 invention and the epitope-bearing fragments thereof described herein can be
combined with one or more fusion partner proteins or peptides, or portions
thereof, including but not limited to GST, His₆, Trx, and portions of the constant
20 domain of immunoglobulins (Ig), resulting in chimeric or fusion polypeptides.
These fusion polypeptides facilitate purification of the polypeptides of the
invention (EP 0 394 827; Traunecker *et al.*, *Nature* 331:84-86 (1988)) for use in
analytical or diagnostic (including high-throughput) format.

25 *Antibodies*

In another aspect, the invention relates to antibodies that recognize and bind
30 to the polypeptides (or epitope-bearing fragments thereof) or nucleic acid
molecules (or portions thereof) of the invention. In a related aspect, the invention
relates to antibodies that recognize and bind to one or more polypeptides encoded
20 by all reading frames of one or more recombination site nucleic acid sequences or
portions thereof, or to one or more nucleic acid molecules comprising one or more
35 recombination site nucleic acid sequences or portions thereof, including but not
limited to *att* sites (including *attB*1, *attB*2, *attP*1, *attP*2, *attL*1, *attL*2, *attR*1, *attR*2
and the like), *lox* sites (*e.g.*, *loxP*, *loxP*511, and the like), FRT, and the like, or
40 mutants, fragments, variants and derivatives thereof. See generally U.S. Patent
No. 5,888,732, which is incorporated herein by reference in its entirety. The
25 antibodies of the present invention may be polyclonal or monoclonal, and may be
prepared by any of a variety of methods and in a variety of species according to
45 methods that are well-known in the art. See, for instance, U.S. Patent No.
5,587,287; Sutcliffe, J.G., *et al.*, *Science* 219:660-666 (1983); Wilson *et al.*, *Cell*
30 37: 767 (1984); and Bittle, F.J., *et al.*, *J. Gen. Virol.* 66:2347-2354 (1985).
Antibodies specific for any of the polypeptides or nucleic acid molecules described
50

5 herein, such as antibodies specifically binding to one or more of the polypeptides
10 encoded by the recombination site nucleotide sequences, or one or more nucleic
acid molecules, described herein or contained in the deposited clones, antibodies
against fusion polypeptides (*e.g.*, binding to fusion polypeptides between one or
5 more of the fusion partner proteins and one or more of the recombination site
polypeptides of the invention, as described herein), and the like, can be raised
15 against the intact polypeptides or polynucleotides of the invention or one or more
antigenic polypeptide fragments thereof.

As used herein, the term "antibody" (Ab) may be used interchangeably with
10 the terms "polyclonal antibody" or "monoclonal antibody" (mAb), except in
specific contexts as described below. These terms, as used herein, are meant to
include intact molecules as well as antibody fragments (such as, for example, Fab
20 and F(ab')₂ fragments) which are capable of specifically binding to a polypeptide
or nucleic acid molecule of the invention or a portion thereof. It will therefore be
25 appreciated that, in addition to the intact antibodies of the invention, Fab, F(ab')₂
and other fragments of the antibodies described herein, and other peptides and
peptide fragments that bind one or more polypeptides or polynucleotides of the
30 invention, are also encompassed within the scope of the invention. Such antibody
fragments are typically produced by proteolytic cleavage of intact antibodies, using
20 enzymes such as papain (to produce Fab fragments) or pepsin (to produce F(ab')₂
35 fragments). Antibody fragments, and peptides or peptide fragments, may also be
produced through the application of recombinant DNA technology or through
synthetic chemistry.

40 Epitope-bearing peptides and polypeptides, and nucleic acid molecules or
25 portions thereof, of the invention may be used to induce antibodies according to
methods well known in the art, as generally described herein (*see, e.g.*, Sutcliffe,
et al., supra; Wilson, *et al., supra*; and Bittle, F. J., *et al., J. Gen. Virol.*
45 66:2347-2354 (1985)).

Polyclonal antibodies according to this aspect of the invention may be made
30 by immunizing an animal with one or more of the polypeptides or nucleic acid
molecules of the invention described herein or portions thereof according to
50 standard techniques (*see, e.g.*, Harlow, E., and Lane, D., *Antibodies: A*

Laboratory Manual, Cold Spring Harbor, NY: Cold Spring Harbor Laboratory Press (1988); Kaufman, P.B., *et al.*, In: *Handbook of Molecular and Cellular Methods in Biology and Medicine*, Boca Raton, Florida: CRC Press, pp. 468-469 (1995)). For producing antibodies that recognize and bind to the polypeptides or nucleic acid molecules of the invention or portions thereof, animals may be immunized with free peptide or free nucleic acid molecules; however, antibody titer may be boosted by coupling of the peptide to a macromolecular carrier, such as albumin, KLH, or tetanus toxoid (particularly for producing antibodies against the nucleic acid molecules of the invention or portions thereof; *see* Harlow and Lane, *supra*, at page 154), or to a solid phase carrier such as a latex or glass microbead. For instance, peptides containing cysteine may be coupled to carrier using a linker such as m-maleimidobenzoyl-N- hydroxysuccinimide ester (MBS), while other peptides may be coupled to carrier using a more general linking agent such as glutaraldehyde. Animals such as rabbits, rats and mice may be immunized with either free (if the polypeptide immunogen is larger than about 25 amino acids in length) or carrier-coupled peptides or nucleic acid molecules, for instance, by intraperitoneal and/or intradermal injection of emulsions containing about 100 µg peptide, polynucleotide, or carrier protein, and Freund's adjuvant. Several booster injections may be needed, for instance, at intervals of about two weeks, to provide a useful titer of antibody which can be detected, for example, by ELISA assay using free peptide or nucleic acid molecule adsorbed to a solid surface. In another approach, cells expressing one or more of the polypeptides or polynucleotides of the invention or an antigenic fragment thereof can be administered to an animal in order to induce the production of sera containing polyclonal antibodies, according to routine immunological methods. In yet another method, a preparation of one or more of the polypeptides or polynucleotides of the invention is prepared and purified as described herein, to render it substantially free of natural contaminants. Such a preparation may then be introduced into an animal in order to produce polyclonal antisera of greater specific activity. The titer of antibodies in serum from an immunized animal, regardless of the method of immunization used, may be increased by selection of anti-peptide or anti-polynucleotide antibodies, for

instance, by adsorption to the peptide or polynucleotide on a solid support and elution of the selected antibodies according to methods well known in the art.

In an alternative method, the antibodies of the present invention are monoclonal antibodies (or fragments thereof which bind to one or more of the polypeptides of the invention). Such monoclonal antibodies can be prepared using hybridoma technology (Köhler *et al.*, *Nature* 256:495 (1975); Köhler *et al.*, *Eur. J. Immunol.* 6:511 (1976); Köhler *et al.*, *Eur. J. Immunol.* 6:292 (1976); Hammerling *et al.*, In: *Monoclonal Antibodies and T-Cell Hybridomas*, Elsevier, N.Y., pp. 563-681 (1981)). In general, such procedures involve immunizing an animal (preferably a mouse) with a polypeptide or polynucleotide of the invention (or a fragment thereof), or with a cell expressing a polypeptide or polynucleotide of the invention (or a fragment thereof). The splenocytes of such mice are extracted and fused with a suitable myeloma cell line. Any suitable myeloma cell line may be employed in accordance with the present invention; however, it is preferable to employ the parent myeloma cell line (SP₂O), available from the American Type Culture Collection, Rockville, Maryland. After fusion, the resulting hybridoma cells are selectively maintained in HAT medium, and then cloned by limiting dilution as described by Wands *et al.* (*Gastroenterol.* 80:225-232 (1981)). The hybridoma cells obtained through such a selection are then assayed to identify clones which secrete antibodies capable of binding one or more of the polypeptides or nucleic acid molecules of the invention, or fragments thereof. Hence, the present invention also provides hybridoma cells and cell lines producing monoclonal antibodies of the invention, particularly that recognize and bind to one or more of the polypeptides or nucleic acid molecules of the invention.

Alternatively, additional antibodies capable of binding to one or more of the polypeptides of the invention, or fragments thereof, may be produced in a two-step procedure through the use of anti-idiotypic antibodies. Such a method makes use of the fact that antibodies are themselves antigens, and that, therefore, it is possible to obtain an antibody which binds to a second antibody. In accordance with this method, antibodies specific for one or more of the polypeptides or polynucleotides of the invention, prepared as described above, are used to immunize an animal, preferably a mouse. The splenocytes of such an

animal are then used to produce hybridoma cells, and the hybridoma cells are screened to identify clones which produce an antibody whose ability to bind to an antibody specific for one or more of the polypeptides or polynucleotides of the invention can be blocked by polypeptides of the invention themselves. Such antibodies comprise anti-idiotypic antibodies to the antibodies recognizing one or more of the polypeptides or polynucleotides of the invention, and can be used to immunize an animal to induce formation of further antibodies specific for one or more of the polypeptides or polynucleotides of the invention.

For use, the antibodies of the invention may optionally be detectably labeled by covalent or non-covalent attachment of one or more labels, including but not limited to chromogenic, enzymatic, radioisotopic, isotopic, fluorescent, toxic, chemiluminescent, or nuclear magnetic resonance contrast agents or other labels.

Examples of suitable enzyme labels include malate dehydrogenase, staphylococcal nuclease, delta-5-steroid isomerase, yeast-alcohol dehydrogenase, alpha-glycerol phosphate dehydrogenase, triose phosphate isomerase, peroxidase, alkaline phosphatase, asparaginase, glucose oxidase, beta-galactosidase, ribonuclease, urease, catalase, glucose-6-phosphate dehydrogenase, glucoamylase, and acetylcholine esterase.

Examples of suitable radioisotopic labels include ^3H , ^{111}In , ^{125}I , ^{131}I , ^{32}P , ^{35}S , ^{14}C , ^{51}Cr , ^{57}Co , ^{58}Co , ^{59}Fe , ^{75}Se , ^{152}Eu , ^{90}Y , ^{67}Cu , ^{217}Bi , ^{211}At , ^{212}Pb , ^{47}Sc , ^{109}Pd , etc. ^{111}In is a preferred isotope where in vivo imaging is used since it avoids the problem of dehalogenation of the ^{125}I or ^{131}I -labeled monoclonal antibody by the liver. In addition, this radionuclide has a more favorable gamma emission energy for imaging (Perkins *et al.*, *Eur. J. Nucl. Med.* 10:296-301 (1985); Carasquillo *et al.*, *J. Nucl. Med.* 28:281-287 (1987)). For example, ^{111}In coupled to monoclonal antibodies with 1-(P-isothiocyanatobenzyl)-DPTA has shown little uptake in non-tumorous tissues, particularly the liver, and therefore enhances specificity of tumor localization (Esteban *et al.*, *J. Nucl. Med.* 28:861-870 (1987)).

Examples of suitable non-radioactive isotopic labels include ^{157}Gd , ^{55}Mn , ^{162}Dy , ^{32}Tr , and ^{56}Fe .

Examples of suitable fluorescent labels include an ^{152}Eu label, a fluorescein label, an isothiocyanate label, a rhodamine label, a phycoerythrin label, a

phycoerythrin label, an allophycocyanin label, an o-phthaldehyde label, a green fluorescent protein (GFP) label, and a fluorescamine label.

Examples of suitable toxin labels include diphtheria toxin, ricin, and cholera toxin.

Examples of chemiluminescent labels include a luminal label, an isoluminal label, an aromatic acridinium ester label, an imidazole label, an acridinium salt label, an oxalate ester label, a luciferin label, a luciferase label, and an aequorin label.

Examples of nuclear magnetic resonance contrasting agents include heavy metal nuclei such as Gd, Mn, and iron.

Typical techniques for binding the above-described labels to the antibodies of the invention are provided by Kennedy *et al.*, *Clin. Chim. Acta* 70:1-31 (1976), and Schurs *et al.*, *Clin. Chim. Acta* 81:1-40 (1977). Coupling techniques mentioned in the latter are the glutaraldehyde method, the periodate method, the dimaleimide method, the m-maleimidobenzyl-N-hydroxy-succinimide ester method, all of which methods are incorporated by reference herein.

It will be appreciated by one of ordinary skill that the antibodies of the present invention may alternatively be coupled to a solid support, to facilitate, for example, chromatographic and other immunological procedures using such solid phase-immobilized antibodies. Included among such procedures are the use of the antibodies of the invention to isolate or purify polypeptides comprising one or more epitopes encoded by the nucleic acid molecules of the invention (which may be fusion polypeptides or other polypeptides of the invention described herein), or to isolate or purify polynucleotides comprising one or more recombination site sequences of the invention or portions thereof. Methods for isolation and purification of polypeptides (and, by analogy, polynucleotides) by affinity chromatography, for example using the antibodies of the invention coupled to a solid phase support, are well-known in the art and will be familiar to one of ordinary skill. The antibodies of the invention may also be used in other applications, for example to cross-link or couple two or more proteins, polypeptides, polynucleotides, or portions thereof into a structural and/or functional complex. In one such use, an antibody of the invention may have two

5 or more distinct epitope-binding regions that may bind, for example, a first polypeptide (which may be a polypeptide of the invention) at one epitope-binding region on the antibody and a second polypeptide (which may be a polypeptide of the invention) at a second epitope-binding region on the antibody, thereby bringing the first and second polypeptides into close proximity to each other such that the first and second polypeptides are able to interact structurally and/or functionally (as, for example, linking an enzyme and its substrate to carry out enzymatic catalysis, or linking an effector molecule and its receptor to carry out or induce a specific binding of the effector molecule to the receptor or a response to the effector molecule mediated by the receptor). Additional applications for the antibodies of the invention include, for example, the preparation of large-scale arrays of the antibodies, polypeptides, or nucleic acid molecules of the invention, or portions thereof, on a solid support, for example to facilitate high-throughput screening of protein or RNA expression by host cells containing nucleic acid molecules of the invention (known in the art as "chip array" protocols; *see, e.g.*, U.S. Patent Nos. 5,856,101, 5,837,832, 5,770,456, 5,744,305, 5,631,734, and 5,593,839, which are directed to production and use of chip arrays of polypeptides (including antibodies) and polynucleotides, and the disclosures of which are incorporated herein by reference in their entireties). By "solid support" is intended any solid support to which an antibody can be immobilized. Such solid supports include, but are not limited to nitrocellulose, diazocellulose, glass, polystyrene, polyvinylchloride, polycarbonate, polypropylene, polyethylene, dextran, Sepharose, agar, starch, nylon, beads and microtitre plates. Preferred are beads made of glass, latex or a magnetic material. Linkage of an antibody of the invention to a solid support can be accomplished by attaching one or both ends of the antibody to the support. Attachment may also be made at one or more internal sites in the antibody. Multiple attachments (both internal and at the ends of the antibody) may also be used according to the invention. Attachment can be via an amino acid linkage group such as a primary amino group, a carboxyl group, or a sulfhydryl (SH) group or by chemical linkage groups such as with cyanogen bromide (CNBr) linkage through a spacer. For non-covalent attachments, addition of an affinity tag sequence to the peptide can be used such as GST

(Smith, D.B., and Johnson, K.S., *Gene* 67:31 (1988)), polyhistidines (Hochuli, E., *et al.*, *J. Chromatog.* 411:77 (1987)), or biotin. Alternatively, attachment can be accomplished using a ligand which binds the Fc region of the antibodies of the invention, *e.g.*, protein A or protein G. Such affinity tags may be used for the reversible attachment of the antibodies to the support. Peptides may also be recognized via specific ligand-receptor interactions or using phage display methodologies that will be familiar to the skilled artisan, for their ability to bind polypeptides of the invention or fragments thereof.

Kits

In another aspect, the invention provides kits which may be used in producing the nucleic acid molecules, polypeptides, vectors, host cells, and antibodies, and in the recombinational cloning methods, of the invention. Kits according to this aspect of the invention may comprise one or more containers, which may contain one or more of the nucleic acid molecules, primers, polypeptides, vectors, host cells, or antibodies of the invention. In particular, a kit of the invention may comprise one or more components (or combinations thereof) selected from the group consisting of one or more recombination proteins (*e.g.*, Int) or auxiliary factors (*e.g.* IHF and/or Xis) or combinations thereof, one or more compositions comprising one or more recombination proteins or auxiliary factors or combinations thereof (for example, GATEWAY™ LR Clonase™ Enzyme Mix or GATEWAY™ BP Clonase™ Enzyme Mix) one or more Destination Vector molecules (including those described herein), one or more Entry Clone or Entry Vector molecules (including those described herein), one or more primer nucleic acid molecules (particularly those described herein), one or more host cells (*e.g.* competent cells, such as *E. coli* cells, yeast cells, animal cells (including mammalian cells, insect cells, nematode cells, avian cells, fish cells, etc.), plant cells, and most particularly *E. coli* DB3, DB3.1 (preferably *E. coli* LIBRARY EFFICIENCY® DB3.1™ Competent Cells; Life Technologies, Inc., Rockville, MD), DB4 and DB5; *see* U.S. Provisional Application No. 60/122,392, filed on March 2, 1999, and the corresponding U.S. Utility Application No. _____ of Hartley *et al.*, entitled "Cells Resistant to Toxic Genes and Uses Thereof," filed

on even day herewith, the disclosures of which are incorporated by reference herein in its entirety), and the like. In related aspects, the kits of the invention may comprise one or more nucleic acid molecules encoding one or more recombination sites or portions thereof, such as one or more nucleic acid molecules comprising a nucleotide sequence encoding the one or more recombination sites (or portions thereof) of the invention, and particularly one or more of the nucleic acid molecules contained in the deposited clones described herein. Kits according to this aspect of the invention may also comprise one or more isolated nucleic acid molecules of the invention, one or more vectors of the invention, one or more primer nucleic acid molecules of the invention, and/or one or more antibodies of the invention. The kits of the invention may further comprise one or more additional containers containing one or more additional components useful in combination with the nucleic acid molecules, polypeptides, vectors, host cells, or antibodies of the invention, such as one or more buffers, one or more detergents, one or more polypeptides having nucleic acid polymerase activity, one or more polypeptides having reverse transcriptase activity, one or more transfection reagents, one or more nucleotides, and the like. Such kits may be used in any process advantageously using the nucleic acid molecules, primers, vectors, host cells, polypeptides, antibodies and other compositions of the invention, for example in methods of synthesizing nucleic acid molecules (*e.g.*, via amplification such as via PCR), in methods of cloning nucleic acid molecules (preferably via recombinational cloning as described herein), and the like.

Optimization of Recombinational Cloning System

The usefulness of a particular nucleic acid molecule, or vector comprising a nucleic acid molecule, of the invention in methods of recombinational cloning may be determined by any one of a number of assay methods. For example, Entry and Destination vectors of the present invention may be assessed for their ability to function (*i.e.*, to mediate the transfer of a nucleic acid molecule, DNA segment, gene, cDNA molecule or library from a cloning vector to an Expression Vector) by carrying out a recombinational cloning reaction as described in more detail in the Examples below and as described in U.S. Application Nos. 08/663,002, filed

5 June 7, 1996 (now U.S. Patent No. 5,888,732), 09/005,476, filed January 12,
1998, 09/177,387, filed October 23, 1998, and 60/108,324, filed November 13,
10 1998, the disclosures of which are incorporated by reference herein in their
entireties. Alternatively, the functionality of Entry and Destination Vectors
5 prepared according to the invention may be assessed by examining the ability of
these vectors to recombine and create cointegrate molecules, or to transfer a
15 nucleic acid molecule of interest, using an assay such as that described in detail
below in Example 19. Analogously, the formulation of compositions comprising
one or more recombination proteins or combinations thereof, for example
10 GATEWAY™ LR Clonase™ Enzyme Mix and GATEWAY™ BP Clonase™
20 Enzyme Mix, may be optimized using assays such as those described below in
Example 18.

25 *Uses*

15 There are a number of applications for the compositions, methods and kits of
the present invention. These uses include, but are not limited to, changing vectors,
targeting gene products to intracellular locations, cleaving fusion tags from desired
30 proteins, operably linking nucleic acid molecules of interest to regulatory genetic
sequences (e.g., promoters, enhancers, and the like), constructing genes for fusion
20 proteins, changing copy number, changing replicons, cloning into phages, and
cloning, e.g., PCR products, genomic DNAs, and cDNAs. In addition, the nucleic
35 acid molecules, vectors, and host cells of the invention may be used in the
production of polypeptides encoded by the nucleic acid molecules, in the
production of antibodies directed against such polypeptides, in recombinational
40 25 cloning of desired nucleic acid sequences, and in other applications that may be
enhanced or facilitated by the use of the nucleic acid molecules, vectors, and host
cells of the invention.

45 In particular, the nucleic acid molecules, vectors, host cells, polypeptides,
antibodies, and kits of the invention may be used in methods of transferring one
30 or more desired nucleic acid molecules or DNA segments, for example one or
more genes, cDNA molecules or cDNA libraries, into a cloning or Expression
50 Vector for use in transforming additional host cells for use in cloning or

5 amplification of, or expression of the polypeptide encoded by, the desired nucleic
acid molecule or DNA segment. Such recombinational cloning methods which
10 may advantageously use the nucleic acid molecules, vectors, and host cells of the
invention, are described in detail in the Examples below, and in commonly owned
5 U.S. Application Nos. 08/486,139, filed June 7, 1995, 08/663,002, filed June 7,
1996 (now U.S. Patent No. 5,888,732), 09/005,476, filed January 12, 1998,
15 09/177,387, filed October 23, 1998, and 60/108,324, filed November 13, 1998,
the disclosures of all of which are incorporated by reference herein in their
entireties.

10
20 It will be understood by one of ordinary skill in the relevant arts that other
suitable modifications and adaptations to the methods and applications described
herein are readily apparent from the description of the invention contained herein
25 in view of information known to the ordinarily skilled artisan, and may be made
15 without departing from the scope of the invention or any embodiment thereof.
Having now described the present invention in detail, the same will be more clearly
30 understood by reference to the following examples, which are included herewith
for purposes of illustration only and are not intended to be limiting of the
invention.

Examples

Example 1: Recombination Reactions of Bacteriophage λ

40 25 The *E. coli* bacteriophage λ can grow as a lytic phage, in which case the host
cell is lysed, with the release of progeny virus. Alternatively, lambda can integrate
into the genome of its host by a process called lysogenization (see Figure 60). In
45 this lysogenic state, the phage genome can be transmitted to daughter cells for
many generations, until conditions arise that trigger its excision from the genome.
30 At this point, the virus enters the lytic part of its life cycle. The control of the
switch between the lytic and lysogenic pathways is one of the best understood
50 processes in molecular biology (M. Ptashne, *A Genetic Switch*, Cell Press, 1992).

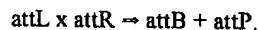
The integrative and excisive recombination reactions of λ , performed *in vitro*, are the basis of Recombinational Cloning System of the present invention. They can be represented schematically as follows:



The four att sites contain binding sites for the proteins that mediate the reactions. The wild type attP, attB, attL, and attR sites contain about 243, 25, 100, and 168 base pairs, respectively. The attB x attP reaction (hereinafter referred to as a "BP Reaction," or alternatively and equivalently as an "Entry Reaction" or a "Gateward Reaction") is mediated by the proteins Int and IHF. The attL x attR reaction (hereinafter referred to as an "LR Reaction," or alternatively and equivalently as a "Destination Reaction") is mediated by the proteins Int, IHF, and Xis. Int (integrase) and Xis (excisionase) are encoded by the λ genome, while IHF (integration host factor) is an *E. coli* protein. For a general review of lambda recombination, see: A. Landy, *Ann. Rev. Biochem.* 58: 913-949 (1989).

Example 2: Recombination Reactions of the Recombinational Cloning System

The LR Reaction -- the exchange of a DNA segment from an Entry Clone to a Destination Vector -- is the *in vitro* version of the λ excision reaction:



There is a practical imperative for this configuration: after an LR Reaction in one configuration of the present method, an att site usually separates a functional motif (such as a promoter or a fusion tag) from a nucleic acid molecule of interest in an Expression Clone, and the 25 bp attB site is much smaller than the attP, attL, and attR sites.

Note that the recombination reaction is conservative, i.e., there is no net synthesis or loss of base pairs. The DNA segments that flank the recombination

5 sites are merely switched. The wild type λ recombination sites are modified for purposes of the GATEWAY™ Cloning System, as follows:

10 To create certain preferred Destination Vectors, a part (43 bp) of attR was removed, to make the excisive reaction irreversible and more efficient (W. Bushman et al., *Science* 230: 906, 1985). The attR sites in preferred Destination Vectors of the invention are 125 bp in length. Mutations were made to the core regions of the att sites, for two reasons: (1) to eliminate stop codons, and (2) to ensure specificity of the recombination reactions (i.e., attR1 reacts only with attL1, attR2 reacts only with attL2, etc.).

15 Other mutations were introduced into the short (5 bp) regions flanking the 15 bp core regions of the attB sites to minimize secondary structure formation in single-stranded forms of attB plasmids, e.g., in phagemid ssDNA or in mRNA. Sequences of attB1 and attB2 to the left and right of a nucleic acid molecule of interest after it has been cloned into a Destination Vector are given in Figure 6.

20 Figure 61 illustrates how an Entry Clone and a Destination Vector recombine in the LR Reaction to form a co-integrate, which resolves through a second reaction into two daughter molecules. The two daughter molecules have the same general structure regardless of which pair of sites, attL1 and attR1 or attL2 and attR2, react first to form the co-integrate. The segments change partners by these reactions, regardless of whether the parental molecules are both circular, one is circular and one is linear, or both are linear. In this example, selection for ampicillin resistance carried on the Destination Vector, which also carries the death gene *ccdB*, provides the means for selecting only for the desired attB product plasmid.

25 **Example 3: Protein Expression in the Recombinational Cloning System**

30 Proteins are expressed *in vivo* as a result of two processes, transcription (DNA into RNA), and translation (RNA into protein). For a review of protein expression in prokaryotes and eukaryotes, see Example 13 below. Many vectors (pUC, BlueScript, pGem) use interruption of a transcribed *lacZ* gene for blue-white screening. These plasmids, and many Expression Vectors, use the *lac* promoter to control expression of cloned genes. Transcription from the *lac*

promoter is turned on by adding the inducer IPTG. However, a low level of RNA is made in the absence of inducer, i.e., the lac promoter is never completely off. The result of this "leakiness" is that genes whose expression is harmful to *E. coli* may prove difficult or impossible to clone in vectors that contain the lac promoter, or they may be cloned only as inactive mutants.

In contrast to other gene expression systems, nucleic acid molecules cloned into an Entry Vector may be designed *not* to be expressed. The presence of the strong transcriptional terminator *rrnB* (Orosz, et al., *Eur. J. Biochem.* 201: 653, 1991) just upstream of the attL1 site keeps transcription from the vector promoters (drug resistance and replication origin) from reaching the cloned gene. However, if a toxic gene is cloned into a Destination Vector, the host may be sick, just as in other expression systems. But the reliability of subcloning by *in vitro* recombination makes it easier to recognize that this has happened -- and easier to try another expression option in accordance with the methods of the invention, if necessary.

Example 4: Choosing the Right Entry Vector

There are two kinds of choices that must be made in choosing the best Entry Vector, dictated by (1) the particular DNA segment that is to be cloned, and (2) what is to be accomplished with the cloned DNA segment. These factors are critical in the choice of Entry Vector used, because when the desired nucleic acid molecule of interest is moved from the Entry Vector to a Destination Vector, all the base pairs between the nucleic acid molecule of interest and the Int cutting sites in attL1 and attL2 (such as in Figure 6) move into the Destination Vector as well. For genomic DNAs that are not expressed as a result of moving into a Destination Vector, these decisions are not as critical.

For example, if an Entry Vector with certain translation start signals is used, those sequences will be translated into amino acids if an amino-terminal fusion to the desired nucleic acid molecule of interest is made. Whether the desired nucleic acid molecule of interest is to be expressed as fusion protein, native protein, or both, dictates whether translational start sequences must be included between the attB sites of the clone (native protein) or, alternatively, supplied by the Destination

5 Vector (fusion protein). In particular, Entry Clones that include translational start sequences may prove less suitable for making fusion proteins, as internal initiation of translation at these sites can decrease the yield of N-terminal fusion protein.
10 These two types of expression afforded by the compositions and methods of the invention are illustrated in Figure 62.

5 No Entry Vector is likely to be optimal for all applications. The nucleic acid molecule of interest may be cloned into any of several optimal Entry Vectors.

15 As an example, consider pENTR7 (Figure 16) and pENTR11 (Figure 20), which are useful in a variety of applications, including (but not limited to):

10
20 •Cloning cDNAs from most of the commercially available libraries. The sites to the left and right of the *ccdB* death gene have been chosen so that directional cloning is possible if the DNA to be cloned does not have two or more of these restriction sites.

15
30 •Cloning of genes directionally: *Sall*, *BamHI*, *XmnI* (blunt), or *KpnI* on the left of *ccdB*; *NotI*, *XhoI*, *XbaI*, or *EcoRV* (blunt), on the right.

20
35 •Cloning of genes or gene fragments with a blunt amino end at the *XmnI* site. The *XmnI* site has four of the six most favored bases for eukaryotic expression (see Example 13, below), so that if the first three bases of the DNA to be cloned are ATG, the open reading frame (ORF) will be expressed in eukaryotic cells (e.g., mammalian cells, insect cells, yeast cells) when it is transcribed in the appropriate Destination Vector. In addition, in pENTR11,
40 25 a Shine-Dalgarno sequence is situated 8 bp upstream, for initiating protein synthesis in a prokaryotic host cell (particularly a bacterial cell, such as *E. coli*) at an ATG.

45
30 50 •Cleaving off amino terminal fusions (e.g., His₆, GST, or thioredoxin) using the highly specific TEV (Tobacco Etch Virus) protease (available from Life Technologies, Inc.). If the nucleic acid molecule of interest is cloned at the

5 blunt *Xmn*I site, TEV cleavage will leave two amino acids on the amino end
of the expressed protein.

10 •Selecting against uncut or singly cut Entry Vector molecules during cloning
5 with restriction enzymes and ligase. If the *ccdB* gene is not removed with a
double digest, it will kill any recipient *E. coli* cell that does not contain a
15 mutation that makes the cell resistant to *ccdB* (see U.S. Provisional
Application No. 60/122,392, filed on March 2, 1999, the disclosure of which
is incorporated by reference herein in its entirety).

10 •Allowing production of amino fusions with ORFs in all cloning sites. There
20 are no stop codons (in the attL1 reading frame) upstream of the *ccdB* gene.

25 In addition, pENTR11 is also useful in the following applications:

15 •Cloning cDNAs that have an *Nco*I site at the initiating ATG into the *Nco*I
site. Similar to the *Xmn*I site, this site has four of the six most favored bases
30 for eukaryotic expression. Also, a Shine-Dalgarno sequence is situated 8 bp
upstream, for initiating protein synthesis in a prokaryotic host cell (particularly
20 a bacterial cell, such as *E. coli*) at an ATG.

35 •Producing carboxy fusion proteins with ORFs positioned in phase with the
reading frame convention for carboxy-terminal fusions (see Figure 20A).

40 25 Table 1 lists some non-limiting examples of Entry Vectors and their
characteristics, and Figures 10-20 show their cloning sites. All of the Entry
Vectors listed in Table 1 are available commercially from Life Technologies, Inc.,
45 Rockville, Maryland. Other Entry Vectors not specifically listed here, which
comprise alternative or additional features may be made by one of ordinary skill
30 using routine methods of molecular and cellular biology, in view of the disclosure
contained herein.

Table 1 Examples of Entry Vectors

Designation	Mnemonic Name	Class of Entry Vector	Distinctive Cloning Sites	Amino Fusions	Native Protein in E. coli	Native Protein in Eukaryotic Cells	Protein Synthesis Features
pENTR-1A, 2B, 3C	Minimal blunt RF A, B, C	Alternative Reading Frame Vectors	Reading frame A, B, or C; blunt cut closest to attL1	Good	Poor	Good	Minimal amino acids between tag and protein; no SD
pENTR4	Minimal Nco	Restr. Enz. Cleavage Vectors	Nco I site (common in euk. cDNAs) closest to attL1	Good	Poor	Good	Good Kozac; no SD
pENTR5	Minimal Nde	Restr. Enz. Cleavage Vectors	Nde I site closest to attL1	Good	Poor	Poor at Nde I, Good at Xmn I	No SD; poor Kozac at Nde, good at Xmn
pENTR6	Minimal Sph	Restr. Enz. Cleavage Vectors	Sph I site closest to attL1	Good	Poor	Poor at Sph I, Good at Xmn I	No SD; poor Kozac at Sph, good at Xmn
pENTR7	TEV Blunt	TEV Cleavage Site Present	Xmn I (blunt) is first cloning site after TEV site	Good	Poor	Good at Xmn I site	TEV protease leaves Gly-Thr on amino end of protein; no SD
pENTR8	TEV Nco	TEV Cleavage Site Present	Nco I is first cloning site after TEV site	Good	Poor	Good	TEV protease leaves Gly-Thr on amino end of protein; no SD

pENTR9	TEV Nde	TEV Cleavage Site Present	Nde I is first cloning site after TEV site	Good	Poor	Poor	TEV protease leaves Gly-Thr on amino end of protein; no SD, poor Kozac
pENTR10	Nde with SD	Good SD for E.coli Expression	Strong SD; Nde I site, no TEV	Poor	Good	Poor	Strong SD, internal starts in amino fusions. Poor Kz. No TEV
pENTR11	2 X SD+Kozac	Good SD for E.coli Expression	Xmn I (blunt) and Nco I sites each preceded by SD and Kozac	Good	Good	Good	Strong SD/Koz Internal starts in amino fusions. No TEV

Entry vectors pENTR1A (Figures 10A and 10B), pENTR2B (Figures 11A and 11B), and pENTR3C (Figures 12A and 12B) are almost identical, except that the restriction sites are in different reading frames. Entry vectors pENTR4 (Figures 13A and 13B), pENTR5 (Figures 14A and 14B), and pENTR6 (Figures 15A and 15B) are essentially identical to pENTR1A, except that the blunt *DraI* site has been replaced with sites containing the ATG methionine codon: *NcoI* in pENTR4, *NdeI* in pENTR5, and *SphI* in pENTR6. Nucleic acid molecules that contain one of these sites at the initiating ATG can be conveniently cloned in these Entry vectors. The *NcoI* site in pENTR4 is especially useful for expression of nucleic acid molecules in eukaryotic cells, since it contains many of the bases that give efficient translation (*see* Example 13, below). (Nucleic acid molecules of interest cloned into the *NdeI* site of pENTR5 are not expected to be highly expressed in eukaryotic cells, because the cytosine at position -3 from the initiating ATG is rare in eukaryotic genes.)

Entry vectors pENTR7 (Figures 16A and 16B), pENTR8 (Figures 17A and 17B), and pENTR9 (Figures 18A and 18B) contain the recognition site for the TEV protease between the attL1 site and the cloning sites. Cleavage sites for *XmnI* (blunt), *NcoI*, and *NdeI*, respectively, are the most 5' sites in these Entry vectors. Amino fusions can be removed efficiently if nucleic acid molecules are cloned into these Entry vectors. TEV protease is highly active and highly specific.

Example 5: Controlling Reading Frame

One of the trickiest tasks in expression of cloned nucleic acid molecules is making sure the reading frame is correct. (Reading frame is important if fusions are being made between two ORFs, for example between a nucleic acid molecule of interest and a His6 or GST domain.) For purposes of the present invention, the following convention has been adopted: The reading frame of the DNA cloned into any Entry Vector must be in phase with that of the attB1 site shown in Figure 16A, pENTR7. Notice that the six As of the attL1 site are split into two lysine codons (aaa aaa). The Destination Vectors that make amino fusions were constructed such that they enter the attR1 site in this reading frame.

Destination Vectors for carboxy terminal fusions were also constructed, including those containing His₆ (pDEST23; Figure 43), GST (pDEST24; Figure 44), or thioredoxin (pDEST25; Figure 45) C-terminal fusion sequences.

Therefore, if a nucleic acid molecule of interest is cloned into an Entry Vector so that the aaa aaa reading frame within the attL1 site is in phase with the nucleic acid molecule's ORF, amino terminal fusions will automatically be correctly phased, for all the fusion tags. This is a significant improvement over the usual case, where each different vector can have different restriction sites and different reading frames.

See Example 15 for a practical example of how to choose the most appropriate combinations of Entry Vector and Destination Vector.

Materials

Unless otherwise indicated, the following materials were used in the remaining Examples included herein:

5X LR Reaction Buffer:

200-250 mM (preferably 250 mM) Tris-HCl, pH 7.5

250-350 mM (preferably 320 mM) NaCl

1.25-5 mM (preferably 4.75 mM) EDTA

12.5-35 mM (preferably 22-35 mM, and most preferably 35 mM)

Spermidine-HCl

1 mg/ml bovine serum albumin

GATEWAY™ LR Clonase™ Enzyme Mix:

per 4 µl of 1X LR Reaction Buffer:

150 ng carboxy-His6-tagged Int (see U.S. Appl. Nos. 60/108,324, filed November 13, 1998, and 09/438,358, filed November 12, 1999, both entirely incorporated by reference herein)

-106-

25 ng carboxy-His6-tagged Xis (see U.S. Appl. Nos. 60/108,324, filed
November 13, 1998, and 09/438,358, filed November 12,
1999, both entirely incorporated by reference herein)

30 ng IHF

50% glycerol

5X BP Reaction Buffer:

125 mM Tris-HCl, pH 7.5

110 mM NaCl

25 mM EDTA

25 mM Spermidine-HCl

5 mg/ml bovine serum albumin

GATEWAY™ BP Clonase™ Enzyme Mix:

per 4 µl of 1X BP Reaction Buffer:

200 ng carboxy-His6-tagged Int (see U.S. Appl. Nos. 60/108,324, filed
November 13, 1998, and 09/438,358, filed November 12,
1999, both entirely incorporated by reference herein)

80 ng IHF

50% glycerol

10X Clonase Stop Solution:

50 mM Tris-HCl, pH 8.0

1 mM EDTA

2 mg/ml Proteinase K

Example 6: LR ("Destination") Reaction

To create a new Expression Clone containing the nucleic acid molecule of
interest (and which may be introduced into a host cell, ultimately for production
of the polypeptide encoded by the nucleic acid molecule), an Entry Clone or
Vector containing the nucleic acid molecule of interest, prepared as described

herein, is reacted with a Destination Vector. In the present example, a β -Gal gene flanked by attL sites is transferred from an Entry Clone to a Destination Vector.

Materials needed:

- 5 X LR Reaction buffer
- Destination Vector (preferably linearized), 75-150 ng/ μ l
- Entry Clone containing nucleic acid molecule of interest, 100-300 ng in $\leq 8 \mu$ l TE buffer
- Positive control Entry Clone (pENTR- β -Gal) DNA (See note, below)
- Positive control Destination Vector, pDEST1 (pTrc), 75 ng/ μ l
- GATEWAY™ LR Clonase™ Enzyme Mix (stored at -80° C)
- 10X Clonase Stop solution
- pUC19 DNA, 10 pg/ μ l
- Chemically competent *E. coli* cells (competence: $> 1 \times 10^7$ CFU/ μ g), 400 μ l.
- LB Plates containing ampicillin (100 μ g/ml) and methicillin (200 μ g/ml) \pm X-gal and IPTG (See below)

Notes:

Preparation of the Entry Clone DNA: Miniprep DNA that has been treated with RNase works well. A reasonably accurate quantitation ($\pm 50\%$) of the DNA to be cloned is advised, as the GATEWAY™ reaction appears to have an optimum of about 100-300 ng of Entry Clone per 20 μ l of reaction mix.

The positive control Entry Clone, pENTR- β -Gal, permits functional analysis of clones based on the numbers of expected blue vs. white colonies on LB plates containing IPTG + Blueo-gal (or X-gal), in addition to ampicillin (100 μ g/ml) and methicillin (200 μ g/ml). Because β -Galactosidase is a large protein, it often yields a less prominent band than many smaller proteins do on SDS protein gels.

In the Positive Control Entry Vector pENTR- β -Gal, the coding sequence of β -Gal has been cloned into pENTR11 (Figures 20A and 20B), with translational start signals permitting expression in *E. coli*, as well as in eukaryotic

cells. The positive control Destination Vector, for example pDEST1 (Figure 21), is preferably linearized.

To prepare X-gal + IPTG plates, either of the following protocols may be used:

A. With a glass rod, spread over the surface of an LB agar plate: 40 μ l of 20 mg/ml X-gal (or Bluo-gal) in DMF plus 4 μ l 200 mg/ml IPTG. Allow liquid to adsorb into agar for 3-4 hours at 37° C before plating cells.

B. To liquid LB agar at ~45°C, add: X-gal (or Bluo-Gal) (20 mg/ml in DMF) to make 50 μ g/ml and IPTG (200 mM in water) to make 0.5-1 mM, just prior to pouring plates. Store X-gal and Bluo-Gal in a light-shielded container.

Colony color may be enhanced by placing the plates at 5°C for a few hours after the overnight incubation at 37°C. Protocol B can give more consistent colony color than A, but A is more convenient when selection plates are needed on short notice.

Recombination in Clonase reactions continues for many hours. While incubations of 45-60 minutes are usually sufficient, reactions with large DNAs, or in which both parental DNAs are supercoiled, or which will be transformed into cells of low competence, can be improved with longer incubation times, such as 2-24 hours at 25°C.

Procedure:

1. Assemble reactions as follows (combine all components at room temperature, except GATEWAY™ LR Clonase™ Enzyme Mix ("Clonase LR"), before removing Clonase LR from frozen storage):

		Tube 1	Tube 2	Tube 3	Tube 4
Component		Neg.	Pos.	Neg.	Test
5	p-Gate-βGal, (Positive control Entry Clone) 75 ng/μl	4 μl	4 μl		
10	pDEST1 (Positive control Destination Vector), 75 ng/μl	4 μl	4 μl		
15	Your Entry Clone (100-300 ng)			1 - 8 μl	1 - 8 μl
20	Destination Vector for your nucleic acid molecule, 75 ng/μl			4 μl	4 μl
25	5 X LR Reaction Buffer	4 μl	4 μl	4 μl	4 μl
30	TE	8 μl	4 μl	To 20 μl	To 16 μl
35	GATEWAY™ LR Clonase™ Enzyme Mix (store at - 80° C, add last)	---	4 μl	---	4 μl
40	Total Volume	20 μl	20 μl	20 μl	20 μl

2. Remove the GATEWAY™ LR Clonase™ Enzyme Mix from the -80° C freezer, place immediately on ice. The Clonase takes only a few minutes to thaw.
3. Add 4 μl of GATEWAY™ LR Clonase™ Enzyme Mix to reactions #2 and #4;
4. Return GATEWAY™ LR Clonase™ Enzyme Mix to - 80° C freezer.
5. Incubate tubes at 25° for at least 60 minutes.
6. Add 2 μl Clonase Stop solution to all reactions. Incubate for 20 min at 37°C. (This step usually increases the total number of colonies obtained by 10-20 fold.)
7. Transform 2 μl into 100 μl competent *E. coli*. Select on plates containing ampicillin at 100 μg/ml.

Example 7: Transformation of *E. coli*

To introduce cloning or Expression Vectors prepared using the recombinational cloning system of the invention, any standard *E. coli* transformation protocol should be satisfactory. The following steps are recommended for best results:

1. Let the mixture of competent cells and Recombinational Cloning System reaction product stand on ice at least 15 minutes prior to the heat-shock step. This gives time for the recombination proteins to dissociate from the DNA, and improves the transformation efficiency.

2. Expect the reaction to be about 1%-5% efficient, i.e., 2 µl of the reaction should contain at least 100 pg of the Expression Clone plasmid (taking into account the amounts of each parental plasmid in the reaction, and the subsequent dilution). If the E. coli cells have a competence of 10^7 CFU/µg, 100 pg of the desired clone plasmid will give about 1000 colonies, or more, if the entire transformation is spread on one ampicillin plate.

3. Always do a control pUC DNA transformation. If the number of colonies is not what you expect, the pUC DNA transformation gives you an indication of where the problem was.

Example 8: Preparation of attB-PCR Product

For preparation of attB-PCR products in the PCR cloning methods described in Example 9 below, PCR primers containing attB1 and attB2 sequences are used. The attB1 and attB2 primer sequences are as follows:

attB1: 5'-GGGGACAAGTTTGTACAAAAAAGCAGGCT-(template-specific sequence)-3'

attB2: 5'-GGGGACCACTTTGTACAAGAAAGCTGGGT-(template-specific sequence)-3'

The attB1 sequence should be added to the amino primer, and the attB2 sequence to the carboxy primer. The 4 guanines at the 5' ends of each of these primers enhance the efficiency of the minimal 25 bp attB sequences as substrates for use in the cloning methods of the invention.

Standard PCR conditions may be used to prepare the PCR product. The following suggested protocol employs PLATINUM *Taq* DNA Polymerase High

Fidelity®, available commercially from Life Technologies, Inc. (Rockville, MD). This enzyme mix eliminates the need for hot starts, has improved fidelity over Taq, and permits synthesis of a wide range of amplicon sizes, from 200 bp to 10 kb, or more, even on genomic templates.

Materials needed:

- PLATINUM Taq DNA Polymerase High Fidelity® (Life Technologies, Inc.)
- attB1- and attB2- containing primer pair (see above) specific for your template
- DNA template (linearized plasmid or genomic DNA)
- 10X High Fidelity PCR Buffer
- 10 mM dNTP mix
- PEG/MgCl₂ Mix (30% PEG 8000, 30 mM MgCl₂)

Procedure:

1.) Assemble the reaction as follows:

Component	Reaction with Plasmid Target	Reaction with Genomic Target
10X High Fidelity PCR Buffer	5 µl	5 µl
dNTP Mix 10 mM	1 µl	1 µl
MgSO ₄ , 50mM	2 µl	2 µl
attB1 Primer, 10 µM	2 µl	1 µl
attB2 Primer, 10 µM	2 µl	1 µl
Template DNA	1-5 ng*	≥ 100 ng
PLATINUM Taq High Fidelity	2 µl	1 µl
Water	to 50 µl	to 50 µl

* Use of higher amounts of plasmid template may permit fewer cycles (10-15) of PCR

2.) Add 2 drops mineral oil, as appropriate.

3.) Denature for 30 sec. at 94°C.

4.) Perform 25 cycles:

94°C for 15 sec-30 sec

55°C for 15 sec-30 sec

68°C for 1 min per kb of template.

5.) Following the PCR reaction, apply 1-2 µl of the reaction mixture to an agarose gel, together with size standards (e.g., 1 Kb Plus Ladder, Life Technologies, Inc.) and quantitation standards (e.g., Low Mass Ladder, Life Technologies, Inc.), to assess the yield and uniformity of the product.

Purification of the PCR product is recommended, to remove attB primer dimers which can clone efficiently into the Entry Vector. The following protocol is fast and will remove DNA <300 bp in size:

6.) Dilute the 50 µl PCR reaction to 200 µl with TE.

7.) Add 100 µl PEG/MgCl₂ Solution. Mix and centrifuge immediately at 13,000 RPM for 10 min at room temperature. Remove the supernatant (pellet is clear and hard to see).

8.) Dissolve the pellet in 50 µl TE and check recovery on a gel.

If the starting PCR template is a plasmid that contains the gene for Kan^r, it is advisable to treat the completed PCR reaction with the restriction enzyme *DpnI*, to degrade the plasmid since unreacted residual starting plasmid is a potential source of false-positive colonies from the transformation of the GATEWAY™ Cloning System reaction. Adding ~5 units of *DpnI* to the completed PCR reaction and incubating for 15 min at 37°C will eliminate this potential problem. Heat inactivate the *DpnI* at 65°C for 15 min, prior to using the PCR product in the GATEWAY™ Cloning System reaction.

Example 9: Cloning attB-PCR products into Entry Vectors via the BP ("Gateway") Reaction

The addition of 5'-terminal attB sequences to PCR primers allows synthesis of a PCR product that is an efficient substrate for recombination with a Donor (attP) Plasmid in the presence of GATEWAY™ BP Clonase™ Enzyme Mix. This reaction produces an Entry Clone of the PCR product (See Figure 8).

The conditions of the Gateway Cloning reaction with an attB PCR substrate are similar to those of the BP Reaction (see Example 10 below), except that the attB-PCR product (see Example 8) substitutes for the Expression Clone, and the attB-PCR positive control (attB-tet^r) substitutes for the Expression Clone Positive Control (GFP).

Materials needed:

- 5 X BP Reaction Buffer
- Desired attB-PCR product DNA, 50-100 ng in $\leq 8 \mu\text{l}$ TE.
- Donor (attP) Plasmid (Figures 49-54), 75 ng/ μl , supercoiled DNA
- attB-tet^r PCR product positive control, 25 ng/ μl
- GATEWAY™ BP Clonase™ Enzyme Mix (stored at -80°C)
- 10x Clonase Stop Solution
- pUC19 DNA, 10 pg/ μl .
- Chemically competent E.coli cells (competence: $\geq 1 \times 10^7$ CFU/ μg), 400 μl

Notes:

- Preparation of attB-PCR DNA: see Example 8.

- The Positive Control attB-tet^r PCR product contains a functional copy of the tet^r gene of pBR322, with its own promoter. By plating the transformation of the control BP Reaction on kanamycin (50 $\mu\text{g/ml}$) plates (if kan^r Donor Plasmids are used; see Figures 49-52) or an alternative selection agent (e.g., gentamycin, if gen^r Donor Plasmids are used; see Figure 54), and then picking about 50 of these colonies onto plates with tetracycline (20 $\mu\text{g/ml}$), the

percentage of Entry Clones containing functional tet^r among the colonies from the positive control reaction can be determined (% Expression Clones = (number of tet^r + kan^r (or gen^r) colonies/kan^r (or gen^r) colonies).

Procedure:

1. Assemble reactions as follows. Combine all components except GATEWAY™ BP Clonase™ Enzyme Mix, before removing GATEWAY™ BP Clonase™ Enzyme Mix from frozen storage.

Component	Neg.	Pos.	Test
	Tube 1	Tube 2	Tube 3
attB-PCR product, 50-100 ng			1 - 8 µl
Donor (attP) Plasmid 75 ng/µl	2 µl	2 µl	2 µl
attB-PCR tet ^r control DNA (75 ng/µl)		4 µl	
5 X BP Reaction Buffer	4 µl	4 µl	4 µl
TE	10 µl	6 µl	To 16 µl
GATEWAY™ BP Clonase™ Enzyme Mix (store at -80° C, add last)	4 µl	4 µl	4 µl
Total Volume	20 µl	20 µl	20 µl

2. Remove the GATEWAY™ BP Clonase™ Enzyme Mix from the -80° C freezer, place immediately on ice. The Clonase takes only a few minutes to thaw.
3. Add 4 µl of GATEWAY™ BP Clonase™ Enzyme Mix to the subcloning reaction, mix.
4. Return GATEWAY™ BP Clonase™ Enzyme Mix to -80° C freezer.
5. Incubate tubes at 25° for at least 60 minutes.

- 5 6. Add 2 μ l Proteinase K (2 μ g/ μ l) to all reactions. Incubate for 20 min at 37°C.
7. Transform 2 μ l into 100 μ l competent *E. coli*, as per 3.2, above. Select on LB
- 10 plates containing kanamycin, 50 μ g/ml.

5 Results:

 In initial experiments, primers for amplifying tetR and ampR from pBR322

15 were constructed containing only the tetR- or ampR-specific targeting sequences,

 the targeting sequences plus attB1 (for forward primers) or attB2 (for reverse

 primers) sequences shown in Figure 9, or the attB1 or attB2 sequences with a 5'

10 tail of four guanines. The construction of these primers is depicted in Figure 65.

20 After PCR amplification of tetR and ampR from pBR322 using these primers and

 cloning the PCR products into host cells using the recombinational cloning system

 of the invention, the results shown in Figure 66 were obtained. These results

25 demonstrated that primers containing attB sequences provided for a somewhat

 higher number of colonies on the tetracycline and ampicillin plates. However,

15 inclusion of the 5' extensions of four or five guanines on the primers in addition

 to the attB sequences provided significantly better cloning results, as shown in

30 Figures 66 and 67. These results indicate that the optimal primers for cloning of

 PCR products using recombinational cloning will contain the recombination site

 sequences with a 5' extension of four or five guanine bases.

20

35 To determine the optimal stoichiometry between attB-containing PCR

 products and attP-containing Donor plasmid, experiments were conducted where

 the amount of PCR product and Donor plasmid were varied during the BP

40 Reaction. Reaction mixtures were then transformed into host cells and plated on

25 tetracycline plates as above. Results are shown in Figure 68. These results

 indicate that, for optimal recombinational cloning results with a PCR product in

 the size range of the tet gene, the amounts of attP-containing Donor plasmids are

45 between about 100-500 ng (most preferably about 200-300 ng), while the optimal

 concentrations of attB-containing PCR products is about 25-100 ng (most

30 preferably about 100 ng), per 20 μ l reaction.

50 Experiments were then conducted to examine the effect of PCR product size

 on efficiency of cloning via the recombinational cloning approach of the invention.

PCR products containing attB1 and attB2 sites, at sizes 256 bp, 1 kb, 1.4 kb, 3.4 kb, 4.6 kb, 6.9 kb and 10.1 kb were prepared and cloned into Entry vectors as described above, and host cells were transformed with the Entry vectors containing the cloned PCR products. For each PCR product, cloning efficiency was calculated relative to cloning of pUC19 positive control plasmids as follows:

$$\text{Cloning Efficiency} = \frac{\text{CFU/ng attB PCR product}}{\text{CFU/ng pUC19 control}} \times \frac{\text{Size (kb) PCR product}}{\text{Size (kb) pUC19 control}}$$

The results of these experiments are depicted in Figures 69A-69C (for 256 bp PCR fragments), 70A-70C (for 1 kb PCR fragments), 71A-71C (for 1.4 kb PCR fragments), 72A-72C (for 3.4 kb PCR fragments), 73A-73C (for 4.6 kb PCR fragments), 74 (for 6.9 kb PCR fragments), and 75-76 (for 10.1 kb PCR fragments). The results shown in these figures are summarized in Figure 77, for different weights and moles of input PCR DNA.

Together, these results demonstrate that attB-containing PCR products ranging in size from about 0.25 kb to about 5 kb clone relatively efficiently in the recombinational cloning system of the invention. While PCR products larger than about 5 kb clone less efficiently (apparently due to slow resolution of cointegrates), longer incubation times during the recombination reaction appears to improve the efficiency of cloning of these larger PCR fragments. Alternatively, it may also be possible to improve efficiency of cloning of large (> about 5 kb) PCR fragments by using lower levels of input attP Donor plasmid and perhaps attB-containing PCR product, and/or by adjusting reaction conditions (*e.g.*, buffer conditions) to favor more rapid resolution of the cointegrates.

Example 10: The BP Reaction

One purpose of the Gateway ("Entry") reaction is to convert an Expression Clone into an Entry Clone. This is useful when you have isolated an individual Expression Clone from an Expression Clone cDNA library, and you wish to transfer the nucleic acid molecule of interest into another Expression Vector, or

to move a population of molecules from an attB or attL library. Alternatively, you may have mutated an Expression Clone and now wish to transfer the mutated nucleic acid molecule of interest into one or more new Expression Vectors. In both cases, it is necessary first to convert the nucleic acid molecule of interest to an Entry Clone.

Materials needed:

- 5 X BP Reaction Buffer
- Expression Clone DNA, 100-300 ng in ≤ 8 μ l TE.
- Donor (attP) Vector, 75 ng/ μ l, supercoiled DNA
- Positive control attB-tet-PCR DNA, 25 ng/ μ l
- GATEWAY™ BP Clonase™ Enzyme Mix (stored at - 80°C)
- Clonase Stop Solution (Proteinase K, 2 μ g/ μ l).

Notes:

Preparation of the Expression Clone DNA: Miniprep DNA treated with RNase works well.

1. As with the LR Reaction (see Example 14), the BP Reaction is strongly influenced by the topology of the reacting DNAs. In general, the reaction is most efficient when one of the DNAs is linear and the other is supercoiled, compared to reactions where the DNAs are both linear or both supercoiled. Further, linearizing the attB Expression Clone (anywhere within the vector) will usually give more colonies than linearizing the Donor (attP) Plasmid. If finding a suitable cleavage site within your Expression Clone vector proves difficult, you may linearize the Donor (attP) Plasmid between the attP1 and attP2 sites (for example, at the *Nco*I site), avoiding the *ccdB* gene. Maps of Donor (attP) Plasmids are given in Figures 49-54.

Procedure:

1. Assemble reactions as follows. Combine all components at room temperature, except GATEWAY™ BP Clonase™ Enzyme Mix, before removing GATEWAY™ BP Clonase™ Enzyme Mix from freezer.

Component	Neg.	Pos.	Test
	Tube 1	Tube 2	Tube 3
Positive Control, attB-tet-PCR DNA, 25 ng/ μ l	4 μ l	4 μ l	
Desired attB Expression Clone DNA (100ng) linearized			1 - 8 μ l
Donor (attP) Plasmid, 75 ng/ μ l	2 μ l	2 μ l	2 μ l
5 X BP Reaction Buffer	4 μ l	4 μ l	4 μ l
TE	10 μ l	6 μ l	To 16 μ l
GATEWAY™ BP Clonase™ Enzyme Mix (store at - 80° C, add last)	---	4 μ l	4 μ l
Total Volume	20 μ l	20 μ l	20 μ l

2. Remove the GATEWAY™ BP Clonase™ Enzyme Mix from the -80°C freezer, place immediately on ice. The mixture takes only a few minutes to thaw.
3. Add 4 μ l of GATEWAY™ BP Clonase™ Enzyme Mix to the subcloning reaction, mix.
4. Return GATEWAY™ BP Clonase™ Enzyme Mix to - 80° C freezer.
5. Incubate tubes at 25° for at least 60 minutes. If both the attB and attP DNAs are supercoiled, incubation for 2-24 hours at 25°C is recommended.
6. Add 2 μ l Clonase Stop Solution. Incubate for 10 min at 37°C.
7. Transform 2 μ l into 100 μ l competent E. coli, as above. Select on LB plates containing 50 μ g/ml kanamycin.

Example 11: Cloning PCR Products into Entry Vectors using Standard Cloning Methods

Preparation of Entry Vectors for Cloning of PCR Products

All of the Entry Vectors of the invention contain the death gene *ccdB* as a stuffer between the "left" and "right" restriction sites. The advantage of this arrangement is that there is virtually no background from vector that has not been cut with both restriction enzymes, because the presence of the *ccdB* gene will kill

all standard E. coli strains. Thus it is necessary to cut each Entry Vector twice, to remove the ccdB fragment.

We strongly recommend that, after digestion of the Entry Vector with the second restriction enzyme, you treat the reaction with phosphatase (calf intestine alkaline phosphatase, CIAP or thermosensitive alkaline phosphatase, TSAP). The phosphatase can be added directly to the reaction mixture, incubated for an additional time, and inactivated. This step dephosphorylates both the vector and ccdB fragments, so that during subsequent ligation there is less competition between the ccdB fragment and the DNA of interest for the termini of the Entry Vector.

Blunt Cloning of PCR products

Generally PCR products do not have 5' phosphates (because the primers are usually 5' OH), and they are not necessarily blunt. (On this latter point, see Brownstein, et al., *BioTechniques* 20: 1006, 1996 for a discussion of how the sequence of the primers affects the addition of single 3' bases.) The following protocol repairs these two defects.

In a 0.5 ml tube, ethanol precipitate about 40 ng of PCR product (as judged from an agarose gel).

1. Dissolve the precipitated DNA in 10 μ l comprising 1 μ l 10 mM rATP, 1 μ l mixed 2 mM dNTPs (i.e., 2 mM each dATP, dCTP, dTTP, and dGTP), 2 μ l 5x T4 polynucleotide kinase buffer (350 mM Tris HCl (pH7.6), 50 mM $MgCl_2$, 500mM KCl, 5 mM 2-mercaptoethanol) 10 units T4 polynucleotide kinase, 1 μ l T4 DNA polymerase, and water to 10 μ l.
2. Incubate the tube at 37° for 10 minutes, then at 65° for 15 minutes, cool, centrifuge briefly to bring any condensate to the tip of the tube.
3. Add 5 μ l of the PEG/ $MgCl_2$ solution, mix and centrifuge at room temperature for 10 minutes. Discard supernatant.
4. Dissolve the invisible precipitate in 10 μ l containing 2 μ l 5x T4 DNA ligase buffer (Life Technologies, Inc.), 0.5 units T4 DNA ligase, and about 50 ng of blunt, phosphatase-treated Entry Vector.

5. Incubate at 25° for 1 hour, then 65° for 10 minutes. Add 90 µl TE, transform 10 µl into 50 - 100 µl competent *E. coli* cells.
6. Plate on kanamycin.

Note: In the above protocol, steps b-c simultaneously polish the ends of the PCR product (through the exonuclease and polymerase activities of T4 DNA polymerase) and phosphorylate the 5' ends (using T4 polynucleotide kinase). It is necessary to inactivate the kinase, so that the blunt, dephosphorylated vector in step e cannot self ligate. Step d (the PEG precipitation) removes all small molecules (primers, nucleotides), and has also been found to improve the yield of cloned PCR product by 50 fold.

Cloning PCR Products after Digestion with Restriction Enzymes

Efficient cloning of PCR products that have been digested with restriction enzymes includes three steps: inactivation of *Taq* DNA polymerase, efficient restriction enzyme cutting, and removal of small DNA fragments.

Inactivation of Taq DNA Polymerase: Carryover of *Taq* DNA polymerase and dNTPs into a RE digestion significantly reduces the success in cloning a PCR product (D. Fox et al., *FOCUS* 20(1):15, 1998), because *Taq* DNA polymerase can fill in sticky ends and add bases to blunt ends. Either TAQQUENCH™ (obtainable from Life Technologies, Inc.; Rockville, Maryland) or extraction with phenol can be used to inactivate the *Taq*.

Efficient Restriction Enzyme Cutting: Extra bases on the 5' end of each PCR primer help the RE cut near ends of PCR products. With the availability of cheap primers, adding 6 to 9 bases on the 5' sides of the restriction sites is a good investment to ensure that most of the ends are digested. Incubation of the DNA with a 5-fold excess of restriction enzyme for an hour or more helps ensure success.

Removal of Small Molecules before Ligation: Primers, nucleotides, primer dimers, and small fragments produced by the restriction enzyme digestion,

can all inhibit or compete with the desired ligation of the PCR product to the cloning vector. This protocol uses PEG precipitation to remove small molecules.

Protocol for cutting the ends of PCR products with restriction enzyme(s):

1. Inactivation of Taq DNA polymerase in the PCR product:

Option A: Extraction with Phenol

A1. Dilute the PCR reaction to 200 μ l with TE. Add an equal volume of phenol:chloroform:isoamyl alcohol, vortex vigorously for 20 seconds, and centrifuge for 1 minute at room temperature. Discard the lower phase.

A2. Extract the phenol from the DNA and concentrate as follows. Add an equal volume of 2-butanol (colored red with "Oil Red O" from Aldrich, if desired), vortex briefly, centrifuge briefly at room temperature. Discard the upper butanol phase. Repeat the extraction with 2-butanol. This time the volume of the lower aqueous phase should decrease significantly. Discard the upper 2-butanol phase.

A3. Ethanol precipitate the DNA from the aqueous phase of the above extractions. Dissolve in a 200 μ l of a suitable restriction enzyme (RE) buffer.

Option B: Inactivation with TaqQuench

B1. Ethanol precipitate an appropriate amount of PCR product (100 ng to 1 μ g), dissolve in 200 μ l of a suitable RE buffer.

B2. Add 2 μ l TaqQuench.

2. Add 10 to 50 units of restriction enzyme and incubate for at least 1 hour. Ethanol precipitate if necessary to change buffers for digestion at the other end of the PCR product.

3. Add ½ volume of the PEG/MgCl₂ mix to the RE digestion. Mix well and immediately centrifuge at room temperature for 10 minutes. Discard the supernatant (pellet is usually invisible), centrifuge again for a few seconds, discard any remaining supernatant.

4. Dissolve the DNA in a suitable volume of TE (depending on the amount of PCR product in the original amplification reaction) and apply an aliquot to an agarose gel to confirm recovery. Apply to the same gel 20-100 ng of the appropriate Entry Vector that will be used for the cloning.

Example 12: Determining The Expected Size of the GATEWAY™ Cloning Reaction Products

If you have access to a software program that will electronically cut and splice sequences, you can create electronic clones to aid you in predicting the sizes and restriction patterns of GATEWAY™ Cloning System recombination products.

The cleavage and ligation steps performed by the enzyme Int in the GATEWAY™ Cloning System recombination reactions mimic a restriction enzyme cleavage that creates a 7-bp 5'-end overhang followed by a ligation step that reseals the ends of the daughter molecules. The recombination proteins present in the Clonase cocktails (see Example 19 below) recognize the 15 bp core sequence present within all four types of att sites (in addition to other flanking sequences characteristic of each of the different types of att sites).

By treating these sites in your software program as if they were restriction sites, you can cut and splice your Entry Clones with various Destination Vectors and obtain accurate maps and sequences of the expected results from your GATEWAY™ Cloning System reactions.

Example 13: Protein Expression
Brief Review of Protein Expression

Transcription: The most commonly used promoters in *E. coli* Expression Vectors are variants of the lac promoter, and these can be turned on by adding

5 IPTG to the growth medium. It is usually good to keep promoters off until
expression is desired, so that the host cells are not made sick by the
10 overabundance of some heterologous protein. This is reasonably easy in the case
of the lac promoters used in *E. coli*. One needs to supply the *lac I* gene (or its
5 more productive relative, the *lac I^q* gene) to make *lac* repressor protein, which
binds near the promoter and keeps transcription levels low. Some Destination
15 Vectors for *E. coli* expression carry their own *lacI^q* gene for this purpose.
(However, lac promoters are always a little "on," even in the absence of IPTG.)

Controlling transcription in eukaryotic cells is not nearly so straightforward
10 or efficient. The tetracycline system of Bujard and colleagues is the most
20 successful approach, and one of the Destination Vectors (pDEST11; Figure 31)
has been constructed to supply this function.

Translation: Ribosomes convert the information present in mRNA into
25 protein. Ribosomes scan RNA molecules looking for methionine (AUG) codons,
15 which begin nearly all nascent proteins. Ribosomes must, however, be able to
distinguish between AUG codons that code for methionine in the middle of
proteins from those at the start. Most often ribosomes choose AUGs that are 1)
30 first in the RNA (toward the 5' end), and 2) have the proper sequence context.
In *E. coli* the favored context (first recognized by Shine and Dalgarno, *Eur. J.*
20 *Biochem.* 57: 221 (1975)) is a run of purines (As and Gs) from five to 12 bases
35 upstream of the initiating AUG, especially AGGAGG or some variant.

In eukaryotes, a survey of translated mRNAs by Kozak (*J. Biol. Chem.*
266: 19867 (1991)) has revealed a preferred sequence context, gcc Acc ATGG,
40 around the initiating methionine, with the A at -3 being most important, and a
25 purine at +4 (where the A of the ATG is +1), preferably a G, being next most
influential. Having an A at -3 is enough to make most ribosomes choose the first
AUG of an mRNA, in plants, insects, yeast, and mammals. (For a review of
45 initiation of protein synthesis in eukaryotic cells, see: Pain, V.M. *Eur. J. Biochem.*
236:747-771, 1996.)

30 *Consequences of Translation Signals for GATEWAY™ Cloning System:*
First, translation signals (Shine-Dalgarno in *E. coli*, Kozak in eukaryotes) have to
50 be close to the initiating ATG. The attB site is 25 base pairs long. Thus if

translation signals are desired near the natural ATG of the nucleic acid molecule of interest, they must be present in the Entry Clone of that nucleic acid molecule of interest. Also, when a nucleic acid molecule of interest is moved from an Entry Clone to a Destination vector, any translation signals will move along. The result is that the presence or absence of Shine-Dalgarno and/or Kozak sequences in the Entry Clone must be considered, with the eventual Destination Vectors to be used in mind.

Second, although ribosomes choose the 5' ATG most often, internal ATGs are also used to begin protein synthesis. The better the translation context around this internal ATG, the more internal translation initiation will be seen. This is important in the GATEWAY™ Cloning System, because you can make an Entry Clone of your nucleic acid molecule of interest, and arrange to have Shine-Dalgarno and/or Kozak sequences near the ATG. When this cassette is recombined into a Destination Vector that transcribes your nucleic acid molecule of interest, you get native protein. If you want, you can make a fusion protein in a different Destination Vector, since the Shine-Dalgarno and/or Kozak sequences do not contain any stop signals in the same reading frame. However, the presence of these internal translation signals may result in a significant amount of native protein being made, contaminating, and lowering the yield of, your fusion protein. This is especially likely with short fusion tags, like His6.

A good compromise can be recommended. If an Entry Vector like pENTR7 (Figure 16) or pENTR8 (Figure 17) is chosen, the Kozak bases are present for native eukaryotic expression. The context for *E. coli* translation is poor, so the yield of an amino-terminal fusion should be good, and the fusion protein can be digested with the TEV protease to make near-native protein following purification.

Recommended Conditions for Synthesis of Proteins in E. coli: When making proteins in *E. coli* it is advisable, at least initially, to incubate your cultures at 30°C, instead of at 37°C. Our experience indicates that proteins are less likely to form aggregates at 30°C. In addition, the yields of proteins from cells grown at 30°C frequently are improved.

5 The yields of proteins that are difficult to express may also be improved
by inducing the cultures in mid-log phase of growth, using cultures begun in the
10 morning from overnight growths, as opposed to harvesting directly from an
overnight culture. In the latter case, the cells are preferably in late log or
5 stationary growth, which can favor the formation of insoluble aggregates.

15 ***Example 14: Constructing Destination Vectors from Existing Vectors***

Destination Vectors function because they have two recombination sites,
10 attR1 and attR2, flanking a chloramphenicol resistance (CmR) gene and a death
gene, ccdB. The GATEWAY™ Cloning System recombination reactions
20 exchange the entire Cassette (except for a few bases comprising part of the attB
sites) for the DNA segment of interest from the Entry Vector. Because attR1,
CmR, ccdB gene, and attR2 are contiguous, they can be moved on a single DNA
25 segment. If this Cassette is cloned into a plasmid, the plasmid becomes a
Destination Vector. Figure 63 shows a schematic of the GATEWAY™ Cloning
System Cassette; attR cassettes in all three reading frames contained in vectors
30 pEZC15101, pEZC15102 and pEZC15103 are shown in Figures 64A, 64B, and
64C, respectively.

20 The protocol for constructing a Destination Vector is presented below.
Keep in mind the following points:

- 35 ◦ Destination Vectors must be constructed and propagated in one of the DB
strains of *E. coli* (e.g., DB3.1, and particularly *E. coli* LIBRARY
EFFICIENCY® DB3.1™ Competent Cells) available from Life
40 25 Technologies, Inc. (and described in detail in U.S. Provisional Application
No. 60/122,392, filed on March 2, 1999, which is incorporated herein by
reference), because the ccdB death gene will kill any *E. coli* strain that has
not been mutated such that it will survive the presence of the ccdB gene.
- 45 30 ◦ If your Destination Vector will be used to make a fusion protein, a
GATEWAY™ Cloning System cassette with the correct reading frame
must be used. The nucleotide sequences of the ends of the cassettes are
50 shown in Figure 78. The reading frame of the fusion protein domain must

be in frame with the core region of the attR1 site (for an amino terminal fusion) so that the six As are translated into two lysine codons. For a C-terminal fusion protein, translation through the core region of the attR2 site should be in frame with -TAC-AAA-, to yield -Tyr-Lys-.

- Note that each reading frame Cassette has a different unique restriction site between the chloramphenicol resistance and *ccdB* genes (*Mlu*I for reading frame A, *Bgl*II for reading frame B, and *Xba*I for reading frame C; see Figure 63).
- Most standard vectors can be converted to Destination Vectors, by inserting the Entry Cassette into the MCS of that vector.

Protocol for Making a Destination Vector

1. If the vector will make an amino fusion protein, it is necessary to keep the "aaa" triplets in attR1 in phase with the triplets of the fusion protein. Determine which Entry cassette to use as follows:

a.) Write out the nucleotide sequence of the existing vector near the restriction site into which the Entry cassette will be cloned. These must be written in triplets corresponding to the amino acid sequence of the fusion domain.

b.) Draw a vertical line through the sequence that corresponds to the restriction site end, after it has been cut and made blunt, i.e., after filling in a protruding 5' end or polishing a protruding 3' end.

c.) Choose the appropriate reading frame cassette:

- If the coding sequence of the blunt end ends after a complete codon triplet, use the reading frame A cassette. See Figures 78, 79 and 80.

•If the coding sequence of the blunt end ends in a single base, use the reading frame B cassette. See Figures 78, 79 and 81.

•If the coding sequence of the blunt end ends in two bases, use the reading frame C cassette. See Figures 78, 79, 82A-B, and 83A-C.

2. Cut one to five micrograms of the existing plasmid at the position where you wish your nucleic acid molecule of interest (flanked by att sites) to be after the recombination reactions. Note: it is better to remove as many of the MCS restriction sites as possible at this step. This makes it more likely that restriction enzyme sites within the GATEWAY™ Cloning System Cassette will be unique in the new plasmid, which is important for linearizing the Destination Vector (Example 14, below).

3. Remove the 5' phosphates with alkaline phosphatase. While this is not mandatory, it increases the probability of success.

4. Make the end(s) blunt with fill-in or polishing reactions. For example, to 1 µg of restriction enzyme-cut, ethanol-precipitated vector DNA, add:

- i. 20 µl 5x T4 DNA Polymerase Buffer (165 mM Tris-acetate (pH 7.9), 330 mM Na acetate, 50 mM Mg acetate, 500 µg/ml BSA, 2.5 mM DTT)
- ii. 5 µl 10mM dNTP mix
- iii. 1 Unit of T4 DNA Polymerase
- iv. Water to a final volume of 100 µl
- v. Incubate for 15 min at 37°C.

5. Remove dNTPs and small DNA fragments: Ethanol precipitate (add three volumes of room temperature ethanol containing 0.1 M sodium acetate, mix well, immediately centrifuge at room temperature 5 - 10 minutes), dissolve wet precipitate in 200 µl TE, add 100 µl 30% PEG 8000, 30 mM MgCl₂, mix well,

immediately centrifuge for 10 minutes at room temperature, discard supernatant, centrifuge again a few seconds, discard any residual liquid.

6. Dissolve the DNA to a final concentration of 10 - 50 ng per microliter. Apply 20 - 100 ng to a gel next to supercoiled plasmid and linear size standards to confirm cutting and recovery. The cutting does not have to be 100% complete, since you will be selecting for the chloramphenicol marker on the Entry cassette.

7. In a 10 µl ligation reaction combine 10 - 50 ng vector, 10 - 20 ng of Entry Cassette (Figure 79), and 0.5 units T4 DNA ligase in ligase buffer. After one hour (or overnight, whichever is most convenient), transform 1 µl into one of the DB strains of competent *E. coli* cells with a *gyrA462* mutation (See U.S. Provisional Application No. 60/122,392, filed on March 2, 1999, which is incorporated herein by reference), preferably DB3.1, and most preferably *E. coli* LIBRARY EFFICIENCY®DB3.1™ Competent Cells. The *ccdB* gene on the Entry Cassette will kill other strains of *E. coli* that have not been mutated so as to survive the presence of the *ccdB* gene.

8. After expression in SOC medium, plate 10 µl and 100 µl on chloramphenicol-containing (30 µg / ml) plates, incubate at 37° C.

9. Pick colonies, make miniprep DNA. Treat the miniprep with RNase A and store in TE. Cut with the appropriate restriction enzyme to determine the orientation of the Cassette. Choose clones with the attR1 site next to the amino end of the protein expression function of the plasmid.

Notes on Using Destination Vectors

- We have found that about ten-fold more colonies result from a GATEWAY™ Cloning System reaction if the Destination Vector is linear or relaxed. If the competent cells you use are highly competent ($>10^8$ per microgram), linearizing the Destination Vector is less essential.

- The site or sites used for the linearization must be within the Entry Cassette. Sites that cut once or twice within each cassette are shown in Figures 80-82.
- Minipreps of Destination Vectors will work fine, so long as they have been treated with RNase. Since most DB strains are *endA*- (See U.S. Provisional Application No. 60/122,392, filed on March 2, 1999, which is incorporated herein by reference), minipreps can be digested with restriction enzymes without a prior phenol extraction.
- Reading the OD₂₆₀ of miniprep DNA is inaccurate unless the RNA and ribonucleotides have been removed, for example, by a PEG precipitation.

Example 15: Some Options in Choosing Appropriate Entry Vectors and Destination Vectors: An Example

In some applications, it may be desirable to express a nucleic acid molecule of interest in two forms: as an amino-terminal fusion in *E. coli*, and as a native protein in eukaryotic cells. This may be accomplished in any of several ways:

Option 1: Your choices depend on your nucleic acid molecule of interest and the fragment that contains it, as well as the available Entry Vectors. For eukaryotic translation, you need consensus bases according to Kozak (*J. Biol. Chem.* 266:19867, 1991) near the initiating methionine (ATG) codon. All of the Entry Vectors offer this motif upstream of the *XmnI* site (blunt cutter). One option is to amplify your nucleic acid molecule of interest, with its ATG, by PCR, making the amino end blunt and the carboxy end containing the natural stop codon followed by one of the "right side" restriction sites (*EcoRI*, *NotI*, *XhoI*, *EcoRV*, or *XbaI* of the pENTR vectors).

If you know your nucleic acid molecule of interest does not have, for example, an *XhoI* site, you can make a PCR product that has this structure:

Xho I

5' ATG nnn nnn --- nnn TAA ctc gag nnn nnn 3'

3' tac nnn nnn --- nnn att gag ctc nnn nnn 5'

After cutting with *Xho*I, the fragment is ready to clone:

```
5' ATG nnn nnn --- nnn TAA c      3'
3' tac nnn nnn --- nnn att gag ct  5'
```

(If you follow this example, don't forget to put a phosphate on the amino oligo.)

Option 2: This PCR product could be cloned into two Entry Vectors to give the desired products, between the *Xmn*I and *Xho*I sites: pENTR1A (Figures 10A, 10B) or pENTR7 (Figures 16A, 16B). If you clone into pENTR1A, amino fusions will have the minimal number of amino acids between the fusion domain and your nucleic acid molecule of interest, but the fusion cannot be removed with TEV protease. The converse is true of clones in pENTR7, i.e., an amino fusion can be cleaved with TEV protease, at the cost of more amino acids between the fusion and your nucleic acid molecule of interest.

In this example, let us choose to clone our hypothetical nucleic acid molecule of interest into pENTR7, between the *Xmn*I and *Xho*I sites. Once this is accomplished, several optional protocols using the Entry Clone pENTR7 may be followed:

Option 3: Since the nucleic acid molecule of interest has been amplified with PCR, it may be desirable to sequence it. To do this, transfer the nucleic acid molecule of interest from the Entry Vector into a vector that has priming sites for the standard sequencing primers. Such a vector is pDEST6 (Figures 26A, 26B). This Destination Vector places the nucleic acid molecule of interest in the opposite orientation to the lac promoter (which is leaky -- see Example 3 above). If the gene product is toxic to *E. coli*, this Destination Vector will minimize its toxicity.

Option 4: While the sequencing is going on, you might wish to check the expression of the nucleic acid molecule of interest in, for example, CHO cells, by recombining the nucleic acid molecule of interest into a CMV promoter vector (pDEST7, Figure 27; or pDEST12, Figure 32), or into a baculovirus vector (pDEST8, Figure 28; or pDEST10, Figure 30) for expression in insect cells. Both

of these vectors will transcribe the coding sequence of your nucleic acid molecule of interest, and translate it from the ATG of the PCR product using the Kozak bases upstream of the *Xmn*I site.

Option 5: If you wish to purify protein, for example to make antibodies, you can clone the nucleic acid molecule of interest into a His6 fusion vector, pDEST2 (Figure 22). Since the nucleic acid molecule of interest is cloned downstream of the TEV protease cleavage domain of pENTR7 (Figure 16), the amino acid sequence of the protein produced will be:

[----- attB1 -----] TEV protease
NH2- MSYYHHHHHHGITSLYKKAGFENLYFQI GTM---COOH

The attB site and the restriction sites used to make the Destination and Entry Vectors are translated into the underlined 11 amino acids (GITSLYKKAGF). Cleavage with TEV protease (arrow) leaves two amino acids, GT, on the amino end of the gene product.

See Figure 55 for an example of a nucleic acid molecule of interest, the chloramphenicol acetyl transferase (CAT) gene, cloned into pENTR7 (Figure 16) as a blunt (amino)-*Xho*I (carboxy) fragment, then cloned by recombination into the His6 fusion vector pDEST2 (Figure 22).

Option 6: If the His6 fusion protein is insoluble, you may go on and try a GST fusion. The appropriate Destination vector is pDEST3 (Figure 23).

Option 7: If you need to make RNA probes and prefer SP6 RNA polymerase, you can make the top strand RNA with your nucleic acid molecule of interest cloned into pSPORT+ (pDEST5 (Figures 25A, 25B)), and the bottom strand RNA with the nucleic acid molecule of interest cloned into pSPORT(-) (pDEST6 (Figures 26A, 26B)). Opposing promoters for T7 RNA polymerase and SP6 RNA polymerase are also present in these clones.

Option 8: It is often worthwhile to clone your nucleic acid molecule of interest into a variety of Destination Vectors in the same experiment. For example, if the number of colonies varies widely when the various recombination reactions are transformed into *E. coli*, this may be an indication that the nucleic acid molecule of interest is toxic in some contexts. (This problem is more clearly evident when a positive control gene is used for each Destination Vector.) Specifically, if many more colonies are obtained when the nucleic acid molecule of interest is recombined into pDEST6 than in pDEST5, there is a good chance that leakiness of the lac promoter is causing some expression of the nucleic acid molecule of interest in pSPORT "+" (which is not harmful in pDEST6 because the nucleic acid molecule of interest is in the opposite orientation).

Example 16: *Demonstration of a One-tube Transfer of a PCR Product (or Expression Clone) to Expression Clone via a Recombinational Cloning Reaction*

In the BxP recombination (Entry or Gateway) reaction described herein, a DNA segment flanked by attB1 and attB2 sites in a plasmid conferring ampicillin resistance was transferred by recombination into an attP plasmid conferring kanamycin resistance, which resulted in a product molecule wherein the DNA segment was flanked by attL sites (attL1 and attL2). This product plasmid comprises an "attL Entry Clone" molecule, because it can react with a "attR Destination Vector" molecule via the LxR (Destination) reaction, resulting in the transfer of the DNA segment to a new (ampicillin resistant) vector. In the previously described examples, it was necessary to transform the BxP reaction products into *E. coli*, select kanamycin resistant colonies, grow those colonies in liquid culture, and prepare miniprep DNA, before reacting this DNA with a Destination Vector in an LxR reaction.

The goal of the following experiment was to eliminate the transformation and miniprep DNA steps, by adding the BxP Reaction products directly to an LxR Reaction. This is especially appropriate when the DNA segment flanked by attB sites is a PCR product instead of a plasmid, because the PCR product cannot give

ampicillin-resistant colonies upon transformation, whereas attB plasmids (in general) carry an ampicillin resistance gene. Thus use of a PCR product flanked by attB sites in a BxP Reaction allows one to select for the ampicillin resistance encoded by the desired attB product of a subsequent LxR Reaction.

Two reactions were prepared: Reaction A, negative control, no attB PCR product, (8 µl) contained 50 ng pEZC7102 (attP Donor plasmid, confers kanamycin resistance) and 2 µl BxP Clonase (22 ng / µl Int protein and 8 ng/µl IHF protein) in BxP buffer (25 mM Tris HCl, pH 7.8, 70 mM KCl, 5 mM spermidine, 0.5 mM EDTA, 250 µg / ml BSA). Reaction B (24 µl) contained 150 ng pEZC7102, 6 µl BxP Clonase, and 120 ng of the attB -tet-PCR product in the same buffer as reaction A. The attB - tet - PCR product comprised the tetracycline resistance gene of plasmid pBR322, amplified with two primers containing either attB1 or attB2 sites, and having 4 Gs at their 5' ends, as described earlier.

The two reactions were incubated at 25°C for 30 minutes. Then aliquots of these reactions were added to new components that comprised LxR Reactions or appropriate controls for the LxR Reaction. Five new reactions were thus produced:

Reaction 1: 5 µl of reaction A was added to a 5 µl LxR Reaction containing 25 ng *Nco*I-cut pEZC8402 (the attR Destination Vector plasmid) in LxR buffer (37.5 mM Tris HCl, pH 7.7, 16.5 mM NaCl, 35 mM KCl, 5 mM spermidine, 375 µg / ml BSA), and 1 µl of GATEWAY™ LR Clonase™ Enzyme Mix (total volume of 10 µl).

Reaction 2: Same as reaction 1, except 5 µl of reaction B (positive) were added instead of reaction A (negative).

Reaction 3: Same as reaction 2, except that the amounts of *Nco*-cut pEZC8402 and GATEWAY™ LR Clonase™ Enzyme Mix were doubled, to 50 ng and 2 µl, respectively.

Reaction 4: Same as reaction 2, except that 25 ng of pEZ11104 (a positive control attL Entry Clone plasmid) were added in addition to the aliquot of reaction B.

Reaction 5: Positive control LxR Reaction, containing 25 ng *Nco*I-cut pEZC8402, 25 ng pEZ11104, 37.5 mM Tris HCl pH 7.7, 16.5 mM NaCl, 35 mM KCl, 5 mM spermidine, 375 µg / ml BSA and 1 µl GATEWAY™ LR Clonase™ Enzyme Mix in a total volume of 5 µl.

All five reactions were incubated at 25°C for 30 minutes. Then, 1 µl aliquots of each of the above five reactions, plus 1 µl from the remaining volume of Reaction B, the standard BxP Reaction, were used to transform 50 µl competent DH5α *E. coli*. DNA and cells were incubated on ice for 15 min., heat shocked at 42°C for 45 sec., and 450 µl SOC were added. Each tube was incubated with shaking at 37°C for 60 min. Aliquots of 100 µl and 400 µl of each transformation were plated on LB plates containing either 50 µg/ml kanamycin or 100 µg/ml ampicillin (see Table 2). A transformation with 10 pg of pUC19 DNA (plated on LB-amp₁₀₀) served as a control on the transformation efficiency of the DH5α cells. Following incubation overnight at 37°C, the number of colonies on each plate was determined.

Results of these reactions are shown in Table 2.

Table 2*

Reaction No.	1	2	3	4	5	6
	Number of Colonies					
Vol. plated:	Neg. Control BxP Reaction	1X pEZC8402 and LR Clonase™	2X pEZC8402 and LR Clonase™	LxR Reaction with Pos. Control DNA	LxR Reaction alone	BxP Reaction alone
100 µl	2	1	8	9	~1000	~1000
400 µl	5	10	35	62	>2000	>2000
Selection:	Kan	Amp	Amp	Amp	Amp	Kan

*(Transformation with pUC 19 DNA yielded 1.4×10^9 CFU/µg DNA.)

34 of the 43 colonies obtained from Reaction 3 were picked into 2 ml Terrific Broth with 100 µg/ml ampicillin and these cultures were grown overnight, with shaking, at 37°C. 27 of the 34 cultures gave at least moderate growth, and of these 24 were used to prepare miniprep DNA, using the standard protocol. These 24 DNAs were initially analyzed as supercoiled (SC) DNA on a 1% agarose gel to identify those with inserts and to estimate the sizes of the inserts. Fifteen of the 24 samples displayed SC DNA of the size predicted (5553 bp) if **tetx7102** had correctly recombined with **pEJC8402** to yield **tetx8402**. One of these samples contained two plasmids, one of ~5500 bp and a one of ~3500 bp. The majority of the remaining clones were approximately 4100 bp in size.

All 15 of the clones displaying SC DNA of predicted size (~5500 bp) were analyzed by two different double digests with restriction endonucleases to confirm the structure of the expected product: **tetx8402**. (See plasmid maps, Figures 57-59) In one set of digests, the DNAs were treated with *Not*I and *Eco*RI, which should cut the predicted product just outside both attB sites, releasing the tet^r insert on a fragment of 1475 bp. In the second set of digests, the DNAs were digested with *Not*I and with *Nru*I. *Nru*I cleaves asymmetrically within the subcloned tet^r insert, and together with *Not*I will release a fragment of 1019 bp.

Of the 15 clones analyzed by double restriction digestion, 14 revealed the predicted sizes of fragments for the expected product.

Interpretation:

The DNA components of Reaction B, **pEJC7102** and attB-tet-PCR, are shown in Figure 56. The desired product of BxP Reaction B is **tetx7102**, depicted in Figure 57. The LxR Reaction recombines the product of the BxP Reaction, **tetx7102** (Figure 57), with the Destination Vector, **pEJC8402**, shown in Figure 58. The LxR Reaction with **tetx7102** plus **pEJC8402** is predicted to yield the desired product **tetx8402**, shown in Figure 59.

Reaction 2, which combined the BxP Reaction and LxR Reaction, gave few colonies beyond those of the negative control Reaction. In contrast, Reaction 3, with twice the amount of **pEJC8402** (Figure 58) and LxR Clonase, yielded a

larger number of colonies. These colonies were analyzed further, by restriction digestion, to confirm the presence of expected product. Reaction 4 included a known amount of attL Entry Clone plasmid in the combined BxP-plus-LxR reaction. But reaction 4 yielded only about 1% of the colonies obtained when the same DNA was used in a LxR reaction alone, Reaction 6. This result suggests that the LxR reaction may be inhibited by components of the BxP reaction.

Restriction endonuclease analysis of the products of Reaction 3 revealed that a sizeable proportion of the colonies (14 of the 34 analyzed) contained the desired tet^r subclone, tetx8402 (Figure 59).

The above results establish the feasibility of performing first a BxP recombination reaction followed by a LxR recombination reaction -- in the same tube -- simply by adding the appropriate buffer mix, recombination proteins, and DNAs to a completed BxP reaction. This method should prove useful as a faster method to convert attB-containing PCR products into different Expression Clones, eliminating the need to isolate first the intermediate attL-PCR insert subclones, before recombining these with Destination Vectors. This may prove especially valuable for automated applications of these reactions.

This same one-tube approach allows for the rapid transfer of nucleic acid molecules contained in attB plasmid clones into new functional vectors as well. As in the above examples, attL subclones generated in a BxP Reaction can be recombined directly with various Destination Vectors in a LxR reaction. The only additional requirement for using attB plasmids, instead of attB-containing PCR products, is that the Destination Vector(s) employed must contain a different selection marker from the one present on the attB plasmid itself and the attP vector.

Two alternative protocols for a one-tube reaction have also proven useful and somewhat more optimal than the conditions described above.

Alternative 1:

Reaction buffer contained 50 mM Tris-HCl (pH 7.5), 50 mM NaCl, 0.25 mM EDTA, 2.5 mM spermidine, and 200 µg/ml BSA. After a 16 (or 3) hour incubation of the PCR product (100 ng) + attP Donor plasmid (100 ng) +

GATEWAY™ BP Clonase™ Enzyme Mix + Destination Vector (100 ng), 2 µl of GATEWAY™ LR Clonase™ Enzyme Mix (per 10 µl reaction mix) was added and the mixture was incubated an additional 6 (or 2) hours at 25°C. Stop solution was then added as above and the mixture was incubated at 37°C as above and transformed by electroporation with 1 µl directly into electrocompetent host cells. Results of this series of experiments demonstrated that longer incubation times (16 hours vs. 3 hours for the BP Reaction, 6 hours vs. 2 hours for the LR Reaction) resulted in about twice as many colonies being obtained as for the shorter incubation times. With two independent genes, 10/10 colonies having the correct cloning patterns were obtained.

Alternative 2:

A standard BP Reaction under the reaction conditions described above for Alternative 1 was performed for 2 hours at 25°C. Following the BP Reaction, the following components were added to the reaction mixture in a total volume of 7 µl:

20 mM Tris-HCl, pH 7.5
100 mM NaCl
5 µg/ml Xis-His6
15% glycerol
~1000 ng of Destination Vector

The reaction mixture was then incubated for 2 hours at 25°C, and 2.5 µl of stop solution (containing 2 µg/ml proteinase K) was added and the mixture was incubated at 37°C for an additional 10 minutes. Chemically competent host cells were then transformed with 2 µl of the reaction mixture, or electrocompetent host cells (e.g., EMax DH10B cells; Life Technologies, Inc.) were electroporated with 2 µl of the reaction mixture per 25-40 µl of cells. Following transformation, mixtures were diluted with SOC, incubated at 37°C, and plated as described above on media selecting for the selection markers on the Destination Vector and the Entry clone (B x P reaction product). Analogous results to those described for Alternative 1 were obtained with these reaction conditions -- a higher level of colonies containing correctly recombined reaction products were observed.

Example 17: Demonstration of a One-tube Transfer of a PCR Product (or Expression Clone) to Expression Clone via a Recombinational Cloning Reaction

Single-tube transfer of PCR product DNA or Expression Clones into Expression Clones by recombinational cloning has also been accomplished using a procedure modified from that described in Example 16. This procedure is as follows:

- Perform a standard BP (Gateway) Reaction (see Examples 9 and 10) in 20 μ l volume at 25°C for 1 hour.

- After the incubation is over, take a 10 μ l aliquot from the 20 μ l total volume and add 1 μ l of Proteinase K (2 mg/ml) and incubate at 37°C for 10 minutes. This first aliquot can be used for transformation and gel assay of BP reaction analysis. Plate BP reaction transformation on LB plates with **Kanamycin** (50 μ g/ml).

- Add the following reagents to the remaining 10 μ l aliquot of the BP reaction:

- 1 μ l of 0.75 M NaCl

- 2 μ l of destination vector (150 ng/ μ l)

- 4 μ l of LR Clonase™ (after thawing and brief mixing)

- Mix all reagents well and incubate at 25°C for 3 hours. Stop the reaction at the end of incubation with 1.7 μ l of Proteinase K (2 mg/ml) and incubate at 37°C for 10 minutes.

- Transform 2 μ l of the completed reaction into 100 μ l of competent cells. Plate 100 μ l and 400 μ l on LB plates with **Ampicilin** (100 μ g/ml).

Notes:

- If your competent cells are less than 10⁸ CFU/ μ g, and you are concerned about getting enough colonies, you can improve the yield several fold by incubating the

BP reaction for 6-20 hours. Electroporation also can yield better colony output than chemical transformation.

•PCR products greater than about 5-6 kb show significantly lower cloning efficiency in the BP reaction. In this case, we recommend using longer incubation times for both BP and LR steps.

•If you want to move your insert gene into several destination vectors simultaneously, then scale up the initial BP reaction volume so that you have a 10 µl aliquot for adding each destination vector.

Example 18: Optimization of GATEWAY™ Clonase™ Enzyme Compositions

The enzyme compositions containing Int and IHF (for BP Reactions) were optimized using a standard functional recombinational cloning reaction (a BP reaction) between attB-containing plasmids and attP-containing plasmids, according to the following protocol:

Materials and Methods:

Substrates:

AttP - supercoiled pDONR201

AttB - linear ~ 1Kb [³H]PCR product amplified from pEZC7501

Proteins:

IntH6 -- His₆-carboxy- tagged λ Integrase

IHF -- Integration Host Factor

Clonase:

50 ng/µl IntH6 and 20 ng/µl IHF, admixed in 25 mM Tris- HCl (pH 7.5), 22 mM NaCl, 5 mM EDTA, 1 mg/ml BSA, 5 mM Spermidine, and 50% glycerol.

Reaction Mixture (total volume of 40 μ l):

1000 ng AttP plasmid

600 ng AttB [3 H] PCR product

8 μ l Clonase (400 ng IntH6, 160 ng IHF) in 25 mM Tris-HCl (pH 7.5),
22 mM NaCl, 5 mM EDTA, 1 mg/ml BSA, 5 mM Spermidine, 5 mM
DTT.

Reaction mixture was incubated for 1 hour at 25°C, 4 μ l of 2 μ g/ μ l
proteinase K was added and mixture was incubated for an additional 20 minutes
at 37°C. Mixture was then extracted with an equal volume of Phenol/Chloroform/
Isoamyl alcohol. The aqueous layer was then collected, and 0.1 volumes of 3 M
sodium acetate and 2 volumes of cold 100% ethanol were added. Tubes were
then spun in a microcentrifuge at maximum RPM for 10 minutes at room
temperature. Ethanol was decanted, and pellets were rinsed with 70% ethanol and
re-centrifuged as above. Ethanol was decanted, and pellets were allowed to air
dry for 5-10 minutes and then dissolved in 20 μ l of 33 mM Tris-Acetate (pH 7.8),
66 mM potassium acetate, 10 mM magnesium acetate, 1 mM DTT, and 1mM
ATP. 2 units of exonuclease V (e.g., Plasmid Safe; EpiCentre, Inc., Madison, WI)
was then added, and the mixture was incubated at 37°C for 30 minutes.

Samples were then TCA-washed by spotting 30 μ l of reaction mixture
onto a Whatman GF/C filter, washing filters once with 10% TCA + 1% NaPPi for
10 minutes, three times with 5% TCA for 5 minutes each, and twice with ethanol
for 5 minutes each. Filters were then dried under a heat lamp, placed into a
scintillation vial, and counted on a β liquid scintillation counter (LSC).

The principle behind this assay is that, after exonuclease V digestion, only
double-stranded circular DNA survives in an acid-insoluble form. All DNA
substrates and products that have free ends are digested to an acid-soluble form
and are not retained on the filters. Therefore, only the 3 H-labeled attB linear DNA
which ends up in circular form after both inter- and intramolecular integration is
complete is resistant to digestion and is recovered as acid-insoluble product.
Optimal enzyme and buffer formulations in the Clonase compositions therefore are
those that give the highest levels of circularized 3 H-labeled attB-containing

sequences, as determined by highest cpm in the LSC. Although this assay was designed for optimization of GATEWAY™ BP Clonase™ Enzyme Mix compositions (Int + IHF), the same type of assay may be performed to optimize GATEWAY™ LR Clonase™ Enzyme Mix compositions (Int + IHF + Xis), except that the reaction mixtures would comprise 1000 ng of AttR (instead of AttP) and 600 ng of AttL (instead of AttB), and 40 ng of His₆-carboxy- tagged Xis (XisH6) in addition to the IntH6 and IHF.

Example 19: Testing Functionality of Entry and Destination Vectors

As part of assessment of the functionality of particular vectors of the invention, it is important to functionally test the ability of the vectors to recombine. This assessment can be carried out by performing a recombinational cloning reaction (as schematized in Figures 2, 4, and 5A and 5B, and as described herein and in commonly owned U.S. Application Nos. 08/486,139, filed June 7, 1995, 08/663,002, filed June 7, 1996 (now U.S. Patent No. 5,888,732), 09/005,476, filed January 12, 1998, and 09/177,387, filed October 23, 1998, the disclosures of all of which are incorporated by reference herein in their entireties), by transforming E. coli and scoring colony forming units. However, an alternative assay may also be performed to allow faster, more simple assessment of the functionality of a given Entry or Destination Vector by agarose gel electrophoresis. The following is a description of such an in vitro assay.

Materials and Methods:

Plasmid templates pEZC1301 (Figure 84) and pEZC1313 (Figure 85), each containing a single wild type att site, were used for the generation of PCR products containing attL or attR sites, respectively. Plasmid templates were linearized with *A*laNI, phenol extracted, ethanol precipitated and dissolved in TE to a concentration of 1 ng/μl.

PCR primers (capital letters represent base changes from wildtype):

attL1 gggg agcct gctttttGtacAaa gttggcatta taaaaaagca ttgc
attL2 gggg agcct gctttCttGtacAaa gttggcatta taaaaaagca ttgc
attL right tgttgccggg aagctagagt aa

attR1 gggg Acaag ttTgtaCaaaaaagc tgaacgaga aacgtaaaat
attR2 gggg Acaag ttTgtaCaaGaaagc tgaacgaga aacgtaaaat
attR right ca gacggcatga tgaacctgaa

PCR primers were dissolved in TE to a concentration of 500 pmol/ μ l. Primer mixes were prepared, consisting of attL1 + attLright primers, attL2 + attLright primers, attR1 + attRright primers, and attR2 + attRright primers, each mix containing 20 pmol/ μ l of each primer.

PCR reactions:

1 μ l plasmid template (1 ng)
1 μ l primer pairs (20 pmoles of each)
3 μ l of H₂O
45 μ l of Platinum PCR SuperMix® (Life Technologies, Inc.)

Cycling conditions (performed in MJ thermocycler):

95°C/2 minutes
94°C/30 seconds
25 cycles of 58°C/30 seconds and 72°C/1.5 minutes
72°C/5 minutes
5°C/hold

The resulting attL PCR product was 1.5 kb, and the resulting attR PCR product was 1.0 kb.

PCR reactions were PEG/MgCl₂ precipitated by adding 150 μ l H₂O and 100 μ l of 3x PEG/ MgCl₂ solution followed by centrifugation. The PCR products were dissolved in 50 μ l of TE. Quantification of the PCR product was performed by gel electrophoresis of 1 μ l and was estimated to be 50-100 ng/ μ l.

Recombination reactions of PCR products containing attL or attR sites with GATEWAY™ plasmids was performed as follows:

8 µl of H₂O

2 µl of attL or attR PCR product (100-200 ng)

2 µl of GATEWAY™ plasmid (100 ng)

4 µl of 5x Destination buffer

4 µl of GATEWAY™ LR Clonase™ Enzyme Mix

20 µl total volume (the reactions can be scaled down to a 5 µl total volume by adjusting the volumes of the components to about ¼ of those shown above, while keeping the stoichiometries the same).

Clonase reactions were incubated at 25°C for 2 hours. 2 µl of proteinase K (2 mg/ml) was added to stop the reaction. 10 µl was then run on a 1 % agarose gel. Positive control reactions were performed by reacting attL1 PCR product (1.0 kb) with attR1 PCR product (1.5 kb) and by similarly reacting attL2 PCR product with attR2 PCR product to observe the formation of a larger (2.5 kb) recombination product. Negative controls were similarly performed by reacting attL1 PCR product with attR2 PCR product and vice versa or reactions of attL PCR product with an attL plasmid, etc.

In alternative assays, to test attB Entry vectors, plasmids containing single attP sites were used. Plasmids containing single att sites could also be used as recombination substrates in general to test all Entry and Destination vectors (*i.e.*, those containing attL, attR, attB and attP sites). This would eliminate the need to do PCR reactions.

Results:

Destination and Entry plasmids when reacted with appropriate att-containing PCR products formed linear recombinant molecules that could be easily visualized on an agarose gel when compared to control reactions containing no attL or attR PCR product. Thus, the functionality of Destination and Entry vectors constructed according to the invention may be determined either by carrying out the Destination or Entry recombination reactions as depicted in

Figures 2, 4, and 5A and 5B, or more rapidly by carrying out the linearization assay described in this Example.

Example 20: PCR Cloning Using Universal Adapter-Primers

As described herein, the cloning of PCR products using the GATEWAY™ PCR Cloning System (Life Technologies, Inc.; Rockville, MD) requires the addition of attB sites (attB1 and attB2) to the ends of gene-specific primers used in the PCR reaction. The protocols described in the preceding Examples suggest that the user add 29 bp (25 bp containing the attB site plus four G residues) to the gene-specific primer. It would be advantageous to high volume users of the GATEWAY™ PCR Cloning System to generate attB-containing PCR product using universal attB adapter-primers in combination with shorter gene-specific primers containing a specified overlap to the adapters. The following experiments demonstrate the utility of this strategy using universal attB adapter-primers and gene-specific primers containing overlaps of various lengths from 6 bp to 18 bp. The results demonstrate that gene-specific primers with overlaps of 10 bp to 18 bp can be used successfully in PCR amplifications with universal attB adapter-primers to generate full-length PCR products. These PCR products can then be successfully cloned with high fidelity in a specified orientation using the GATEWAY™ PCR Cloning System.

Methods and Results:

To demonstrate that universal attB adapter-primers can be used with gene-specific primers containing partial attB sites in PCR reactions to generate full-length PCR product, a small 256 bp region of the human hemoglobin cDNA was chosen as a target so that intermediate sized products could be distinguished from full-length products by agarose gel electrophoresis.

The following oligonucleotides were used:

B1-Hgb: GGGG ACA AGT TTG TAC AAA AAA GCA GGC T-5' -Hgb*
B2-Hgb: GGGG ACC ACT TTG TAC AAG AAA GCT GGG T-3' -Hgb**

-145-

5

10

15

20

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18B1-Hgb: TG TAC AAA AAA GCA GGC T-5'-Hgb
 18B2-Hgb: TG TAC AAG AAA GCT GGG T-3'-Hgb
 15B1-Hgb: AC AAA AAA GCA GGC T-5'-Hgb
 15B2-Hgb: AC AAG AAA GCT GGG T-3'-Hgb
 12B1-Hgb: AA AAA GCA GGC T-5'-Hgb
 12B2-Hgb: AG AAA GCT GGG T-3'-Hgb
 11B1-Hgb: A AAA GCA GGC T-5'-Hgb
 11B2-Hgb: G AAA GCT GGG T-3'-Hgb
 10B1-Hgb: AAA GCA GGC T-5'-Hgb
 10B2-Hgb: AAA GCT GGG T-3'-Hgb
 9B1-Hgb: AA GCA GGC T-5'-Hgb
 9B2-Hgb: AA GCT GGG T-3'-Hgb
 8B1-Hgb: A GCA GGC T-5'-Hgb
 8B2-Hgb: A GCT GGG T-3'-Hgb
 7B1-Hgb: GCA GGC T-5'-Hgb
 7B2-Hgb: GCT GGG T-3'-Hgb
 6B1-Hgb: CA GGC T-5'-Hgb
 6B2-Hgb: CT GGG T-3'-Hgb

attB1 adapter: GGGG ACA AGT TTG TAC AAA AAA GCA GGC T
 attB2 adapter: GGGG ACC ACT TTG TAC AAG AAA GCT GGG T

* -5'-Hgb = GTC ACT AGC CTG TGG AGC AAG A
 ** -3'-Hgb = AGG ATG GCA GAG GGA GAC GAC A

The aim of these experiments was to develop a simple and efficient universal adapter PCR method to generate attB containing PCR products suitable for use in the GATEWAY™ PCR Cloning System. The reaction mixtures and thermocycling conditions should be simple and efficient so that the universal adapter PCR method could be routinely applicable to any PCR product cloning application.

PCR reaction conditions were initially found that could successfully amplify predominately full-length PCR product using gene-specific primers containing 18bp and 15 bp overlap with universal attB primers. These conditions are outlined below:

10 pmoles of gene-specific primers
10 pmoles of universal attB adapter-primers
1 ng of plasmid containing the human hemoglobin cDNA.
100 ng of human leukocyte cDNA library DNA.
5 µl of 10x PLATINUM Taq HiFi® reaction buffer (Life Technologies, Inc.)
2 µl of 50 mM MgSO₄
1 µl of 10 mM dNTPs
0.2 µl of PLATINUM Taq HiFi® (1.0 unit)
H₂O to 50 µl total reaction volume

Cycling conditions:

25 x

95°C/5 min
94°C/15 sec
50°C/30 sec
68°C/1 min
68°C/5 min
5°C/hold

To assess the efficiency of the method, 2 µl (1/25) of the 50 µl PCR reaction was electrophoresed in a 3 % Agarose-1000 gel. With overlaps of 12 bp or less, smaller intermediate products containing one or no universal attB adapter predominated the reactions. Further optimization of PCR reaction conditions was obtained by titrating the amounts of gene-specific primers and universal attB adapter-primers. The PCR reactions were set up as outlined above except that the amounts of primers added were:

0, 1, 3 or 10 pmoles of gene-specific primers
0, 10, 30 or 100 pmoles of adapter-primers

Cycling conditions:

25 x $\left\{ \begin{array}{l} 95^{\circ}\text{C}/3 \text{ min} \\ 94^{\circ}\text{C}/15 \text{ sec} \\ 50^{\circ}\text{C}/45 \text{ sec} \\ 68^{\circ}\text{C}/1 \text{ min} \\ 68^{\circ}\text{C}/5 \text{ min} \\ 5^{\circ}\text{C}/\text{hold} \end{array} \right.$

The use of limiting amounts of gene-specific primers (3 pmoles) and excess adapter-primers (30 pmoles) reduced the amounts of smaller intermediate products. Using these reaction conditions the overlap necessary to obtain predominately full-length PCR product was reduced to 12 bp. The amounts of gene-specific and adapter-primers was further optimized in the following PCR reactions:

0, 1, 2 or 3 pmoles of gene-specific primers
0, 30, 40 or 50 pmoles of adapter-primers

Cycling conditions:

25 x $\left\{ \begin{array}{l} 95^{\circ}\text{C}/3 \text{ min} \\ 94^{\circ}\text{C}/15 \text{ sec} \\ 48^{\circ}\text{C}/1 \text{ min} \\ 68^{\circ}\text{C}/1 \text{ min} \\ 68^{\circ}\text{C}/5 \text{ min} \\ 5^{\circ}\text{C}/\text{hold} \end{array} \right.$

The use of 2 pmoles of gene-specific primers and 40 pmoles of adapter-primers further reduced the amounts of intermediate products and generated predominately full-length PCR products with gene-specific primers containing an 11 bp overlap. The success of the PCR reactions can be assessed in any PCR application by performing a no adapter control. The use of limiting amounts of gene-specific primers should give faint or barely visible bands when 1/25 to 1/10 of the PCR reaction is electrophoresed on a standard agarose gel. Addition of the

universal attB adapter-primers should generate a robust PCR reaction with a much higher overall yield of product.

PCR products from reactions using the 18 bp, 15 bp, 12 bp, 11 bp and 10 bp overlap gene-specific primers were purified using the CONCERT® Rapid PCR Purification System (PCR products greater than 500 bp can be PEG precipitated). The purified PCR products were subsequently cloned into an attP containing plasmid vector using the GATEWAY™ PCR Cloning System (Life Technologies, Inc.; Rockville, MD) and transformed into *E. coli*. Colonies were selected and counted on the appropriate antibiotic media and screened by PCR for correct inserts and orientation.

Raw PCR products (unpurified) from the attB adapter PCR of a plasmid clone of part of the human beta-globin (Hgb) gene were also used in GATEWAY™ PCR Cloning System reactions. PCR products generated with the full attB B1/B2-Hgb, the 12B1/B2, 11B1/B2 and 10B1/B2 attB overlap Hgb primers were successfully cloned into the GATEWAY™ pENTR21 attP vector (Figure 49). 24 colonies from each (24 x 4 = 96 total) were tested and each was verified by PCR to contain correct inserts. The cloning efficiency expressed as cfu/ml is shown below:

Primer Used	cfu/ml
Hgb full attB	8,700
Hgb 12 bp overlap	21,000
Hgb 11 bp overlap	20,500
Hgb 10 bp overlap	13,500
GFP control	1,300

Interestingly, the overlap PCR products cloned with higher efficiency than did the full attB PCR product. Presumably, and as verified by visualization on agarose gel, the adapter PCR products were slightly cleaner than was the full attB PCR product. The differences in colony output may also reflect the proportion of PCR product molecules with intact attB sites.

Using the attB adapter PCR method, PCR primers with 12 bp attB overlaps were used to amplify cDNAs of different sizes (ranging from 1 to 4 kb)

from a leukocyte cDNA library and from first strand cDNA prepared from HeLa total RNA. While three of the four cDNAs were able to be amplified by this method, a non-specific amplification product was also observed that under some conditions would interfere with the gene-specific amplification. This non-specific product was amplified in reactions containing the attB adapter-primers alone without any gene-specific overlap primers present. The non-specific amplification product was reduced by increasing the stringency of the PCR reaction and lowering the attB adapter PCR primer concentration.

These results indicate that the adapter-primer PCR approach described in this Example will work well for cloned genes. These results also demonstrate the development of a simple and efficient method to amplify PCR products that are compatible with the GATEWAY™ PCR Cloning System that allows the use of shorter gene-specific primers that partially overlap universal attB adapter-primers. In routine PCR cloning applications, the use of 12 bp overlaps is recommended. The methods described in this Example can thus reduce the length of gene-specific primers by up to 17 residues or more, resulting in a significant savings in oligonucleotide costs for high volume users of the GATEWAY™ PCR Cloning System. In addition, using the methods and assays described in this Example, one of ordinary skill can, using only routine experimentation, design and use analogous primer-adapters based on or containing other recombination sites or fragments thereof, such as attL', attR, attP, lox, FRT, etc.

Example 21: Mutational Analysis of the Bacteriophage Lambda attL and attR Sites: Determinants of att Site Specificity in Site-specific Recombination

To investigate the determinants of att site specificity, the bacteriophage lambda attL and attR sites were systematically mutagenized. As noted herein, the determinants of specificity have previously been localized to the 7 bp overlap region (TTTATAC, which is defined by the cut sites for the integrase protein and is the region where strand exchange takes place) within the 15 bp core region (GCTTTTATATACTAA) which is identical in all four lambda att sites, attB, attP, attL and attR. This core region, however, has not heretofore been systematically

mutagenized and examined to define precisely which mutations produce unique changes in *att* site specificity.

Therefore, to examine the effect of *att* sequence on site specificity, mutant *attL* and *attR* sites were generated by PCR and tested in an *in vitro* site-specific recombination assay. In this way all possible single base pair changes within the 7 bp overlap region of the core *att* site were generated as well as five additional changes outside the 7 bp overlap but within the 15 bp core *att* site. Each *attL* PCR substrate was tested in the *in vitro* recombination assay with each of the *attR* PCR substrates.

Methods

To examine both the efficiency and specificity of recombination of mutant *attL* and *attR* sites, a simple *in vitro* site-specific recombination assay was developed. Since the core regions of *attL* and *attR* lie near the ends of these sites, it was possible to incorporate the desired nucleotide base changes within PCR primers and generate a series of PCR products containing mutant *attL* and *attR* sites. PCR products containing *attL* and *attR* sites were used as substrates in an *in vitro* reaction with GATEWAY™ LR Clonase™ Enzyme Mix (Life Technologies, Inc.; Rockville, MD). Recombination between a 1.5 kb *attL* PCR product and a 1.0 kb *attR* PCR product resulted in a 2.5 kb recombinant molecule that was monitored using agarose gel electrophoresis and ethidium bromide staining.

Plasmid templates pEZC1301 (Figure 84) and pEZC1313 (Figure 85), each containing a single wild type *attL* or *attR* site, respectively, were used for the generation of recombination substrates. The following list shows primers that were used in PCR reactions to generate the *attL* PCR products that were used as substrates in L x R Clonase reactions (capital letters represent changes from the wild-type sequence, and the underline represents the 7 bp overlap region within the 15 bp core *att* site; a similar set of PCR primers was used to prepare the *attR* PCR products containing matching mutations):

GATEWAY™ sites (note: attL2 sequence in GATEWAY™ plasmids begins "accga" while the attL2 site in this example begins "agcct" to reflect wild-type attL outside the core region.):

attL1: gggg agcct gcttttttGtacAaa gttggcatta taaaaa-
agca ttgc

attL2: gggg agcct gcttttCttGtacAaa gttggcatta taaaaa-
agca ttgc

Wild-type:

attL0: gggg agcct gctttttttataactaa gttggcatta taaaaa-
agca ttgc

Single base changes from wild-type:

attLT1A: gggg agcct gcttttAttataactaa gttggcatta taaaaa-
agca ttgc

attLT1C: gggg agcct gcttttCttataactaa gttggcatta taaaaa-
agca ttgc

attLT1G: gggg agcct gcttttGttataactaa gttggcatta taaaaa-
agca ttgc

attLT2A: gggg agcct gcttttAtataactaa gttggcatta taaaaa-
agca ttgc

attLT2C: gggg agcct gcttttCtataactaa gttggcatta taaaaa-
agca ttgc

attLT2G: gggg agcct gcttttGtataactaa gttggcatta taaaaa-
aagca ttgc

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attLT3A: gggg agcct gctttttAataactaa gttggcatta taaaa-
aagca ttgc

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attLT3C: gggg agcct gctttttCataactaa gttggcatta taaaa-
aagca ttgc

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attLT3G: gggg agcct gctttttGataactaa gttggcatta taaaa-
aagca ttgc

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attLA4C: gggg agcct gcttttttCtactaa gttggcatta taaaa-
aagca ttgc

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attLA4G: gggg agcct gcttttttGtactaa gttggcatta taaaa-
aagca ttgc

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attLA4T: gggg agcct gcttttttTtactaa gttggcatta taaaa-
aagca ttgc

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attLT5A: gggg agcct gcttttttaAactaa gttggcatta taaaa-
aagca ttgc

attLT5C: gggg agcct gcttttttaCactaa gttggcatta taaaa-
aagca ttgc

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attLT5G: gggg agcct gcttttttaGactaa gttggcatta taaaa-
aagca ttgc

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attLA6C: gggg agcct gcttttttatCctaa gttggcatta taaaa-
aagca ttgc

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attLA6G: gggg agcct gctttttttatGctaa gttggcatta taaaa-
aagca ttgc

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5 attLA6T: gggg agcct gctttttttatTctaa gttggcatta taaaa-
aagca ttgc

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10 attLC7A: gggg agcct gctttttttataAataa gttggcatta taaaa-
aagca ttgc

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attLC7G: gggg agcct gctttttttataGtaa gttggcatta taaaa-
aagca ttgc

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attLC7T: gggg agcct gctttttttataTtaa gttggcatta taaaa-
aagca ttgc

Single base changes outside of the 7 bp overlap:

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20 attL8: gggg agcct Actttttttataactaa gttggcatta taaaa-
aagca ttgc

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25 attL9: gggg agcct gcCttttttataactaa gttggcatta taaaaa-
agca ttgc

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attL10: gggg agcct gcttCttttataactaa gttggcatta taaaaa-
agca ttgc

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attL14: gggg agcct gctttttttataacCaa gttggcatta taaaaa-
agca ttgc

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35 attL15: gggg agcct gctttttttataactaG gttggcatta taaaaa-
agca ttgc

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Note: additional vectors wherein the first nine bases are gggg agcca (*i.e.*, substituting an adenine for the thymine in the position immediately preceding the 15-bp core region), which may or may not contain the single base pair substitutions (or deletions) outlined above, can also be used in these experiments.

Recombination reactions of *attL*- and *attR*-containing PCR products was performed as follows:

8 µl of H₂O
2 µl of attL PCR product (100 ng)
2 µl of attR PCR product (100 ng)
4 µl of 5x buffer
4 µl of GATEWAY™ LR Clonase™ Enzyme Mix
20 µl total volume

Clonase reactions were incubated at 25°C for 2 hours.

2 µl of 10X Clonase stop solution (proteinase K, 2 mg/ml) were added to stop the reaction.

10 µl were run on a 1 % agarose gel.

Results

Each *attL* PCR substrate was tested in the *in vitro* recombination assay with each of the *attR* PCR substrates. Changes within the first three positions of the 7 bp overlap (TTTATAC) strongly altered the specificity of recombination. These mutant *att* sites each recombined as well as the wild-type, but only with their cognate partner mutant; they did not recombine detectably with any other *att* site mutant. In contrast, changes in the last four positions (TTTATAC) only partially altered specificity; these mutants recombined with their cognate mutant as well as wild-type *att* sites and recombined partially with all other mutant *att* sites except for those having mutations in the first three positions of the 7 bp

overlap. Changes outside of the 7 bp overlap were found not to affect specificity of recombination, but some did influence the efficiency of recombination.

Based on these results, the following rules for *att* site specificity were determined:

- Only changes within the 7 bp overlap affect specificity.
- Changes within the first 3 positions strongly affect specificity.
- Changes within the last 4 positions weakly affect specificity.

Mutations that affected the overall efficiency of the recombination reaction were also assessed by this method. In these experiments, a slightly increased (less than 2-fold) recombination efficiency with *attLT1A* and *attLC7T* substrates was observed when these substrates were reacted with their cognate *attR* partners. Also observed were mutations that decreased recombination efficiency (approximately 2-3 fold), including *attLA6G*, *attL14* and *attL15*. These mutations presumably reflect changes that affect Int protein binding at the core *att* site.

The results of these experiments demonstrate that changes within the first three positions of the 7 bp overlap (TTTATAC) strongly altered the specificity of recombination (*i.e.*, *att* sequences with one or more mutations in the first three thymidines would only recombine with their cognate partners and would not cross-react with any other *att* site mutation). In contrast, mutations in the last four positions (TTTATAC) only partially altered specificity (*i.e.*, *att* sequences with one or more mutations in the last four base positions would cross-react partially with the wild-type *att* site and all other mutant *att* sites, except for those having mutations in one or more of the first three positions of the 7 bp overlap). Mutations outside of the 7 bp overlap were not found to affect specificity of recombination, but some were found to influence (*i.e.*, to cause a decrease in) the efficiency of recombination.

Example 22: Discovery of Att Site Mutations That Increase the Cloning Efficiency of GATEWAY™ Cloning Reactions

In experiments designed to understand the determinants of *att* site specificity, point mutations in the core region of *attL* were made. Nucleic acid molecules containing these mutated *attL* sequences were then reacted in an LR

reaction with nucleic acid molecules containing the cognate *attR* site (*i.e.*, an *attR* site containing a mutation corresponding to that in the *attL* site), and recombinational efficiency was determined as described above. Several mutations located in the core region of the *att* site were noted that either slightly increased (less than 2-fold) or decreased (between 2-4-fold) the efficiency of the recombination reaction (Table 3).

Table 3. *Effects of attL mutations on Recombination Reactions.*

Site	Sequence	Effect on Recombination
attL0	agcctgcttttttataactaagttggcatta	
attL5	agcctgctttAttataactaagttggcatta	slightly increased
attL6	agcctgcttttttataTtaagttggcatta	slightly increased
attL13	agcctgctttttttatGctaagttggcatta	decreased
attL14	agcctgctttttttatacCaagttggcatta	decreased
attL15	agcctgctttttttatactaGgttggcatta	decreased
consensus	CAACTTnnTnnnAnnAAGTTG	

It was also noted that these mutations presumably reflected changes that either increased or decreased, respectively, the relative affinity of the integrase protein for binding the core *att* site. A consensus sequence for an integrase core-binding site (CAACTTNNNT) has been inferred in the literature but not directly tested (see, *e.g.*, Ross and Landy, *Cell* 33:261-272 (1983)). This consensus core integrase-binding sequence was established by comparing the sequences of each of the four core *att* sites found in *attP* and *attB* as well as the sequences of five non-*att* sites that resemble the core sequence and to which integrase has been shown to bind *in vitro*. These experiments suggest that many more *att* site mutations might be identified which increase the binding of integrase to the core *att* site and thus increase the efficiency of GATEWAY™ cloning reactions.

Example 23: Effects of Core Region Mutations on Recombination Efficiency

To directly compare the cloning efficiency of mutations in the att site core region, single base changes were made in the attB2 site of an attB1-TET-attB2 PCR product. Nucleic acid molecules containing these mutated *attB2* sequences were then reacted in a BP reaction with nucleic acid molecules containing non-cognate *attP* sites (*i.e.*, wildtype *attP2*), and recombinational efficiency was determined as described above. The cloning efficiency of these mutant attB2 containing PCR products compared to standard attB1-TET-attB2 PCR product are shown in Table 4.

Table 4. Efficiency of Recombination With Mutated attB2 Sites.

<u>Site</u>	<u>Sequence</u>	<u>Mutation</u>	<u>Cloning Efficiency</u>
attB0	tcaagttagtataaaaaagcaggct		
attB1	ggggacaagttgtacaaaaagcaggct		
attB2	ggggaccactttgtacaagaagctgggt		100%
attB2.1	ggggaAcactttgtacaagaagctgggt	C→A	40%
attB2.2	ggggacAactttgtacaagaagctgggt	C→A	131%
attB2.3	ggggaccCctttgtacaagaagctgggt	A→C	4%
attB2.4	ggggaccaAttgtacaagaagctgggt	C→A	11%
attB2.5	ggggaccacGttgtacaagaagctgggt	T→G	4%
attB2.6	ggggaccactGgtacaagaagctgggt	T→G	6%
attB2.7	ggggaccactGgtacaagaagctgggt	T→G	1%
attB2.8	ggggaccacttTtacaagaagctgggt	G→T	0.5%

As noted above, a single base change in the attB2.2 site increased the cloning efficiency of the attB1-TET-attB2.2 PCR product to 131% compared to the attB1-TET-attB2 PCR product. Interestingly, this mutation changes the integrase core binding site of attB2 to a sequence that matches more closely the proposed consensus sequence.

Additional experiments were performed to directly compare the cloning efficiency of an attB1-TET-attB2 PCR product with a PCR product that contained attB sites containing the proposed consensus sequence (see Example 22) of an integrase core binding site. The following attB sites were used to amplify attB-TET PCR products:

attB1 ggggacaagtttgtacaaaaaagcaggct
attB1.6 ggggacaaCtttgtacaaaaaagTTggct
attB2 ggggaccactttgtacaagaaagctgggt
attB2.10 ggggacAactttgtacaagaaagTtgggt

BP reactions were carried out between 300 ng (100 fmoles) of pDONR201 (Figure 49A) with 80 ng (80 fmoles) of attB-TET PCR product in a 20 µl volume with incubation for 1.5 hrs at 25°C, creating pENTR201-TET Entry clones. A comparison of the cloning efficiencies of the above-noted attB sites in BP reactions is shown in Table 5.

Table 5. Cloning efficiency of BP Reactions.

PCR product	CFU/ml	Fold Increase
B1-tet-B2	7,500	
B1.6-tet-B2	12,000	1.6 x
B1-tet-B2.10	20,900	2.8 x
B1.6-tet-B2.10	30,100	4.0 x

These results demonstrate that attB PCR products containing sequences that perfectly match the proposed consensus sequence for integrase core binding sites can produce Entry clones with four-fold higher efficiency than standard Gateway attB1 and attB2 PCR products.

The entry clones produced above were then transferred to pDEST20 (Figure 40A) via LR reactions (300 ng (64 fmoles) pDEST20 mixed with 50 ng (77 fmoles) of the respective pENTR201-TET Entry clone in 20 µl volume; incubated for 1 hr incubation at 25°C). The efficiencies of cloning for these reactions are compared in Table 6.

Table 6. Cloning Efficiency of LR Reactions.

pENTR201-TET x pDEST20	CFU/ml	Fold Increase
L1-tet-L2	5,800	
L1.6-tet-L2	8,000	1.4
L1-tet-L2.10	10,000	1.7
L1.6-tet-L2.10	9,300	1.6

These results demonstrate that the mutations introduced into attB1.6 and attB2.10 that transfer with the gene into entry clones slightly increase the efficiency of LR reactions. Thus, the present invention encompasses not only mutations in attB sites that increase recombination efficiency, but also to the corresponding mutations that result in the attL sites created by the BP reaction.

To examine the increased cloning efficiency of the attB1.6-TET-attB2.10 PCR product over a range of PCR product amounts, experiments analogous to those described above were performed in which the amount of attB PCR product was titrated into the reaction mixture. The results are shown in Table 7.

Table 7. Titration of attB PCR products.

Amount of attB PCR product (ng)	PCR product	CFU/ml	Fold Increase
20	attB1-TET-attB2	3,500	6.1
	attB1.6-TET-attB2.10	21,500	
50	attB1-TET-attB2	9,800	5.0
	attB1.6-TET-attB2.10	49,000	
100	attB1-TET-attB2	18,800	2.8
	attB1.6-TET-attB2.10	53,000	
200	attB1-TET-attB2	19,000	2.5
	attB1.6-TET-attB2.10	48,000	

These results demonstrate that as much as a six-fold increase in cloning efficiency is achieved with the attB1.6-TET-attB2.10 PCR product as compared to the standard attB1-TET-attB2 PCR product at the 20 ng amount.

Example 24: Determination of attB Sequence Requirements for Optimum Recombination Efficiency

To examine the sequence requirements for attB and to determine which attB sites would clone with the highest efficiency from populations of degenerate attB sites, a series of experiments was performed. Degenerate PCR primers were designed which contained five bases of degeneracy in the B-arm of the attB site. These degenerate sequences would thus transfer with the gene into Entry clone in BP reactions and subsequently be transferred with the gene into expression clones in LR reactions. The populations of degenerate attB and attL sites could thus be cycled from attB to attL back and forth for any number of cycles. By altering the reaction conditions at each transfer step (for example by decreasing the reaction time and/or decreasing the concentration of DNA) the reaction can be made increasingly more stringent at each cycle and thus enrich for populations of attB and attL sites that react more efficiently.

The following degenerate PCR primers were used to amplify a 500 bp fragment from pUC18 which contained the lacZ alpha fragment (only the attB portion of each primer is shown):

attB1	GGGG ACAAGTTTGTACAAA AAAGC AGGCT
attB1n16-20	GGGG ACAAGTTTGTACAAA nnnnn AGGCT
attB1n21-25	GGGG ACAAGTTTGTACAAA AAAGC nnnnn
attB2	GGGG ACCACTTTGTACAAG AAAGC TGGGT
attB2n16-20	GGGG ACCACTTTGTACAAG nnnnn TGGGT
attB2n21-25	GGGG ACCACTTTGTACAAG AAAGC nnnnn

The starting population size of degenerate att sites is 4^5 or 1024 molecules. Four different populations were transferred through two BP reactions and two LR reactions. Following transformation of each reaction, the population of transformants was amplified by growth in liquid media containing the appropriate selection antibiotic. DNA was prepared from the population of clones by alkaline

lysis miniprep and used in the next reaction. The results of the BP and LR cloning reactions are shown below.

BP-1, overnight reactions

	cfu/ml	percent of control
attB1-LacZa-attB2	78,500	100 %
attB1n16-20-LacZa-attB2	1,140	1.5 %
attB1n21-25-LacZa-attB2	11,100	14 %
attB1-LacZa-attB2n16-20	710	0.9 %
attB1-LacZa-attB2n21-25	16,600	21 %

LR-1, pENTR201-LacZa x pDEST20/*EcoRI*, 1hr reactions

	cfu/ml	percent of control
attL1-LacZa-attL2	20,000	100 %
attL1n16-20-LacZa-attL2	2,125	11 %
attL1n21-25-LacZa-attL2	2,920	15 %
attL1-LacZa-attL2n16-20	3,190	16 %
attL1-LacZa-attL2n21-25	1,405	7 %

BP-2, pEXP20-LacZa/*ScaI* x pDONR 201, 1hr reactions

	cfu/ml	percent of control
attB1-LacZa-attB2	48,600	100 %
attB1n16-20-LacZa-attB2	22,800	47 %
attB1n21-25-LacZa-attB2	31,500	65 %
attB1-LacZa-attB2n16-20	42,400	87 %
attB1-LacZa-attB2n21-25	34,500	71 %

LR-2, pENTR201-LacZa x pDEST6/*NcoI*, 1hr reactions

	cfu/ml	percent of control
attL1-LacZa-attL2	23,000	100 %
attL1n16-20-LacZa-attL2	49,000	213 %
attL1n21-25-LacZa-attL2	18,000	80 %
attL1-LacZa-attL2n16-20	37,000	160 %
attL1-LacZa-attL2n21-25	57,000	250 %

These results demonstrate that at each successive transfer, the cloning efficiency of the entire population of att sites increases, and that there is a great deal of flexibility in the definition of an attB site. Specific clones may be isolated from the above reactions, tested individually for recombination efficiency, and

sequenced. Such new specificities may then be compared to known examples to guide the design of new sequences with new recombination specificities. In addition, based on the enrichment and screening protocols described herein, one of ordinary skill can easily identify and use sequences in other recombination sites, e.g., other *att* sites, *lox*, FRT, etc., that result in increased specificity in the recombination reactions using nucleic acid molecules containing such sequences.

Example 25: Design of att Site PCR Adapter-Primers

Additional studies were performed to design gene-specific primers with 12bp of attB1 and attB2 at their 5'-ends. The optimal primer design for *att*-containing primers is the same as for any PCR primers: the gene-specific portion of the primers should ideally have a T_m of $> 50^\circ\text{C}$ at 50 mM salt (calculation of T_m is based on the formula $59.9 + 41(\%GC) - 675/n$).

Primers:

12bp attB1: AA AAA GCA GGC TNN - forward gene-specific primer

12bp attB2: A GAA AGC TGG GTN - reverse gene-specific primer

attB1 adapter primer: GGGGACAAGTTTGTACAAAAAAGCAGGCT

attB2 adapter primer: GGGGACCACTTTGTACAAGAAAGCTGGGT

Protocol:

(1) Mix 200 ng of cDNA library or 1 ng of plasmid clone DNA (alternatively, genomic DNA or RNA could be used) with 10 pmoles of gene specific primers in a 50 μl PCR reaction, using one or more polypeptides having DNA polymerase activity such as those described herein. (The addition of greater than 10 pmoles of gene-specific primers can decrease the yield of attB PCR product. In addition, if RNA is used, a standard reverse transcriptase-PCR (RT-

PCR) protocol should be followed; *see, e.g.*, Gerard, G.F., *et al.*, *FOCUS* 11:60 (1989); Myers, T.W., and Gelfand, D.H., *Biochem.* 30:7661 (1991); Freeman, W.N., *et al.*, *BioTechniques* 20:782 (1996); and U.S. Application No. 09/064,057, filed April 22, 1998, the disclosures of all of which are incorporated herein by reference.)

1st PCR profile:

(a) 95°C for 3 minutes

(b) 10 cycles of:

(i) 94°C for 15 seconds

(ii) 50°C* for 30 seconds

(iii) 68°C for 1 minute/kb of target amplicon

(c) 68°C for 5 minutes

(d) 10°C hold

*The optimal annealing temperature is determined by the calculated T_m of the gene-specific part of the primer.

(2) Transfer 10 µl to a 40 µl PCR reaction mix containing 35 pmoles each of the attB1 and attB2 adapter primers.

2nd PCR profile:

(a) 95°C for 1 minute

(b) 5 cycles of:

(i) 94°C for 15 seconds

(ii) 45°C* for 30 seconds

(iii) 68°C for 1 minute/kb of target amplicon

(c) 15-20 cycles** of:

(i) 94°C for 15 seconds

(ii) 55°C* for 30 seconds

(iii) 68°C for 1 minute/kb of target amplicon

(d) 68°C for 5 minutes

(e) 10°C hold

*The optimal annealing temperature is determined by the calculated T_m of the gene-specific part of the primer.

**15 cycles is sufficient for low complexity targets.

Notes:

1. It is useful to perform a no-adaptor primer control to assess the yield of attB PCR product produced.
2. Linearized template usually results in slightly greater yield of PCR product.

Example 26: One-Tube Recombinational Cloning Using the GATEWAY™ Cloning System

To provide for easier and more rapid cloning using the GATEWAY™ cloning system, we have designed a protocol whereby the BP and LR reactions may be performed in a single tube (a "one-tube" protocol). The following is an example of such a one-tube protocol; in this example, an aliquot of the BP reaction is taken before adding the LR components, but the BP and LR reactions may be performed in a one-tube protocol without first taking the BP aliquot:

<u>Reaction Component</u>	<u>Volume</u>
attB DNA (100-200 ng/25 µl reaction)	1-12.5 µl
attP DNA (pDONR201) 150 ng/µl	2.5 µl
5X BP Reaction Buffer	5.0 µl
Tris-EDTA	(to 20 µl)
<u>BP Clonase</u>	<u>5.0 µl</u>
Total vol.	25 µl

After the above components were mixed in a single tube, the reaction mixtures were incubated for 4 hours at 25°C. A 5 µl aliquot of reaction mixture was removed, and 0.5 µl of 10X stop solution was added to this reaction mixture and incubated for 10 minutes at 37°C. Competent cells were then transformed with 1-2 µl of the BP reaction per 100 µl of cells; this transformation yielded colonies of Entry Clones for isolation of individual Entry Clones and for quantitation of the BP Reaction efficiency.

To the remaining 20 µl of BP reaction mixture, the following components of the LR reaction were added:

<u>Reaction Component</u>	<u>Final Concentration</u>	<u>Volume Added</u>
NaCl	0.75 M	1 µl
Destination Vector	150 ng/ul	3 µl
<u>LR Clonase</u>		<u>6 µl</u>
Total vol.		30 µl

After the above components were mixed in a single tube, the reaction mixtures were incubated for 2 hours at 25°C. 3 µl of 10X stop solution was added, and the mixture was incubated for 10 minutes at 37°C. Competent cells were then transformed with 1-2 µl of the reaction mixture per 100 µl of cells

Notes:

1. If desired, the Destination Vector can be added to the initial BP reaction.
2. The reactions can be scaled down by 2x, if desired.
3. Shorter incubation times for the BP and/or LR reactions can be used (scaled to the desired cloning efficiencies of the reaction), but a lower number of colonies will typically result.
4. To increase the number of colonies obtained by several fold, incubate the BP reaction for 6-20 hours and increase the LR reaction to 3 hours. Electroporation also works well with 1-2 ul of the PK-treated reaction mixture.

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- 10
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5. PCR products greater than about 5 kb may show significantly lower cloning efficiency in the BP reaction. In this case, we recommend using a one-tube reaction with longer incubation times (e.g., 6-18 hours) for both the BP and LR steps.

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Example 27: Relaxation of Destination Vectors During the LR Reaction

To further optimize the LR Reaction, the composition of the LR Reaction buffer was modified from that described above and this modified buffer was used in a protocol to examine the impact of enzymatic relaxation of Destination Vectors during the LR Reaction.

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LR Reactions were set up as usual (see, e.g., Example 6), except that 5X BP Reaction Buffer (see Example 5) was used for the LR Reaction. To accomplish Destination Vector relaxation during the LR Reaction, Topoisomerase I (Life Technologies, Inc., Rockville, MD; Catalogue No. 38042-016) was added to the reaction mixture at a final concentration of ~15U per µg of total DNA in the reaction (for example, for reaction mixtures with a total of 400ng DNA in the 20 µl LR Reaction, ~6units of Topoisomerase I was added). Reaction mixtures were set up as follows:

<u>Reaction Component</u>	<u>Volume</u>
ddH ₂ O	6.5 µl
4X BP Reaction Buffer	5 µl
100ng single chain/linear pENTR CAT, 50 ng/µl	2 µl
300ng single chain/linear pDEST6, 150ng/µl	2 µl
Topoisomerase I, 15 U/ml	0.5 µl
LR Clonase	4 µl

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Reaction mixtures were incubated at 25°C for 1 hour, and 2 µl of 2 µg/µl Proteinase K was then added and mixtures incubated for 10 minutes at 37°C to stop the LR Reaction. Competent cells were then transformed as described in the preceding examples. The results of these studies demonstrated that relaxation of

substrates in the LR reaction using Topoisomerase I resulted in a 2- to 10-fold increase in colony output compared to those LR reactions performed without including Topoisomerase I.

Having now fully described the present invention in some detail by way of illustration and example for purposes of clarity of understanding, it will be obvious to one of ordinary skill in the art that the same can be performed by modifying or changing the invention within a wide and equivalent range of conditions, formulations and other parameters without affecting the scope of the invention or any specific embodiment thereof, and that such modifications or changes are intended to be encompassed within the scope of the appended claims.

All publications, patents and patent applications mentioned in this specification are indicative of the level of skill of those skilled in the art to which this invention pertains, and are herein incorporated by reference to the same extent as if each individual publication, patent or patent application was specifically and individually indicated to be incorporated by reference.

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INDICATIONS RELATING TO DEPOSITED MICROORGANISM
OR OTHER BIOLOGICAL MATERIAL
(PCT Rule 13bis)

REC'D 17 APR 2000

WIPO PCT

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A. The indications made below relate to the microorganism referred to in the description on page 52, line 31.

20

B. IDENTIFICATION OF DEPOSIT

Further deposits are identified on an additional sheet ☒

Name of depositary institution
Agricultural Research Culture Collection (NRRL)
International Depository Authority

Address of depositary institution (including postal code and country)

1815 N. University Street
Peoria, Illinois 61604
United States of America

25

Date of deposit
February 27, 1999

Accession Number
NRRL B-30099

C. ADDITIONAL INDICATIONS (leave blank if not applicable)

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30

Escherichia coli DB3.1(pAHPKan) or Escherichia coli DB3.1(pAnPKan)

35

D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)

40

E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)

The indications listed below will be submitted to the international Bureau later (specify the general nature of the indications, e.g., "Accession Number of Deposit")

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**INDICATIONS RELATING TO DEPOSITED MICROORGANISM
OR OTHER BIOLOGICAL MATERIAL**
(PCT Rule 13bis)

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A. The indications made below relate to the microorganism referred to in the description on page 16, line 16.		REC'D 1 1 2000 WIPO PCT
B. IDENTIFICATION OF DEPOSIT		Further deposits are identified on an additional sheet <input checked="" type="checkbox"/>
Name of depositary institution Agricultural Research Culture Collection (NRRL) International Depository Authority		
Address of depositary institution (including postal code and country) 1815 N. University Street Peoria, Illinois 61604 United States of America		
Date of deposit February 27, 1999	Accession Number NRRL B-30101	
C. ADDITIONAL INDICATIONS (leave blank if not applicable)		This information is continued on an additional sheet <input type="checkbox"/>
Escherichia coli DB3.1(pENTR-2B)		
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)		
E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)		
The indications listed below will be submitted to the international Bureau later (specify the general nature of the indications, e.g., "Accession Number of Deposit")		

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167.4

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INDICATIONS RELATING TO DEPOSITED MICROORGANISM
OR OTHER BIOLOGICAL MATERIAL
(PCT Rule 13bis)

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A. The indications made below relate to the microorganism referred to in the description on page <u>55</u> , line <u>16</u> .	
B. IDENTIFICATION OF DEPOSIT Further deposits are identified on an additional sheet <input checked="" type="checkbox"/>	
Name of depositary institution Agricultural Research Culture Collection (NRRL) International Depository Authority	
Address of depositary institution (including postal code and country) 1815 N. University Street Peoria, Illinois 61604 United States of America	
Date of deposit February 27, 1999	Accession Number NRRL B-30102
C. ADDITIONAL INDICATIONS (leave blank if not applicable) This information is continued on an additional sheet <input type="checkbox"/>	
Escherichia coli DB3.1(pENTR-3C)	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)	
E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable) The indications listed below will be submitted to the international Bureau later (specify the general nature of the indications, e.g., "Accession Number of Deposit")	

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INDICATIONS RELATING TO DEPOSITED MICROORGANISM
OR OTHER BIOLOGICAL MATERIAL
(PCT Rule 13bis)

A. The indications made below relate to the microorganism referred to in the description on page 8.	REC'D 547 APR 2000 WIPO PCT
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B. IDENTIFICATION OF DEPOSIT Further deposits are identified on an additional sheet ☒

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Agricultural Research Culture Collection (NRRL)
International Depository Authority

Address of depositary institution (including postal code and country)

1815 N. University Street
Peoria, Illinois 61604
United States of America

Date of deposit
February 27, 1999

Accession Number
NRRL B-30103

C. ADDITIONAL INDICATIONS (leave blank if not applicable) This information is continued on an additional sheet ☐

Escherichia coli DB3.1(pEZC15101)

D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)

E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)

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INDICATIONS RELATING TO DEPOSITED MICROORGANISM
OR OTHER BIOLOGICAL MATERIAL (PCT Rule 13bis)

REC'D 17

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A. The indications made below relate to the microorganism referred to in the description on page 54, line 9.

B. IDENTIFICATION OF DEPOSIT

Further deposits are identified on an additional sheet ☒

Name of depositary institution
Agricultural Research Culture Collection (NRRL)
International Depository Authority

Address of depositary institution (including postal code and country)

1815 N. University Street
Peoria, Illinois 61604
United States of America

Date of deposit
February 27, 1999

Accession Number
NRRL B-30104

C. ADDITIONAL INDICATIONS (leave blank if not applicable)

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Escherichia coli DB3.1(pEZC15102)

D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)

E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)

The indications listed below will be submitted to the international Bureau later (specify the general nature of the indications, e.g., "Accession Number of Deposit")

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Authorized officer Diana Harris	Authorized officer

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Applicant's or agent's file reference number	0942.468PC03	International application No. PCT/US	00/05432
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**INDICATIONS RELATING TO DEPOSITED MICROORGANISM
OR OTHER BIOLOGICAL MATERIAL**
(PCT Rule 13bis)

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A. The indications made below relate to the microorganism referred to in the description on page 54, line 9.

B. IDENTIFICATION OF DEPOSIT

Further deposits are identified on an additional sheet ☒

Name of depositary institution
Agricultural Research Culture Collection (NRRL)
International Depository Authority

Address of depositary institution (including postal code and country)

1815 N. University Street
Peoria, Illinois 61604
United States of America

Date of deposit
February 27, 1999

Accession Number
NRRL B-30105

C. ADDITIONAL INDICATIONS (leave blank if not applicable)

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Escherichia coli DB3.1(pEZC15103)

D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)

E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)

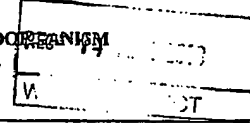
The indications listed below will be submitted to the international Bureau later (specify the general nature of the indications, e.g., "Accession Number of Deposit")

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167.8

Applicant's or agent's file reference number	0942.408PC03	International application No. tl PCT/US 00/05432
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INDICATIONS RELATING TO DEPOSITED MICROORGANISM
OR OTHER BIOLOGICAL MATERIAL
(PCT Rule 13bis)



A. The indications made below relate to the microorganism referred to in the description on page 51, line 20-21

B. IDENTIFICATION OF DEPOSIT

Further deposits are identified on an additional sheet ☒

Name of depositary institution
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International Depository Authority

Address of depositary institution (including postal code and country)

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United States of America

Date of deposit
February 27, 1999

Accession Number
NRRL B-30108

C. ADDITIONAL INDICATIONS (leave blank if not applicable)

This information is continued on an additional sheet ☐

Escherichia coli DB10B(pCMVSPORT6)

D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)

E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)

The indications listed below will be submitted to the international Bureau later (specify the general nature of the indications, e.g., "Accession Number of Deposit")

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Wanda Fitch

Authorized officer

Claims

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WHAT IS CLAIMED IS:

1. An isolated nucleic acid molecule comprising a nucleotide sequence selected from the group of nucleotide sequences consisting of an attB1 nucleotide sequence as set forth in Figure 9, an attB2 nucleotide sequence as set forth in Figure 9, an attP1 nucleotide sequence as set forth in Figure 9, an attP2 nucleotide sequence as set forth in Figure 9, an attL1 nucleotide sequence as set forth in Figure 9, an attL2 nucleotide sequence as set forth in Figure 9, an attR1 nucleotide sequence as set forth in Figure 9, an attR2 nucleotide sequence as set forth in Figure 9, a polynucleotide complementary thereto, and a mutant, fragment, or derivative thereof.

2. An isolated nucleic acid molecule comprising an attB1 nucleotide sequence as set forth in Figure 9, a polynucleotide complementary thereto, or a mutant, fragment, variant or derivative thereof.

3. An isolated nucleic acid molecule comprising an attB2 nucleotide sequence as set forth in Figure 9, a polynucleotide complementary thereto, or a mutant, fragment, variant or derivative thereof.

4. An isolated nucleic acid molecule comprising an attP1 nucleotide sequence as set forth in Figure 9, a polynucleotide complementary thereto, or a mutant, fragment, variant or derivative thereof.

5. An isolated nucleic acid molecule comprising an attP2 nucleotide sequence as set forth in Figure 9, a polynucleotide complementary thereto, or a mutant, fragment, variant or derivative thereof.

6. An isolated nucleic acid molecule comprising an attL1 nucleotide sequence as set forth in Figure 9, a polynucleotide complementary thereto, or a mutant, fragment, variant or derivative thereof.

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7. An isolated nucleic acid molecule comprising an attL2 nucleotide sequence as set forth in Figure 9, a polynucleotide complementary thereto, or a mutant, fragment, variant or derivative thereof.

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8. An isolated nucleic acid molecule comprising an attR1 nucleotide sequence as set forth in Figure 9, a polynucleotide complementary thereto, or a mutant, fragment, variant or derivative thereof.

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9. An isolated nucleic acid molecule comprising an attR2 nucleotide sequence as set forth in Figure 9, a polynucleotide complementary thereto, or a mutant, fragment, variant or derivative thereof.

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10. The isolated nucleic acid molecule of claim 1, further comprising one or more functional or structural nucleotide sequences selected from the group consisting of one or more multiple cloning sites, one or more localization signals, one or more transcription termination sites, one or more transcriptional regulatory sequences, one or more translational signals, one or more origins of replication, one or more fusion partner peptide-encoding nucleic acid molecules, one or more protease cleavage sites, and one or more 5' polynucleotide extensions.

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11. The nucleic acid molecule of claim 10, wherein said transcriptional regulatory sequence is a promoter, an enhancer, or a repressor.

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12. The nucleic acid molecule of claim 10, wherein said fusion partner peptide-encoding nucleic acid molecule encodes glutathione S-transferase (GST), hexahistidine (His₆), or thioredoxin (Trx).

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13. The nucleic acid molecule of claim 10, wherein said 5' polynucleotide extension consists of from one to five nucleotide bases.

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14. The nucleic acid molecule of claim 13, wherein said 5' polynucleotide extension consists of four or five guanine nucleotide bases.

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15. A primer nucleic acid molecule suitable for amplifying a target nucleotide sequence, comprising the isolated nucleic acid molecule of claim 1 or a portion thereof linked to a target-specific nucleotide sequence useful in amplifying said target nucleotide sequence.

16. The primer nucleic acid molecule of claim 15, wherein said primer comprises an attB1 nucleotide sequence having the sequence shown in Figure 9 or a portion thereof, or a polynucleotide complementary to the sequence shown in Figure 9 or a portion thereof.

17. The primer nucleic acid molecule of claim 15, wherein said primer comprises an attB2 nucleotide sequence having the sequence shown in Figure 9 or a portion thereof, or a polynucleotide complementary to the sequence shown in Figure 9 or a portion thereof.

18. The primer nucleic acid molecule of claim 15, further comprising a 5' terminal extension of four or five guanine bases.

19. A vector comprising the isolated nucleic acid molecule of claim 1.

20. The vector of claim 19, wherein said vector is an Expression Vector.

21. A host cell comprising the isolated nucleic acid molecule of claim 1 or the vector of claim 19.

22. A method of synthesizing or amplifying one or more nucleic acid molecules comprising:

- (a) mixing one or more nucleic acid templates with at least one polypeptide having polymerase or reverse transcriptase activity and at least a first primer comprising a template-specific sequence that is complementary to or capable of hybridizing to said

5 templates and at least a second primer comprising all or a portion
of a recombination site wherein said at least a portion of said
10 second primer is homologous to or complementary to at least a
portion of said first primer; and

- 5 (b) incubating said mixture under conditions sufficient to synthesize or
amplify one or more nucleic acid molecules complementary to all
15 or a portion of said templates and comprising one or more
recombination sites or portions thereof at one or both termini of
said molecules.

20 23. A method of synthesizing or amplifying one or more nucleic acid
molecules comprising:

- 25 (a) mixing one or more nucleic acid templates with at least one
polypeptide having polymerase or reverse transcriptase activity
15 and at least a first primer comprising a template-specific sequence
that is complementary to or capable of hybridizing to said
30 templates and at least a portion of a recombination site, and at
least a second primer comprising all or a portion of a
20 recombination site wherein said at least a portion of said
35 recombination site on said second primer is complementary to or
homologous to at least a portion of said recombination site on said
first primer; and

- 40 (b) incubating said mixture under conditions sufficient to synthesize or
25 amplify one or more nucleic acid molecules complementary to all
or a portion of said templates and comprising one or more
recombination sites or portions thereof at one or both termini of
said molecules.

45 24. A method of amplifying or synthesizing one or more nucleic acid
30 molecules comprising:

- 50 (a) mixing one or more nucleic acid templates with at least one
polypeptide having polymerase or reverse transcriptase activity

5 and one or more first primers comprising at least a portion of a recombination site and a template-specific sequence that is complementary to or capable of hybridizing to said template;

10 (b) incubating said mixture under conditions sufficient to synthesize or amplify one or more first nucleic acid molecules complementary to all or a portion of said templates wherein said molecules comprise at least a portion of a recombination site at one or both termini of said molecules;

15 (c) mixing said molecules with one or more second primers comprising one or more recombination sites, wherein said recombination sites of said second primers are homologous to or complementary to at least a portion of said recombination sites on said first nucleic acid molecules; and

20 (d) incubating said mixture under conditions sufficient to synthesize or amplify one or more second nucleic acid molecules complementary to all or a portion of said first nucleic acid molecules and which comprise one or more recombination sites at one or both termini of said molecules.

25 25. A polypeptide encoded by the isolated nucleic acid molecule of any one of claims 1-10.

30 26. An isolated nucleic acid molecule comprising one or more *att* recombination sites comprising at least one mutation in its core region that increases the specificity of interaction between said recombination site and a second *att* recombination site.

35 27. The isolated nucleic acid molecule of claim 26, wherein said mutation is at least one substitution mutation of at least one nucleotide in the seven basepair overlap region of said core region of said recombination site.

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28. The isolated nucleic acid molecule of claim 26, wherein said nucleic acid molecule comprises the sequence NNNATAC, wherein "N" refers to any nucleotide with the proviso that if one of the first three nucleotides in the consensus sequence is a T/U, then at least one of the other two of the first three nucleotides is not a T/U.

29. An isolated nucleic acid molecule comprising one or more mutated *att* recombination sites comprising at least one mutation in its core region that enhances the efficiency of recombination between a first nucleic acid molecule comprising said mutated *att* recombination site and a second nucleic acid molecule comprising a second recombination site that interacts with said mutated *att* recombination site.

30. The isolated nucleic acid molecule of claim 29, wherein said mutated *att* recombination site is a mutated *attL* site comprising a core region having the nucleotide sequence caactnntnnnannaagttg, wherein "n" represents any nucleotide.

31. The isolated nucleic acid molecule of claim 30, wherein said mutated *attL* recombination site comprises a core region having a nucleotide sequence selected from agcctgctttattataactaagttggcatta (*attL5*) and agcctgctttttatattaagttggcatta (*attL6*).

32. The isolated nucleic acid molecule of claim 29, wherein said mutated *att* recombination site comprises a core region having a nucleotide sequence selected from the group consisting of ggggacaactttgtacaaaaagttggct (*attB1.6*), ggggacaactttgtacaagaaagctgggt (*attB2.2*), and ggggacaactttgtacaagaaagttgggt (*attB2.10*).

33. A vector selected from the group consisting of pENTR1A, pENTR2B, pENTR3C, pENTR4, pENTR5, pENTR6, pENTR7, pENTR8, pENTR9, pENTR10, pENTR11, pDEST1, pDEST2, pDEST3, pDEST4,

pDEST5, pDEST6, pDEST7, pDEST8, pDEST9, pDEST10, pDEST11, pDEST12.2 (also known as pDEST12), pDEST13, pDEST14, pDEST15, pDEST16, pDEST17, pDEST18, pDEST19, pDEST20, pDEST21, pDEST22, pDEST23, pDEST24, pDEST25, pDEST26, pDEST27, pDEST28, pDEST29, pDEST30, pDEST31, pDEST32, pDEST33, pDEST34, pDONR201 (also known as pENTR21 attP vector or pAttPkan Donor Vector), pDONR202, pDONR203 (also known as pEZ15812), pDONR204, pDONR205, pDONR206 (also known as pENTR22 attP vector or pAttPgen Donor Vector), pDONR207, pMAB58, pMAB62, pMAB85 and pMAB86.

34. A host cell comprising the vector of claim 33.

35. A polypeptide encoded by the vector of claim 33.

36. A kit for use in synthesizing a nucleic acid molecule, said kit comprising the isolated nucleic acid molecule of any one of claims 1-10, 26 and 29.

37. A kit for use in synthesizing a nucleic acid molecule, said kit comprising the primer of claim 15 or claim 18.

38. A kit for use in cloning a nucleic acid molecule, said kit comprising the vector of claim 19 or claim 33.

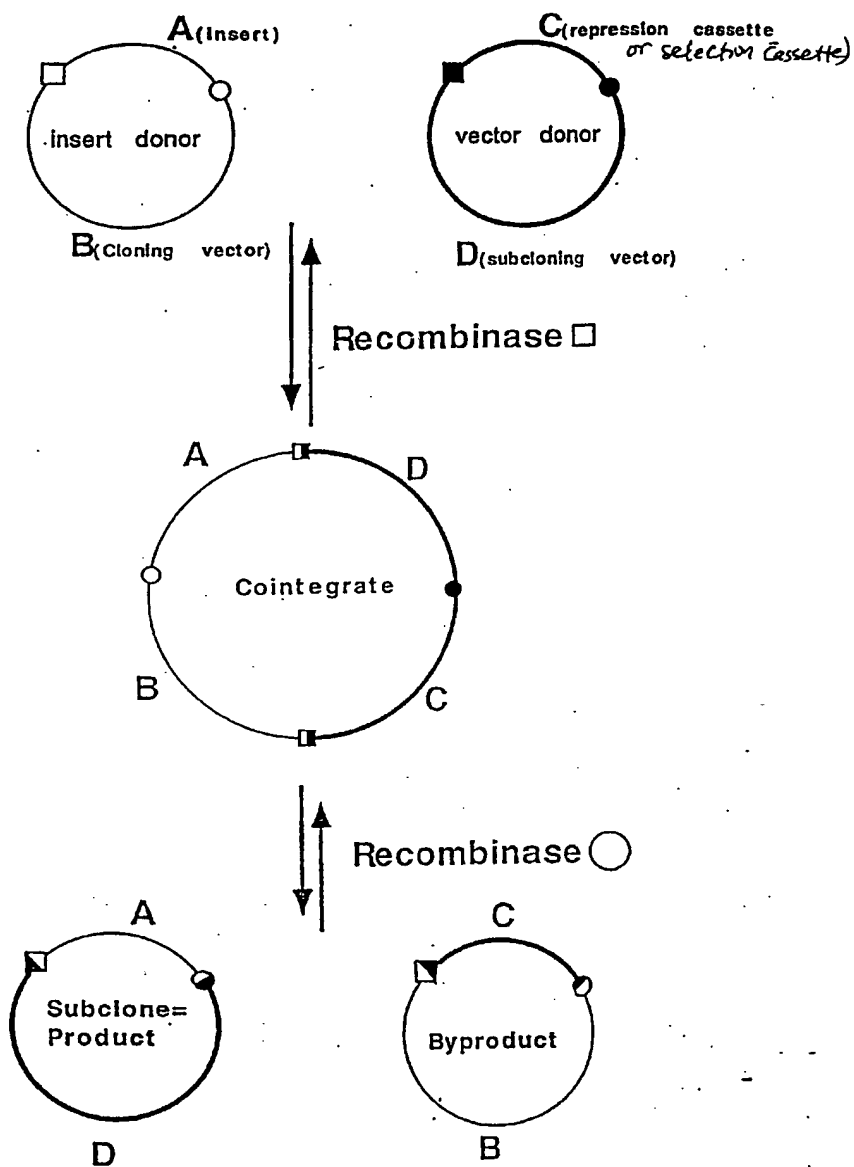


Figure 1

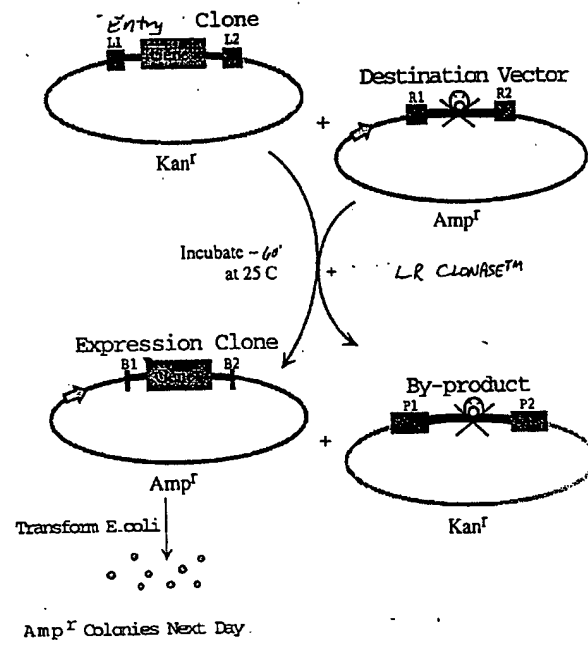


FIGURE 2

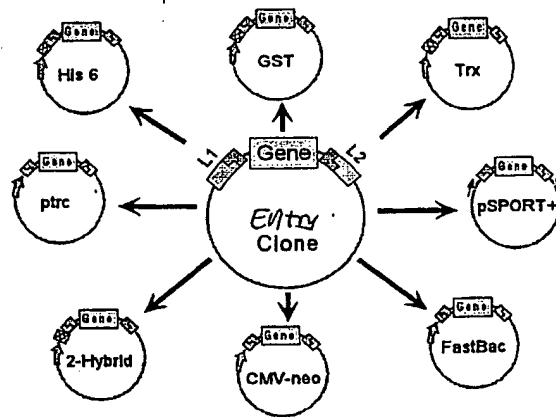


FIGURE 3

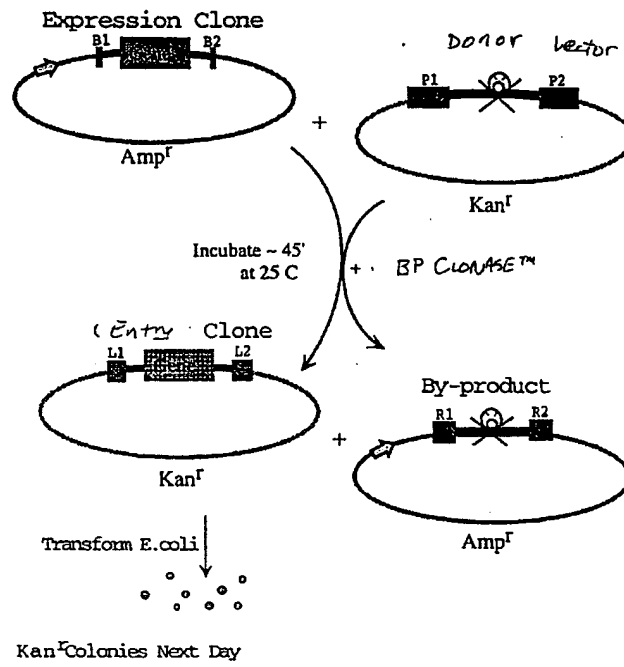


FIGURE 4

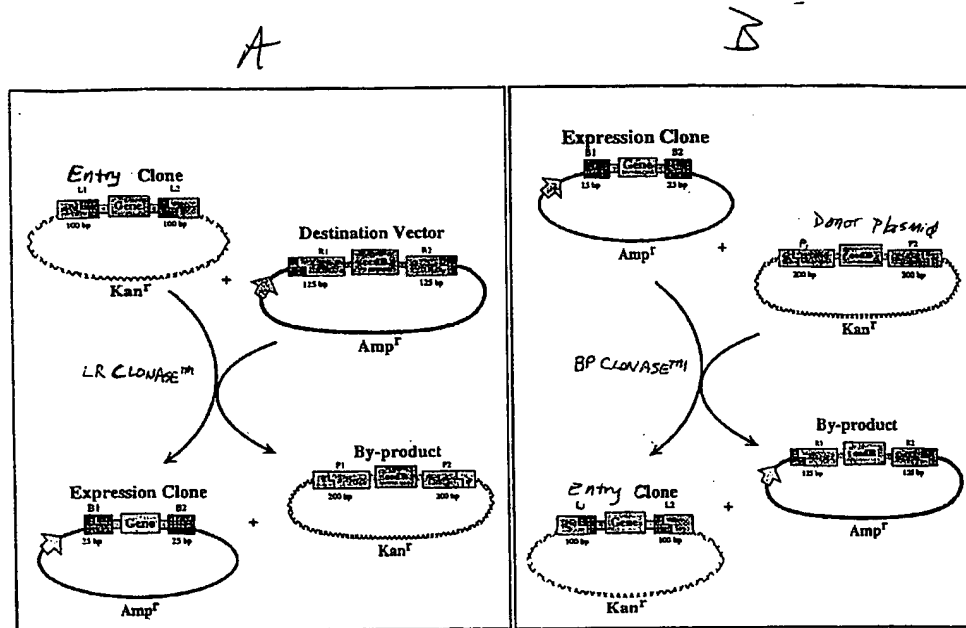


FIGURE 5

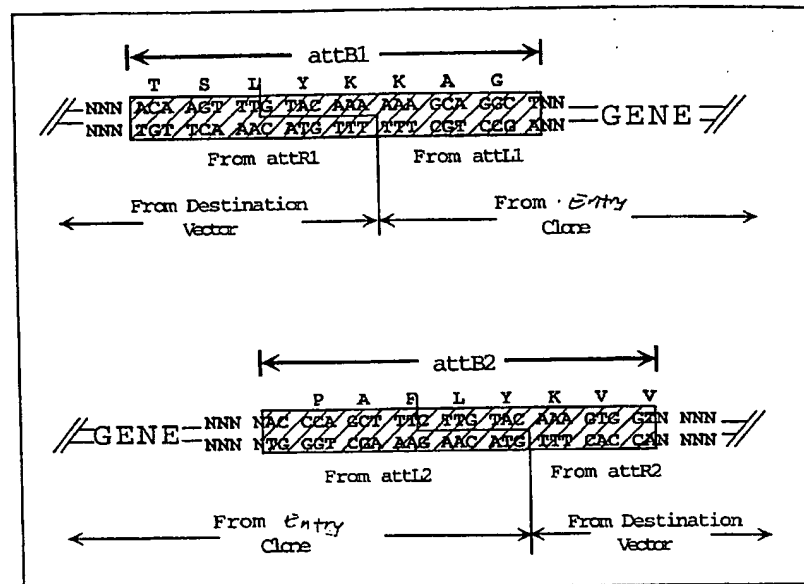


FIGURE 6

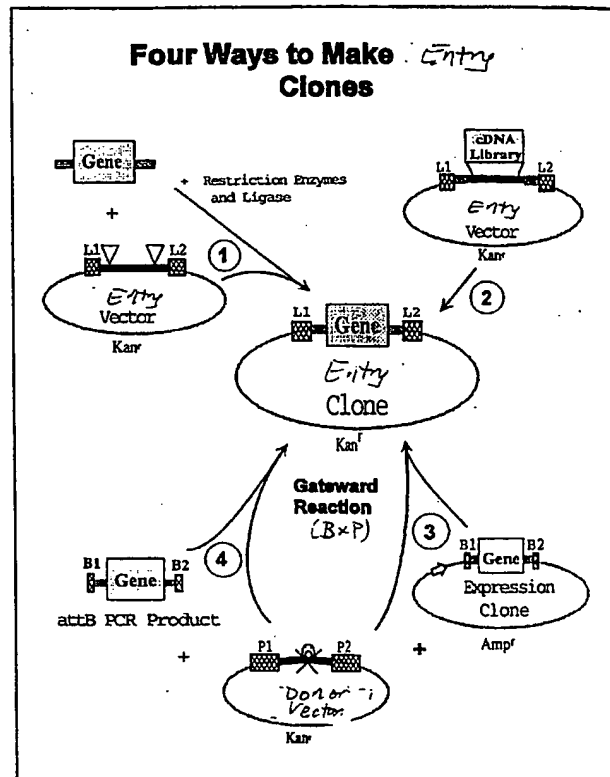


FIGURE 7

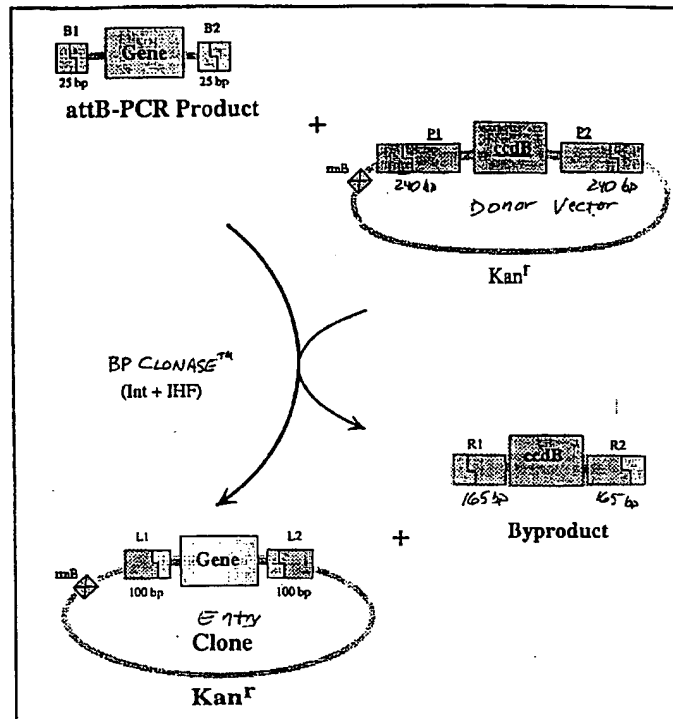


FIGURE 8

Recombination Site Nucleotide Sequences

attB1: 5'-ACAAGTTTGTACAAAAAAGCAGGCT-3'

attB2: 5'-ACCCAGCTTTCTTGTACAAAGTGGT-3'

attP1: 5'-TACAGGTCCTAATACCATCTAAGTAGTTGATTCATAGTACTGGATATG-
TTGTGTTTTACAGTATTATGTAGTCTGTTTTTATGCAAAATCTAATTTA-
ATATATTGATATTTATATCATTTCACGTTTCTCGTTCAGCTTTTTGTAC-
AAAGTTGGCATTATAAAAAAGCATTGCTCATCAATTTGTTGCAACGAACA-
GGTCACTATCAGTCAAAATAAAATCATTATTTG-3'

attP2: 5'-CAAATAATGATTTTATTTTGAAGTATAGTGACCTGTTGTTGCAACAAAT-
TGATAAGCAATGCTTTCTTATAATGCCAACTTTGTACAAGAAAGCTGAAC-
GAGAAACGTAAAATGATATAAATATCAATATATTAAATTAGATTTTGCAT-
AAAAACAGACTACATAATACTGTAAACACAACATATCCAGTCACTATGA-
ATCAACTACTTAGATGGTATTAGTGACCTGTA-3'

attR1: 5'-ACAAGTTTGTACAAAAAAGCTGAACGAGAAACGTAAAATGATATAAA-
TATCAATATATTAAATTAGATTTTGCATAAAAAACAGACTACATAATAC-
TGTAACACAACATATCCAGTCACTATG-3'

attR2: 5'-GCAGGTCGACCATAGTGACTGGATATGTTGTGTTTTACAGTATTAT-
GTAGTCTGTTTTTATGCAAAATCTAATTTAATATATTGATATTT-
ATATCATTTTACGTTTCTCGTTCAGCTTTCTTGTACAAAGTGGT-3'

attL1: 5'-CAAATAATGATTTTATTTTGAAGTATAGTGACCTGTTGTTGCAAC-
AAATTGATAAGCAATGCTTTTTTATAATGCCAACTTTGTACAAAAAA-
GCAGGCT-3'

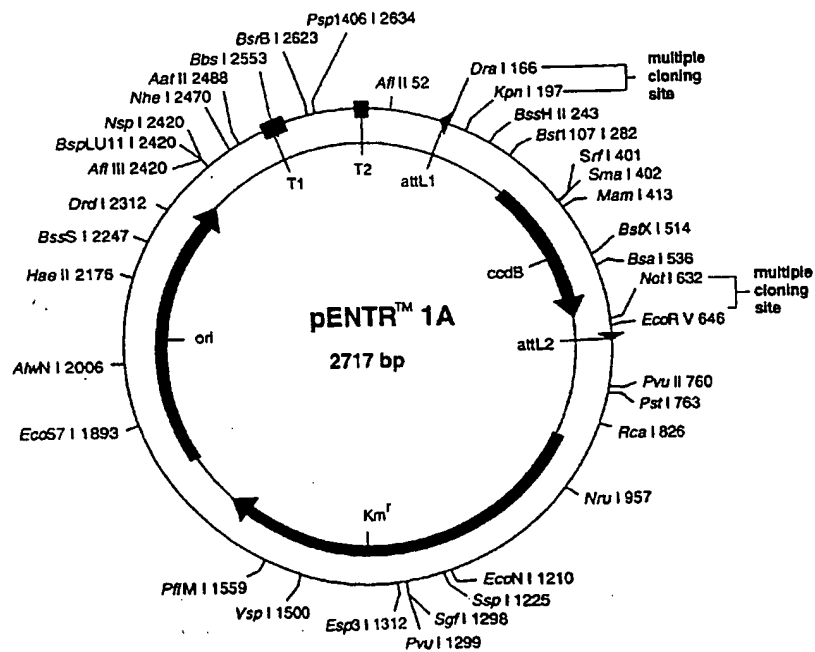
attL2: 5'-CAAATAATGATTTTATTTTGAAGTATAGTGACCTGTTGTTGCAACAA-
ATTGATAAGCAATGCTTTCTTATAATGCCAACTTTGTACAAGAAAGCTGGGT-3'

Figure 9

Figure 10A: Cloning sites of the Entry Vector pENTRTM1A (reading frame A)

ACT TTG TAC AAA AAA GCA GGC TTT AAA GGA ACC AAT TCA GTC GAC TGG ATC CGG TAC CGA ATT C
 TGA AAC ATG TTT TTT CGT CCG AAA TTT CCT TGG ITTA AGT CAG CTG ACC TAG GCC ATG GCT TAA G
 thr leu tyr lys lys ala gly phe lys gly thr asn ser val asp trp ile arg tyr arg ile

EcoR I Not I Xho I EcoR V
 G AAT TCG CCG CCG CAC TCG AGA TAT CTA GAC CCA GCT TTC TTG TAC AAA
 C TTA AGC GCC GGC GTG AGC TCT ATA GAT CTG GGT CGA AAG AAC ATG TTT



pENTR1A 2717 bp

Base Nos.	Gene Encoded
67..166	attL1
321..626	ccdB
655..754	attL2
877..1686	KmR
1791..2364	ori

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1 CTGACGGATG GCCTTTTTCG GTTCTACAA ACTCTTCCTG TTAGTTAGTT ACTTAAGCTC
61 GGGCCCCAAA TAATGATTTT ATTTTGACTG ATAGTGACCT GTTCGTTGCA ACAAATTGAT
121 AAGCAATGCT TTTTATAAT GCCAACTTTG TACAAAAAAG CAGGCTTTAA AGGAACCAAT
181 TCAGTCGACT GGATCCGGTA CCGAATTCGC TTACTAAAAG CCAGATAACA GTATGCGTAT
241 TTGCGCGCTG ATTTTTCGGG TATAAGAATA TATACTGATA TGTATACCCG AAGTATGTCA
301 AAAAGAGGTG TGCTTCTAGA ATGCAGTTTA AGGTTTACAC CTATAAAGA GAGAGCCGTT
361 ATCGTCTGTT TGTGGATGTA CAGAGTGATA TTATTGACAC GCCCGGGCGA CGGATAGTGA
421 TCCCCCTGGC CAGTGACAGT CTGCTGTGAG ATAAAGTCTC CCGTGAACTT TACCCGGTGG
481 TGCATATCGG GGATGAAAGC TGGCGCATGA TGACCACCGA TATGGCCAGT GTGCCGGTCT
541 CCGTTATCGG GGAAGAAGTG GCTGATCTCA GCCACCGCGA AAATGACATC AAAAACGCCA
601 TTAACCTGAT GTTCTGGGGA ATATAGAATT CGCGGCCGCA CTCGAGATAT CTAGACCCAG
661 CTTTCTTGTA CAAAGTTGGC ATTATAAGAA AGCATTGCTT ATCAATTGTT TGCAACGAAC
721 AGGTCACAT CAGTCAAAAT AAAATCATT TTTGCCATCC AGCTGCAGCT CTGGCCCGTG
781 TCTCAAAATC TCTGATGTTA CATTGCACAA GATAAAATA TATCATCATG AACATATAAA
841 CTGCTCTGCT ACATAAACAG TAATACAAGG GGTGTTATGA GCCATATTCA ACGGGAAACG
901 TCGAGGCCGC GATTAAATTC CAACATGGAT GCTGATTAT ATGGGTATAA ATGGGCTCGC
961 GATAATGTCG GGCAATCAGG TGGGACAATC TATCGCTTGT ATGGGAAGCC CGATGCGCCA
1021 GAGTTGTTTC TGAACATAGG CAAAGGTAGC GTTGCCAATG ATGTTACAGA TGAGATGTGC
1081 AGACTAAACT GGCTGACGGA ATTTATGCCT CTTCCGACCA TCAAGCAATT TATCGTACT
1141 CCTGATGATG CATGGTTACT CACCACTGCG ATCCCCGAA AAACAGCAIT CCAGGTATTA
1201 GAAGAAATAT CTGATTCAGG TGAATAATAT GTTGATGCGC TGGCAGTGTG CCTGCGCCGG
1261 TTGCATTGCA TTCCTGTTTG TAATTGTCCT TTTAACAGCG ATCGCGTATT TCGTCTCGCT
1321 CAGCGCAAT CACGAATGAA TAACGGTTTG GTTGATGCGA GTGATTTTGA TGACGAGCGT
1381 AATGGCTGGC CTGTTGAACA AGTCTGGAAG GAAATGCATA AACTTTTGCC ATTTCTACCG
1441 GATTGAGTCG TCACTCATGG TGATTTCTCA CTTGATAACC TTATTTTGA CGAGGGGAAA
1501 TTAATAGGTT GTATTGATGT TGGACGAGTC GGAATCGCAG ACCGATACCA GGATCTTGCC
1561 ATCCTATGGA ACTGCCCTCG TGAGTTTCTT CCTTCATTAC AGAAACGGCT TTTTCAAAAA
1621 TATGGTATTG ATAATCCTGA TATGAATAAA TTGCAGTTTC ATTTGATGCT CGATGAGTTT
1681 TTCTAATCAG AATTGGTTAA TTGGTTGTAA CATTATTGAG ATTTGGCCCC GTTCCACTGA
1741 GCGTCAGACC CCGTAGAAAA GATCAAAGGA TCTTCTTGAG ATCCTTTTTT TCTGCGCGTA
1801 ATCTGCTGCT TGCAAAACAA AAAACCAACG CTACCAGCGG TGGTTTGTGTT GCCGGATCAA
1861 GAGCTACCAA CTCTTTTTCG GAAGGTAAGT GGCTTCAGCA GAGCGCAGAT ACCAAATACT
1921 GTTCTTCTAG TGTAGCCGTA GTTAGGCCAC CACTTCAAGA ACTCTGTAGC ACCGCCATCA
1981 TACCTCGCTC TGCTAATCCT GTTACCAGTG GCTGCTGCCA GTGGCGATAA GTCGTGTCTT
2041 ACCGGGTTGG ACTCAAGACG ATAGTTACCG GATAAGGCGC AGCGGTGCGG CTGAACGGGG
2101 GGTTCGTGCA CACAGCCAGC CTTGGAGCGA ACGACCTACA CCGAACTGAG ATACCTACAG
2161 CGTGAGCTAT GAGAAAGCGC CACGCTTCCC GAAGGGAGAA AGGCGGACAG GTATCCGGTA
2221 AGCGGCAGGG TCGGAACAGG AGAGCGCAGC AGGGAGCTTC CAGGGGAAAA CGCCTGGTAT
2281 CTTTATAGTC CTGTGCGGTT TCGCCACCTC TGACTTGAGC GTCGATTTT GTGATGCTCG
2341 TCAGGGGGGC GGAGCCTATG GAAAAACGCC AGCAACGCGG CCTTTTACG GTTCTGCGC
2401 TTTTGCTGGC CTTTGTCTCA CATGTTCTTT CCTGCGTTAT CCCCTGATTG TGTGGATRAA
2461 CGTATTACCG CTAGCATGGA TCTCGGGGAC GTCTAACTAC TAAGCGAGAG TAGGGAACTG
2521 CCAGGCATCA AATAAACGGA AAGGCTCAGT CGGAAGACTG GGCCTTTCGT TTTATCTGTT
2581 GTTTGTGCGT GAACGCTCTC CTGAGTAGGA CAAATCCGCC GGGAGCGGAT TTGAACGTTG
2641 TGAAGCAACG GCCCGGAGG TGGCGGGCAG GACGCCCCGC ATAAACTGCC AGGCATCAAA
2701 CTAAGCAGAA GGCCATC

```

FIGURE 10B

Figure 11A: Cloning Sites of the Entry Vector pENTR2B (reading frame B)

Int	attL1	EheI	XmnI	Sall	BamHI											
TTG	TAC	AAA	AAA	GCA	GGC	TPG	CPC	CGG	AAC	CAA	TTC	AGT	CGA	CTG	GAT	CCG
AAC	ATG	TTT	TTT	CGT	CCG	ACC	GCG	GCC	TTG	GTT	AAG	TCA	GCT	GAC	CTA	GGC
Leu	Tyr	Lys	Lys	Ala	Gly	Trp	Arg	Arg	Asn	Gln	Phe	Ser	Arg	Leu	Asp	Pro

KpnI	EcoRI	EcoRI	NotI	XhoI	EcoRV	XbaI										
GTA	CCG	AAT	TC-	ccdB	--G	AAT	TCG	CCG	CCG	CAC	TCG	AGA	TAT	CTA	GAC	CCA
CAT	GGC	TTA	AG			C	TTA	AGC	GCC	GGC	GTG	AGC	TCT	ATA	GAT	CTG
Val	Pro	Asn				Asn	Ser	Arg	Pro	His	Ser	Arg	Tyr	Leu	Asp	Pro

Int	attL2
GCT	TTT
CGA	AAG
AAC	ATG
TTT	C
Ala	Phe
Leu	Tyr
Lys	

pENTR2B 2718 bp

Location (Base Nos.)	Gene Encoded
67..166	attL1
322..627	ccdB
656..755	attL2
878..1687	KmR
1792..2365	ori

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1 CTGACGGATG GCCTTTTTCG GTTCTACAA ACTCTTCTG TTAGTTAGTT ACTTAAGCTC
61 GGGCCCCAAA TAATGATTTT ATTTTGACTG ATAGTGACCT GTTCGTTGCA ACAAAATTGAT
121 AAGCAATGCT TTTTATAAAT GCCAACTTTG TACAAAAAAG CAGGCTGGCG CCGGAACCAA
181 TTCAGTCGAC TGGATCCGGT ACCGAATTCT CTTACTAAAA GCCAGATAAC AGTATGCGTA
241 TTTGCGCGCT GATTTTTCGG GTATAAGAAT ATATACTGAT ATGTATACCC GAAGTATGTC
301 AAAAAAGAGGT GTGCTTCTAG AATGCAGTTT AAGGTTTACA CCTATAAAAG AGAGAGCCGT
361 TATCGTCTGT TTGTGGATGT ACAGAGTGAT ATTATTGACA CGCCCGGGCG ACGGATGGTG
421 ATCCCCCTGG CCAGTGCACG TCTGCTGTCA GATAAAGTCT CCCGTGAAC TACCCCGGTG
481 GTGCATATCG GGGATGAAAG CTGGCGCATG ATGACCACCG ATATGGCCAG TGTGCCGGTC
541 TCCGTTATCG GGAAGAAGT GGCTGATCTC AGCCACCGCG AAAATGACAT CAAAAACGCC
601 ATTAACCTGA TGTCTGGGG AATATAGAAT TCGCGGCCGC ACTCGAGATA TCTAGACCCA
661 GCTTCTTGT ACAAAGTTGG CATTATAAGA AAGCATTTGCT TATCAATTTG TTGCAACGAA
721 CAGGTCACTA TCAGTCAAAA TAAATCATT ATTGCCATC CAGCTGCAGC TCTGGCCCGT
781 GTCTCAAAAT CTCTGATGTT ACATTGCACA AGATAAAAT ATATCATCAT GAACAATAAA
841 ACTGTCTGCT TACATAAACA GTAATACAAG GGGTGTATG AGCCATATT CACGGGAAC
901 GTCGAGGCCG CGATTAAAT CCAACATGGA TGCTGATTTA TATGGGTATA AATGGGCTCG
961 CGATAATGTC GGGCAATCAG GTGCGACAAT CTATCGCTTG TATGGGAAGC CCGATGCGCC
1021 AGAGTTGTTT CTGAAACATG GCAAAGGTAG CGTTGCCAAT GATGTTACAG ATGAGATGGT
1081 CAGACTAAAC TGGCTGACGG AATTTATGCC TCTCCGACC ATCAAGCAAT TTATCCGTAC
1141 TCCTGATGAT GCATGGTTAC TCACCACTGC GATCCCCGGA AAAACAGCAT TCCAGGTATT
1201 AGAAGAATAT CCTGATTCAG GTGAAAATAT TGTTGATGCG CTGGCAGTGT TCCTGCGCCG
1261 GTTGCAATCG ATTCTGTGTT GTAATTGTCC TTTTAACAGC GATCGCGTAT TTCGTCCTCG
1321 TCAGGCGCAA TCACGAATGA ATAACGGTTT GGTGATGCG AGTGATTTTG ATGACGAGCG
1381 TAATGGCTGG CCTGTGAAC AAGTCTGGAA AGAAATGCAT AAATTTTTCG CATTCTCACC
1441 GGATTCAGTC GTCACTCATG GTGATTTCTC ACTTGATAAC CTTATTTTTC ACGAGGGGAA
1501 ATTAATAGGT TGTATTGATG TTGGACGAGT CGGAATCGCA GACCGATACC AGGATCTTGC
1561 CATCCTATGG AACTGCCTCG GTGAGTTTTC TCCTTCATTA CAGAAACGGC TTTTCAAAA
1621 ATATGGTATT GATAATCCTG ATATGAATAA ATTGCAGTTT CATTGTATGC TCGATGAGTT
1681 TTTCTAATCA GAATTGGTTA APTGGTTGTA ACATTATTCG GATTTGGGCC CGTTCCAATG
1741 AGCGTCAGAC CCCGTAGAAA AGATCAAAGG ATCTTCTTGA GATCCTTTTT TTCTGCGCGT
1801 AATCTGCTGC TTGCAAAACA AAAAACCCAC GCTACCAGCG GTGGTTTGTG TGCCGGATCA
1861 AGAGCTACCA ACTCTTTTTT CGAAGGTAAC TGGCTTCAGC AGAGCGCAGA TACCAAAATAC
1921 TGTTCTTCTA GTGTAGCCGT AGTTAGGCCA CCACITCAAG AACTCTGTAG CACCGCTTAC
1981 ATACCTCGCT CTGCTAATCC TGTACCAGT GGCTGCTGCC AGTGGCGATA AGTCGTGTCT
2041 TACCGGGTTG GACTCAAGAC GATAGTTACC GGATAAGGCG CAGCGGTCCG GCTGAACGGG
2101 GGGTTCGTGC ACACAGCCCA GCTTGGAGCG AACGACCTAC ACCGAACTGA GATACCTACA
2161 GCGTGAGCTA TGAGAAAGCG CCACGCTTCC CGAAGGGAGA AAGGCGGACA GGTATCCGGT
2221 AAGCGGCAGG GTCGGAACAG GAGAGCGCAC GAGGGAGCTT CCAGGGGGAA ACGCCTGGTA
2281 TCTTTATAGT CCTGTGCGGT TTCGCCACCT CTGACTTGAG CGTCGATTTT TGTGATGCTC
2341 GTCAGGGGGG CGGAGCCTAT GGA AAAACGC CAGCAACGCG GCCTTTTAC GGTTCCTGGC
2401 CTTTGTCTGG CTTTGTCTC ACATGTTCTT TCCTGCGTTA TCCCTTGATT CTGTGGATAA
2461 CCGTATTACC GCTAGCATGG ATCTCGGGGA CGTCTAATA CTAAGCGAGA GTAGGGAAC
2521 GCCAGGCATC AAATAAAACG AAAGGCTCAG TCGGAAGACT GGGCCTTTTC TTTTATCTGT
2581 TGTTTGTGCG TGAACGCTCT CCTGAGTAGG ACAAATCCGC CGGGAGCGGA TTTGAACGTT
2641 GTGAAGCAAC GGCCCGGAGG GTGCGGGCA GAGCGCCGC CATAACTGC CAGGCATCAA
2701 ACTAAGCAGA AGGCCATC

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Figure 1B

Figure 24: Cloning Sites of the Entry Vector pENTR3C (reading frame C)

Int	attL1	DraI	XmnI	Sall	BamHI												
TTC	TAC	AAA	AAA	GCA	GGC	TCT	TTA	AAG	GAA	CCA	ATT	CAG	TCG	ACT	GSA	TCC	GGT
AAC	ATG	TTT	TTT	CGT	CCG	AGA	AAT	TTC	CTT	GGT	TAA	GTC	AGC	TGA	CCT	AGG	CCA
							↓			↓				↓		↓	↓
Leu	Tyr	Lys	Lys	Ala	Gly	Ser	Leu	Lys	Glu	Pro	Ile	Gln	Ser	Thr	Gly	Ser	Gly

KpnI	EcoRI	PvuI	EcoRI	NotI	XhoI	EcoRV	XbaI									
ACC	GAA	TTC	GAT	CSC--	ccdB	--G	AAT	TCG	CCG	CCG	CAC	TCG	AGA	TAT	CTA	
TGG	CTT	AAG	CTA	GCG			C	TTA	AGC	GCC	GGC	GTG	AGC	TCT	ATA	GAT
Thr	Glu	Phe					Asn	Ser	Arg	Pro	His	Ser	Arg	Tyr	Leu	

attL2	Int						
GAC	CCA	GCT	TTC	TTG	TAC	AAA	G
CTG	GGT	CGA	AAG	AAC	ATG	TTT	C
Asp	Pro	Ala	Phe	Leu	Tyr	Lys	

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pENTR3C 2723 bp

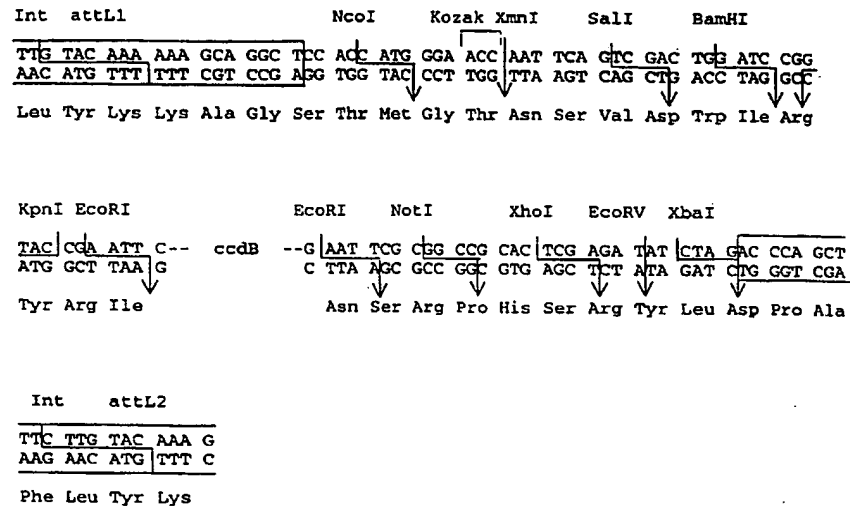
<u>Location (Base Nos.)</u>	<u>Gene Encoded</u>
67..166	attL1
327..632	ccdB
661..760	attL2
883..1692	KmR
1797..2370	ori

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1 CTGACGGATG GCCTTTTTCG GTTCTACAA ACTCTTCTCG TTAGTTAGTT ACTTAAGCTC
61 GGGCCCCAAA TAATGATTTT ATTTTGACTG ATAGTGACCT GTTCGTTGCA ACAAAATTGAT
121 AAGCAATGCT TTTTATAAT GCCAACTTTG TACAAAAAAG CAGGCTCTTT AAAGGAACCA
181 ATTCAAGTCG CTGGATCCGG TACCGAATTC GATCGCTTAC TAAAGCCAG ATAACAGTAT
241 GCGTATTTGC GCGCTGATTT TTGCGGTATA AGAATATATA CTGATATGTA TACCCGAAGT
301 ATGTCAAAAA GAGGTGTGCT TCTAGAATGC AGTTTAAAGT TTACACCTAT AAAAGAGAGA
361 GCCGTTATCG TCTGTTTGTG GATGTACAGA GTGATATTAT TGACACGCCC GGGCGACGGA
421 TGGTGATCCC CTTGGCCAGT GCACGTCTGC TGTGAGATAA AGTCTCCCGT GAACTTTACC
481 CCGTGGTGCA TATCGGGGAT GAAAGCTGGC GCATGATGAC CACCGATATG GCCAGTGTGC
541 CCGTCTCCGT TATCGGGGAA GAAGTGGCTG ATCTCAGCCA CCGCGAAAAT GACATCAAAA
601 ACGCCATTAA CCTGATGTTT TGGGGAATAT AGAATTCGCG GCCGCACTCG AGATATCTAG
661 ACCCAGCTTT CTGTGACAAA GTTGGCATT TAAGAAAGCA TTGCTTATCA ATTTGTTGCA
721 ACGAACAGGT CACTATCAGT CAAATAAAAA TCATTATTG CCATCCAGCT GCAGCTCTGG
781 CCCGTGTCTC AAAATCTCTG ATGTTACATT GCACAAGATA AAAATATATC ATCATGAACA
841 ATAAACTGTG CTGCTTACAT AAACAGTAAT ACAAGGGGTG TTATGAGCCA TATCAACGG
901 GAAACGTCGA GCGCGCGATT AAATTCCAAC ATGGATGCTG ATTTATATGG GTATAAATGG
961 GCTCGCGATA ATGTCGGGCA ATCAGGTGCG ACAATCTATC GCTTGTATGG GAAGCCCGAT
1021 GCGCCAGAGT TGTTCCTGAA ACATGGGCAA GGTAGCGTTG CCAATGATG TACAGATGAG
1081 ATGGTCAGAC TAAACTGGCT GACGGAATTT ATGCTCTCTC CGACCATCAA GCATTTTATC
1141 CGTACTCCTG ATGATGCATG GTTACTCACC ACTGCGATCC CCGGAAAAAC AGCATTCCAG
1201 GTATTAGAAG AATATCTCGA TTCAGGTGAA AATATTGTTG ATGCGCTGGC AGTGTTCCTG
1261 CGCCGCTTGC ATTCGATTCC TGTTTGTAAT TGTCCTTTTA ACAGCGATCG CGTATTTCGT
1321 CTCGCTCAGG CGCAATCAGC AATGAATAAC GGTTCGTTG ATGCGAGTGA TTTTGATGAG
1381 GAGCGTAATG GCTGGCCTGT TGAACAGTC TGGAAAGAAA TGCATAAACT TTTGCCATTC
1441 TCACCGGATT CAGTCGTAC TCATGGTGAT TTCTCACTTG ATAACCTTAT TTTTGACGAG
1501 GGGAAATTAA TAGGTTGTAT TGATGTTGGA CGAGTCGGAA TCGCAGACCG ATACCAGGAT
1561 CTGCCCATCC TATGGAACG CCTCGGTGAG TTTCTCCTT CATTACAGAA ACGGCTTTTT
1621 CAAAAATATG GTATTGATA TCCTGATATG AATAAATTGC AGTTTCATTG GATGCTCGAT
1681 GAGTTTCTCT AATCAGAATT GGTTAATTGG TTGTAACATT ATTCAGATTG GGGCCCGTTC
1741 CACTGAGCGT CAGACCCCGT AGAAAAAGATC AAAGGATCTT CTTGAGATCC TTTTCTCTG
1801 CGCGTAATCT GCTGCTTGCA AACAAAAAAA CCACCGCTAC CAGCGGTGGT TTGTTTGGCG
1861 GATCAAGAGC TACCAACTCT TTTTCCGAAG GTAACCTGGT TCAGCAGAGC GCAGATACCA
1921 AATACTGTTT TTCTAGTGTA GCCGTAGTTA GGCCACCACT TCAAGAACTC TGTAGCACCG
1981 CCTACATACC TCGCTCTGCT AATCCTGTTA CCAGTGGCTG CTGCCAGTGG CGATAAGTCTG
2041 TGTCTTACCG GGTGGAATC AAGACGATAG TTACCGGATA AGGCGCAGCG GTCGGGCTGA
2101 ACGGGGGGTT CGTGACACAC GCCCAGCTTG GAGCGAACGA CCTACACCGA ACTGAGATAC
2161 CTACAGCGTG AGCTATGAGA AAGCGCCACG CTTCCCGAAG GGAGAAAGGC GGACAGGTAT
2221 CCGGTAAGCG GCAGGGTCTG AACAGGAGAG CGCACGAGGG AGCTTCCAGG GGGAAACGCC
2281 TGGTATCTTT ATAGTCTGTG CGGGTTTCGC CACCTCTGAC TTGAGCGTGG ATTTTGTGTA
2341 TGCTCGTCAG GGGGGCGGAG CCTATGGAAA AACGCCAGCA ACGCGGCCTT TTTACGGTTC
2401 CTGGCCTTTT GCTGGCCTTT TGCTCACATG TTCCTTCTG CGTTATCCCC TGATTCTGTG
2461 GATAACCGTA TTACCGCTAG CATGGATCTC GGGGACGTCT AACTACTAAG CGAGAGTAGG
2521 GAAGTCCGAG GCATCAATAA AAACGAAAGG CTCAGTCGGA AGACTGGGCC TTTGCTTTTA
2581 TCTGTTGTTT GTCGGTGAAC GCTCTCCTGA GTAGGACAAA TCCGCCGGGA GCGGATTGTA
2641 ACGTTGTGAA GCAACGGGCC GGAGGGTGGC GGGCAGGACG CCCGCCATAA ACTGCCAGGC
2701 ATCAAACTAA GCAGAAGGCC ATC

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Figure 12B

Figure 13A: Cloning Sites of the Entry Vector pENTR4 :

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pENTR4 2720 bp

Location (Base Nos.)	Gene Encoded
67..166	attL1
324..629	ccdB
658..757	attL2
880..1689	KmR
1794..2367	ori

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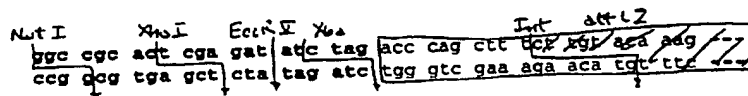
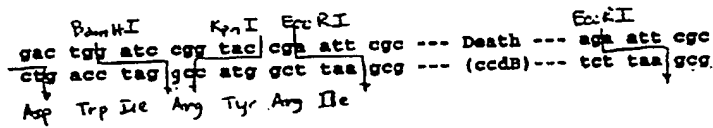
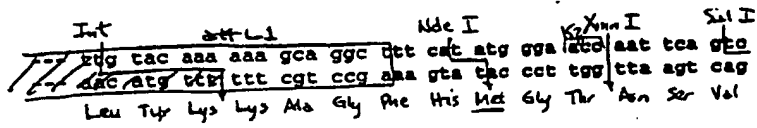
1 CTGACGGATG GCCTTTTTCG GTTCTACAA ACTCTTCTG TTAGTTAGTT ACTTAAGCTC
61 GGGCCCCAAA TAATGATTTT ATTTTGACTG ATAGTGACCT GTTCGTTGCA ACAAATTGAT
121 AAGCAATGCT TTTTATAAT GCCAACTTTG TACAAAAAAG CAGGCTCCAC CATGGGAACC
181 AATTCAGTCG ACTGGATCCG GTACCGAATT CGCTTACTAA AAGCCAGATA ACAGTATGCG
241 TATTTCGCGC CTGATTTTTG CGGTATAAGA ATATATACTG ATATGTATAC CCGAAGTATG
301 TCAAAAAGAG GTGTGCTTCT AGAATGCAGT TTAAGGTTTA CACCTATAAA AGAGAGAGCC
361 GTTATCGTCT GTTTGTGGAT GTACAGAGTG ATATTATGTA CACGCCCGGG CGACGGATGG
421 TGATCCCCCT GGGCAGTGCA CGTCTGCTGT CAGATAAAGT CTCCCGTGAA CTTTACCCGG
481 TGSTGCATAT CGGGGATGAA AGCTGGCGCA TGATGACCAC CGATATGGCC AGTGTGCCGG
541 TCTCCGTTAT CGGGGAAGAA GTGGCTGATC TCAGCCACCG CGAAAAATGAC ATCAAAAACG
601 CCATTAACCT GATGTTCTGG GGAATATAGA ATTCGCGGCC GCACCTGAGA TATCTAGACC
661 CAGCTTTCTT GTACAAAGTT GGCATTATAA GAAAGCATTG CTTATCAATT TGTGCAACG
721 AACAGGTCAC TATCAGTCAA AATAAAATCA TTATTGCCA TCCAGCTGCA GCTCTGGCCC
781 GTGTCTCAAA ATCTCTGATG TTACATTGCA CAAGATAAAA ATATATCATC ATGAACAATA
841 AAACCTGTCTG CTTACATAAA CAGTAATACA AGGGGTGTTA TGAGCCATAT TCAACGGGAA
901 ACGTCGAGGC CGCGATTAAA TTCCAACATG GATGCTGATT TATATGGGTA TAAATGGGCT
961 CGCGATAATG TCGGGCAATC AGGTGCGACA ATCTATCGCT TGTATGGGAA GCCCGATGCG
1021 CCAGAGTTGT TTCTGAACA TGGCAAAGGT AGCGTTGCCA ATGATGTTAC AGATGAGATG
1081 GTCAGACTAA ACTGGCTGAC GGAATTTATG CCTCTTCCGA CCATCAAGCA TTTTATCCGT
1141 ACTCCTGGTG ATGCATGGTT ACTCACCAC GCGATCCCGG GAAAAACAGC ATTCCAGGTA
1201 TTAGAAGAAT ATCCTGATTC AGGTGAAAT ATTGTTGATG CGCTGGCAGT GTTCTGCGC
1261 CGGTTGCATT CGATTCTCTG TTGTARTTGT CCTTTTAAAC GCGATCGCGT ATTTCTGCTC
1321 GCTCAGGCGC AATCAGCAAT GAATAACGGT TTGGTTGATG CGAGTGATTT TGATGACGAG
1381 CGTAATGGCT GGCCTGTGTA ACAAGTCTGG AAAGAAATGC ATAACTTTT GCCATTCTCA
1441 CCGGATTGAG TCCTCACTCA TGGTGATTTT TCACTTGATA ACCTTATTTT TGACGAGGGG
1501 AAATTAATAG GTTGATTTGA TGTGGACGA GTCCGAATCG CAGACCGATA CCAGGATCTT
1561 GCCATCCTAT GGAAGTGCCT CGGTGAGTTT TCTCTTCAT TACAGAAACG GCTTTTTCAG
1621 AAATATGGTA TTGATAATCC TGATATGAAT AAATTGCAAT TTCATTTGAT GCTCGATGAG
1681 TTTTCTAAT CAGAATGGT TAATTGGTTG TAACATTATT CAGATTGGGC CCCGTTCCAC
1741 TGAGCGTCAG ACCCGTAGA AAAGATCAAA GGATCTTCTT GAGATCCTTT TTTTCTGCGC
1801 GTAATCTGCT GCTTGCAAAC AAAAAACCA CCGCTACCAG CGGTGGTTTG TTTGCCGGAT
1861 CAAGAGCTAC CAACTCTTTT TCCGAAGGTA ACTGGCTTCA GCAGAGCGCA GATACCAAT
1921 ACTGTTCTTC TAGTGTAGCC GTAGTTAGGC CACCCTTCA AGAATCTGT AGCACCCTCT
1981 ACATACCTCG CTCTGCTAAT CCTGTTACCA GTGGCTGCTG CCAGTGCGCA TAAGTCGTGT
2041 CTTACCGGGT TGGACTCAAG ACGATAGTTA CCGGATAAGG CGCAGCGGTC GGGCTGAACG
2101 GGGGGTTCGT GCACACAGCC CAGCTTGGAG CGAACGACCT ACACCGAAT GAGATACCTA
2161 CAGCGTCAGC TATGAGAAAG CGCCACGCTT CCGAAGGGA GAAAGGCGGA CAGGTATCCG
2221 GTAAGCGGCA GGGTCGGAAC AGGAGAGCGC ACGAGGGAGC TTCCAGGGGG AAACGCTCGG
2281 TATCTTTATA GTCCGTGTCG GTTTCGCCAC CTCTGACTTG AGCGTCGATT TTTGTGATGC
2341 TCGTCAGGGG GGGCGAGCCT ATGGA AAAAC GCCAGCAACG CGGCCTTTT ACGGTTCTCTG
2401 GCCTTTTGCT GGCCTTTTGC TCACATGTTT TTTCTGCGT TATCCCTGA TTTCTGGAT
2461 AACCGTATTA CCGCTAGCAT GGATCTCGGG GACGTCTAAC TACTAAGCGA GAGTAGGGAA
2521 CTGCCAGGCA TCAAAATAAA CGAAAGGCTC AGTCGAAGA CTGGGCCTTT CGTTTTATCT
2581 GTTGTGAGCA GGTGAACGCT CTCCTGAGTA GGACAAATCC GCCGGGAGCG GATTTGAACG
2641 TTGTGAAGCA ACGGCCCGGA GGGTGGCGGG CAGGACGCCC GCCATAAACT GCCAGGCATC
2701 AAATAAGCA GAAGGCCATC

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FIGURE 13B

18/240

Figure 1A: Cloning sites of the Entry Vector pENTR5



19/240

pENTR5 2720 bp

Location (Base Nos.)	Gene Encoded
67..166	attL1
324..629	ccdB
658..757	attL2
880..1689	KmR
1794..2367	ori

```

1 CTGACGGATG GCCTTTTTCG GTTCTACAA ACTCTTCCTG TTAGTTAGTT ACTTAAGCTC
61 GGGCCCCAAA TAATGATTTT ATTTTGACTG ATAGTGACCT GTTCGTTGCA ACAATTTGAT
121 AAGCAATGCT TTTTATAAT GCCAACTTTG TACAAAAAAG CAGGCTTTCA TATGGGAACC
181 AATTCAGTCG ACTGGATCCG GTACCGAATT CGCTTACTAA AAGCCAGATA ACAGTATGCG
241 TATTTGCGCG CTGATTTTTC CGGTATAAGA ATATATACTG ATATGTATAC CCGAAGTATG
301 TCAAAAAGAG GTGTGCTTCT AGAATGCAGT TTAAGGTTTA CACCTATAAA AGAGAGAGCC
361 GTTATCGTCT GTTTGTGGAT GTACAGAGTG ATATTATTGA CACGCCCGGG CGACGGATGG
421 TGATCCCCCT GGCCAGTGCA CGTCTGCTGT CAGATAAAGT CTCCCGTGAA CTTTACCCGG
481 TGGTGACATAT CGGGGATGAA AGCTGGCGCA TGATGACCCG CGATATGGCC AGTGTGCCGG
541 TCTCCGTTAT CGGGGAAGAA GTGGCTGATC TCAGCCACCG CGAAAATGAC ATCAAAAACG
601 CCATTAACTT GATGTTCTGG GGAATATAGA ATTCGCGGGC GCATCGGAGA TATCTAGACC
661 CAGCTTTCTT GTACAAAGTT GGCATTATAA GAAAGCATTG CTTATCAATT TGTTCGAACG
721 AACAGGTCAC TATCAGTCAA AATAAAATCA TTATTTGCCA TCCAGCTGCA GCTCTGGCCC
781 GTGTCTCAAA ATCTCTGATG TTACATTGCA CAAGATAAAA ATATATCATC ATGAACAATA
841 AAAGTGTCTG CTACATAAAA CAGTAATACA AGGGGTGTTA TGAGCCATAT TCAACGGGAA
901 ACGTCGAGGC CGCGATTAAA TTCCAACATG GATGCTGATT TATATGGGTA TAAATGGGCT
961 CGCGATAATG TCGGGCAATC AGGTGCGACA ATCTATCGCT TGTATGGGAA GCCCGATGCG
1021 CCAGAGTTGT TTCTGAAACA TGGCAAAGGT AGCGTTGCCA ATGATGTTAC AGATGAGATG
1081 GTCAGACTAA ACTGGCTGAC GGAATTTATG CCTCTTCCGA CCATCAAGCA TTTTATCCGT
1141 ACTCCTGATG ATGCATGGTT ACTCACCACG GCGATCCCGG GAAAACACAGC ATTCAGGTA
1201 TTAGAAGAAT ATCCTGATTC AGGTGAAAAT ATTGTTGATG CGCTGGCAGT GTTCCTGCGC
1261 CGGTGTCATT CGATTCTCTG TTGTAATTGT CCTTTTAAAC GCGATCGCGT ATTTCTGCTC
1321 GCTCAGGCGC AATCAGCAAT GAATAACGGT TTGGTTGATG CGAGTGATTG TGATGACGAG
1381 CGTAATGGCT GGCCTGTTGA ACAAGTCTGG AAAGAAATGC ATAAACTTTT GCCATTCTCA
1441 CCGGATTGAG TCGTCACTCA TGGTGATTTC TCACTTGATA ACCTTATTTT TGACGAGGGG
1501 AAATTAATAG GTTGTATTGA TGTGGACGA GTCCGAATCG CAGACCGATA CCAGGATCTT
1561 GCCATCCTAT GGAACCTGCT CGGTGAGTTT TCTCCTTCAT TACAGAAACG GCTTTTCAA
1621 AAATATGGTA TTGATAATCC TGATATGAAT AAATTGCAGT TTCAATTGAT GCTCGATGAG
1681 TTTTCTAAT CAGAATTGGT TAATTGGTTG TAACATTATT CAGATTGGGC CCCGTTCCAC
1741 TGAGCGTCAG ACCCCGTAGA AAAGATCAAA GGATCTTCTT GAGATCCTTT TTTTCTGCGC
1801 GTAATCTGCT GCTTGCAAC AAAAAACCA CCGCTACCAG CGGTGGTTTG TTTGCCGAT
1861 CAAGAGCTAC CAACTCTTTT TCCGAAGGTA ACTGGCTTCA GCAGAGCGCA GATACCAAT
1921 ACTGTTCTTC TAGTGTAGCC GTAGTTAGGC CACCACTTCA AGAACTCTGT AGCACCGCCT
1981 ACATACCTCG CTCTGCTAAT CCTGTTACCA GTGGCTGCTG CCAGTGGCGA TAAGTCGTGT
2041 CTTACCGGGT TGGACTCAAG ACGATAGTTA CCGGATAAGG CGCAGCGGTC GGGCTGAACG
2101 GGGGGTTCGT GCACACAGCC CAGCTTGGAG CGAACGACCT ACACCGAACT GAGATACCTA
2161 CAGCGTGAGC TATGAGAAAG CGCCACGCTT CCCGAAGGGA GAAAGGCGGA CAGGTATCCG
2221 GTAAAGCGCA GGGTCGGAAC AGGAGAGCGC ACGAGGGAGC TTCCAGGGGG AAACGCCCTG
2281 TATCTTTATA GTCTGTGCG GTTTCGCCAC CTCTGACTTG AGCGTCGATT TTTGTGATGC
2341 TCGTCAGGGG GGCAGAGCCT ATGGAAAAAC GCCAGCAACG CGGCCTTTT ACGGTTCCTG
2401 GCCTTTTGCT GGCCTTTTGC TCACATGTTT TTTCTGCGT TATCCCTGA TTCTGTGGAT
2461 AACCGTATTA CCGCTAGCAT GGATCTCGGG GACGTCTAAC TACTAAGCGA GAGTAGGGAA
2521 CTGCCAGGCA TCGAATAAAA CGAAAGGCTC AGTCGGAAGA CTGGGCCTTT CGTTTTATCT
2581 GTTGTGTTGTC GGTGAACGCT CTCTGAGTA GGACAAATCC GCCGGGAGCG GATTTGAACG
2641 TTGTGAAGCA ACGGCCCGGA GGGTGGCGGG CAGGACGCCC GCCATAAACT GCCAGGCATC
2701 AAACTAAGCA GAAGGCCATC

```

Figure 14B

20/240

Figure 1A: Cloning sites of the Entry Vector pENTR6

Int attL1 Sph I Kpn I Xmn I Sal I
~~--- ttt tac aaa aaa gca ggc tgc atg cga tgc aat tca gtc~~
~~--- aac atg tct ttt cgt ccg atg taa gct tgg tta agt cag~~
 Leu Tyr Lys Lys Ala Gly Cys Met Arg Thr Asn Ser Val

BamH I Kpn I EcoR I EcoR I
 gac tgg atc cgg tac cga att cgc --- Death --- aga att cgc
 cgg acc tag ggc atg gct taa ggc --- (codB) --- tct taa ggc
 Asp Trp Ile Arg Tyr Arg Ile

Not Xho I EcoR I Xba I Int attL2
 ggc cgc act cga gat atc tag acc cag att tgc tgc aga aag ---
 cgg ggc tga gct cta tag atc tgg gtc gaa aga aca tgc ttc ---

pENTR6 2717 bp

Location (Base Nos.)	Gene Encoded
67..166	attL1
321..626	ccdB
655..754	attL2
877..1686	KmR
1791..2364	ori

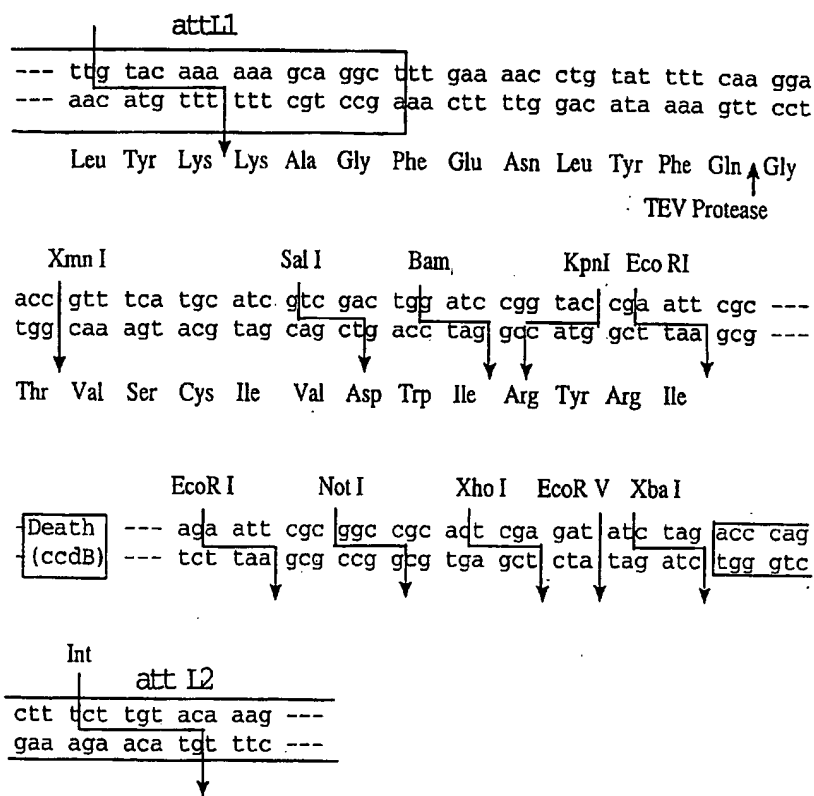
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1 CTGACGGATG GCCTTTTTCG GTTCTACAA ACTCTTCTCG TTAGTTAGTT ACTTAAGCTC
61 GGGCCCCAAA TAATGATTTT ATTTTGACTG ATAGTGACCT GTTCGTTGCA ACAAATTGAT
121 AAGCAATGCT TTTTATAAT GCCAACTTTG TACAAAAAAG CAGGCTGCAT GCGAACCAAT
181 TCAGTCGACT GGATCCGGTA CCGAATTCCG TTACTAAAAG CCAGATAACA GTATGCGTAT
241 TTGCGCGCTG ATTTTTCGCG TATAAGAATA TATACTGATA TGTATACCCG AAGTATGTCA
301 AAAAGAGGTG TGCTTCTAGA ATGCAGTTTA AGGTTTACAC CTATAAAAGA GAGAGCCGTT
361 ATCGTCTGTT TGTGGATGTA CAGAGTGATA TTATTGACAC GCCCGGGCGA CGGATGGTGA
421 TCCCCCTGGC CAGTGCAAGT CTGCTGTGAG ATAAAGTCTC CCGTGAACCT TACCCGCTGG
481 TGCATATCGG GGATGAAAGC TGGCGCATGA TGACCACCGA TATGGCCAGT GTGCCGCTCT
541 CCGTTATCGG GGAAGAAGTG GCTGATCTCA GCCACCGCGA AAATGACATC AAAAACGCCA
601 TTAACCTGAT GTTCTGGGGA ATATAGAATT CGCGGCCGCA CTCGAGATAT CTAGACCCAG
661 CTTTCTTGTA CAAAGTTGGC ATTATAAGAA AGCATTGCTT ATCAATTGTT TGCAACGAAC
721 AGGTCACTAT CAGTCAAAAT AAAATCATT TTTGCCATCC AGCTGCAGCT CTGGCCCGTG
781 TCTCAAAATC TCTGATGTTA CATTGCACAA GATAAAAATA TATCATCATG AACATAAAAA
841 CTGTCTGCTT ACATAAACAG TAATACAAGG GGTGTTATGA GCCATATTCA ACGGGAAACG
901 TCGAGGGCCG GATTAAATTC CAACATGGAT GCTGATTAT ATGGGTATAA ATGGGCTCGC
961 GATAATSTCG GGCAATCAGG TCGGACAATC TATCGCTTGT ATGGGAAGCC CGATGCGCCA
1021 GAGTTGTTTC TGAAACATGG CAAAGGTAGC GTTGCCAAATG ATGTTACAGA TGAGATGGTC
1081 AGACTAAACT GGCTGACGGA ATTATGCCT CTTCGACCA TCAAGCATT TATCCGTACT
1141 CCTGATGATG CATGGTTACT CACCACTGCG ATCCCCGGAA AAACAGCATT CCAGGTATTA
1201 GAAGAATATC CTGATTGAGG TGAAATATTT GTTGATGCGC TGGCAGTGT CTGCGCCGCG
1261 TTGCATTGCA TTCCTGTTTG TAATTGTCCT TTTAACAGCG ATCGCGTATT TCGTCTCGCT
1321 CAGGCGCAAT CACGAATGAA TAACGGTTTG GTTGATGCGA GTGATTTTGA TGACGAGCGT
1381 AATGGCTGGC CTGTTGAACA AGTCTGGAAG GAAATGCATA AACTTTTGCC ATTCTCACC
1441 GATTGAGTCG TCACTCATGG TGATTCTCA CTTGATAACC TTATTTTGA CGAGGGGAAA
1501 TTAATAGGTT GTATTGATGT TGGACGAGTC GGAATCGCAG ACCGATACCA GGATCTTGCC
1561 ATCCTATGGA ACTGCCTCGG TGAGTTTCT CCTTCATTAC AGAAACGGCT TTTTCAAAAA
1621 TATGGTATTG ATAATCTGTA TATGAATAAA TTGCAGTTTC ATTTGATGCT CGATGAGTTT
1681 TTCTAATCAG AATTGGTTAA TTGGTTGTAA CATTATTGAG ATTGGGCCCC GTTCCACTGA
1741 GCGTCAGACC CCGTAGAAAA GATCAAAGGA TCTTCTTGAG ATCCTTTTTC TCTGCGCGTA
1801 ATCTGCTGCT TGCAAAACAA AAAACCAACG CTACCAGCGG TGGTTTGTTT GCCGGATCAA
1861 GAGCTACCAA CTCTTTTCC GAAGGTAACT GGCTTCAGCA GAGCGCAGAT ACCAAATACT
1921 GTTCTTCTAG TGTAGCCGTA GTTAGGCCAC CACTTCAAGA ACTCTGTAGC ACCGCTTACA
1981 TACCTCGCTC TGCTAATCCT GTTACCAGTG GCTGCTGCCA GTGGCGATAA GTCGTGCTTT
2041 ACCGGGTTGG ACTCAAGACG ATAGTTACCG GATAAGGCGC AGCGGTCCGG CTGAACGGGG
2101 GGTTCGTGCA CACAGCCGAG CTTGGAGCGA ACGACCTACA CCGAAGTGA ATACCTACAG
2161 CGTGAGCTAT GAGAAAGCGC CACGCTTCCC GAAGGGAGAA AGCGGACAG GTATCCGGTA
2221 AGCGGCAGGG TCGGAACAGG AGAGCGCACG AGGGAGCTTC CAGGGGGAAG CGCTTGATAT
2281 CTTTATAGTC CTGTGCGGTT TCGCCACCTC TGACTTGAGC GTCGATTTTT GTGATGCTCG
2341 TCAGGGGGGC GGAGCCTATG GAAAAACGCC AGCAACGCGG CCTTTTACG GTTCTCGGCC
2401 TTTTGCTGGC CTTTGTCTCA CATGTTCTTT CTGCGTTAT CCCCTGATTC TGTGGATAAC
2461 CGTATTACCG CTAGCATGGA TCTCGGGGAC GTCTAACTAC TAAGCGAGAG TAGGGAACTG
2521 CCAGGCATCA AATAAAACGA AAGGCTCAGT CGGAAGACTG GGCCTTTCGT TTTATCTGTT
2581 GTTGTGCGGT GAACGCTCTC CTGAGTAGGA CAAATCCGCC GGGAGCGGAT TTGAACGTTG
2641 TGAAGCAACG GCCCGGAGGG TGGCGGGCAG GACGCCGCC ATAAACTGCC AGGCATCAAA
2701 CTAAGCAGAA GGCCATC

```

Figure 15B

Figure 16A: Cloning sites of the Entry Vector pENTR7



pENTR7 2738 bp

Location (Base Nos.)	Gene Encoded
67..166	attL1
342..647	ccdB
676..775	attL2
898..1707	KmR
1812..2385	ori

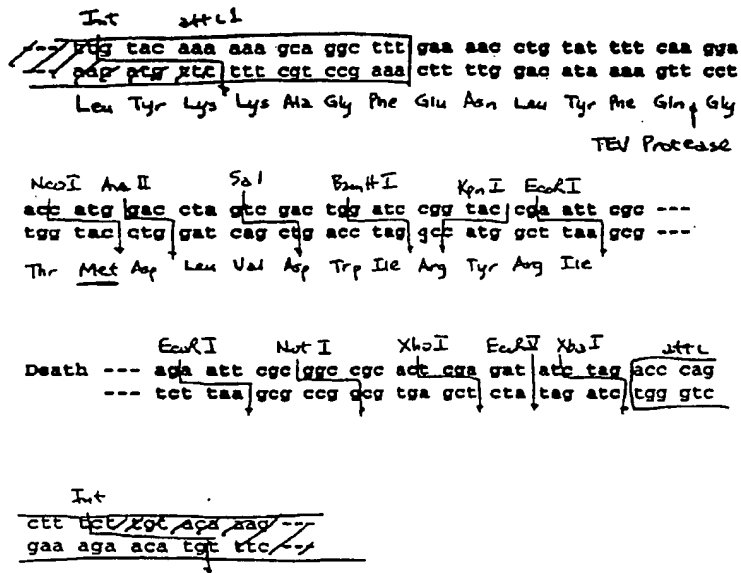
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1 CTGACGGATG GCCTTTTTCG GTTCTACAA ACTCTTCTCG TTAGTTAGTT ACTTAAGCTC
61 GGGCCCCAAA TAATGATTTT ATTTTGACTG ATAGTGACCT GTTCGTTGCA ACAAATTGAT
121 AAGCAATGCT TTTTATAAAT GCCAACTTTG TACAAAAAAG CAGGCTTTGA AAACCTGTAT
181 TTTCAAGGAA CCGTTTCATG CATCGTCGAC TGGATCCGGT ACCGAATTCT CTTACTAAAA
241 GCCAGATAAC AGTATGCGTA TTTGCGCGCT GATTTTTCGG GTATAAGAAT ATATACTGAT
301 ATGTATACCC GAAGTATGTC AAAAAGAGGT GTGCTTCTAG AATGCAGTTT AAGGTTTACA
361 CCTATAAAG AGAGAGCCGT TATCGTCTGT TTGTGGATGT ACAGAGTGAT ATTATTGACA
421 CGCCCGGGCG ACGGATATGT ATCCCTCTGG CCAGTGACAG TCTGCTGTCA GATAAAGTCT
481 CCCGTGAAC TTACCCGGTG GTGCATATCG GGGATGAAAG CTGGCGCATG ATGACCCCG
541 ATATGGCCAG TGTGCCGGTC TCCGTTATCG GGAAGAAGT GGCTGATCTC AGCCACCGCG
601 AAAATGACAT CAAAAACGCC ATTAACCTGA TGTTCGCGG AATATAGAAT TCGCGGCCGC
661 ACTCGAGATA TCTAGACCCA GCTTCTTGT ACAAAGTTGG CATTATAAGA AAGCATTGCT
721 TATCAATTG TTGCAACGAA CAGGTCACCTA TCAGTCAAAA TAAATCATT ATTTGCCATC
781 CAGCTGCAGC TCTGGCCCGT GTCTCAAAAT CTCTGATGTT ACATTGCACA AGATAAAAAAT
841 ATATCATCAT GAACAATAAA ACTGTCTGCT TACATAAACA GTAATACAAG GGGTGTATG
901 AGCCATATTC AACGGGAAAC GTCGAGGCCG CGATTAAATT CCAACATGGA TGCTGATTTA
961 TATGGGTATA AATGGGCTCG CGATAATGTC GGGCAATCAG GTGCGACAAT CTATCGCTTG
1021 TATGGGAAGC CCGATGCGCC AGAGTTGTTT CTGAAACATG GCAAAGGTAG CGTTGCCAAT
1081 GATGTTACAG ATGAGATGCT CAGACTAAAC TGGCTGACGG AATTTATGCC TCTTCCGACC
1141 ATCAAGCATT TTATCCGTAC TCCTGATGAT GCATGGTTAC TCACCACTGC GATCCCCGGA
1201 AAAACAGCAT TCCAGGTATT AGAAGAATAT CCTGATTGAG GTGAAAATAT TGTGTATGCG
1261 CTGGCAGTGT TCCTGCGCCG GTTGCAATCG ATTCTGTTT GTAATTGTCC TTTTAAACAGC
1321 GATCGCGTAT TTCGTCTCGC TCAGGCGCAA TCACGAATGA ATAACGGTTT GGTGTATGCG
1381 AGTGATTTTG ATGACGAGCG TAATGGCTGG CCTGTTGAAC AAGTCTGGAA AGAAATGCAT
1441 AAACCTTTGC CATCTCACC GGATTCACTC GTCACCTATG GTGATTCTC ACTTGATAAC
1501 CTTATTTTTC ACGAGGGGAA ATTAATAGGT TGTATTGATG TTGGACGAGT CGGAATCGCA
1561 GACCGATACC AGGATCTTGC CATCTATG GAACTGCCCTG GTGAGTTTTC TCCTTCATTA
1621 CAGAAACGGC TTTTCAAAA ATATGGTATT GATAATCCTG ATATGAATAA ATTGCAAGTT
1681 CATTGATGC TCGATGAGTT TTTCTAATCA GAATTGGTTA ATTGGTTGTA ACATTATTCA
1741 GATTGGGCCC CGTTCACCTG AGCGTCAGAC CCCGTAGAAA AGATCAAAGG ATCTTCTTGA
1801 GATCCTTTT TCTGCGCGT AATCTGCTGC TTGCAACAAA AAAAACCACC GCTACCAGCG
1861 GTGGTTTGTG TGCCGGATCA AGAGCTACCA ACTCTTTTTC CGAAGGTAAC TGGCTTCAGC
1921 AGAGCGCAGA TACCAAAATC TGTCTTCTA GTGTAGCCGT AGTTAGGCCA CCACTTCAAG
1981 AACTCTGTAG CACCGCTTAC ATACCTCGCT CTGCTAATCC TGTTACCAGT GGCTGCTGCC
2041 AGTGCGGATA AGTCGTGTCT TACCGGGTTG GACTCAAGAC GATAGTTACC GGATAAGGCG
2101 CAGCGGTCGG GCTGAACGGG GGGTTCGTGC ACACAGCCCA GCTTGGAGCG AACGACCTAC
2161 ACCGAACCTGA GATACCTACA GCGTGAGCTA TGAGAAAGCG CCACGCTTCC CGAAGGAGAG
2221 AAGGCGGACA GGTATCCGGT AAGCGGCAGG GTCGGAACAG GAGAGCGCAC GAGGGAGCTT
2281 CCAGGGGAAA ACGCCTGGTA TCTTTATAGT CCTGTGCGGT TTCGCCACCT CTGACTTGAG
2341 CGTCGATTTT TGTGATGCTC GTCAGGGGGG CGGAGCCTAT GAAAAAACGC CAGCAACGCG
2401 GCCTTTTAC GGTTCCTGGC CTTTGTCTGG CTTTGTCTC ACATGTCTT TCCTGCGTTA
2461 TCCCTGATT CTGTGGATAA CCGTATTACC GCTAGCATGG ATCTCGGGGA CGTCTAACTA
2521 CTAAGCGAGA GTAGGGAAGT GCCAGGCATC AAATAAAACG AAAGGCTCAG TCGGAAGACT
2581 GGGCCTTTTC TTTTATCTGT TGTGTGTCGG TGAACGCTCT CCTGAGTAGG ACAAATCCGC
2641 CGGGAGCGGA TTTGAACGTT GTGAAGCAAC GGCCCGGAGG GTGGCGGGCA GGACGCCCGC
2701 CATAAACTGC CAGGCATCAA ACTAAGCAGA AGGCCATC

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FIGURE 16B

24/240

Figure 17A: Cloning Sites of the *ENTY* Vector, *PEUTRB*

25/240

pENTR8 2735 bp

Location (Base Nos.)	Gene Encoded
67..166	attL1
339..644	ccdB
673..772	attL2
895..1704	KmR
1809..2382	ori

```

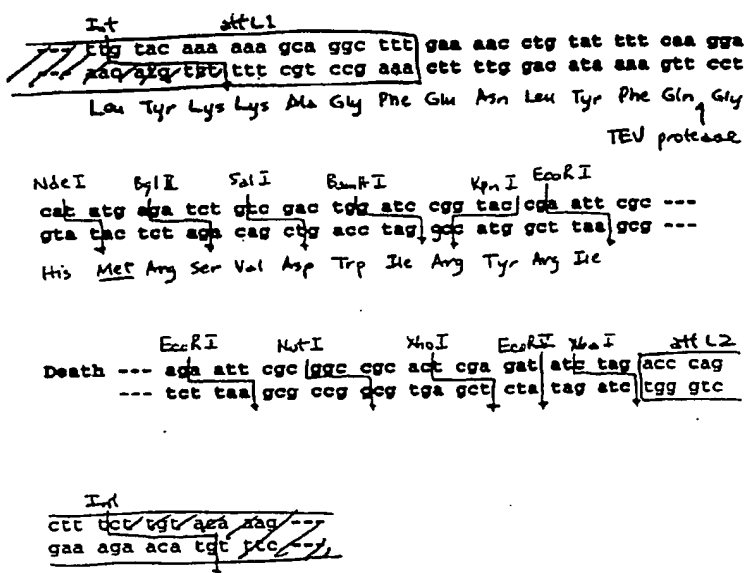
1 CTGACGGATG GCCTTTTTCG GTTCTACAA ACTCTTCCTG TTAGTTAGTT ACTTAAGCTC
61 GGGCCCCCAA TAATGATTTT ATTTTGA CTG ATAGTGACCT GTTCGTTGCA ACAATTTGAT
121 AAGCAATGCT TTTTATAAT GCCAACTTTG TACAAAAAAG CAGGCTTTGA AAACCTGTAT
181 TTTCAAGGAA CCATGGACCT AGTCGACTGG ATCCGGTACC GAATTCGCTT ACTAAAAGCC
241 AGATAACAGT ATGCGTATTT GCGCGCTGAT TTTTGGCGTA TAAGAATATA TACTGATATG
301 TATACCCGAA GTATGTCRAA AAGAGGTGTG CTTCTAGAAT GCAGTTTAAG GTTTACACCT
361 ATAAAAGAGA GAGCCGTTAT CGTCTGTTTG TGGATGTACA GAGTGATATT ATTGACACGC
421 CCGGGCGACG GATAGTGATC CCCCTGGCCA GTGCACGTCT GCTGTGATG AAAGTCTCCC
481 GTGAACCTTA CCGCGTGGTG CATATCGGGG ATGAAAGCTG GCGCATGATG ACCACCGATA
541 TGGCCAGTGT GCCGGTCTCC GTTATCGGGG AAGAAGTGGC TGATCTCAGC CACCGCGAAA
601 ATGACATCAA AAACGCCATT AACCTGATGT TCTGGGGAAT ATAGAATTCC GCGCGCACT
661 CGAGATATCT AGACCCAGCT TTCTGTGACA AAGTTGGCAT TATAAGAAAG CATTGCTTAT
721 CAATTTGTTG CAACGAACAG GTCACATCA GTCAAAATAA AATCATTATT TGCCATCCAG
781 CTGCAGCTCT GCGCCGTGTC TCAAAATCTC TGATGTTACA TTGCACAAGA TAAAAATATA
841 TCATCATGAA CAATAAACT GTCTGCTTAC ATAAACAGTA ATACAAGGGG TGTATGAGC
901 CATATTCAAC GGGAAACGTC GAGGCCGCGA TTAATTTCCA ACATGGATGC TGATTTATAT
961 GGGTATAAAT GGGCTCGCGA TAATGTCGGG CAATCAGGTG CGACAATCTA TCGCTTGTAT
1021 GGGAAAGCCCG ATGCGCCAGA GTTGTTCCTG AAACATGGCA AAGGTAGCGT TGCCATGAT
1081 GTTACAGATG AGATGGTCAG ACTAACTGG CTGACGGAAT TTATGCTCTT TCCGACCATC
1141 AAGCATTTTA TCCGTACTCC TGATGATGCA TGGTTACTCA CCACTGCGAT CCCCAGAAA
1201 ACAGCATTCC AGGTATTAGA AGAATATCCT GATTCAGGTG AAAATATTGT TGATGCGCTG
1261 GCAGTGTCCC TGCGCCGGTT GCATTGATT CTTGTTTGTG ATTGTCCTTT TAACAGCGAT
1321 CGCGTATTTT GTCTGCTCA GCGCAATCA CGAATGAATA ACGGTTTGGT TGATGCGAGT
1381 GATTTTGATG ACGAGCGTAA TGGCTGCGCT GTTGAACAAG TCTGGAAAGA AATGCATAAA
1441 CTTTGGCCAT TCTCACCGBA TTCAGTCGTC ACTCATGGTG ATTTCTCACT TGATAACCTT
1501 ATTTTGTACG AGGGGAAATT AATAGGTTGT ATTGATGTTG GACGAGTCGG AATCGCAGAC
1561 CGATACCAGG ATCTTGCCAT CCTATGGAAC TGCTCGGTG AGTTTCTTCC TTCATTACAG
1621 AAACGGCTTT TTCAAAAATA TGGTATTGAT AATCCTGATA TGAATAAATT GCAGTTTCAT
1681 TTGATGCTCG ATGAGTTTTT CTAATCAGAA TTGGTTAATT GGTGTGAACA TTATTCAGAT
1741 TGGGCCCCGT TCCACTGAGC GTCAGACCCC GTAGAAAAGA TCAAAAGGATC TTCTTGAGAT
1801 CCTTTTTTTC TGCGCGTAAT CTGCTGCTTG CAAACAAAAA AACCACCGCT ACCAGCGGTG
1861 GTTTGTTTGC CGGATCAAGA GCTACCAACT CTTTTTCCGA AGGTAACCTG CTTCAGCAGA
1921 GCGCAGATAC CAAATACTGT TCTTCTAGTG TAGCCGTAGT TAGGCCACCA CTTCAAGAAC
1981 TCTGTAGCAC CGCTACATA CCTCGCTCTG CTAATCCTGT TACCACTGGC TGCTGCCAGT
2041 GCGGATAAGT CGTGTCTTAC CGGGTTGGAC TCAAGACGAT AGTTACCGGA TAAGGCGCAG
2101 CGGTGCGGCT GAACGGGGGG TTCGTGCACA CAGCCGAGCT TGGAGCGAAC GACCTACACC
2161 GAACTGAGAT ACCTACAGCG TGAGCTATGA GAAAGCGCCA CGCTTCCCGA AGGGAGAAAG
2221 GCGGACAGGT ATCCGTAAG CCGCAGGGTC GGAACAGGAG AGCGCACGAG GGAGCTTCCA
2281 GGGGGAAACG CCTGGTATCT TTATAGTCCT GTCGGGTTTC GCCACCTCTG ACTTGAGCGT
2341 CGATTTTGTG GATGCTCGTC AGGGGGGCGG AGCCTATGGA AAAACGCCAG CAACGCGGCC
2401 TTTTACGGT TCTTGGCCTT TTGCTGGCCT TTTGCTCACA TGTTCTTTCC TGCGTTATCC
2461 CCTGATTCTG TGGATAACCG TATTACCGCT AGCATGGATC TCGGGGACGT CTAACCTACTA
2521 AGCGAGAGTA GGGAACTGCC AGGCATCAAA TAAAACGAAA GGCTCAGTCG GAAGACTGGG
2581 CCTTTCGTTT TATCTGTTGT TTGTCGGTGA ACGCTCTCCT GAGTAGGACA AATCCGCGG
2641 GAGCGGATTT GAACGTTGTG AAGCAACGGC CCGGAGGGTG GCGGGCAGGA CGCCCGCCAT
2701 AAACGCCCAG GCATCAAACT AAGCAGAAGG CCATC

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Figure 17B

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Figure 18: Cloning sites of the ENTRY Vector pENTRY



27/240

pENTR9 2735 bp

Location (Base Nos.)	Gene Encoded
67..166	attL1
339..644	ccdB
673..772	attL2
895..1704	KmR
1809..2382	ori

```

1 CTGACGGATG GCCTTTTTCG GTTCTTACAA ACTCTTCCTG TTAGTTAGTT ACTTAAGCTC
61 GGGCCCCCAA TAATGATTTT ATTTTGACTG ATAGTGACCT GTTCGTTGCA ACAAAATTGAT
121 AAGCAATGCT TTTTATAAT GCCAACTTTC TACRAAAAAG CAGGCTTTGA AAACCTGTAT
181 TTTCAAGGAC ATATGAGATC TGTCGACTGG ATCCGGTACC GAATTCGCTT ACTAAAGGCC
241 AGATAACAGT ATGCGTATTT GCGCGCTGAT TTTTGGCGTA TAAGAAATATA TACTGATATG
301 TATACCCGAA GTATGTCAAA AAGAGGTGTG CTTCTAGAAT GCAGTTTAAG GTTTACACCT
361 ATAAAAGAGA GAGCCGTTAT CGTCTGTTTG TGGATGTACA GAGTGATATT ATTGACACGC
421 CCGGGCGACG GATAGTGATC CCCCTGGCCA GTGCACGCTC GCTGTCAGAT AAAGTCTCCC
481 GTGAACCTTA CCCGGTGGTG CATATCGGGG ATGAAAGCTG GCGCATGATG ACCACCGATA
541 TGGCCAGTGT GCCGGTCTCC GTTATCGGGG AAGAAGTGGC TGATCTCAGC CACCCGCAAA
601 ATGACATCAA AAACGCCATT AACCTGATGT TCTGGGGAAT ATAGAATTCG CGGCCGCAC
661 CGAGATATCT AGACCCAGCT TTCTTGATCA AAGTTGGCAT TATAAGAAAG CATTTGCTTAT
721 CAATTGTGTT CAACGAACAG GTCACATCA GTCAAAATAA AATCATTATT TGCCATCCAG
781 CTGCAGCTCT GGGCCGCTGC TCAAAATCTC TGATGTTACA TTGCACAAGA TAAAAATATA
841 TCATCATGAA CAATAAAACT GTCTGCTTAC ATAAACAGTA ATACAAGGGG TGTATGAGC
901 CATATTCAAC GGGAAACGTC GAGGCCGCGA TTAAATTCCA ACATGGATGC TGATTTATAT
961 GGGTATAAAT GGGCTCGCGA TAATGTCGGG CAATCAGGTG CGACAATCTA TCGCTTGTAT
1021 GGGAAAGCCC ATGCGCCAGA GTTGTTCCTG AAACATGGCA AAGGTAGCGT TGCCAATGAT
1081 GTTACAGATG AGATGGTCAG ACTAACTGG CTGACGGAAT TTATGCTCTC TCCGACCATC
1141 AAGCATTTTA TCCGTACTCC TGATGATGCA TGGTTACTCA CCATGCGCAT CCCCAGGAAA
1201 ACAGCATTCC AGGTATTAGA AGAATATCCT GATTCAGGTG AAAATATTGT TGATGCGCTG
1261 GCAGTGTCCC TGCGCCGTTT GCATTGCTG CCTGTTTGTA ATTGTCCTTT TAACAGCGAT
1321 CGCGTATTTT GTCTCGCTCA GCGCAATCA CGAATGAATA ACGGTTTGGT TGATGCGAGT
1381 GATTTTGATG ACGAGCGTAA TGGCTGGCCT GTTGAACAAG TCTGGAAAGA AATGCATAAA
1441 CTTTTGCCAT TCTCACCAGA TTCAGTCGTC ACTCATGGTG ATTTCTCACT TGATAACCTT
1501 ATTTTGGACG AGGGGAAATT AATAGGTTGT ATTGATGTTG GACGAGTCGG AATCGCAGAC
1561 CGATACCAGG ATCTTGCCAT CCTATGGAAC TGCCTCGGTG AGTTTTCTCC TTCATTACAG
1621 AAACGGCTTT TTCAAAAATA TGGTATTGAT AATCCTGATA TGAATAAATT GCAGTTTCAT
1681 TTGATGCTCG ATGAGTTTTC CTAATCAGAA TTGGTTAATT GGTGTGAACA TTATTCAGAT
1741 TGGGCCCCGT TCCACTGAGC GTCAGACCCC GTAGAAAAGA TCAAAGGATC TTCTTGAGAT
1801 CCTTTTTTTC TGCGCGTAAT CTGCTGCTTG CAAACAAAAA AACCACCGCT ACCAGCGGTG
1861 GTTTGTTTGC CGGATCAAGA GCTACCAACT CTTTTCCGA AGGTAAGTGG CTTCAGCAGA
1921 GCGCAGATAC CAAATACTGT TCTTCTAGTG TAGCCGTAGT TAGGCCACCA CTTCAAGAAC
1981 TCTGTAGCAC CGCCTACATA CCTCGCTCTG CTAATCCTGT TACCAGTGGC TGCTGCCAGT
2041 GCGGATAAGT CGTGCTTAC CGGGTTGGAC TCAAGACGAT AGTTACCGGA TAAGGCGCAG
2101 CGGTCGGGCT GAACGGGGGG TTCTGTGACA CAGCCCAGCT TGGAGCGAAC GACCTACACC
2161 GAACTGAGAT ACCTACAGCG TGAGCTATGA GAAAGCGCCA CGCTTCCCGA AGGGAGAAAG
2221 GCGGACAGGT ATCCGTAAG CGGCAGGGTC GGAACAGGAG AGCGCACGAG GGAGCTTCCA
2281 GGGGAAACG CCTGGTATCT TTATAGTCCT GTCGGGTTTC GCCACCTCTG ACTTGAGCGT
2341 CGATTTTGTG GATGCTCGTC AGGGGGGCGG AGCCTATGGA AAAACGCCAG CAACCGCGCC
2401 TTTTACGGT TCCTGGCCTT TTGCTGGCCT TTTGCTCACA TGTTCTTTCC TGCGTTATCC
2461 CCTGATTCTG TGGATAACCG TATTACCGCT AGCATGGATC TCGGGGACGT CTAACCTACTA
2521 AGCGAGAGTA GGGAACTGCC AGGCATCAAA TAAAACGAAA GGCTCAGTCG GAAGACTCGGG
2581 CCTTTCTGTT TATCTGTTGT TTGTCGGTGA ACGCTCTCCT GAGTAGGACA AATCCGCGGG
2641 GAGCGGATTT GAACGTTGTG AAGCAACGGC CCGGAGGGTG GCGGGCAGGA CGCCCGCCAT
2701 AAATGCCAG GCATCAAACT AAGCAGAAGG CCATC

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FIGURE 18B

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Figure 19A: Cloning sites of the ENTRY Vector pENTRY10

Int attC1 S.D. - 12 Nde

--- ~~cdg tac aaa aaa gca ggc gtc gaa cta agg aaa tac tta cgt~~ ---

--- ~~aac atg gtc ttt cgt ccg agc ctt gat tcc ttt atg aat gta~~ ---

Leu Tyr Lys Lys Ala Gly Phe Glu Leu Arg Lys Tyr Leu His

K3 Xba Sal Bam Kpn EcoRI

atg gga acc aat tca gtc gac tgg atc cgg tac cga att cgc ---

tac cct tgg tta agt cag cgg acc tag gcp atg gct taa gcg ---

Met Gly Thr Asn Ser Val Asp Trp Ile Arg Tyr Arg Ile

EcoRI Not Xho EcoRII Xba attC2

Death --- aga att cgc ggc cgc act cga gat atc tag acc cag

(ccdB) --- tct taa gcg cgg ggg tga gct cta tag atc tgg gtc

Int

cct ttt ggg aca gag ---

gaa aga aca tgg ttc ---

pENTR10 2738 bp

Location (Base Nos.)	Gene Encoded
67..166	attL1
342..647	ccdB
676..775	attL2
898..1707	KmR
1812..2385	ori

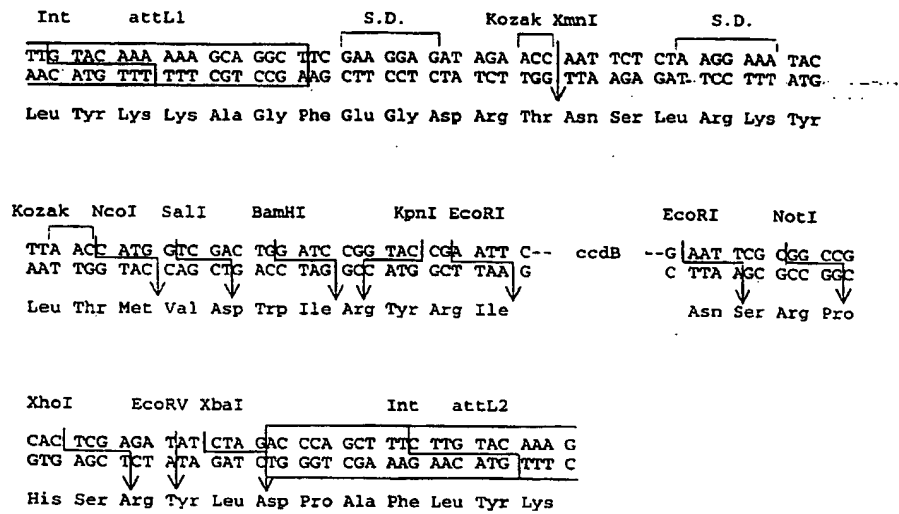
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61 GGGCCCCAAA TAATGATTTT ATTTTGACTG ATAGTGACCT GTTCGTTGCA ACAATTTGAT
121 AAGCAATGCT TTTTATAAT GCCAACTTTG TACAAAAAAG CAGGCTTCGA ACTAAGGAAA
181 TACTTACATA TGGGAACCAA TTCAGTCGAC TGGATCCGGT ACCGAATTCG CTTACTAAAA
241 GCCAGATAAC AGTAGTCGTA TTTGCGCGCT GATTTTTCG GTATAAGAAT ATATACTGAT
301 ATGTATACCC GAAGTATGTC AAAAAGAGGT GTGCTTCTAG AATGCAGTTT AAGGTTTACA
361 CCTATAAAG AGAGAGCCGT TATCGTCTGT TTGTGGATGT ACAGAGTGAT ATTATTGACA
421 CGCCCCGGCG ACGGATGGTG ATCCCCCTGG CCAGTGCACG TCTGCTGTCA GATAAAGTCT
481 CCCGTGAAC TTAACCGGTG GTGCATATCG GGGATGAAAG CTGGCGCATG ATGACCACCG
541 ATATGGCCAG TGTGCGGTC TCCGTTATCG GGAAGAAGT GGCTGATCTC AGCCACCGCG
601 AAAATGACAT CAAAAACGCC ATTAACCTGA TGTCTGGGG AATATAGAAT TCGCGGCCGC
661 ACTCGAGATA TCTAGACCCA GCTTCTTGT ACAAAGTTGG CATTATAAGA AAGCATTGCT
721 TATCAATTTG TTGCAACGAA CAGGTCAC TAAGTCAAAA TAAATCATT ATTTGCCATC
781 CAGCTGCAGC TCTGCGCCGT GTCTCAAAAT CTCTGATGTT ACATTGCACA AGATAAAAAAT
841 ATATCATCAT GAACAATAAA ACTGTCTGCT TACATAACA GTAATACAAG GGGTGTATG
901 AGCCATATTC AACGGGAAAC GTCGAGGCCG CGATTAAAT CCAACATGGA TGCTGATTTA
961 TATGGGTATA AATGGGCTCG CGATAATGTC GGGCAATCAG GTGCGACAAT CTATCGCTTG
1021 TATGGGAAGC CCGATGCGCC AGAGTTGTTT CTGAAACATG GCAAAGGTAG CGTTGCCAAT
1081 GATGTTACAG ATGAGATGGT CAGACTAAAC TGGCTGACCG AATTTATGCC TCTCCGACC
1141 ATCAAGCATT TTATCCGTAC TCCTGATGAT GCATGGTTAC TCACCACTGC GATCCCCGGA
1201 AAAACAGCAT TCCAGGTATT AGAAGAATAT CCTGATTCAG GTGAAAAATAT TGTGATGCG
1261 CTGGCAGTGT TCCTGCGCCG GTTGCAATCG ATTCTGTTT GTAATTGTCC TTTTAACAGC
1321 GATCGCGTAT TTCGTCTCGC TCAGGCGCAA TCACGAATGA ATAACGGTTT GGTGATGCG
1381 AGTGATTTTG ATGACGAGCG TAATGGCTGG CCTGTGAAC AAGTCTGGAA AGAATGTCAT
1441 AAACTTTTCG CATTCTCACC GGATTCAATG GTCACATCAT GTGATTCTC ACTTGATAAC
1501 CTTATTTTTC ACGAGGGGAA ATTAATAGGT TGTATTGATG TTGGACGAGT CGGAATCGCA
1561 GACCGATACC AGGATCTTGC CATCCTATGG AACTGCCTCG GTGAGTTTTC TCCTTCATTA
1621 CAGAAACGGC TTTTCAAAA ATATGGTATT GATAATCCTG ATATGAATAA ATTGCAGTTT
1681 CATTTGATGC TCGATGAGTT TTTCTAATCA GAATTGGTTA ATTGGTTGTA ACAATTATTC
1741 GATTGGGCCC CGTTCCACTG AGCGTCAGAC CCCGTAGAAA AGATCAAAGG ATCTTCTTGA
1801 GATCCTTTTT TTCTGCGCGT AATCTGCTGC TTGCAACAA AAAAACCACC GCTACCAGCG
1861 GTGGTTTGTG TGCCGGATCA AGAGCTACCA ACTCTTTTTC CGAAGGTAAC TGGCTTCAGC
1921 AGAGCGCAGA TACCAATATC TGTCTTCTA GTGTAGCCGT AGTTAGGCCA CCACTTCAAG
1981 AACTCTGTAG CACCGCCTAC ATACCTCGCT CTGCTAATCC TGTTACCAGT GGCTGCTGCC
2041 AGTGGCGATA AGTCGTGTCT TACCGGGTTG GACTCAAGAC GATAGTTACC GGATAAGGCG
2101 CAGCGGTTCG GCTGAACGGG GGGTTCGTGC ACACAGCCCA GCTTGGAGCG AACGACCTAC
2161 ACCGAACCTG GATACCTACA GCGTGAGCTA TGAGAAAGCG CCACGCTTCC CGAAGGGAGA
2221 AAGGCGGACA GGTATCCGGT AAGCGGCAGG GTCGGAACAG GAGAGCGCAC GAGGGAGCTT
2281 CCAGGGGGAA ACGCCTGGTA TCTTTATAGT CCTGTCCGGT TTCGCCACCT CTGACTTGAG
2341 CGTCGATTTT TGTGATGCTC GTCAGGGGGG CGGAGCCTAT GGAACAAACG CAGCAACGCG
2401 GCCTTTTCAC GGTTCCTGGC CTTTGTCTGG CCTTTGCTC ACATGTTCTT TCCTGCGTTA
2461 TCCCCTGATT CTGTGGATAA CCGTATTACC GCTAGCATGG ATCTCGGGGA CGTCTAATA
2521 CTAAGCGAGA GTAGGGAACT GCCAGGCATC GAATAAAACG AAAGGCTCAG TCGGAAGACT
2581 GGGCCTTTTC TTTTATCTGT TGTTTGTCGG TGAACGCTCT CCTGAGTAGG ACAATCCGC
2641 CGGGAGCGGA TTTGAACGTT GTGAAGCAAC GGCCCGGAGG GTGGCGGGCA GGACGCCCGC
2701 CATAAATGTC CAGGCATCAA ACTAAGCAGA AGGCCATC

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FIGURE 19B

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Figure 20A: Cloning Sites of the Entry Vector pENTR11

pENTR11 2744 bp (rotated to position 2578)

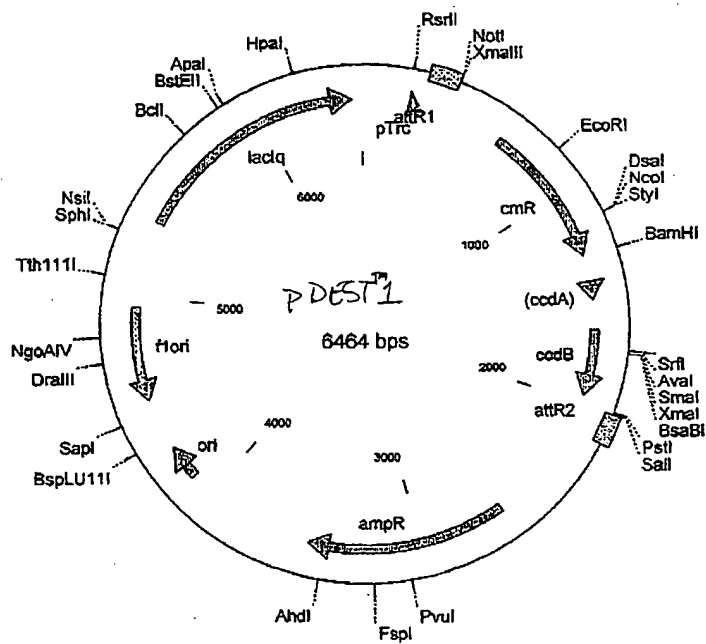
<u>Location (Base Nos.)</u>		<u>Gene Encoded</u>
67..166		attL1
348..653		ccdB
683..781		attL2
904..1713		KmR
1818..2391		ori

1	CTGACGGATG	GCCTTTTTCG	GTTTCTACAA	ACTCTTCCTG	TTAGTTAGTT	ACTTAAGCTC
61	GGGCCCCAAA	TAATGATTTT	ATTTTGACTG	ATAGTGACCT	GTTTCGTTGCA	ACAAATTGAT
121	AAGCAATGCT	TTTTTATAAT	GCCAACTTTG	TACAAAAAAG	CAGGCTTCGA	AGGAGATAGA
181	ACCAATTCTC	TAAGGAAATA	CTTAACCATG	GTCGACTGGA	TCCGGTACCG	AATTTCGTTA
241	CTAAAAGCCA	GATAACAGTA	TGCGTATTTG	CGCGCTGATT	TTTGCGGTAT	AAGAAATATAT
301	ACTGATATGT	ATACCCGAAG	TATGTCAAAA	AGAGGTGTGC	TTCTAGAATG	CAGTTTAAGG
361	TTTACACCTA	TAAAAGAGAG	AGCCGTTATC	GTCTGTTTGT	GGATGTACAG	AGTGATATTA
421	TTGACACGCC	CGGGCGACGG	ATAGTGATCC	CCCTGGCCAG	TGCACGTCGT	CTGTGAGATA
481	AAGTCTCCCG	TGAACTTTAC	CCGGTGGTGC	ATATCGGGGA	TGAAAGCTGG	CGCATGATGA
541	CCACCGATAT	GGCCAGTGTG	CCGGTCTCCG	TTATCGGGGA	AGAAGTGGCT	GATCTCAGCC
601	ACCGCGAAAA	TGACATCAAA	AACGCCATTA	ACCTGATGTT	CTGGGGAATA	TAGAATTCGC
661	GGCCGCACTC	GAGATATCTA	GACCCAGCTT	TCTTGATCAA	AGTTGGCATT	ATAAGAAAGC
721	ATTGCTTATC	AAITTTGTTC	AACGAACAGG	TCACATATCAG	TCAAAAATAA	ATCATTATTT
781	GCCATCCAGC	TGCAGCTCTG	GCCCGTGTCT	CAAAATCTCT	GATGTTACAT	TGCACAAGAT
841	AAAAATATAT	CATCATGAAC	AATAAAACTG	TCTGCTTACA	TAAACAGTAA	TACAAGGGGT
901	GTTATGAGCC	ATATTCACAG	GGAAACGTCG	AGGCCGCGAT	TAAATTCCAA	CATGGATGCT
961	GATTTATATG	GGTATAAATG	GGCTCGCGAT	AATGTCGGGC	AATCAGGTGC	GACAACTTAT
1021	CGCTTGTATG	GGAAAGCCCGA	TGCGCCAGAG	TTGTTTCTGA	AACATGGCAA	AGGTAGCGTT
1081	GCCAATGATG	TTACAGATGA	GATGGTCAGA	CTAACTGGC	TGACGGAAT	TATGCCTCTT
1141	CCGACCATCA	AGCATTTTAT	CCGTACTCCT	GATGATGCAT	GGTTACTCAC	CACCTGCGATC
1201	CCCGGAAAAA	CAGCATTCCA	GGTATTAGAA	GAATATCCTG	ATTTCAGGTGA	AAATATTGTT
1261	GATGCGCTGG	CAGTGTTCCT	GCGCCGGTTG	CATTGCAATC	CTGTTTGTAA	TTGTCCTTTT
1321	AACAGCGATC	GCCTATTTCG	TCTCGCTCAG	GCGCAATCAC	GAATGAATAA	CGGTTTGGTT
1381	GATGCGAGTG	ATTTTGATGA	CGAGCGTAAT	GGCTGGCCTG	TTGAACAAGT	CTGGAAAGAA
1441	ATGCATAAAC	TTTTTGCCATT	CTCACCGGAT	TCAGTCGTCA	CTCATGGTGA	TTTCTCACTT
1501	GATAACCTTA	TTTTTGACGA	GGGGAAATTA	ATAGGTTGTA	TTGATGTTGG	ACGAGTCGGA
1561	ATCGCAGACC	GATACCAGGA	TCTTGCCATC	CTATGGAAT	GCCTCGGTGA	GTTTTCTCCT
1621	TCATTACAGA	AACGGCTTTT	TCAAAAATAT	GGTATTGATA	ATCCTGATAT	GAATAAATTG
1681	CAGTTTCATT	TGATGCTCGA	TGAGTTTTC	TAATCAGAAT	TGGTTAATTG	GTTGTAACAT
1741	TATTCAGATT	GGGCCCCGTT	CCACTGAGCG	TCAGACCCCG	TAGAAAAGAT	CAAAGGATCT
1801	TCTTGAGATC	CTTTTTCCT	GCGCGTAATC	TGCTGCTTGC	AAACAAAAAA	ACCACCGCTA
1861	CCAGCGGTGG	TTTGTTCGCC	GGATCAAGAG	CTACCAACTC	TTTTTCCGAA	GGTAAGTGGC
1921	TTCAGCAGAG	CGCAGATACC	AAATACTGTT	CTTCTAGTGT	AGCCGTAGTT	AGGCCACCAC
1981	TTCAAGAACT	CTGTAGCACC	GCCTACATAC	CTCGCTCTGC	TAATCCTGTT	ACCAAGTGGCT
2041	GCTGCCAGTG	GCGATAAGTC	GTGTCTTACC	GGGTTGGAAT	CAAGACGATA	GTTACCGGAT
2101	AAGGCGCAGC	AGTCGGGCTG	AACGGGGGGT	TCGTGCACAC	AGCCAGCTT	GGAGCGAACG
2161	ACCTACACCG	AACAGAGATA	CCTACAGCGT	GAGCTATGAG	AAAGCGCCAC	GCCTCCCGAA
2221	GGGAGAAAGG	CGGACAGGTA	TCCGGTAAGC	GGCAGGTCG	GAACAGGAGA	GCGCACGAGG
2281	GAGCTTCAG	GGGGAAACGC	CTGGTATCTT	TATAGTCCTG	TCGGGTTTCG	CCACCTCTGA
2341	CTTGAGCGTC	GATTTTGTG	ATGCTCGTCA	GGGGGGCGGA	GCCTATGGAA	AAACGCCAGC
2401	AACGCGGCCCT	TTTTACGGTT	CCTGGCCCTT	TGCTGGCCCT	TTGCTCACAT	GTTCTTTCCT
2461	GCGTTATCCC	CTGATTCTGT	GGATAACCGT	ATTACCGCTA	GCATGGATCT	CGGGGACGTC
2521	TAACTACTAA	GCGAGAGTAG	GGAAGTCCCA	GGCATCAAT	AAAACGAAAG	GCTCAGTCGG
2581	AAGACTGGGC	CTTTCGTTTT	ATCTGTTGTT	TGTCGGTGAA	CGCTCTCCTG	AGTAGGACAA
2641	ATCCGCCGGG	AGCGGATTG	AACGTTGTGA	AGCAACGGCC	CGGAGGGTGG	CGGGCAGGAC
2701	GCCCCCATA	AACAGCCAGG	CATCAACTA	AGCAGAAGGC	CATC	

FIGURE 20B

Figure 2: LA:pDEST1 Native Protein Expression in E. coli

⁻³⁵ Trc promoter ⁻¹⁰ ^{RNA}
 1 atgagctgtt gacgattaat catccggctc gataaattg tgggaattgtg agcggataac
 tactcgacaa ctgttaatta gtaggcggag catattacac accttaacac tcgcttattg
 61 aatttcacac aggaacaga caggatatagg atcacaagtt ttttgaada agctgaaga
 ttaaagtgtg tcccttgctc gtccatatcc tagtttcaa acatgtttt tgcatttct



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pDEST1 6464 bp

<u>Location (Base Nos.)</u>	<u>Gene Encoded</u>
216..257	Trc promoter
397..273	attR1
647..1306	CmR
1426..1510	inactivated ccdA
1648..1953	ccdB
1994..2118	attR2
2598..3503	ampR
4104..4264	ori
4504..4941	flori (fl intergenic region)
5340..6420	lacIq

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1  GTTTGACAGC TTATCATCGA CTGCACGGTG CACCAATGCT TCTGGCGTCA GGCAGCCATC
61  GGAAGCTGTG GTATGGCTGT GCAGGTCGTA AATCACTGCA TAATTCGTGT CGCTCAAGGC
121 GCACTCCCGT TCTGGATAAT GTTTTTTGCG CCGACATCAT AACGGTTCTG GCAAAATATTC
181 TGAATGAGC TGTTGACAAT TAATCATCCG GTCGGTATAA TCTGTGGAAT TGTGAGCGGG
241 ATAACAATTT CATCGCGAGG TACCAAGCTA TCACAAGTTT GTACAAAAAA GCTGAACGAG
301 AAACGTAAAA TGATATAAAT ATCAATATAT TAAATTAGAT TTTGCATAAA AAACAGACTA
361 CATAATACTG TAAAAACAAA CATATCCAGT CACTATGGCG GCCGCTAAGT TGGCAGCATC
421 ACCCGACGCA CTTTGGCGCG AATAAATACC TGTGACGGAA GATCACTTCG CAGAATAAAT
481 AAATCCTGGT GTCCCTGTTG ATACCGGGAA GCCCTGGGCC AACTTTTGGC GAAATAGAGA
541 CGTTGATCGG CACGTAAAGG GTTCCAACTT TCACCATAAT GAAATAAGAT CACTACCGGG
601 CGTATTTTTT GAGTTATCGA GATTTTCAGG AGCTAAGGAA GCTAAAATGG AGAAAAAAT
661 CACTGGATAT ACCACCGTTG ATATATCCCA ATGGCATCGT AAAGAACATT TTGAGGCATT
721 TCAGTCAGTT GCTCAATGTA CCTATAACCA GACCGTTCAG CTGGATATTA CGGCCTTTTT
781 AAAGACCGTA AAAAAAATA AGCACAAGTT TTATCCGGCC TTTATTACCA TTCTTGCCCG
841 CCTGATGAAT GCTCATCCGG AATTCCGTAT GGCAATGAAA GACGGTGAGC TGGTGATATG
901 GGATAGTGTT CACCCTTGTT ACACCGTTTT CCATGAGCAA ACTGAAACGT TTTTCATCGT
961 CTGGAGTGAA TACCACGAGC ATTTCCGGCA GTTTCTACAC ATATATTGCG AAGATGTGGC
1021 GTGTTACGGT GAAAACTCGG CCTATTTCCC TAAAGGGTTT ATTGAGAATA TGTTTTTCTG
1081 CTCAGCCAAT CCCTGGGTGA GTTTCACCAG TTTTGATTTA AACGTGGCCA ATATGGACAA
1141 CTTCTTGCCG CCGGTTTTCA CCATGGGCAA ATATTATACG CAAGGCGACA AGGTGCTGAT
1201 GCCGCTGGCG ATTCAAGTTC ATCATGCCGT CTGTGATGGC TTCCATGTGC GCAGAATGCT
1261 TAATGAATTA CAACAGTACT GCGATGAGTG GCAGGGCGGG GCGTAAACGC GTGGATCCGG
1321 CTTACTAAAA GCCAGATAAC AGTATGCGTA TTTGCGCGCT GATTTTTCG GATATAAGAT
1381 ATATACTGAT ATGTATACCC GAAGTATGTC AAAAAAGAGT GTGCTATGAA GCAGCCTATT
1441 ACAGTGACAG TTGACACGGA CAGCTATCAG TTGCTCAAGG CATATATGAT GTCAATATCT
1501 CCGGCTTGGT AAGCACAACC ATGCAGAATG AAGCCCGTCG TCTGCGTGCC GAACGCTGGA
1561 AAGCGGAAAA TCAGGAAGGG ATGGCTGAGG TCGCCCGGTT TATTGAAATG AACGGCTCTT
1621 TTGCTGACGA GAACAGGGAC TGGTGAAATG CAGTTTAAAG TTTACACCTA TAAAGAGAG
1681 AGCCGTTATC GTCGTGTTGT GGATGTACAG AGTGATATTA TTGACACGCC CGGGCGACGG
1741 ATSGTGATCC CCCTGGCCAG TGCACGCTCG CTGTCAGATA AAGTCTCCCG TGAACCTTAC
1801 CCGGTGGTGC ATATCGGGGA TGAAAGCTGG CGCATGATGA CCACCGATAT GGCCAGTGTG
1861 CCGGTCTCCG TTATCGGGGA AGAAGTGGCT GATCTCAGCC ACCCGGAAAA TGACATCAAA
1921 AACGCCATTA ACCTGATGTT CTGGGGAATA TAAATGTCAG GCTCCCTTAT ACACAGCCAG
1981 TCTGCAGGTC GACCATAGTG ACTGGATATG TTGTGTTTTA CAGTATTATG TAGTCTGTTT
2041 TTTATGCAAA ATCTAATTTA ATATATTGAT ATTTATATCA TTTTACGTTT CTCGTTTCAGC
2101 TTTCTTGTAC AAAGTGGTGA TAGCTTGGCT GTTTTGGCGG ATGAGAGAAG ATTTTCAGCG
2161 TGATACAGAT TAAATCAGAA CGCAGAAGCG GTCTGATAAA ACAGAATTTG CCTGGCGGCA
2221 GTAGCGCGGT GGTCCACCT GACCCCATGC CGAACTCAGA AGTGAAACGC CGTAGCGCCG
2281 ATGGTAGTGT GGGGTCTCCC CATGCCGAGG TAGGGAACCT CCAGGCATCA AATAAAACGA
2341 AAGGCTCAGT CGAAAGACTG GGCCTTTCGT TTTATCTGTT GTTTGTGCGT GAACGCTCTC
2401 CTGAGTAGGA CAATCCGCC GGGAGCGGAT TTGAACGTTG CGAAGCAACG GCCCGGAGGG
2461 TGGCGGGCAG GACGCGCGCC ATAAACTGCC AGGCATCAAA TTAAGCAGAA GGCCATCTCTG
2521 ACGGATGGCC TTTTTCGTT TCTACAACT CTTTTCGTTT ATTTTCTCAA ATACATTCAA-

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FIGURE 2/B

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2581 ATATGTATCC GCTCATGAGA CAATAACCCT GATAAATGCT TCAATAATAT TGAAAAAGGA
 2641 AGAGTATGAG TATTCAACAT TTCCGTGTGCG CCTTATTCC CTTTTTTGCG GCATTTTGCC
 2701 TTCCTGTTTT TGCTCACCCA GAAACGCTGG TGAAAGTAAA AGATGCTGAA GATCAGTTGG
 2761 GTGCACGAGT GGGTTACATC GAACTGGATC TCAACAGCGG TAAGATCCTT GAGAGTTTTC
 2821 GCCCCGAAGA ACGTTTTCCA ATGATGAGCA CTTTTAAAGT TCTGCTATGT GCGCGGTAT
 2881 TATCCCGTGT TGACGCCGGG CAAGAGCAAC TCGGTGCGCG CATACACTAT TCTCAGAATG
 2941 ACTTGGTGTA GTACTCACCA GTCACAGAAA AGCATCTTAC GGATGGCATG ACAGTAAGAG
 3001 AATTATGCAG TGCTGCCATA ACCATGAGTG ATAACACTGC GGCCAACTTA CTCTGACAA
 3061 CGATCGGAGG ACCGAAGGAG CTAACCGCTT TTTTGCACAA CATGGGGGAT CATGTAACCTC
 3121 GCCTTGATCG TTGGGAACCG GAGCTGAATG AAGCCATACC AAACGACGAG CGTGACACCA
 3181 CGATGCCCTAC AGCAATGGCA ACAACGTTGC GCAACTATT AACTGGCGAA CTACTTACTC
 3241 TAGCTTCCCG GCAACAATTA ATAGACTGGA TGGAGGCGGA TAAAGTTGCA GGACCACTTC
 3301 TGCGCTCGGC CCTTCCGGCT GGCTGGTTTA TTGCTGATAA ATCTGGAGCC GGTGAGCGTG
 3361 GGTCTCGCGG TATCATTGCA GCCTGGGGC CAGATGGTAA GCCCTCCCGT ATCGTAGTTA
 3421 TCTACACGAC GGGGAGTCAG GCAACTATGG ATGAACGAAA TAGACAGATC GCTGAGATAG
 3481 GTGCCTCACT GATTAAGCAT TGGTAACTGT CAGACCAAGT TACTCATAT ATACTTTAGA
 3541 TTGATTTAAA ACTTCATTTT TAATTTAAAA GGATCTAGGT GAAGATCCTT TTTGATAATC
 3601 TCATGACCAA AATCCCTTAA CGTGAGTTTT CGTCCACTG AGCGTCAGAC CCTGTAGAAA
 3661 AGATCAAAGG ATCTTCTTGA GATCCTTTTT TTCTGCGCGT AATCTGCTGC TTGCAAAACA
 3721 AAAAACCACC GCTACCAGCG GTGGTTTGTG TGCCGGATCA AGAGCTACCA ACTCTTTTTT
 3781 CGAAGGTAAC TGGCTTCAGC AGAGCGCAGA TACCAATAC TGTCTTCTA GTGTAGCCGT
 3841 AGTTAGGCCA CCACTTCAAG AACTCTGTAG CACCGCCTAC ATACCTCGCT CTGCTAATCC
 3901 TGTTACCACT GGCTGCTGCC AGTGGCGATA AGTCGTGTCT TACCGGGTTG GACTCAAGAC
 3961 GATAGTTACC GGATAAGGCG CAGCGGTGCG GCTGAACGGG GGGTTCGTGC ACACAGCCCA
 4021 GCTTGAGCGG AACGACCTAC ACCGAACCTGA GATACCTACA GCGTGAGCTA TGAGAAAGCG
 4081 CCACGCTTCC CGAAGGGAGA AAGGCGGACA GGTATCCGGT AAGCGGCAGG GTCGGAACAG
 4141 GAGAGCGCAC GAGGGAGCTT CCAGGGGGAA ACGCTGTTA TCTTTATAGT CTTGTCGGGT
 4201 TTCCGCACCT CTGACTTGAG CGTCGATTTT TGTGATGCTC GTCAGGGGGG CGGAGCCTAT
 4261 GGAATAACGC CAGCAACGCG GCCTTTTCAC GGTTCCTGGC CTTTGTCTGG CTTTGTCTC
 4321 ACATGTTCTT TCCTGCGTTA TCCCCTGATT CTGTGGATAA CCGTATTACC GCCTTTGAGT
 4381 GAGCTGATAC CGCTCGCCGC AGCCGAACGA CCGAGCGCAG CGAGTCAGTG AGCGAGGAAG
 4441 CGGAAGAGCG CCTGATGCGG TATTTTCTCC TTACGCATCT GTGCGGTATT TCACACCGCA
 4501 TAATTTTGTG AAAATTTCGC TTAATTTTGT GTTAATACAG CTCATTTTTT AACCAATAGG
 4561 CCGAAATCGG CAAAATCCCT TATAAATCAA AAGAATAGAC CGAGATAGGG TTGAGTGTG
 4621 TTCCAGTTTG GAACAAGAGT CCACTATTAA AGAACGTGGA CTCCAACGTC AAAGGGCGAA
 4681 AAACCGTCTA TCAGGGCGAT GGCCCACTAC GTGAACCATC ACCCTAATCA AGTTTTTTGG
 4741 GGTGAGGTTG CCGTAAAGCA CTAAATCGGA ACCCTAAAGG GAGCCCCCGA TTTAGAGCTT
 4801 GACGGGGAAA GCCGGCGAAC GTGGCGAGAA AGGAAGGGAA GAAAGCGAAA GGAGCGGGCG
 4861 CTAGGGCGCT GGCAAGTGTA GCGGTACGCG TCGCGGTAAC CACCACACCC GCCGCGCTTA
 4921 ATGCGCCGCT ACAGGGCGCG TCCATTCCGC ATTGAGGCTG CTATGGTGCA CTCTCAGTAC
 4981 AATCTGCTCT GATGCGCAT AGTTAAGCCA GTACCAGTCA CGTAGCGATA TCGGAGTGTA
 5041 TACACTCCGC TATCGCTACG TGACTGGGTC ATGGCTGCGC CCCGACACCC GCCAACACCC
 5101 GCTGACGCGC CCTGACGGGC TTGTCTGCTC CCGGCATCCG CTTACAGACA AGCTGTGACC
 5161 GTCTCCGGGA GCTGCATGTG TCAGAGGTTT TCACCGTCAT CACCGAAACG CGCGAGGCAG
 5221 CAGATCAATT CGCGCGCGAA GGCGAAGCGG CATGCATTTA CGTTGACACC ATCGAATGGT
 5281 GCAAAACCTT TCGCGGTATG GCATGATAGC GCCCGGAAGA GAGTCAATTG AGGGTGGTGA
 5341 ATGTGAAACC AGTAACGTTA TACGATGTCG CAGAGTATGC CGGTGTCTCT TATCAGACCG
 5401 TTCCCGCGT GGTGAACCA GCGAGCCACG TTTCTGCGAA AACGCGGGAA AAAGTGGAG
 5461 CGCGATGGC GGAGCTGAAT TACATTCCCA ACCGCGTGGC ACAACAACCT GCGGGCAAAAC
 5521 AGTCGTTGCT GATTGGCGTT GCCACCTCCA GTCTGGCCCT GCACGCGCCG TCGCAAAATTG
 5581 TCGCGGCGAT TAAATCTCGC GCCGATCAAC TGGGTGCCAG CGTGGTGGTG TCGATGGTAG
 5641 AACGAAGCGG CGTCGAAGCC TGTAAAGCGG CGGTGCACAA TCTTCTCGCG CAACGCGCTT
 5701 GTGGGCTGAT CATTAACTAT CCGCTGGATG ACCAGGATGC CATTGCTGTG GAAGCTGCCT
 5761 GCACTAATGT TCCGGCGTTA TTTCTTGATG TCTCTGACCA GACACCCATC AACAGTATTA
 5821 TTTTCTCCCA TGAAGACGGT ACGCGACTGG GCGTGGAGCA TCTGGTGCBA TTGGGTCAAC
 5881 AGCAAAATCG GCTGTTAGCG GGCCCATTA GTTCTGTCTC GGCGCGTCTG CGTCTGGCTG
 5941 GCTGGCATAA ATATCTCACT CGCAATCAAA TTCAGCCGAT AGCGGAACGG GAAGGCGACT
 6001 GGAGTGCCAT GTCCGGTTTT CAACAAACCA TGCAAAATGCT GAATGAGGGC ATCGTTCCCA-

Figure 21C

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6061 CTGCGATGCT GGTGCCAAC GATCAGATGG CGTGGGCGC AATGCGGCC ATTACCGAGT
6121 CCGGGCTGCG CGTTGGTGCG GATATCTCGG TAGTGGGATA CGACGATACC GAAGACAGCT
6181 CATGTTATAT CCCGCCGTTA ACCACCATCA AACAGGATT TCGCCTGCTG GGGCAAACCA
6241 GCGTGGACCG CTTGCTGCAA CTCTCTCAGG GCCAGGCGGT GAAGGGCAAT CAGCTGTTGC
6301 CCGTCTCACT GGTGAAAAGA AAAACCACCC TGGCACCCAA TACGCAAACC GCCTCTCCCC
6361 GCGCGTTGCG CGATTCAATTA ATGCAGCTGG CACGACAGGT TTCCCGACTG GAAAGCGGGC
6421 AGTGAGCGCA ACGCAATTAA TGTGAGTTAG CGCGAATTGA TCTG

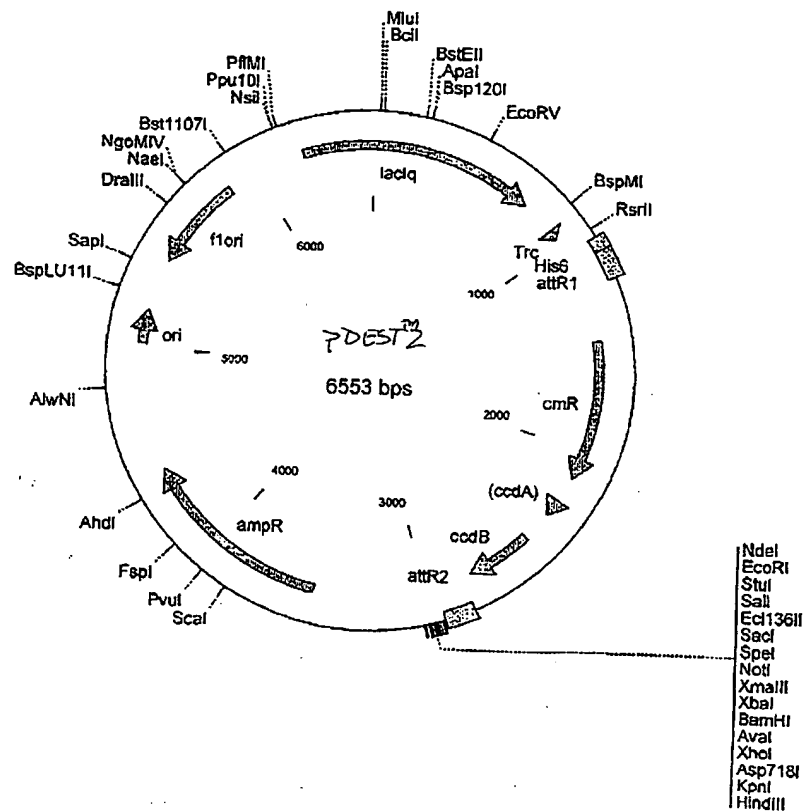
FIGURE 21D

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Figure 22A: pDEST2

His6 fusions in E. coli

970 aat att ctg aaa tga gct ⁻³⁵ gtt gac aat taa tca tcc ggt ccg ⁻¹⁰ cat aat ctg
 tta taa gac ttt act cga caa ctg tta att agt agg cca ggc ata tta gac
 1021 tgg ^{RNA} aat tgt gag cgg ata aca att tca cac agg aaa cag acc ^{Met Ser Tyr} atg tcg tac
 acc tta aca ctc gcc tat tgt taa agt gtg tcc ttt gtc tgg tac agc atg
 1072 ^{Tyr His His His His His His} tac cat cac cat cat cat cat ^{Glu Ile} gaa agt tgg ^{cap} ^{attR1} ~~aaa gca gca~~
 atg gta gtg gta gtg gta gtg ccg tag ~~tgt tca aac atg ttt cty caa cxt~~



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pDEST2 6553 bp

<u>Location (Base Nos.)</u>		<u>Gene Encoded</u>
912..962		Trc
1223..1009		attR1
1473..2132		CmR
2252..2336		inactivated ccdA
2474..2779		ccdB
2820..2944		attR2
3509..4414		ampR
5015..5175		ori
5415..5852		flori (f1 intergenic region)
6225..752		lacIq

1	GGCGGTGCAC	AATCTTCTCG	CGCAACGCGT	CAGTGGGCTG	ATCATTAACT	ATCCGCTGGA
61	TGACCAGGAT	GCCATTGCTG	TGGAAGCTGC	CTGCACTAAT	GTTCCGGCGT	TATTTCTTGA
121	TGTCTCTGAC	CAGACACCCA	TCAACAGTAT	TATTTTCTCC	CATGAAGACG	GTACGCGACT
181	GGCGGTGGAG	CATCTGGTCC	CATTGGGTCA	CCAGCAAATC	GCGCTGTTAG	CGGGCCCAAT
241	AAGTTCTGTC	TCGGCGCGTC	TGCGTCTGGC	TGGCTGGCAT	AAATATCTCA	CTCGCAATCA
301	AATTCAGCCG	ATAGCGGAAC	GGGAAGGCGA	CTGGAGTGCC	ATGTCCGGTT	TTCACAAAC
361	CATGCAAATG	CTGAATGAGG	GCATCGTTCC	CACTGCGATG	CTGGTTGCCA	ACGATCAGAT
421	GGCGCTGGGC	GCAATGCGCG	CCATTACCGA	GTCGGGCGTG	GCGCTTGGTG	CGGATATCTC
481	GGTAGTGGGA	TACGACGATA	CCGAAGACAG	CTCATGTTAT	ATCCCGCCGT	CAACCACCAT
541	CAACACGGAT	TTTCGCCTGC	TGGGGCAAAC	CAGCGTGGAC	CGCTTGCTGC	AACTCTCTCA
601	GGGCCAGGCG	GTGAAGGGCA	ATCAGCTGTT	GCCCGTCTCA	CTGGTGAAAA	GAAAAACCAC
661	CCTGGCACCC	AATACGCAAA	CCGCCTCTCC	CCGCGCGTTG	GCCGATTCAT	TAATGCAGCT
721	GGCACGACAG	GTTTCCCGAC	TGGAAGCGG	GCAGTGAGCG	CAACGCAATT	AATGTGAGTT
781	AGCGCGAATT	GATCTGGTTT	GACAGCTTAT	CATCGACTGC	ACGGTGCAAC	AATGCTTCTG
841	CGTTCAGGCA	GCCATCGGAA	GCTGTGGTAT	GGCTGTGCAG	GTCGTAATC	ACTGCATAAT
901	TCGTGTGCTC	CAAGGCGCAC	TCCCGTCTCG	GATAATGTTT	TTTGCGCCGA	CATCATAACG
961	GTTCTGGCAA	ATATTCTGAA	ATGAGCTGTT	GACAAATTAAT	CATCCGGTCC	TATATATCTG
1021	TGGAATTGTG	AGCGGATAAC	AATTTCACAC	AGGAAACAGA	CCATGTCGTA	CTACCATCAC
1081	CATCACCATC	ACGGCATCAC	AAGTTGTAC	AAAAAAGCTG	AACGAGAAAC	GTAATAATGAT
1141	ATAAATATCA	ATATATTAAA	TTAGATTTTG	CATAAAAAAC	AGACTACATA	ATACTGTAAA
1201	ACACAACATA	TCCAGTCACT	ATGGCGGCCG	CTAAGTTGGC	AGCATCACCC	GACGCACTTT
1261	GCGCCGAATA	AATACCTGTG	ACGGAAGATC	ACTTCGCAGA	ATAAATAAAT	CCTGGTGTCC
1321	CTGTTGATAC	CGGGAAGCCC	TGGGCCAACT	TTTGGCGAAA	ATGAGACGTT	GATCGGCACG
1381	TAAGAGGTTT	CAACTTTCAC	CATAATGAAA	TAAGATCACT	ACCGGGCGTA	TTTTTTGAGT
1441	TATCGAGATT	TTTCCAGGCT	AAGGAAGCTA	AAATGGAGAA	AAAAATCACT	GGATATACCA
1501	CCGTTGATAT	ATCCCAATGG	CATCGTAAAG	AACATTTTGA	GGCATTTCAG	TCAGTTGCTC
1561	AATGTACCTA	TAACCAAGAC	GTTTCTGCTG	ATATTACGGC	CTTTTAAAG	ACCGTAAAGA
1621	AAAATAAGCA	CAAGTTTAT	CCGGCCTTTA	TTCACATTCT	TGCCCGCCTG	ATGAATGCTC
1681	ATCCGGAATT	CCGTATGGCA	ATGAAAGACG	GTGAGCTGGT	GATATGGGAT	AGTGTTCACC
1741	CTTGTTACAC	CGTTTTCCAT	GAGCAAACTG	AAACGTTTTT	ATCGCTCTGG	AGTGAATACC
1801	ACGACGATTT	CCGGCAGTTT	CTACACATAT	ATTCGCAAGA	TGTGGCGTGT	TACGGTGAAA
1861	ACCTGGCCTA	TTTCCCTAAA	GGGTTTATG	AGAATATGTT	TTTCGTCTCA	GCCAAATCCCT
1921	GGGTGAGTTT	CACCAAGTTT	GATTTAAACG	TGGCCAATAT	GGACAACTTC	TTCCGCCCCG
1981	TTTTCAACAT	GGGCAATAT	TATACGCAAG	GCGACAAGGT	GCTGATGCCG	CTGGCGATTG
2041	AGGTTTATCA	TGCCGTCTGT	GATGGCTTCC	ATGTCGGCAG	AATGCTTAAT	GAATTACAAC
2101	AGTACTGCGA	TGAGTGGCAG	GGCGGGGCGT	AAACGCGTGG	ATCCGCGTTA	CTAAAAGCCA
2161	GATAACAGTA	TGCGTATTTG	CGCGCTGATT	TTTGCCTGAT	AAGAATATAT	ACTGATATGT
2221	ATACCCGAAG	TATGTCAAAA	AGAGGTGTGC	TATGAAGCAG	CGTATTACAG	TGACAGTTGA
2281	CAGCGACAGC	TATCAGTTGC	TCAAGGCATA	TATGATGTCA	ATATCTCCGG	TCTGGTAAGC
2341	ACAACCATGC	AGAATGAAGC	CCGTCTGCTG	CGTGCCGAAC	GCTGGAAAGC	SGAAAATCAG
2401	GAAGGGATGG	CTGAGGTGCG	CCGTTTATT	GAAATGAACG	GCTCTTTTGC	TGACGAGAAG
2461	AGGGACTGGT	GAAATGCAGT	TTAAGGTTTA	CACCTATAAA	AGAGAGAGCC	GTTATCGTCT
2521	GTTTGTGGAT	GTACAGAGTG	ATATTATTGA	CACGCCCGGG	CGACGGATGG	TGATCCCCCT

FIGURE 22B

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2581 GGCCAGTGCA CGTCTGCTGT CAGATAAAGT CTCCTCGTGAA CTTTACCCGG TGGTGCATAT
2641 CGGGGATGAA AGCTGGCGCA TGATGACCAC CGATATGGCC AGTGTGCCGG TCTCCGTTAT
2701 CGGGGAAGAA GTGGCTGATC TCAGCCACCG CGAAAATGAC ATCAAAAACG CCAATTAACCT
2761 GATGTTCTGG GGAATATAAA TGTCAAGGCTC CCTTATACAC AGCCAGTCTG CAGGTGCGCC
2821 ATAGTGACTG GATATGTTGT GTTTTACAGT ATTATGTAGT CTGTTTTTTA TGCAAAATCT
2881 AATTTAATAT ATTGATATTT ATATCATTTT ACGTTTCTCG TTCAGCTTTC TTGTACAAAG
2941 TGGTGATGCC CATATGGGAA TTCAAAGGCC TACGTCGACG AGCTCACTAG TCGCGGCCCG
3001 TTCTAGAGGA TCCCTCGAGG CATGCGGTAC CAAGCTTGCC TGTTTTGGCG GATGAGAGAA
3061 GATTTTCAGC CTGATACAGA TTAATCAGA ACGCAGAAGC GGTCTGATAA AACAGAATTT
3121 GCCTGGCGGC AGTAGCGCGG TGGTCCCACC TGACCCCATG CCGAACTCAG AAGTGAAACG
3181 CCGTAGCGCC GATGGTAGTG TGGGGTCTCC CCATGCGAGA GTAGGGAACG GCCAGGCATC
3241 AAATAAAACG AAAGGCTCAG TCGAAAGACT GGGCCTTTTCG TTTTATCTGT TGTTTGTGCG
3301 TGAACGCTCT CCTGAGTAGG ACAAATCCGC CGGGAGCGGA TTTGAACGTT GCGAAGCAAC
3361 GGCCCGGAGG GTGGCGGCCA GGACGCCCGC CATAACTGCG CAGGCATCAA ATTAAGCAGA
3421 AGGCCATCCT GACGGATGGC CTTTTTTCGT TTCTACAAAC TCTTTTGTGT TATTTTCTA
3481 AATACATTCA AATATGTATC CGCTCATGAG ACAATAACCC TGATAAATGC TTCAATAATA
3541 TTGAAAAAGG AAGAGTATGA GTATTCAACA TTTCCGTGTC GCCCTTATTC CCTTTTTCG
3601 GGCATTTTGC CTTCCCTGTT TTGCTCACCC AGAAACGCTG GTGAAAGTAA AAGATGCTCA
3661 ASATCAGTTG GGTGCACGAG TGGGTACAT CGAACTGGAT CTCAACAGCG GTAAGATCCT
3721 TGAGAGTTTT CGCCCGAAG AACGTTTTCC AATGATGAGC ACTTTTAAAG TTCTGCTATG
3781 TGGCGCGGTA TTATCCCGTG TTGACGCCGG GCAAGAGCAA CTCGGTCGCC GCATACACTA
3841 TTCTCAGAA GACTTGGTTG AGTACTCACC AGTCACAGAA AAGCATCTTA CGGATGGCAT
3901 GACAGTAAGA GAATTATGCA GTGCTGCCAT AACCATGAGT GATAACACTG CGGCCAACTT
3961 ACTTCTGACA ACGATCGGAG GACCGAAGGA GCTAACCGCT TTTTGCACA ACATGGGGGA
4021 TCATGTAACT CGCCTTGATC GTTGGGAACC GGAGCTGAAT GAAGCCATAC CAAACGACGA
4081 CGCTGACACC ACGATGCCTA CAGCAATGGC AACAACGTTG CGCAAACTAT TAACTGGCGA
4141 ACTACTTACT CTAGCTTCCC GGCAACAATT AATAGACTGG ATGGAGGCGG ATAAAGTTGC
4201 AGGACCACTT CTGCGCTCGG CCCTTCCGGC TGGCTGGTTT ATTGCTGATA AATCTGGAGC
4261 CGGTGAGCGT GGGTCTCGCG GTATCATTGC AGCACTGGGG CCAGATGGTA AGCCCTCCCG
4321 TATCGTAGTT ATCTACACGA CGGGGAGTCA GGCAACTATG GATGAACGAA ATAGACAGAT
4381 CGCTGAGATA GGTGCCTCAC TGATTAAGCA TTGGTAACTG TCAGACCAAG TTTACTATA
4441 TATACTTTAG ATTGATTAA AACTTCATT TTAATTTAAA AGGATCTAGG TGAAGATCCT
4501 TTTTGATAAT CTCATGACCA AAATCCCTTA ACGTGAGTTT TCGTTCCACT GAGCGTCAGA
4561 CCCCAGTAA AAGATCAAAG GATCTTCTG AGATCCTTTT TTTCTGCGCG TAATCTGCTG
4621 CTTGCAACA AAAAAACCA CGCTACCAGC GGTGGTTTGT TTGCCGGATC AAGAGCTACC
4681 AACTCTTTT CCGAAGGTAA CTGGCTTCAG CAGAGCGCAG ATACCAAATA CTGCTCCTCT
4741 AGTGTAGCCG TAGTTAGGCC ACCACTTCAA GAACTCTGTA GCACCCGCTA CATACCTCGC
4801 TCTGCTAATC CTGTTACCAG TGGCTGCTGC CAGTGGCGAT AAGTCGTGTC TTACCGGGTT
4861 GGACTCAAGA CGATAGTTAC CGGATAAGGC GCAGCGGTCC GGTGAAACGG GGGGTTCCGT
4921 CACACAGCCC AGCTTGGAGC GAACGACCTA CACCGAACTG AGATACCTAC AGCGTGAGCT
4981 ATGAGAAAGC GCCACGCTTC CCGAAGGGAG AAAGGCGGAC AGGTATCCGG TAAGCGGCAG
5041 GGTGCGAACA GGAGAGCGCA CGAGGGAGCT TCCAGGGGGA AACGCCCTGG ATCTTTATAG
5101 TCCTGTGCGG TTTCCGCCACC CTGACTTGA GCGTCGATTT TTGTGATGCT CGTCAGGGGG
5161 GCGGAGCCTA TGGAAAAACG CCAGCAACGC GGCCTTTTTA CGGTTCCTGG CCTTTTGTG
5221 GCCTTTTGCT CACATGTTCT TTCTGCGTT ATCCCTGAT TCTGTGGATA ACCGTATTAC
5281 CGCCTTTGAG TGAGCTGATA CCGCTCGCG CAGCCGAACG ACCGAGCGCA GCGAGTCAGT
5341 GAGCGAGGAA GCGGAAGAGC GCCTGATGCG GTATTTTCTC CTTACGCATC TGTGCGGTAT
5401 TTCACACCGC ATAATTTTGT TAAATTCGC GTTAAATTT TGTAAATCA GCTCATTTT
5461 TAACCAATAG GCCGAAATCG GCAAAATCCC TTATAAATCA AAAGAATAGA CCGAGATAGG
5521 GTTGAGTGTT GTTCCAGTTT GGAACAAGAG TCCACTATTA AAGAACGTGG ACTCCAACGT
5581 CAAAGGGCGA AAAACCGTCT ATCAGGGCGA TGGCCCACTA CGTGAACCAT CACCCTAATC
5641 AAGTTTTTTG GGGTCGAGGT GCCGTAAAGC ACTAAATCGG AACCTTAAAG GGAGCCCCCG
5701 ATTTAGAGCT TGACGGGGAA AGCCGGCGAA CGTGGCGAGA AAGGAAGGGA AGAAAGCGAA
5761 AGGAGCGGCG GCTAGGGCGC TGGCAAGTGT AGCGGTCACG CTGCGCGTAA CCACCACACC
5821 CGCCGCGCTT AATGCGCCGC TACAGGGCGC GTCCCATTCG CCATTCAGGC TGCTATGGTG
5881 CACTCTCAGT ACAAATCTGCT CTGATGCCGC ATAGTTAAGC CAGTATACAC TCCGCTATCG
5941 CTACGTGACT GGGTCATGGC TGGCCCCGA CACCCGCCAA CACCCGCTGA CGCGCCCTGA
6001 CGGGCTTGTC TGCTCCCGGC ATCCGCTTAC AGACAAGCTG TGACCGTCTC CGGGAGCTGC

FIGURE 22C

6061 ATGTGTCAGA GGTTCACACC GTCATCACCG AACGCGCGA GGCAGCAGAT CAATTCGCGC
6121 GCGAAGGCGA AGCGGCATGC ATTTACGTTG ACACCATCGA ATGGTGCAAA ACCTTCGCG
6181 GTATGGCATG ATAGCGCCCG GAAGAGAGTC AATTCAGGGT GGTGAATGTG AAACCAGTAA
6241 CGTTATACGA TGTCGCAGAG TATGCCGCTG TCTCTTATCA GACCGTTTCC CGCGTGGTGA
6301 ACCAGGCCAG CCACGTTTCT GCGAAAACGC GGGAAAAAGT GGAAGCGGCG ATGGCGGAGC
6361 TGAATTACAT TCCCAACCGC GTGGCACAAC AACTGGCGGG CAAACAGTCG TTGCTGATTG
6421 GCGTTGCCAC CTCCAGTCTG GCCCTGCACG CGCCGTCGCA AATTGTCGCG GCGATTAAAT
6481 CTCGCGCCGA TCAACTGGGT GCCAGCGTGG TGGTGTGAT GGTAGAACGA AGCGGCGTCG
6541 AAGCCTGTAA AGC

FIGURE 22D

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Figure 23A: pDEST3

GST fusions in E. coli

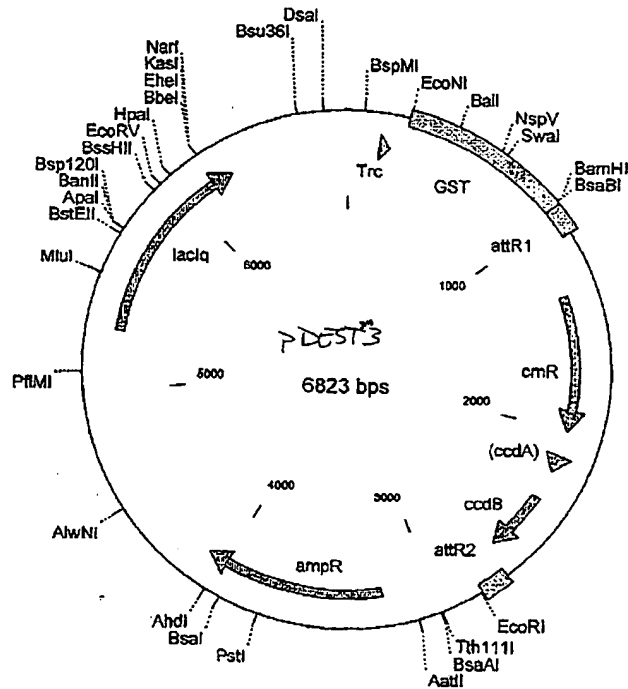
154 cgg ttc tgg caa ata ttc tga aat gag ctg ⁻³⁵ ttg aca att aat cat cgg ctc
 gcc aag acc gtt tat aag act tta ctc gac aac tgt taa tta gta gcc gag

205 ⁻¹⁰ gta taa tgt gtg gaa ttg tga gcg gat aac aat ttc aca cag gaa aca gta
 cat att aca cac ctt aac act cgc cta ttg tta aag tgt gtc ctt tgt cat

256 ^{M S P I L} ttc atg tcc cct ata cta ^{GST} ggt tat tgg aaa att aag ggc ctt gtg caa ccc
 aag tac agg gga tat gat cca ata acc ttt taa ttc ccg gaa cac gtt ggg

919 ^{" GST} ctg gtt ccg cgt gga tct cgt cgt gca tct gtt gga tcc cca tca ^{T S} aca agt
 gac caa ggc gca cct aga gca gca cgt aga caa cct agg ggt agt tgt tca

970 ^{H Y K K} ~~tgg ttc aag aag gct gaa cga gaa acg tga aat gat ata aat acc aat ata~~
~~aac atg ttt ttc cga gct gct cct tgc att tta cta tat tta tag tta tat~~



pDEST3 6823 bp

Location (Base Nos.)		Gene Encoded
150..200		Trc
1087..963		attR1
1337..1996		CmR
2116..2200		inactivated ccdA
2338..2643		ccdB
2684..2808		attR2
3231..4091		ampR
5295..6254		lacIq
1	ACGTTATCGA CTGCACGGTG CACCAATGCT TCTGGCGTCA GGCAGCCATC GGAAGCTGTG	
61	GTATGGCTGT GCAGGTCGTA AATCACTGCA TAATTCGTGT CGCTCAAGGC GCACCTCCCGT	
121	TCTGGATAAT GTTTTTTGCG CCGACATCAT AACGGTTCGT GCAAATATTC TGAAATGAGC	
181	TGTTGACAAT TAATCATCGG CTCGTATAAT GTGTGGAATT GTGAGCGGAT AACAAATTCA	
241	CACAGGAAAC AGTATTCATG TCCCTATAC TAGGTATTG GAAAAATTAAG GGCCTTGTGC	
301	AACCCACTCG ACTTCTTTTG GAATATCTTG AAGAAAAATA TGAAGAGCAT TTGTATGAGC	
361	GCGATGAAGG TGATAAATGG CGAAACAAAA AGTTTGAATT GGGTTTGGAG TTTCCCAATC	
421	TTCTTTATTA TATTGATGGT GATGTTAAAT TAACACAGTC TATGGCCATC ATACGTTATA	
481	TAGCTGACAA GCACAACATG TTGGGTGGTT GTCCAAAAGA CCGTGCAGAG ATTTCAATGC	
541	TTGAAGGAGC GGTTTTGGAT ATTAGATACG GTGTTTCGAG AATTGCATAT AGTAAAGACT	
601	TTGAACTCTT CAAAGTTGAT TTTCTTAGCA AGCTACCTGA AATGCTGAAA ATGTTCTGAAG	
661	ATCGTTTATG TCATAAAACA TATTTAAATG GTGATCATGT AACCCATCCT GACTTCATGT	
721	TGTATGACGC TCTTGATGTT GTTTTATACA TGGACCCAAT GTGCCTGGAT GCGTTCCCAA	
781	AATTAGTTTG TTTTAAAAAA CGTATTGAAG CTATCCCAACA AATTGATAAG TACTTGAAAT	
841	CCAGCAAGTA TATAGCATGG CCTTTGCAGG GCTGGCAAGC CACGTTTGGT GGTGGCGACC	
901	ATCCTTCAAA ATCGGATCTG GTTCCGCGTG GATCTCGTCG TGCATCTGTT GGATCCCCAT	
961	CAACAAGTTT GTACAAAAAA GCTGAACGAG AAACGTAAAA TGATATAAAT ATCAATATAT	
1021	TAAATTAGAT TTTGCATAAA AAACAGACTA CATAACTGCT TAAACACAA CATATCCAGT	
1081	CACTATGGCG GCCGCTAAGT TGGCAGCATC ACCCGACGCA CTTTGCGCCG AATAAATACC	
1141	TGTGACGGAA GATCACTTCG CAGAATAAAT AAATCCTGGT GTCCCTGTTG ATACCGGGAA	
1201	GCCCTGGGCC AACTTTTGGC GAAATGAGA CGTTGATCGG CACGTAAGAG GTTCCAACCT	
1261	TCACCATAAT GAAATAAGAT CACTACCGGG CGTATTTTTC GAGTTATCGA GATTTTCAGG	
1321	AGCTAAGGAA GCTAAATGG AGAAAAAAT CACTGGATAT ACCACCGTTG ATATATCCCA	
1381	ATGGCATCGT AAAGAACATT TTGAGGCATT TCAGTCAGTT GCTCAATGTA CCTATAACCA	
1441	GACCGTTCAG CTGGATATTA CGGCCCTTTT AAAGACCGTA AAGAAAAATA AGCACAAGTT	
1501	TTATCCGGCC TTATTTCACT TTCTTGCCCG CCTGATGAAT GCTCATCCGG AATTCCGTAT	
1561	GGCAATGAAA GACGGTGAGC TGGTGATATG GGTAGTGTT CACCCTTGTT ACACCGTTTT	
1621	CCATGAGCAA ACTGAAACGT TTTTCATCGCT CTGGAGTGAA TACCACGACG ATTTCCGGCA	
1681	GTCTCTACAC ATATATTTCG AAGATGTGGC GTGTTACGGT GAAAAACCTGG CCTATTTCCC	
1741	TAAAGGGTTT ATTGAGAATA TGTTTTTCGT CTCAGCCAAT CCCTGGGTGA GTTTCACCAG	
1801	TTTTGATTTA AACGTGGCCA ATATGGACAA CTCTTCGCC CCCGTTTTCA CCATGGGCAA	
1861	ATATTATACG CAAGGCGACA AGGTGCTGAT GCCGCTGGCG ATTCAGGTTT ATCATGCCGT	
1921	CTGTGATGGC TTCCATGTCG GCAGAATGCT TAATGAATTA CAACAGTACT GCGATGAGTG	
1981	GCAGGGCGGG GCGTAAAGAT CTGGATCCGG CTACTAAAA GCCAGATAAC AGTATGCGTA	
2041	TTTGGCGCGT GATTTTTCGG GTATAAGAAT ATATACTGAT ATGTATACCC GAAGTATGTC	
2101	AAAAAGAGGT GTGCTATGAA GCAGCGTATT ACAGTGACAG TTGACAGCGA CAGCTATCAG	
2161	TTGCTCAAGG CATATATGAT GTCAATATCT CCGGTCTGGT AAGCACAAAC ATGCAGAATG	
2221	AAGCCCGTCG TCTGCGTGCC GAACGCTGGA AAGCGGAAAA TCAGGAAGGG ATGGCTGAGG	
2281	TCGCCCCGTT TATTGAAATG AACGGCTCTT TTGCTGACGA GAACAGGGAC TGGTGAAATG	
2341	CAGTTTAAGG TTTACACCTA TAAAAGAGAG AGCCGTTATC GTCTGTTTGT GGATGTACAG	
2401	AGTGATATTA TTGACACGCC CCGGCGACGG ATGGTGATCC CCCTGGCCAG TGCACGTCTG	
2461	CTGTCAGATA AAGTCTCCCG TGAACCTTAC CCGGTGGTGC ATATCGGGGA TGAAAGCTGG	
2521	CGCATGATGA CCACCGATAT GGCAGTGTG CCGTCTCCG TTATCGGGGA AGAAGTGGCT	
2581	GATCTCAGCC ACCGCGAAAA TGACATCAAA AACGCCATTA ACCTGATGTT CTGGGGAATA	
2641	TAAATGTCAG GCTCCCTTAT ACACAGCCAG TCTGCAGGTC GACCATAGTG ACTGGATATG	

FIGURE 23B

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2701 TTGTGTTTTA CAGTATTATG TAGTCTGTTT TTTATGCAAA ATCTAATTTA ATATATTGAT
2761 ATTTATATCA TTTTACGTTT CTCGTTCAGC TTTCTTGTA AAAGTGGTTG ATGGGAATTC
2821 ATCGTGACTG ACTGACGATC TGCCTCGCGC GTTTCGGTGA TGACGGTGAA AACCTCTGAC
2881 ACATGCAGCT CCCGGAGACG GTCACAGCTT GTCGTGAAGC GGATGCCGGG AGCAGACAAG
2941 CCCGTCAGGG CGCGTCAGCG GGTGTTGGCG GGTGTCGGGG CGCAGCCATG ACCCAGTCAC
3001 GTAGCGATAG CGGAGTGAT AATTCTTGAA GACGAAAGGG CCTCGTGATA CGCCTATTTT
3061 TATAGGTTAA TGTCATGATA ATAATGTTTT CTAGACGTC AGGTGGCACT TTTCGGGGAA
3121 ATGTGCGCGG AACCCTTATT TGTTTATTTT TCTAAATACA TTCAAATATG TATCCGCTCA
3181 TGAGACAATA ACCCTGATAA ATGCTTCAAT AATATTGAAA AAGGAAGAGT ATGAGTATTC
3241 AACATTTCCG TGTGCGCCTT ATTCCCTTTT TTGCGGCATT TTGCCTTCCT GTTTTTGTCTC
3301 ACCCAGAAAC GCTGGTGAAA GTAAAAGATG CTGAAGATCA GTTGGGTGCA CGAGTGGGTT
3361 ACATCGAACT GGATCTCAAC AGCGGTAAGA TCCTTGAGAG TTTTCGCCCC GAAGAACGTT
3421 TTCCAATGAT GAGCACTTTT AAAGTTCTGC TATGTGGCGC GGTATTATCC CGTGTGACG
3481 CCGGGCAAGA GCAACTCGGT CGCCGCATAC ACTATTCTCA GAATGACTTG GTTGAGTACT
3541 CACCAGTCAC AGAAAAGCAT CTTACGGATG GCATGACAGT AAGAGAATTA TGCAGTGTCTG
3601 CCATAACCAT GAGTGATAAC ACTGCGGCCA ACTTACTTCT GACAACGATC GGAGGACCGA
3661 AGGAGCTAAC CGCTTTTTTG CACAACATGG GGGATCATGT AACTCGCCTT GATCGTTGGG
3721 AACCGGAGCT GAATGAAGCC ATACCAAACG ACGAGCGTGA CACCACGATG CCTGCAGCAA
3781 TGGCAACAAC GTTGGCGCAA CTATTAACATG GCGAACTACT TACTCTAGCT TCCCGGCAAC
3841 AATTAATAGA CTGGATGGAG GCGGATAAAG TTGCGAGGAC ACTTCTGCGC TCGGCCCTTC
3901 CGGCTGGCTG GTTTATTGCT GATAAATCTG GAGCCGGTGA GCGTGGGTCT CGCGGTATCA
3961 TTGCAGCACT GGGGCCAGAT GGTAAGCCCT CCCGTATCGT AGTTATCTAC ACGACGGGGA
4021 GTCAGGCAAC TATGGATGAA CGAAATAGAC AGATCGCTGA GATAGGTGCC TCACTGATTA
4081 AGCATTGGTA ACTGTCAGAC CAAGTTTACT CATATATACT TTAGATTGAT TTAAAACTTC
4141 ATTTTAAATT TAAAAGGATC TAGGTGAAGA TCCTTTTTGA TAATCTCATG ACCAAAATCC
4201 CTTAACGTGA GTTTTCGTTT CACTGAGCGT CAGACCCCGT AGAAAAGATC AAAGGATCTT
4261 CTTGAGATCC TTTTTCGTTT CGCGTAATCT GCTGCTTGA AAAAAAAGG CACCGCTAC
4321 CAGCGGTGGT TTGTTTGCCG GATCAAGAGC TACCAACTCT TTTTCCGAAG GTAAGTGGCT
4381 TCAGCAGAGC GCAGATACCA AATACTGTCC TTCTAGTGA GCCGTAGTTA GGCCACCCT
4441 TCAAGAACTC TGTCAGCCG CCTACATACC TCGCTCTGCT AATCCTGTTA CCAAGTGGCTG
4501 CTGCCAGTGG CGATAAGTCG TGTCTTACC GGTGGGACTC AAGACGATAG TTACCGGATA
4561 AGGCCGACG GTCGGGCTGA ACGGGGGGTT CGTGACACA GCCCAGCTTG GAGCGAACCA
4621 CCTACACCGA ACTGAGATAC CTACAGCGTG AGCTATGAGA AAGCGCCACG CTTCCCGAAG
4681 GGAGAAAAGC GACAGGTAT CCGGTAAGCG GCAGGGTGG AACAGGAGAG CGCAGGAGGG
4741 AGCTTCCAGG GGGAAACGCC TGGTATCTTT ATAGTCTCTG CGGGTTTCGC CACCTCTGAC
4801 TTGAGCGTCG ATTTTGTGA TGCTCGTCAG GGGGGCGGAG CCTATGGAAA AACGCCAGCA
4861 ACGCGGCCTT TTTACGGTTC CTGGCCTTTT GCTGGCCTTT TGCTCACATG TTCTTTCCTG
4921 CGTTATCCCC TGATTCTGTG GATAACCGTA TTACCGCCTT TGAGTGAGCT GATACCGCTC
4981 GCGCGAGCCG AACGACCGAG CGCAGCGAGT CAGTGAGCGA GGAAGCGGAA GAGCGCCTGA
5041 TGGGATATT TCTCCTTACG CATCTGTGCG GTATTTTACA CCGCATAAAT TCCGACACCA
5101 TCGAATGGTG CAAAACCTTT CGCGGTATGG CATGATAGCG CCCGGAAGAG AGTCAATTCA
5161 GGGTGGTGAA TGTGAAACCA GTAACGTTAT ACGATGTCGC AGAGTATGCC GGTGTCTCTT
5221 ATCAGACCGT TTCCGCGGTG GTGAACCAAG CCAGCCACGT TTCTGCGAAA ACGCGGAAA
5281 AAGTGGAAGC GCGGATGGCG GAGCTGAATT ACATTCCTAA CCGCGTGGCA CAACAACCTG
5341 CGGGCAACA GTCGTTGCTG ATTGGCGTTG CCACCTCCAG TCTGGCCCTG CACGCGCCGT
5401 CGCAAAATGT CCGGCGGATT AAATCTCGCG CCGATCAACT GGGTGCCAGC GTGGTGGTGT
5461 CGATGGTAGA ACGAAGCGGC GTCGAAGCCT GTAAAGCGGC GGTGCACAA CTCTCGCGC
5521 AACGCGTCAG TGGGCTGATC ATTAACATAT CGCTGGATGA CCAGGATGCC ATTGCTGTGG
5581 AAGCTGCCCTG CACTAATGTT CCGGCGTTAT TTCTTGATGT CTCTGACCAG ACACCCATCA
5641 ACAGTATTAT TTTCTCCCAT GAAGACGTA GCGACTGGG CGTGGAGCAT CTGGTCGCAT
5701 TGGGTCACCA GCAAAATCGCG CTGTTAGCGG GCCCATTAAG TTCTGTCTCG GCGCGTCTGC
5761 GTCTGGCTGG CTGGCATAAA TATCTCACTC GCAATCAAA TACAGCGGATA GCGGAACGGG
5821 AAGGCGACTG GAGTGCCATG TCCGGTTTTT AACAAACCAT GCAAAATGCTG AATGAGGGCA
5881 TCGTTCCAC TGCGATGCTG GTTGCCAACG ATCAGATGGC GCTGGGCGCA ATGCGCGCCA
5941 TTACCGAGTC CCGGCTGCGG GTTGGTGGCG ATATCTCGGT AGTGGGATAC GACGATACCG
6001 AAGACAGCTC ATGTTATATC CCGCGTTAA CCACCATCAA ACAGGATTTT CGCTGCTGG
6061 GGCAAAACCAG CGTGGACCGC TTGCTGCAAC TCTCTCAGGG CCAGGCGGTG AAGGGCAATC
6121 AGCTGTTGCC CGTCTCACTG GTGAAAAGAA AAACCAACCT GCGGCCCAAT ACGCAAACCG-

FIGURE 23C

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6181 CCTCTCCCG CGCGTTGGCC GATTCATTAA TGCAGCTGGC ACGACAGGTT TCCCGACTGG
6241 AAAGCGGGCA GTGAGCGCAA CGCAATTAAT GTGAGTTAGC TCACTCATTG GGCACCCAG
6301 GCTTTTACACT TTATGCTTCC GGCTCGTATG TTGTGTGGAA TTGTGAGCGG ATAACAATT
6361 CACACAGGAA ACAGCTATGA CCATGATTAC GGATTCAGTG GCCGTGCTTT TACAACGTCG
6421 TGACTGGGAA AACCCTGGCG TTACCCAAC TAATCGCCTT GCAGCACATC CCCCTTTCGC
6481 CAGCTGGCGT AATAGCGAAG AGGCCCGCAC CGATCGCCCT TCCCAACAGT TGCGCAGCCT
6541 GAATGGCGAA TGGCGCTTTG CCTGGTTTCC GGCACCAGAA GCGGTGCCCG AAAGCTGGCT
6601 GGAGTGGGAT CTTCTGAGG CCGATACTGT CGTCGTCCCC TCAAACGTCG AGATGCACGG
6661 TTACGATGCG CCCATCTACA CCAACGTAAC CTATCCCATT ACGGTCAATC CGCCGTTTGT
6721 TCCCACGGAG AATCCGACGG GTTGTACTC GCTCACATT AATGTTGATG AAAGCTGGCT
6781 ACAGGAAGGC CAGACGCGAA TTATTTTGA TGGCGTTGGA ATT

FIGURE 23D

His6-thioredoxin fusions in *E. coli*

919 gca aat att ctg aaa tga gct gct gac aat taa tca tcc ggt ccg cat aat
cgt tta taa gac ttt act cga cta ctg tta att agt agg cca ggc ata tta

978 ctg tgg ^{→ mfaA} aat tgt gag cgg ata aca att tca cac agy aaa cay acc Met Glt
gac acc tta aca ctc gcc tat tgt taa agt gty tcc ttt gtc tgg tac cca

Hs6

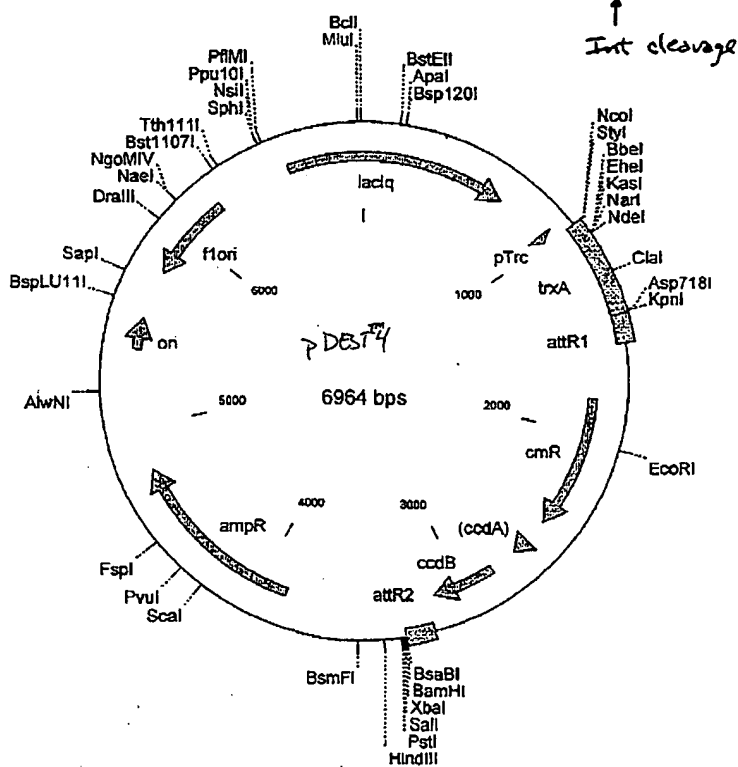
1021 Hs1 Hs2 Hs3 Hs4 Hs5 Hs6 As Tyr Asa Ile Asa Thr Thr Gln Asa Leu Tyr
cat cat cat cat cat cat gat tgc gat atc cca acg acc gaa aac ctg tgc
gta gta gta gta gta gta cta atg cta tag ggt tgc tgg ctt ttg gac ata

TEV protease → Thioredoxin - (~150 amino acids)

1072 Pro Gln Gly Asa Met Ser Arg Ile Ile Ile His Leu Arg Asa Ser
ttt cag ggt gcc cat atg agc gat aac att cat ctg act gac gat agt
aaa gtc ccg cgg gta tac tcy cta ttt taa taa gtg gac tga ctg ctg tca

attR1

1429 Gat Asp Asp Lys Val Pro Ile Thr Ser Leu Tyr Lys Lys
gat gat gat gat aat gta ccc atc tgc tca aac atg ttt tct aga gct gct
cta ctg cta ctg ttc cat cgg tag tgt tca aac atg ttt tct aga gct gct



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pDEST4 6964 bp

<u>Location (Base Nos.)</u>		<u>Gene Encoded</u>
964..1003		Trc
1577..1453		attR1
1827..2486		CmR
2606..2690		inactivated ccdA
2828..3133		ccdB
3174..3298		attR2
3872..4777		ampR
5378..5538		ori
5778..6215		flori (f1 intergenic region)
6587..704		lacIq

1	CTATCCGCTG	GATGACCAGG	ATGCCATTGC	TGTGGAAGCT	GCCTGCACTA	ATGTTCCGGC
61	GTTATTCTCT	GATGCTCTCG	ACCAGACACC	CATCAACAGT	ATTATTTTCT	CCCATGAAGA
121	CGGTACGCGA	CTGGGCGTGG	AGCATCTGGT	CGCATTGGGT	CACCAGCAAA	TGCGCGTGTT
181	AGCGGGCCCA	TTAAGTTCTG	TCTCGGCGCG	TCTGCGTCTG	GCTGGCTGGC	ATAAATATCT
241	CACTCGCAAT	CAAAATCAGC	CGATAGCGGA	ACGGGAAGGC	GACTGGAGTG	CCATGTCCGG
301	TTTTCAACAA	ACCATGCAAA	TGCTGAATGA	GGGCATCGTT	CCCACTGCGA	TGCTGGTTGC
361	CAACGATCAG	ATGGCGCTGG	GCGCAATGCG	CGCCATTACC	GAGTCCGGGC	TGCGCGTTGG
421	TGCGGATATC	TCGGTAGTGG	GATACGACGA	TACCGAAGAC	AGCTCATGTT	ATATCCCGCC
481	GTCAACCACC	ATCAAAACAGG	ATTTTCGCCT	GCTGGGGCAA	ACCAGCGTGG	ACCGCTTGCT
541	GCAACTCTCT	CAGGGCCAGG	CGGTGAAGGG	CAATCAGCTG	TTGCCGTCTC	CACTGGTGAA
601	AAGAAAAACC	ACCCTGGCAC	CCAATACGCA	AACCGCCTCT	CCCCCGCGGT	TGGCCGATTG
661	ATTAATGCAG	CTGGCAGCAC	AGGTTTCCCG	ACTGGAAGC	GGGCAGTGAG	CGCAACGCAA
721	TTAATGTGAG	TTAGCGCGAA	TTGATCTGGT	TTGACAGCTT	ATCATCGACT	GCACGGTGCA
781	CCAATGCTTC	TGGCGTCAGG	CAGCCATCGG	AAGCTGTGGT	ATGGCTGTGC	AGGTGCTAAA
841	TCATGCGATA	ATTCTGTCTG	CTCAAGGCGC	ACTCCCGTTC	TGGATAATGT	TTTTCGCGCC
901	GACATCATAA	CGGTTCTGGC	AAATATTCTG	AAATGAGCTG	TTGACAATTA	ATCATCCGGT
961	CCGTATAATC	TGTGGAATTG	TGAGCGGATA	ACAATTTTAC	ACAGGAAACA	GACCATGGGT
1021	CATCATCATC	ATCATCACGA	TTACGATATC	CCAACGACCG	AAAACCTGTA	TTTTCAGGGC
1081	GCCCATATGA	GCGATAAAAT	TATTCACCTG	ACTGACGACA	GTTTGTGAC	GGATGTACTC
1141	AAAGCGGACG	GGGCGATCCT	CGTCGATTTT	TGGGCAGAGT	GGTGCGGTCC	GTGCAAAATG
1201	ATCGCCCGGA	TTCTGGATGA	AATCGCTGAC	GAATATCAGG	GCAAACCTGAC	CGTTGCAAAA
1261	CTGAACATCG	ATCAAAACCC	TGGCACTGCG	CCGAAATATG	GCATCCGTGG	TATCCCGACT
1321	CTGCTGCTGT	TCAAAAACGG	TGAAGTGGCG	GCAACCAAAG	TGGGTGCACT	GTCTAAAGGT
1381	CAGTTGAAAG	AGTTCCCTCGA	CGCTAACCTG	GCCGGTTCTG	GTTCTGGTGA	TGACGATGAC
1441	AAGGTACCCA	TCACAAGTTT	GTACAAAAAA	GCTGAACGAG	AAACGTAAAA	TGATATAAAT
1501	ATCAATATAT	TAAATTAGAT	TTTGCATAAA	AAACAGACTA	CATAATACTG	TAAAACACAA
1561	CATATCCAGT	CACTATGGCG	GCCGCTAAGT	TGGCAGCATC	ACCCGACGCA	CTTTGCGCCG
1621	AATAAATACC	TGTGACGGAA	GATCACTTCG	CAGAATAAAT	AAATCCTGGT	GTCCTGTGTT
1681	ATACCGGGAA	GCCCTGGGCC	AACCTTTTGGC	GAAAATGAGA	CGTTGATCGG	CACGTAAGAG
1741	GTTCCAACCT	TCACCATAAT	GAAATAAGAT	CACTACCGGG	CGTATTTTCT	GAGTTATCGA
1801	GATTTTCAGG	AGCTAAGGAA	GCTAAATGAG	AGAAAAAAT	CACTGGATAT	ACCACCGTTG
1861	ATATATCCCA	ATGGCATCGT	AAAGAACATT	TTGAGGCATT	TCAGTCAGTT	GCTCAATGTA
1921	CCTATAACCA	GACCGTTTCA	CTGGATATTA	CGGCCTTTT	AAAGACCGTA	AAGAAAAATA
1981	AGCACAAAGT	TTATCCGGCC	TTTATTCACA	TTCTTGCCCG	CCTGATGAAT	GCTCATCCGG
2041	AATTCCGTAT	GGCAATGAAA	GACGGTGAGC	TGGTGATATG	GGATAGTGT	CACCTTGTG
2101	ACACCGTTTT	CCATGAGCAA	ACTGAAACGT	TTTCATCGCT	CTGGAGTGAA	TACCACGACG
2161	ATTTCCGGCA	GTTTCTACAC	ATATATTCCG	AAGATGTGGC	GTGTTACGGT	GAAAACCTGG
2221	CCTATTTCCT	TAAAGGGTTT	ATTGAGAATA	TGTTTTTCGT	CTCAGCCAAT	CCCTGGGTGA
2281	GTTTCACCA	TTTTGATTTA	AACGTGGCCA	ATATGGACAA	CTTCTTCGCC	CCCGTTTTCA
2341	CCATGGGCAA	ATATTATACG	CAAGGCGACA	AGGTGCTGAT	GCCGCTGGCG	ATTCAGSTTC
2401	ATCATGCCGT	CTGTGATGGC	TTCCATGTGC	GCAGAATGCT	TAATGAATTA	CAACAGTACT
2461	GCGATGAGTG	GCAGGCGGGG	GCGTAAACGC	GTGGATCCGG	CTTACTAAAA	GCCAGATAAC
2521	AGTATGCGTA	TTTGCGCGCT	GATTTTTCGG	GTATAAGAAT	ATATACTGAT	ATGTATACCC

FIGURE 24B

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2581 GAAGTATGTC AAAAAGAGGT GTGCTATGAA GCAGCGTATT ACAGTGACAG TTGACAGCGA
 2541 CAGCTATCAG TTGCTCAAGG CATATATGAT GTCAATATCT CCGGTCTGGT AAGCACAACC
 2701 ATGCAGAAATG AAGCCCGTCG TCTGCGTGCC GAACGCTGGA AAGCGGAAAA TCAGGAAGGG
 2761 ATGGCTGAGG TCGCCCGGTT TATTGAAATG AACGGCTCTT TTGCTGACGA GAACAGGGAC
 2821 TGGTGAAATG CAGTTTAAAG TTTACACCTA TAAAAGAGAG AGCCGTTATC GTCTGTTTGT
 2881 GGATGTACAG AGTGATATTA TTGACACGCC CGGGCGACGG ATGGTGATCC CCCTGGCCAG
 2941 TGCACGCTCG CTGTCAGATA AAGTCTCCCG TGAACCTTAC CCGGTGGTGC ATATCGGGGA
 3001 TGAAGCTGG CGCATGATGA CCACCGATAT GGCCAGTGTG CCGGTCTCCG TTATCGGGGA
 3061 AGAAGTGGCT GATCTCAGCC ACCGCGAAAA TGACATCAAA AACGCCATTA ACCTGATGTT
 3121 CTGGGGAATA TAAATGTCAG GCTCCCTTAT ACACAGCCAG TCTGCAGGTC GACCATAGTG
 3181 ACTGGATATG TTGTGTTTTA CAGTATTATG TAGTCTGTTT TTTATGCAAA ATCTAAITTA
 3241 ATATATTGAT ATTTATATCA TTTTACGTTT CTCGTTTACG TTTCTGTAC AAAGTGGTGA
 3301 TGGGGATCCT CTAGAGTCGA CCTCCAGTAA TCGTACAGGG TAGTACAAAT AAAAAGGCA
 3361 CGTCAGATGA CGTGCCTTTT TCTTTGTGAG CAGTAAGCTT GGCTGTTTTG GCGGATGAGA
 3421 GAAGATTTC AGCCTGATAC AGATTAAATC AGAACGCGAGA AGCGGTCTGA TAAAACAGAA
 3481 TTTGCTTGGC GGCAGTAGCG CGGTGGTCCC ACCTGACCCC ATGCCGAAC CAGAAGTGAA
 3541 ACGCCGTAGC GCCGATGGTA GTGTGGGGTC TCCCATGCG AGAGTAGGGA ACTGCCAGGC
 3601 ATCAAATAAA ACGAAAGGCT CAGTCGAAAG ACTGGGCTT TCGTTTTATC TGTGTTTTGT
 3661 CGGTGAACGC TCTCTGAGT AGGACAAATC CGCCGGGAGC GGATTTGAAC GTTCCGAAGC
 3721 AACGGCCCGG AGGGTGGCGG GCAGGACGCC CGCCATAAAC TGCCAGGCAT CAAATTAAGC
 3781 AGAAGGCCAT CTTGACGGAT GGCCCTTTTG CGTTTCTACA AACTCTTTT GTTTATTTTT
 3841 CTAAATACAT TCAAATATGT ATCCGCTCAT GAGACAATAA CCCTGATAAA TGCTTCAATA
 3901 ATATTGAAAA AGGAAGAGTA TGAGTATTCA ACATTTCCGT GTCCGCCCTTA TTCCCTTTTT
 3961 TGCGGCATTT TGCCCTCCTG TTTTGTCTCA CCCAGAAACG CTGGTGAAG TAAAAGATGC
 4021 TGAAGATCAG TTGGGTGCAC GAGTGGGTTA CATCGAACTG GATCTCAACA CCGGTAAGAT
 4081 CCTTGAGAGT TTTCCGCCCG AAGAACGTTT TCCAATGATG AGCACTTTTA AAGTCTGCT
 4141 ATGTGGCGCG GTATTATCCC GTGTTGACGC CGGGCAAGAG CAACTCGGTC GCCGCATACA
 4201 CTATTCTCAG AATGACTTGG TTGAGTACTC ACCAGTCACA GAAAAGCATC TTACGGATGG
 4261 CATGACAGTA AGAGAATTAT GCAGTGCTGC CATACCATG AGTGATAACA CTGCGGCCAA
 4321 CTTACTTCTG ACAACGATCG GAGGACCGAA GGAGCTAACC GCTTTTTTGC ACAACATGGG
 4381 GGATCATGTA ACTCGCCTTG ATCGTTGGGA ACCGGAGCTG AATGAAGCCA TACCAAACGA
 4441 CGAGCGTGAC ACCACGATGC CTACAGCAAT GGCAACAACG TTGCGCAAC TATTAAGTGG
 4501 CGAACTACTT ACTCTAGCTT CCCGCAACA ATTAATAGAC TGGATGGAGG CGGATAAAGT
 4561 TGCAGGACCA CTTCTGCGCT CGGCCCTTCC GGCTGGCTGG TTTATGCTG ATAATCTGG
 4621 AGCCGGTGAG CGTGGGTCTC GCGGTATCAT TGCAGCACTG GGGCCAGATG GTAAGCCCTC
 4681 CCGTATCGTA GTTATCTACA CGACGGGAG TCAGGCAACT ATGGATGAAC GAAATAGACA
 4741 GATCGCTGAG ATAGTGCTC CACTGATTAA GCATTGGTAA CTGTCAGACC AAGTTTACTC
 4801 ATATATACTT TAGATTGATT TAAAACITCA TTTTAAATTT AAAAGGATCT AGGTGAAGAT
 4861 CCTTTTTGAT AATCTCATGA CCAAATCCC TTAACGTGAG TTTTCGTTCC ACTGAGCGTC
 4921 AGACCCCGTA GAAAAGATCA AAGGATCTTC TTGAGATCCT TTTTTTCTGC GCGTAATCTG
 4981 CTGCTTGCAA ACAAAAAAAC CACCGCTACC AGCGGTGGTT TGTTTGCCGG ATCAAGAGCT
 5041 ACCAACTCTT TTTCCGAAGG TAACTGGCTT CAGCAGAGCG CAGATACCAA ATACTGTCTC
 5101 TCTAGTGTAG CCGTAGTTAG GCCACCACTT CAAGAACTCT GTAGCACC GC CATACACTT
 5161 CGCTCTGCTA ATCCTGTTAC CAGTGGCTGC TGCCAGTGGC GATAAGTCGT GTCTTACCGG
 5221 GTTGGACTCA AGACGATAGT TACCGGATAA GGCGCAGCGG TCGGGCTGAA CGGGGGGTTT
 5281 GTGCACACAG CCCAGCTTGG AGCGAACGAC CTACACCGAA CTGAGATACC TACAGCGTGA
 5341 GCTATGAGAA AGCGCCACGC TTCCCGAAGG GAGAAAGGCG GACAGGTATC CGGTAAGCGG
 5401 CAGGGTCGGA ACAGGAGAGC GCACGAGGGA GCTTCCAGGG GGAAACGCCT GGTATCTTTA
 5461 TAGTCTGTG GGGTTTCGCC ACCTCTGACT TGAGCGTCTGA TTTTGTGAT GCTCTGTCAGG
 5521 GGGGCGGAGC CTATGGAAAA ACGCCAGCAA CGCGGCTTTT TTACGGTTCC TGGCCTTTTG
 5581 CTGGCCTTTT GCTCACATGT TCTTTCCTGC GTTATCCCTT GATTCTGTGG ATAACCGTAT
 5641 TACCGCCTTT GAGTGAGCTG ATACCGCTCG CCGCAGCCGA ACGACCGAGC GCAGCGAGTC
 5701 AGTGAGCGAG GAAGCGGAAG AGCGCTGAT GCGGTATTTT CTCCTTACGC ATCTGTGCGG
 5761 TATTTACAC CGCATAATTT TGTAAAAATT CGCGTTAAAT TTTTGTAAAA TCAGCTCAT
 5821 TTTTAACCAA TAGGCCGAAA TCGGCAAAAT CCCTTATAAA TCAAAAGAA AGACCGAGAT
 5881 AGGGTTGAGT GTTGTTCAG TTTGGAACAA GAGTCCACTA TTAAGAAACG TGGACTCCAA
 5941 CGTCAAAGGG CGAAAAACCG TCTATCAGGG CGATGGCCCA CTACGTGAAC CATCACCCCTA
 6001 ATCAAGTTTT TTGGGTGCGA GGTGCCGTAA AGCACTAAAT CGGAACCCCTA AAGGAGGCC-

FIGURE 24C

6061 CCGATTTAGA GCTTGACGGG GAAAGCCGGC GAACGTGGCG AGAAAGGAAG GGAAGAAAGC
6121 GAAAGGAGCG GGCGCTAGGG CGCTGGCAAG TGTCGCGTC ACGCTGCGCG TAACCACCAC
6181 ACCCGCCGCG CTTAATGCGC CGCTACAGGG CGCGTCCATT CGCCATTCAG GCTGCTATGG
6241 TGCACTCTCA GTACAATCTG CTCTGATGCC GCATAGTTAA GCCAGTATAC ACTCCGCTAT
6301 CGCTACGTGA CTGGGTCATG GCTGCGCCCC GACACCCGCC AACACCCGCT GACGCGCCCT
6361 GACGGGCTTG TCTGCTCCCG GCATCCGCTT ACAGACAAGC TGTGACCGTC TCCGGGAGCT
6421 GCATGTGTCA GAGGTTTCA CCGTCATCAC CGAAACGCGC GAGGCAGCAG ATCAATTTCG
6481 GCGCGAAGGC GAAGCGGCAT GCATTTACGT TGACACCATC GAATGGTGCA AAACCTTTCG
6541 CGGTATGGCA TGATAGCGCC CGGAAGAGAG TCAATTCAGG GTGGTGAATG TGAAACCACT
6601 AACGTTATAC GATGTGCGAG AGTATGCCGG TGTCTCTTAT CAGACCGTTT CCCGCGTGGT
6661 GAACCAGGCC AGCCACGTTT CTGCGAAAAC GCGGGAAAAA GTGGAAGCGG CGATGGCGGA
6721 GCTGAATTAC ATTCCCAACC GCGTGGCACA ACAACTGGCG GGCAACAGT CGTTGCTGAT
6781 TGGCGTTGCC ACCTCCAGTC TGGCCCTGCA CGCGCCGTCG CAAATTGTG CGGCGATTAA
6841 ATCTCGCGCC GATCAACTGG GTGCCAGCGT GGTGGTGTG ATGGTAGAAC GAAGCGGCGT
6901 CGAAGCCTGT AAAGCGGCGG TGCACAATCT TCTCGCGCAA CGCGTCAGTN GGGCTGATCA
6961 TTAA

FIGURE 24b

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Figure 25b pDEST5

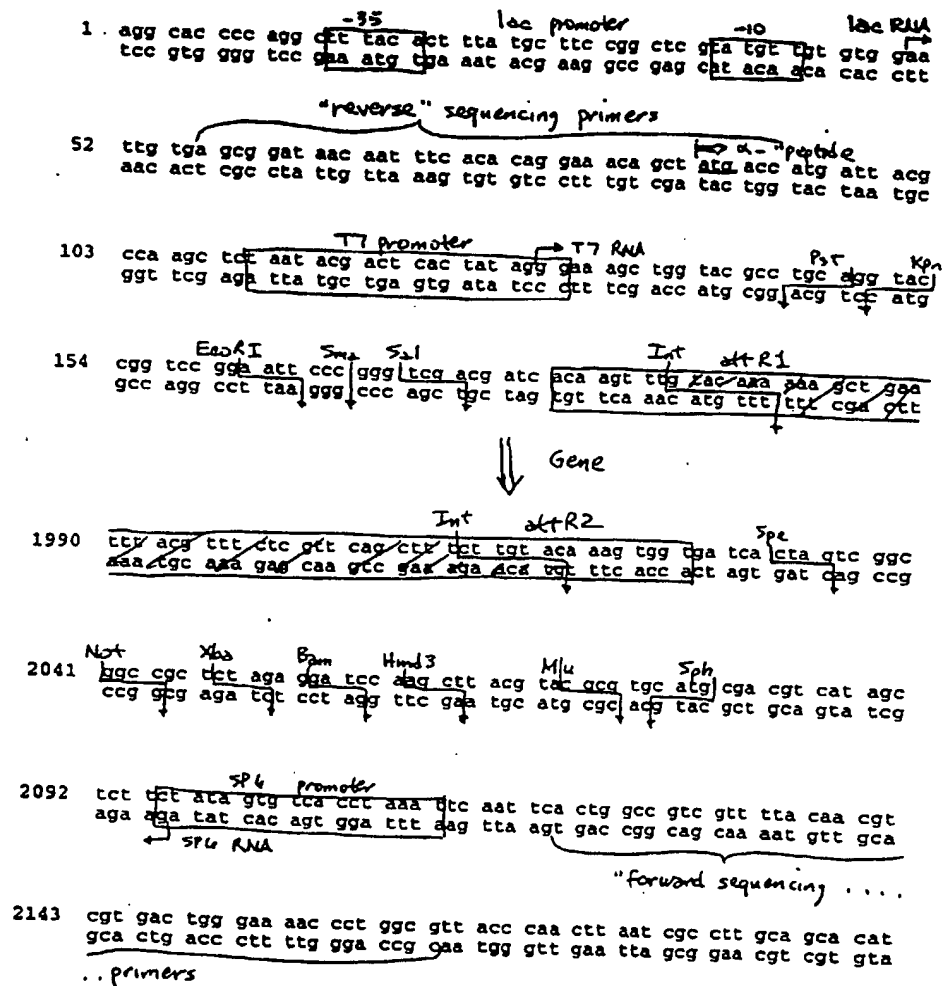
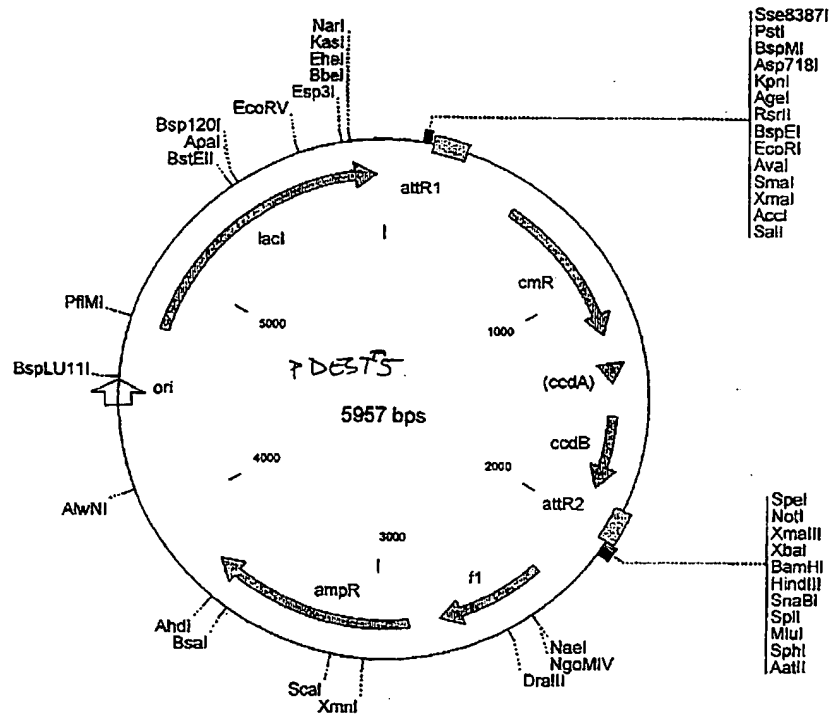
pSPORT '+' (for sequencing, probes,
phagemid)

Figure 25B γ DEST5 (cont'd)



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pDEST5 5957 bp

Location (Base Nos.)	Gene Encoded
305..181	attR1
555..1214	CmR
1334..1418	inactivated ccdA
1556..1861	ccdB
1902..2026	attR2
2278..2733	f1 (f1 intergenic region)
2865..3722	ampR
5378..5538	ori
4756..5922	lacI

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1 AGGCACCCCA GGCTTTACAC TTTATGCTTC CGGCTCGTAT GTTGTGTGGA ATTGTGAGCG
61 GATAACAATT TCACACAGGA AACAGCTATG ACCATGATTA CGCCAAGCTC TAATACGACT
121 CACTATAGGG AAAGCTGGTA CGCCTGCAGG TACCGGTCCG GAATTCCTCG GTCCAGCATC
181 ACAAGTTTGT ACAAAAAAGC TGAACGAGAA ACGTAAATG ATATAAATAT CAATATATTA
241 AATTAGATTT TGCATAAAAA ACAGACTACA TAATACTGTA AAACACAACA TATCCAGTCA
301 CTATGGCGGC CGCTAAGTTG GCAGCATCAC CCGACGCACT TTGCGCCGAA TAAATACCTG
361 TGACGGAAGA TCACTTCGCA GAATAAATAA ATCCTGGTGT CCCTGTGTAT ACCGGGAAGC
421 CCTGGGCCAA CTTTTGGCGA AAATGAGACG TTGATCGGCA CGTAAGAGGT TCCAACTTTC
481 ACCATAATGA AATAAGATCA CTACCGGGCG TATTTTTTGA GTTATCGAGA TTTTCAGGAG
541 CTAAGGAAGC TAAATGGAG AAAAAAATCA CTGGATATAC CACCGTTGAT ATATCCCAAT
601 GGCATCGTAA AGAACATTTT GAGGCATTTC AGTCAGTTGC TCAATGTACC TATAACCGAG
661 CCGTTCAGCT GGATATTACG GCCTTTTAA AGACCGTAAA GAAAAATAAG CACAAGTTT
721 ATCCGGCCTT TATTCACATT CTGCCCCGCC TGATGAATGC TCATCCGGAA TTCCGTATGG
781 CAATGAAAGA CGGTGAGCTG GTGATATGGG ATAGTGTTC A CCTTGTTC ACCGTTTTC
841 ATGAGCAAAC TGAAACGTTT TCATCGCTCT GGAGTGAATA CCACGACGAT TTCCGGCAGT
901 TTCTACACAT ATATTGCAA GATGTGGCGT GTTACGGTGA AAACCTGGCC TATTTCCCTA
961 AAGGGTTTAT TGAGAATATG TTTTTCGTCT CAGCCAATCC CTGGGTGAGT TTCACCAAGT
1021 TTGATTTAAA CGTGGCCAAT ATGGACAAC TCTTCGCCCC CGTTTTCACC ATGGGCAAA
1081 ATTATACGCA AGGCGACAAG GTGCTGATGC CGCTGGCGAT TCAGGTTTCAT CATGCCGTCT
1141 GTGATGGCTT CCAATGTGGC AGAATGCTTA ATGAATTACA ACAGTACTGC GATGAGTGGC
1201 AGGGCGGGGC GTAAACGCGT GGATCCGGCT TACTAAAAGC CAGATAACAG TATGCGTATT
1261 TGGCGCGCTG TTTTTCGGGT ATAAGAATAT ATACTGATAT GTATACCCGA AGTATGTCAA
1321 AAAGAGGTGT GCTATGAAGC AGCGTATTAC AGTGACAGTT GACAGCGACA GCTATCAGTT
1381 GCTCAAGGCA TATATGATGT CAATATCTCC GGTCTGGTAA GCACAACCAT GCAGAATGAA
1441 GCGCGTCGTC TGGCTGCCGA ACGCTGGAAA GCGGAAAATC AGGAAGGGAT GGCTGAGGTC
1501 GCGCGGTTTA TTGAAATGAA CGGCTCTTTT GCTGACGAGA ACAGGGACTG GTGAAATGCA
1561 GTTTAAGGTT TACACCTATA AAAGAGAGAG CCGTTATCGT CTGTTTGTGG ATGTACAGAG
1621 TGATATTATT GACACGCCCG GCGGACGGAT GGTGATCCCC CTGGCCAGTG CACGCTGCT
1681 GTCAGATAAA GTCTCCCGTG AACTTTACCC GGTGGTGCAT ATCGGGGATG AAAGCTGGCG
1741 CATGATGACC ACCGATATGG CCAGTGTGCC GGTCTCCGTT ATCGGGGAAG AAGTGGCTGA
1801 TCTCAGCCAC CGCGAAATG ACATCAAAAA CGCCATTAACT CTGATGTTCT GGGGAATATA
1861 AATGTCAGGC TCCCTTATAC ACAGCCAGTC TGCAGGTCGA CCAATAGTAC TGGATATGTT
1921 GTGTTTTTACA GTATTATGTA GTCTGTTTTT TATGCAAAAT CTAATTTAAT ATATTGATAT
1981 TTATATCATT TTACGTTTCT CGTTCAGCTT TCTTGTAACA AGTGGTGATC ACTAGTCGGC
2041 GCGCGTCTTA GAGGATCCAA GCTTACGTAC GCGTGCATGC GACGTCATAG CTCTTCTATA
2101 GTGTCACTTA AATTCAATTC ACTGGCCGTC GTTTTACAAC GTCGTGACTG GGGAAACCTT
2161 GCGGTTACCC AACTTAATCG CCTTGACGCA CATCCCCCTT TCGCCAGCTG GCGTAATAGC
2221 GAAGAGGCCC GCACCGATCG CCCTTCCCAA CAGTTGCGCA GCCTGAATGG CGAATGGACG
2281 CGCCCTGTAG CGGCGCATTA AGCGCGGCGG GTGTGGTGGT TACGCGCAGC GTGACCGCTA
2341 CACTTGCCAG CGCCCTAGCG CCCGCTCCTT TCGCTTTCTT CCCTTCCTTT CTCGCCACGT
2401 TCGCCGGCTT TCCCGTCAA GCTCTAAATC GGGGGCTCCC TTTAGGGTTC CGATTAGTGT
2461 CTTTACGGCA CCTCGACCCC AAAAAACTTG ATTAGGGTGA TGGTTCACGT AGTGGGCCAT
2521 CGCCCTGATA GACGGTTTTT CGCCCTTGA CGTTGGAGTC CACGTTCTTT AATAGTGGAC
2581 TCTTGTCCA AACTGGAACA AACTCAACC CTATCTCGGT CTATTCCTTT GATTTATAAG-

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FIGURE 25C

2641 GGATTTTGCC GATTTCGGCC TATTGGTTAA AAAATGAGCT GATTTAACAA AAATTTAACG
2701 CGAATTTTAA CAAAATATTA ACGTTTACAA TTTCAGGTGG CACTTTTCGG GGAATGTGC
2761 GCGGAACCCC TATTTGTTTA TTTTCTAAA TACATTCAAA TATGTATCCG CTGATGAGAC
2821 AATAACCCCTG ATAAATGCTT CAATAATATT GAAAAAGGAA GAGTATGAGT ATTCAACATT
2881 TCCGTGTCGC CCTTATTCCT TTTTTCGGG CATTTTGCCT TCCTGTTTTT GCTCACCCAG
2941 AAACGCTGGT GAAAGTAAAA GATGCTGAAG ATCAGTTGGG TGCACGAGTG GGTACATCG
3001 AACTGGATCT CAACAGCGGT AAGATCCTTG AGAGTTTTCG CCCGGAAGAA CGTTTTCCAA
3061 TGATGAGCAC TTTTAAAGTT CTGCTATGTG GCGCGGTATT ATCCCGTATT GACGCCGGGC
3121 AAGAGCAACT CGGTCGCCGC ATACACTATT CTCAGAATGA CTTGGTTGAG TACTCACCAG
3181 TCACAGAAAA GCATCTTACG GATGGCATGA CAGTAAGAGA ATTATGCAST GCTGCCATAA
3241 CCATGAGTGA TAACACTGCG GCCAACTTAC TTCTGACAAC GATCGGAGGA CCGAAGGAGC
3301 TAACCGCTTT TTTGCACAAC ATGGGGGATC ATGTAACCTG CCTTGATCGT TGGGAACCGG
3361 AGCTGAATGA AGCCATACCA AACGACGAGC GTGACACCAC GATGCTCTGA GCAATGGCAA
3421 CAACGTTGCG CAAACTATTA ACTGGCGAAC TACTTACTCT AGCTTCCCGG CAACAATTAA
3481 TAGACTGGAT GGAGGCGGAT AAGTTGCGAG GACCACTTCT GCGCTCGGCC CTTCCGCTG
3541 GCTGGTTTAT TGCTGATAAA TCTGGAGCCG GTGAGCGTGG GTCTCGCGGT ATCATTGCGAG
3601 CACTGGGGCC AGATGGTAAG CCCTCCCGTA TCGTAGTTAT CTACACGACG GGGAGTCAGG
3661 CAACTATGGA TGAACGAAAT AGACAGATCG CTGAGATAGG TGCCTCACTG ATTAAGCATT
3721 GGTAACGTG AGACCAAGTT TACTCATATA TACTTTAGAT TGATTAAAA CTTCAATTTT
3781 AATTAAAAAG GATCTAGGTG AAGATCCTTT TTGATAATCT CATGACCAAA ATCCCTTAAC
3841 GTGAGTTTTC GTTCCACTGA CCGTCAGACC CCGTAGAAAA GATCAAAGGA TCTTCTTGAG
3901 ATCCTTTTTT TCTGCGCGTA ATCTGCTGCT TGCAACAAAA AAAACCACCG CTACCAGCGG
3961 TGGTTTGT TTGCGGATCAA GAGCTACCAA CTCTTTTTCG GAAGGTAACG GGCTTCAGCA
4021 GAGCGCAGAT ACCAAATACT GTCCCTCTAG TGTAGCCGTA GTTAGGCCAC CACTTCAAGA
4081 ACTCTGTAGC ACCGCCTACA TACCTCGCTC TGCTAATCCT GTTACCAGTG GCTGCTGCCA
4141 GTGGCGATAA GTCGTGTCTT ACCGGGTGGG ACTCAAGACG ATAGTTACCG GATAAGGCGC
4201 AGCGGTCGGG CTGAACGGGG GGTTCGTGCA CACAGCCACG CTTGGAGCGA ACGACCTACA
4261 CCGAACTGAG ATACCTACAG CGTGAGCATT GAGAAAGCGC CACGCTTCCC GAAGGAGAGAA
4321 AGGCGGACAG GTATCCGGTA AGCGGCAGGG TCGGAACAGG AGAGCGCAGC AGGGAGCTTC
4381 CAGGGGAAAA CGCCTGGTAT CTTTATAGTC CTGTCGGGTT TCGCCACCTC TGACTTGAGC
4441 GTCGAATTTT GTGATGCTCG TCAGGGGGGC GGAGCCTATG GAAAAACGCC AGCAACGCGG
4501 CCTTTTACG GTTCCTGGCC TTTTGTGCGC CTTTGTCTCA CATGTTCTTT CTTGCGTTAT
4561 CCCCTGATTC TGTGGATAAC CGTATTACCG CCTTTGAGTG AGCTGATACC GCTCGCCGCA
4621 GCCGAACGAC CGAGCGCAGC GAGTCAGTGA GCGAGGAAGC GGAAGAGCGC CCAATACGCA
4681 AACCGCTCTT CCCCAGCGGT TGGCCGATTG ATTAATGCAG AGCTTGCAAT TCGCGCGCGA
4741 AGGCGAAGCG GCATTTACGT TGACACCATC GAATGGCGCA AAACCTTTTCG CGGTATGGCA
4801 TGATAGCGCC CGGAAGAGAG TCAATTCAGG GTGGTGAATG TGAACACAGT AACGTATATC
4861 GATGTCGCG AGTATGCCCG TGTCTCTTAT CAGACCGTTT CCCGCGTGGT GAACGAGGCC
4921 AGCCACGTTT CTGCGAAAC GCGGGAAGAA GTGGAAGCGG CGATGGCGGA GCTGAATTAC
4981 ATTCCCAACC GCGTGGCACA ACAACTGGCG GGCACACAGT CGTTGCTGAT TGGCGTTGCC
5041 ACCTCCAGTC TGGCCCTGCA CGCGCCGTCG CAAATTGTG CGGCGATTAA ATCTCGCGCC
5101 GATCAACTGG GTGCCAGCGT GGTGGTGTG ATGGTAGAAC GAAGCGGCGT CGAAGCCTGT
5161 AAAGCGGCGG TGCACAATCT TCTCGCGCAA CCGGTGAGTG GGCTGATCAT TAACTATCCG
5221 CTGGATGACC AGGATGCCAT TGCTGTGAA GCTGCCGTGA CTAATGTTCC GGCCTTATTT
5281 CTTGATGTCT CTGACCAGAC ACCCATCAAC AGTATTATTT TCTCCCATGA AGACGGTACG
5341 CGACTGGGCG TGGAGCATCT GGTGCGATTG GGTCAACAGC AAATCGCGCT GTTAGCGGGC
5401 CCATTAAATT CTGTCTCGGC GCGTCTGCGT CTGGCTGGCT GGCATAAATA TCTCACTCGC
5461 AATCAAATTC AGCCGATAGC GGAACGGGAA GCGCACTGGA GTGCCATGTC CGGTTTTCAA
5521 CAAACCATGC AAATGCTGAA TGAGGGCATC GTTCCCACTG CGATGCTGGT TGCCACCGAT
5581 CAGATGGCGC TGGGCGCAAT GCGCGCCATT ACCGAGTCCG GGCTGCGCGT TGGTGGCGAT
5641 ATCTCGGTAG TGGGATACGA CGATACCGAA GACAGCTCAT GTTATATCCC GCCGTCAACC
5701 ACCATCAAAC AGGATTTTCG CCTGCTGGGG CAAACGAGCG TGGACCGCTT GCTGCAACTC
5761 TCTCAGGGCC AGGCGGTGAA GGGCAATCAG CTGTTGCCCG TCTCACTGGT GAAAAAGAAA
5821 ACCACCTGG CGCCCAATAC GCAAACCGCC TCTCCCGCG CGTTGCGCGA TTCATTAATG
5881 CAGCTGGCAC GACAGGTTTC CCGACTGGAA AGCGGCGAGT GAGCGCAACG CAATTAATGT
5941 GAGTTAGCTC ACTCAT

FIGURE 25D

Figure 2Left

pDST6

pSPORT "4"
(opposite strand)

"forward" sequencing primers

1 taa/cgc cag ggt ttt ccc agt cac gac gtt gta aaa cga cgg cca ctg aat
att gcg gtc cca aaa ggg tca gtg ctg caa cat ttt gct gcc ggt cac tta

52 tga att tag gtg aca cta tag aag agc tat gac gtc gca tgc acg cgt acg
act tca atc cac tgt gat atc ttc tgg ata ctg cag ggt acg tgc gca tgc

103 tga gct tgg atc ctc tag agc ggc cgc cga cta gtg atc aca agt tgg tgc
att cga acc tag gag atc tcg ccg ggc gct gat gac tag tgt tca aac atg

154 aaa daa gct gaa cga gaa acg taa aat gat ata aat atc aac ata ttd aat
ttt tat cga ctt gct ctt tgc att tta cta tat tta tag cta tat aat tta

↓
Gene

1939 tat tta tat tat ttt acg att ctc gtt tag ctt gct tgt aca aag tgg tga
ata aat ata gta aaa tgc aaa gag aaa gtc gaa aga aca tgc ttc acc att

1990 tcg tgg acc cgg gaa ttc cgg acc ggt act tgc tgg cgt acc agc ttt ccc
agc agc tgg gcc ctt aag gcc tgg gca tgg acg tcc gca tgg tgg aaa ggg

T7 RNA

2041 tat agt gag tgg tat tag agc ttg ggg taa tca tgg tca tag ctg ttt cct
ata tca ctc agc ata atc tgc aac cgc att agt acc agt atc gac aaa gga

T7 promoter

α-peptide

"reverse .."

2092 gtg tga aat tgt tat ccg ctc aca att cca cac aac ata cga gct gga agc
cac act tta aca ata ggc gag tgt taa ggt gtg tgg tat gct cgg cct tgc

... sequencing primers

lac RNA

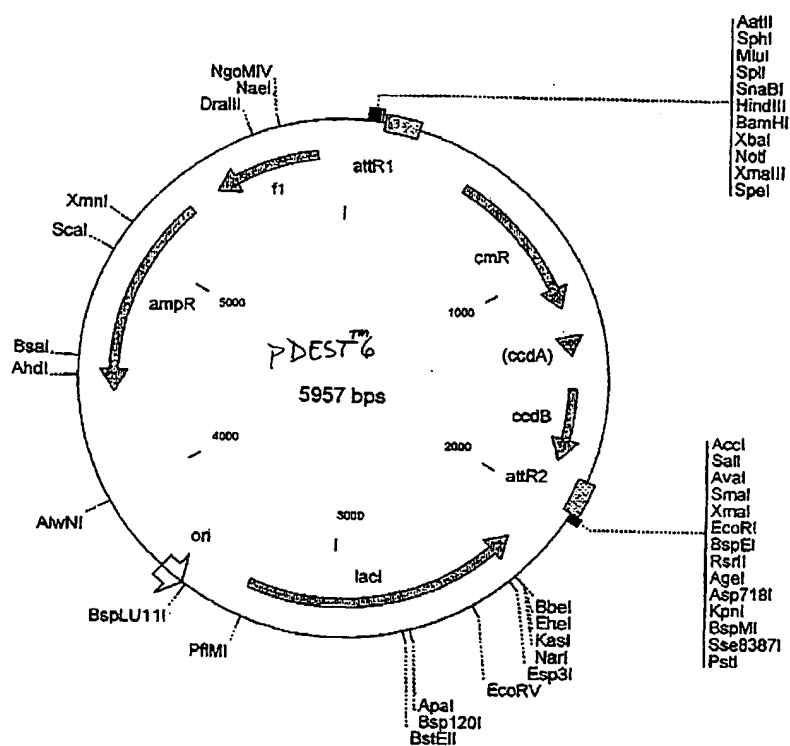
2143 ata aag tgt aaa gcc tgg ggt gcc taa tga gtg agc taa ctc aca tta att
tat ttc aca ttc cgg acc cca cgg att act cac tgg att gag tgt aat taa

53/240

Figure 26B

PDEST6

(cont'd)



pDEST6 5957 bp

Location (Base Nos.)	Gene Encoded
266..142	attR1
516..1175	CmR
1295..1379	inactivated ccdA
1517..1822	ccdB
1863..1987	attR2
2203..3369	lacI
4403..5260	ampR
5392..5847	f1 (f1 intergenic region)

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1 TAACGCCAGG GTTTTCCAG TCACGACGTT GTAAAACGAC GGCCAGTGAA TTGAATTTAG
61 GTGACACTAT AGAAGAGCTA TGACGTCGCA TGCACGCGTA CGTAAGCTTG GATCCTCTAG
121 AGCGGCCGCG GACTAGTGAT CACAAGTTTG TACAAAAAAG CTGAACGAGA AACGTAAAT
181 GATATAAATA TCAATATATT AAATTAGATT TTGCATAAAA AACAGACTAC ATAATACTGT
241 AAAACACAAC ATATCCAGTC ACTATGGCGG CCGCTAAGTT GGCAGCATCA CCCGACGCAC
301 TTTGCGCCGA ATAAATACCT GTGACGGAAG ATCACTTCGC AGAATAAATA AATCCTGGTG
361 TCCCTGTGTA TACCGGGAAG CCCTGGGCCA ACTTTTGGCG AAAATGAGAC GTTGATCGGC
421 ACGTAAGAGG TTCCAACCTT CACCATAATG AAATAAGATC ACTACCGGGC GTATTTTTTG
481 AGTTATCGAG ATTTTCAGGA GCTAAGGAAG CTAAAATGGA GAAAAAATC ACTGGATATA
541 CCACCGTTGA TATATCCCAA TGGCATCGTA AAGAACATTT TGAGGCATTT CAGTCAGTTG
601 CTCAATGTAT CTATAACAG ACCGTTGAGC TGGATATTAC GGCCTTTTTA AAGACCGTAA
661 AGAAAAATAA GCACAAGTTT TATCCGGCCT TTATTACAT TCTTGCCCGC CTGATGAATG
721 CTCATCCGGA ATTCGGTATG GCAATGAAAG ACGGTGAGCT GGTGATATGG GATAGTGTTC
781 ACCCTTGTTA CACCGTTTTC CATGAGCAAA CTGAAACGTT TTCATCGCTC TGGAGTGAAT
841 ACCACGACGA TTTCCGGCAG TTTCTACACA TATATTCGCA AGATGTGGCG TGTACGGTG
901 AAAACCTGGC CTATTTCCCT AAAGGGTTTA TTGAGAATAT GTTTTTCGTC TCAGCCAATC
961 CCGTGGGTGAG TTTCAACAGT TTTGATTTAA ACGTGGCCAA TATGGACAAC TTCTTCGCCC
1021 CCGTTTTTAC CATGGGCCAA TATTATACGC AAGGCGACAA GGTGCTGATG CCGCTGGCGA
1081 TTCAGGTTCA TCATGCCGTC TGTGATGGCT TCCATGTCGG CAGAAATGCTT AATGAATTAC
1141 AACAGTACTG CGATGAGTGG CAGGCGGGGG CGTAAACGCG TGGATCCGGC TTAATAAAG
1201 CCAGATAACA GTATGCGTAT TTGCGCGCTG ATTTTTCGCG TATAAGAATA TATACTGATA
1261 TGTATACCCG AAGTATGTCA AAAAGAGGTG TGCTATGAAG CAGCGTATTA CAGTGACAGT
1321 TGACAGCGAC AGCTATCAGT TGCTCAAGGC ATATATGATG TCAATATCTC CGTCTCGGTA
1381 AGCACAAACA TGCAGAATGA AGCCCGTCGT CTGCGTGGCG AACGCTGGAA AGCGGAAAAAT
1441 CAGGAAGGGA TGGCTGAGGT CGCCCGGTTT ATTGAAATGA ACGGCTCTTT TGCTGACGAG
1501 AACAGGGACT GGTGAAATCC AGTTTAAGGT TTACACCTAT AAAAGAGAGA GCCGTTATCG
1561 TCTGTTTGTG GATGTACAGA GTGATATTAT TGACACGCCC GGGCGACGGA TGGTATCCCC
1621 CCTGGCCAGT GCACGTCTGC TGTGAGATAA AGTCTCCCGT GAACITTTACC CGGTGGTGCA
1681 TATCGGGGAT GAAAGCTGGC GCATGATGAC CACCGATATG GCCAGTGTGC CGGTCTCCGT
1741 TATCGGGGAA GAAGTGGCTG ATCTCAGCCA CCGCGAAAAT GACATCAAAA ACGCCATTAA
1801 CCTGATGTTT TGGGGAATAT AAATGTCAGG CTCCCTTATA CACAGCCAGT CTGCAGGTG
1861 ACCATAGTGA CTGGATATGT TGTGTTTAC AGTATTATGT AGTCTGTTTT TTATGCAAAA
1921 TCTAATTATA TATATTGATA TTTATATCAT TTTACGTTTC TCGTTACAGT TTCTTTGACA
1981 AAGTGGTGAT CGTCGACCCG GGAATTCCGG ACCGTTACCT GCAGGCGTAC CAGCTTTCCC
2041 TATAGTGAGT CGTATTAGAG CTTGGCGTAA TCATGGTCAT AGCTGTTTCC TGTGTGAAAT
2101 TGTATATCCG TCACAATTCC ACACAACATA CGAGCCGGA GCATAAAGTG TAAAGCCTGG
2161 GGTGCTAAT GAGTGAGCTA ACTCACATTA ATTGCGTTGC GCTCACTGCC CGCTTTCCAG
2221 TCGGGAAACC TGTGTTGCA GCTGCATTAA TGAATCGGCC AACGCGCGGG GAGAGGCGGT
2281 TTGCGTATTG GCGCGCAGGG TGGTTTTTCT TTTCAACAGT GAGACGGGCA ACAGCTGATT
2341 GCCCTTCACC GCCTGGCCCT GAGAGAGTTG CAGCAAGCGG TCCACGCTGG TTTGCCCCAG
2401 CAGGCGAAAA TCCTGTTTGA TGGTGGTTGA CGGCGGGATA TAACATGAGC TGTCTTCGGT
2461 ATCGTCGTAT CCCACTACCG AGATATCCGC ACCAACGCGC AGCCCGGACT CGGTAATGGC
2521 GCGCATTCGG CCCAGCGCCA TCTGATCGTT GGCAACCAGC ATCGCAGTGG GAACGATGCC
2581 CTCATTGAGC ATTTGCAATG TTTGTTGAAA ACCGGACATG GCACCTCCAGT CGCTTCCCG
2641 TTCCGCTATC GGCTGAATTT GATTGCGAGT GAGATATTTA TGCCAGCCAG CCAGACGCAG-

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FIGURE 26C

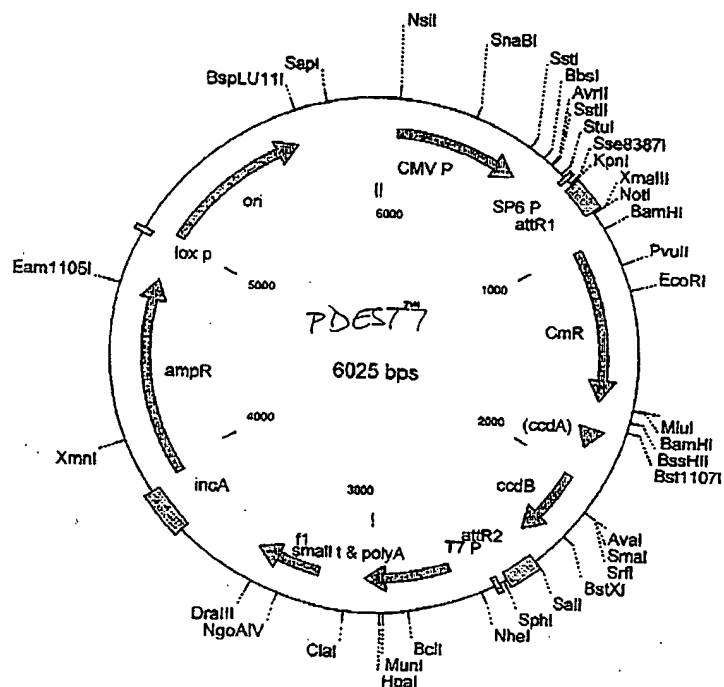
2701 ACGCGCCGAG ACAGAACTTA ATGGGCCCCG TAACAGCGCG ATTTGCTGGT GACCCAATGC
2761 GACCAAGATGC TCCACGCCCA GTCGCGTACC GTCTTCATGG GAGAAAATAA TACTGTTGAT
2821 GGGTGTCTGG TCAGAGACAT CAAGAAATAA CGCCGGAACA TTAGTGCAAG CAGCTTCCAC
2881 AGCAATGGCA TCCTGGTCAT CCAGCGGATA GTTAATGATC AGCCCACTGA CCCGTGCGC
2941 GAGAAGATTG TGCACCGCCG CTTTACAGGC TTCGACGCGG CTTGCTTCTA CCATCGACAC
3001 CACCACGCTG GCACCCAGTT GATCGGCGCG AGATTTAATC GCGCGACAA TTGCGACCG
3061 CGCGTGCAGG GCCAGACTGG AGGTGGCAAC GCCAATCAGC AACGACTGTT TGCCCGCCAG
3121 TTGTTGTGCC ACGCGGTTGG GAATGTAATT CAGCTCCGCC ATCGCCGCTT CCACCTTTTC
3181 CCGCGTTTTT GCAGAAACGT GGCTGGCCTG GTTACCACG CGGAAACGG TCTGATAAGA
3241 GACACCGGCA TACTCTGCGA CATCGTATAA CGTTACTGGT TTCACATTCA CCACCTGAA
3301 TTGACTCTCT TCCGGGCGCT ATCATGCCAT ACCGCGAAAG GTTTTGGGCC ATTCGATGGT
3361 GTCAACGTAA ATGCCGCTTC GCCTTGGCGC GCGAATTGCA AGCTCTGCAT TAATGAATCG
3421 GCCAACGCGC GGGGAGAGGC GGTTTGCGTA TTGGGCGCTC TTCCGCTTCC TCGCTCACTG
3481 ACTCGCTGCG CTCGGTCGTT CGGCTGCGGC GAGCGGTATC AGCTCACTCA AAGGCGGTAA
3541 TACGGTTATC CACAGAATCA GGGGATAACG CAGGAAAGAA CATGTGAGCA AAAGGCCAGC
3601 AAAAGGCCAG GAACCGTAAA AAGGCCGCGT TGCTGGCGTT TTTCCATAGG CTCGCCCCCC
3661 CTGACGAGCA TCACAAAAAT CGACGCTCAA GTCAGAGGTG GCGAAACCCG ACAGGACTAT
3721 AAAGATACCA GCGGTTTCCC CCTGGAAGCT CCTCGTGGC CTCTCTGTTT CCGACCTGCG
3781 CGCTTACCGG ATACCTGTCC GCCTTTCTCC CTTGCGGAAG CGTGGCGCTT TCTCAATGCT
3841 CACGCTGTAG GTATCTCAGT TCGGTGTAGG TCGTTCGCTC CAAGCTGGGC TGTGTGCACG
3901 AACCCCCCGT TCAGCCCGAC CGCTGCGCCT TATCCGGTAA CTATCGTCTT GAGTCCAACC
3961 CGGTAAGACA CGACTTATCG CCACTGGCAG CAGCCACTGG TAACAGGATT AGCAGAGCGA
4021 GGTATGTAGG CGGTGCTACA GAGTCTTGA AGTGGTGGCC TAACTACGGC TACACTAGAA
4081 GGACAGTATT TGGTATCTGC GCTCTGCTGA AGCCAGTTAC CTTGCGAAAA AGAGTTGGTA
4141 GCTCTTGATC CGGCAACAA ACCACCGCTG GTAGCGGTGG TTTTITTTGT TGCAAGCAGC
4201 AGATTACGCG CAGAAAAAAA GGTATCTCAAG AAGATCCTTT GATCTTTTCT ACGGGGTCTG
4261 ACGCTCAGTG GAACGAAAAAC TCACGTTAAG GGATTTTGGT CATGAGATTA TCAAAAAGGA
4321 TCTTCACCTA GATCCTTTTA AATTAAAAAT GAAGTTTAA ATCAATCTAA AGTATATATG
4381 AGTAAACTTG GTCTGACAGT TACCAATGCT TAATCAGTGA GGCACCTATC TCAGCGATCT
4441 GTCTATTTCG TCTCATCTCA GTTGCTGAC TCCCGTCTGT GTAGATAACT ACAGTACGGG
4501 AGGGCTTACC ATCTGGCCCC AGTGCTGCAA TGATACCGCG AGACCCACGC TCACCGGCTC
4561 CAGATTATAT AGCAATAAAC CAGCCAGCCG GAAGGGCCGA GCGCAGAAAT GGTCTGCAA
4621 CTTTATCCGC CTCCATCCAG TCTATTAAAT GTTGCCGGGA AGCTAGAGTA AGTAGTTCCG
4681 CAGTTAATAG TTGCGCAAC GTTGTGCGCA TTGCTACAGG CATCGTGGT TCACGCTCGT
4741 CGTTTGGTAT GGCTTCATTC AGCTCCGCTT CCAACGATC AAGGCGAGTT ACATGATCCC
4801 CCATGTTGTG CAAAAAGCG GTTAGCTCCT TCGGTCTCTC GATCGTTGTC AGAAGTAAGT
4861 TGGCCGCGAG GTTATCACTC ATGGTTATGG CAGCACTGCA TAATTCTCTT ACTGTCATGC
4921 CATCCGTAAG ATGCTTTTCT GTGACTGGTG AGTACTCAAC CAAGTCATTC TGAGAATAGT
4981 GTATGCGGCG ACCGAGTTGC TCTTGCCCGG CGTCAATACG GGATAATACC GCGCCACATA
5041 GCAGAACTTT AAAAGTGCTC ATCATTGGAA AACGTTCTTC GGGCGGAAAA CTCTCAAGGA
5101 TCTTACCGCT GTTGAGATCC AGTTCGATGT AACCCACTCG TGCACCCAAC TGATCTTCAG
5161 CATCTTTTAC TTTACCCAGC GTTTCTGGGT GAGCAAAAAC AGGAAGGCAA AATGCCGCAA
5221 AAAAGGGAAT AAGGGCGACA CGGAAATGTT GAATACTCAT ACTCTTCTT TTTCAATATT
5281 ATTGAAGCAT TTATCAGGGT TATTGTCTCA TGAGCGGATA CATATTGAA TGTATTTAGA
5341 AAAATAAACA AATAGGGGTT CCGCGCACAT TTCCCGGAAA AGTGCCACCT GAAATTTGTA
5401 ACGTTAATAT TTTGTTAAAA TTCGCGTTAA ATTTTGTGTA AATCAGCTCA TTTTAAACC
5461 AATAGGCCGA AATCGGCAAA ATCCCTTATA AATCAAAGA ATAGACCGAG ATAGGGTTGA
5521 GTGTTGTTCC AGTTTGGAAC AAGAGTCCAC TATTAAAGAA CGTGGACTCC AACGTCAAAG
5581 GCGGAAAAAC CGTCTATCAG GCGGATGGCC CACTACGTGA ACCATCACCC TAATCAAGTT
5641 TTTTGGGGTC GAGGTGCCGT AAAGCACTAA ATCGGAACCC TAAAGGGAGC CCCCATTATA
5701 GAGCTTGACG GGGAAAGCCG GCGAACGTGG CGAGAAAGGA AGGGAAGAAA GCGAAAGGAG
5761 CCGGCGCTAG GCGCTGGCA AGTGTAGCGG TCACGCTGGC CGTAACCAAC ACACCCGCGG
5821 CGCTTAATGC GCGCTACAG GCGCGCTCCA TTCGCCATTC AGGCTGCGCA ACTGTTGGGA
5881 AGGGCGATCG GTGCGGGCCT CTTGCTATT ACGCCAGCTG GCGAAAGGGG GATGTGCTGC
5941 AAGGCGATTA AGTTGGG

FIGURE 26b

Figure 27A: PDEST7

CMV promoter for eukaryotic expression

970 cca ttg acg caa atg ggc ggt agg cgt gta cgg tgg gag gtc tat ata agc
 ggt aac tgc gtt tac ccg cca tcc gca cat gcc acc ctc cag ata tat tcc
 1021 aga gct cgt tta gtg aac cgt cag atc gcc tgg aga cgc cat cca cgc tgt
 tct cga gca aat cac ttg gca gtc tag cgg acc tct gcg gta ggt gcg aca
 1072 ttt gac ctc cat aga aga cac cgg gac cga tcc agc ctc cgg act cta gcc
 aaa ctg gag gta tct tct gtg gcc ctg gct agg tcc gag ggc tga gat cgg
 1123 tag gcc gcg gag cgg ata aca att tca cac agg aaa cag cta tga cca cta
 atc cgg cgc ctc gcc tat tgt taa agt gtg tcc ttt gtc gat act ggt gat
 1174 ggc ttt tgc aaa aag cta ttt agg tga cac tat aga agg tac gcc tgc agg
 ccg aaa acg ttt ttc gat aaa tcc act gtg ata tct tcc atg cgg acg tct
 1225 ^{Kpn} ^{EcoRI} ^{NotI} ^{attR1}
 tac/cgg tcc gga att ccc atc aca agt tgg tag aaa ggt gaa cga gaa
 atg gcc agg cct taa/ggg tag tgt tca aac atg ttt tct cga ctc gct ctc



pDEST7 6025 bp (rotated to position 2800)

Location (Base Nos.)	Gene Encoded
67..589	CMV promoter
906..782	attR1
1015..1674	CmR
1794..1878	inactivated ccdA
2016..2321	ccdB
2362..2486	attR2
2671..3033	small t & polyA
3227..3502	f1
3962..4822	ampR
5022..5661	ori

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1  ATTATCATGA  CATTAACTTA  TAAAAATAGG  CGTAGTACGA  GGCCCTTTCA  CTCATTAGAT
61  GCATGTCGTT  ACATAACTTA  CGGTAAATGG  CCCGCCTGGC  TGACCGCCCA  ACGACCCCGG
121  CCCATTGACG  TCAATAATGA  CGTATGTTCC  CATAGTAACG  CCAATAGGGA  CTTTCCATTG
181  ACGTCAATGG  GTGGAGTATT  TACGGTAAAC  TGCCCACTTG  GCAGTACATC  AAGTGTATCA
241  TATGCCAAGT  ACGCCCCCTA  TTGACGTCAA  TGACGGTAAA  TGGCCCCGCT  GGCATTATGC
301  CCAGTACATG  ACCTTATGGG  ACTTTCCTAC  TTGGCAGTAC  ATCTACGTAT  TAGTCATCGC
361  TATTACCATG  GTGATGCGGT  TTTGGCAGTA  CATCAATGGG  CGTGGATAGC  GGTTTGACTC
421  ACGGGGATTT  CCAAGTCTCC  ACCCCATTGA  CGTCAATGGG  AGTTTGTITT  GGCACCAAAA
481  TCAACGGGAC  TTTCCAAAAT  GTCGTAACAA  CTCGCCCCCA  TTGACGCAAA  TGGGCGGTAG
541  GCGTGTACGG  TGGGAGGTCT  ATATAAGCAG  AGCTCGTTTA  GTGAACCGTC  AGATCGCCTG
601  GAGACGCCAT  CCACGCTGTT  TTGACCTCCA  TAGAAGACAC  CGGGACCGAT  CCAGCCTCCG
661  GACTCTAGCC  TAGGCCGCGG  AGCGGATAAC  AATTTACAC  AGGAAACAGC  TATGACCATT
721  AGGCCCTTGC  AAAAAGCTAT  TTAGGTGACA  CTATAGAAGG  TACGCCTGCA  GGTACCGGAT
781  CACAAGTTTG  TACAAAAAAG  CTGAACGAGA  AACGTAAAAT  GATATAAATA  TCAATATATT
841  AAATTAGATT  TTGCATAAAA  AACAGACTAC  ATAATACTGT  AAAACACAAC  ATATCCAGTC
901  ACTATGGCGG  CCGCATTAGG  CACCCAGGCG  TTTACACTTT  ATGCTTCCGG  CTCGTATAAT
961  GTGTGGATTT  TGAGTTAGGA  TCCGTCGAGA  TTTTCAGGAG  CTAAGGAAGC  TAAAATGGAG
1021  AAAAAAATCA  CTGGATATAC  CACCGTTGAT  ATATCCCAAT  GGCATCGTAA  AGAACATTTT
1081  GAGGCATTTC  AGTCAGTTGC  TCAATGTACC  TATAACCAGA  CCGTTCAGCT  GGATATTACG
1141  GCCTTTTTAA  AGACCGTAAA  GAAAAATAAG  CACAAGTTTT  ATCCGGCCTT  TATTACATT
1201  CTGCCCCGCC  TGATGAATGC  TCATCCGGAA  TTCCGTATGG  CAATGAAAGA  CGGTGAGCTG
1261  GTGATATGGG  ATAGTGTTCA  CCCTTGTTAC  ACCGTTTCC  ATGAGCAAA  TGAAACGTTT
1321  TCATCGCTCT  GGAGTGAATA  CCACGACGAT  TTCCGGCAGT  TTCTACACAT  ATATTGCAAA
1381  GATGTGGCGT  GTTACGGTGA  AAACCTGGCC  TATTTCCCTA  AAGGGTTTAT  TGAGAAATATG
1441  TTTTTCGTCT  CAGCCAATCC  CTGGGTGAGT  TTCACCACTT  TTGATTTAAA  CGTGGCCAAT
1501  ATGGACAAC  TCTTCGCCCC  CGTTTTCAAC  ATGGGCAAA  ATTATACGCA  AGGCACAAG
1561  GTGCTGATGC  CGCTGGCGAT  TCAGGTTTCA  CATGCCGTCT  GTGATGGCTT  CCATGTCGGC
1621  AGAATGCTTA  ATGAATTACA  ACAGTACTGC  GATGAGTGGC  AGGGCGGGGC  GTAAACGCGT
1681  GGATCCGGCT  TACTAAAAGC  CAGATAACAG  TATGCGTATT  TGCGCGCTGA  TTTTTCGGGT
1741  ATAAGAATAT  ATACTGATAT  GTATACCCGA  AGTATGTCAA  AAAGAGGTGT  GCTATGAAGC
1801  AGCGTATTAC  AGTGACAGTT  GACAGCGACA  GCTATCAGTT  GCTCAAGGCA  TATATGATGT
1861  CAATATCTCC  GGTCTGGTAA  GCACAACCAT  GCAGAATGAA  GCCCGTCGTC  TGCGTGCCGA
1921  ACGCTGGAAG  GCGGAAAATC  AGGAAGGGAT  GGCTGAGGTC  GCCCGGTTTA  TTGAAATGAA
1981  CGGCTCTTTT  GCTGACGAGA  ACAGGGACTG  GTGAAATGCA  GTTTAAGGTT  TACACCTATA
2041  AAAGAGAGAG  CCGTTATCGT  CTGTTTGTGG  ATGTACAGAG  TGATATTATT  GACACGCCCCG
2101  GCGGACGGAT  GGTGATCCCC  CTGGCCAGTG  CACGTCTGCT  GTCAGATAAA  GTCTCCCGTG
2161  AACTTTACCC  GGTGTGTCAT  ATCGGGGATG  AAAGCTGGCG  CATGATGACC  ACCGATATGG
2221  CCAGTGTGCC  GGTCTCCGTT  ATCGGGGAAG  AAGTGGCTGA  TCTCAGCCAC  CGCGAAAATG
2281  ACATCAAAAA  CGCCATTAA  CTGATGTTCT  GGGGAATATA  AATGTCAGGC  TCCCTTATAC
2341  ACAGCCAGTC  TGCAGGTCGA  CCATAGTGAC  TGGATATGTT  GTGTTTACA  GTATTATGTA
2401  GTCTGTTTTT  TATGCAAAAT  CTAATTTAAT  ATATTGATAT  TTATATCATT  TTACGTTTCT
2461  CGTTCAGCTT  TCTTGACAA  AGTGGTGATC  GCGTGATGC  GACGTCATAG  CTCTCTCCCT
2521  ATAGTGAGTC  GTATTATAAG  CTAGGCACTG  GCCGTCGTTT  TACAACGTCG  TGACTGGGAA

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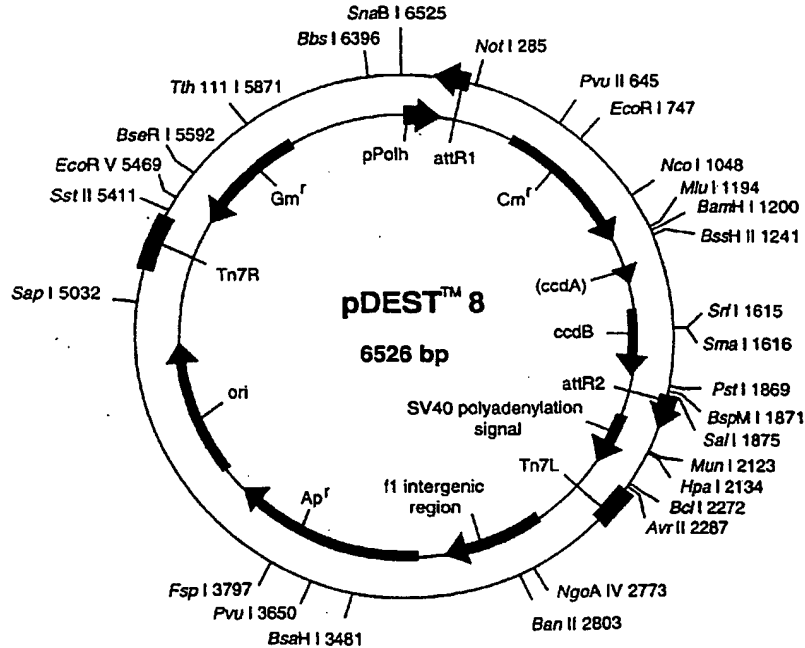
FIGURE 27B

2581 AACTGCTAGC TTGGGATCTT TGTGAAGGAA CCTTACTTCT GTGGTGTGAC ATAATTGGAC
2641 AAACCTACCTA CAGAGATTTA AAGCTCTAAG GTAAATATAA AATTTTTAAG TGTATAATGT
2701 GTTAAACTAG CTGCATATGC TTGCTGCTTG AGAGTTTTGC TTAGTGAGTA TGATTTATGA
2761 AAATATTATA CACAGGAGCT AGTGATTCTA ATTGTTTGTG TATTTTAGAT TCACAGTCCC
2821 AAGGCTCATT TCAGGCCCCCT CAGTCCTCAC AGTCTGTTCA TGATCATAAT CAGCCATACC
2881 ACATTTGTAG AGGTTTTACT TGCTTTAAAA AACCTCCAC ACCTCCCCCT GAACCTGAAA
2941 CATAAAATGA ATGCAATTGT TGTGTTAAAC TTGTTTATTG CAGCTTATAA TGGTTACAAA
3001 TAAAGCAATA GCATCACAAA TTTCACAAAT AAAGCATTIT TTTCACTGCA TTCCTAGTTGT
3061 GGTTTGTCCA AACTCATCAA TGTATCTTAT CATGTCGGA TCGATCCTGC ATTAATGAAT
3121 CGGCCAACGC GCGGGGAGAG GCGGTTTGCG TATTGGCTGG CGTAATAGCG AAGAGGCCCC
3181 CACCGATCGC CTTTCCCAAC AGTTGCGCAG CCTGAATGGC GAATGGGACG CGCCTGTAG
3241 CGGCGCATT AAGCGCGCGG GTGTGGTGGT TACGCGCAGC GTGACCGCTA CACTTGCCAG
3301 CGCCCTAGCG CCGGCTCCTT TCCTTTCTTT CCCTTCCTTT CTCGCCACGT TCGCCGGCTT
3361 TCCCGTCAA GCTCTAAATC GGGGGCTCCC TTTAGGGTTC CGATTTAGTG CTTTACGGCA
3421 CCTCGACCCC AAAAACTTG ATTAGGGTGA TGGTTCACGT AGTGGGCCAT CGCCCTGATA
3481 GACGGTTTTT CGCCCTTTGA CGTTGGAAGT CACGTTCTTT AATAGTGGAC TCTTGTTCGA
3541 AACTTGAACA ACACCTCAAC CTATCTCGGT CTATCTTTT GATTTATAAG AGCTTTTGCC
3601 GATTTGCGCC TATTGGTTAA AAAATGAGCT GATTTAAACA AAATTTAAGC CGAATTTTAA
3661 CAAAATATTA ACGTTTACAA TTTCAGGTGG CACTTTTCGG GGAATGTGC GCGGAACCCC
3721 TATTTGTTTA TTTTCTAAA TACATTCAA TATGTATCCG CTCATGCCAG GTCTTGGAT
3781 GGTGAGAACG GCTTGCTCGG CAGCTTCGAT GTGTGCTGGA GGGAGAATAA AGGTCTAAGA
3841 TGTGCGATAG AGGGAAGTCG CATTGAATTA TGTGCTGTGT AGGGATCGCT GGTATCAAAT
3901 ATGTGTGCCC ACCCTGTGCA TGAGACAATA ACCCTGATAA ATGCTTCAAT AATATTGAAA
3961 AAGGAAGAGT ATGAGTATTC AACATTTCCG TGTGCGCCTT ATTCCTTTT TTGCGGCATT
4021 TTGCTTCTCT GTTTTTGCTC ACCCAGAAAC GCTGGTGAAA GTAAAGATG CTGAAGATCA
4081 GTTGGGTGCA CGAGTGGGTT ACATCGAACT GGATCTCAAC AGCGGTAAGA TCCTTGAGAG
4141 TTTTCGCCCC GAAGAACGTT TTCCAATGAT GAGCACTTTT AAAGTTCCTG TATGTGGCGC
4201 GGTATTATCC CGTATTGACG CCGGGCAAGA GCAACTCGGT CGCCGCATAC ACTATTCTCA
4261 GAATGACTTG GTTGAGTACT CACCACTCAC AGAAAAGCAT CTTACGGATG GCATGACAGT
4321 AAGAGAATTA TGCAGTGCTG CCATAACCAT GAGTGATAAC ACTGCGGCCA ACTTACTTCT
4381 GACAACGATC GGAGGACCGA AGGAGCTAAC CGCTTTTTTG CACAACATGG GGGATCATGT
4441 AACTCGCCTT GATCGTTGGG AACCAGGAGT GAATGAAGCC ATACCAAACG ACGAGCGTGA
4501 CACCACGATG CCGTAGCAA TGGCAACAAC GTTGCACAAA CTATTAAGTCT GCGAAGTACT
4561 TACTCTAGCT TCCCGGCAAC AATTAATAGA CTGGATGGAG GCGGATAAAG TTGAGGAC
4621 ACTTCTGCGC TCGGCCCTTC CGGCTGGCTG GTTTATTGCT GATAAATCTG GAGCGGTGA
4681 GCGTGGGTCT CGCGGTATCA TTGCAGCACT GGGGCCAGAT GGTAAAGCCT CCCGTATCGT
4741 AGTTATCTAC ACGACGGGGA GTCAGGCAAC TATGGATGAA CGAAATAGAC AGCTCGCTGA
4801 GATAGGTGCC TCACTGATTA AGCATTGGTA ACTGTGAGC CAAGTTTACT CATATATACT
4861 TTAGATTGAT TTAACCTTC ATTTTAAAT TAAAGGATC TAGGTGAAGA TCCTTTTGA
4921 TAATCTCATG CCATAACTTC GTATAATGTA TGCTATACGA AGTTATGGCA TGACCAAAAT
4981 CCCTTAACGT GAGTTTTCGT TCCACTGAGC GTCAGACCCC GTAGAAAAGA TCAAAGGATC
5041 TTCTTGAGAT CCTTTTTTTC TGCGCGTAAT CTGCTGCTTG CAAACAAAAA AACCACCGCT
5101 ACCAGCGGTG GTTTGTTTGC CGGATCAAGA GCTACCAACT CTTTTCCGA AGGTAACCTG
5161 CTTCAAGACA TCTGTAGCAC CGCTACATA CCTCGCTCTG CTAATCCTGT TACCAGTGGC
5281 TGCTGCCAGT GGCGATAAGT CGTGTCTTAC CGGGTTGGAC TCAAGACGAT AGTTACCGGA
5341 TAAGGCGCAG CGGTGCGGCT GAACGGGGGG TTCGTGCACA CAGCCAGCT TGGAGCGAAC
5401 GACCTACACC GAAGTGAAT ACCTACAGCG TGAGCATTGA GAAAGCGCCA CGCTTCCCGA
5461 AGGGAGAAAG GCGGACAGGT ATCCGGTAAG CGGCAGGGTC GGAACAGGAG AGCGCACGAG
5521 GGAGCTTCCA GGGGAAACG CCTGGTATCT TTATAGTCTT GTGCGGTTTC GCCACCTCTG
5581 ACTTGAGCGT CGATTTTTGT GATGCTCGTC AGGGGGGCGG AGCCTATGGA AAAACGCCAG
5641 CAACGCGGCC TTTTACGGT TCCTGGCCTT TTGCTGGCCT TTTGCTCACA TGTCTTTCC
5701 TGGCTTATCC CTGTATTCTG TGGATAACCG TATTACCGCC TTTGAGTGAG CTGATACCGC
5761 TCGCCGAGC CGAACGACCG AGCGCAGCGA GTCAGTGAGC GAGGAAGCGG AAGAGCGCCC
5821 AATACGCAAA CGCCTCTCC CGCGGTTTG GCGGATTCAT TAATGCAGAG CTTGCAATTC
5881 GCGCGTTTTT CAATATTATT GAAGCATTTA TCAGGGTTAT TGTCTCATGA GCGGATACAT
5941 ATTTGAATGT ATTTAGAAAA ATAAACAAAT AGGGGTTCCG CGCACATTTC CCCGAAAAGT
6001 GCCACCTGAC GTCTAAGAAA CCAAT

FIGURE 27C

Figure 28A: pDEST8 Polyhedron Promoter, Baculovirus Transfer Plasmid

1 *AacI*
 cgt ata ctc cgg aat att aat aga tca tgg aga taa tta aaa tga taa cca
 gca tat gag gcc tta taa tta tct agt acc tct att aat ttt act att ggt
 52 tct cgc aaa taa ata agt att tta ctg ttt tgg taa cag ttt tgt aat aaa
 aga gcg ttt att tat tca taa aat gac aaa agc att gtc aaa aca tta ttt
 103 aaa acc tat aaa tat tcc gga tta ttc ata ccg tcc cac cat cgg gcg cgg
 ttt tgg ata ttt ata agg cct aat aag tat ggc agg gtg gta gcc cgc gcc
 154 *Bam* *Int* *attR1*
 atc atc aca agt ttg tgg gaa aag gct gaa cga gaa agg taa aat aat ata
 tag tag tgt tca aac atg ttt ttc cga ctt gct ctt tgc att tta cta tat



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pDEST8 6526 bp

Location (Base Nos.)	Gene Encoded
23..152	Ppolh
284..160	attR1
534..1193	CmR
1313..1397	inactivated ccdA
1535..1840	ccdB
1881..2005	attR2
2766..3146	f1
3240..4090	ampR
4289..4869	ori
5564..6496	genR

1	CGTATACTCC	GGAATATTAA	TAGATCATGG	AGATAATTAA	AATGATAACC	ATCTCGCAAA
61	TAAATAAGTA	TTTTACTGTT	TTCGTAACAG	TTTTGTAATA	AAAAAACCTA	TAAATATTCC
121	GGATTATTCA	TACCGTCCCA	CCATCGGGCG	CGGATCATCA	CAAGTTTGTA	CAAAAAAGCT
181	GAACGAGAAA	CGTAAATGA	TATAAATATC	AATATATTAA	ATTAGATTTT	GCATAAAAAA
241	CAGACTACAT	AATACTGTAA	AACACAACAT	ATCCAGTCAC	TATGGCGGCC	GCTAAGTTGG
301	CAGCATCACC	CGACGCACCT	TGCGCCGAAT	AAATACCTGT	GACGGAAGAT	CACCTTCGCAG
361	AATAAATAAA	TCCTGGTGTC	CCTGTTGATA	CCGGGAAGCC	CTGGGCCAAC	TTTTGGCGAA
421	AATGAGACGT	TGATCGGCAC	GTAAGAGGTT	CCAACCTTCA	CCATAATGAA	ATAAGATCAC
481	TACCGGGCGT	ATTTTTTGAG	TTATCGAGAT	TTTCAGGAGC	TAAGGAAGCT	AAAATGGAGA
541	AAAAAATCAC	TGGATATACC	ACCGTTGATA	TATCCCAATG	GCATCGTAAA	GAACATTTTG
601	AGGCATTTCA	GTCAAGTTGCT	CAATGTACCT	ATAACCGAC	CGTTCAGCTG	GATATTACGG
661	CCTTTTTTAA	GACCGTAAAG	AAAAATAAGC	ACAAGTTTAA	TCCGGCCTTT	ATTCACATTTC
721	TTGCCCGCCT	GATGAATGCT	CATCCGGAAT	TCCGTATGGC	AATGAAAGAC	GGTGAGCTGG
781	TGATATGGGA	TAGTGTTCAC	CCTTGTTACA	CCGTTTTCCA	TGAGCAAACT	GAACGTTTTT
841	CATCGCTCTG	GAGTGAATAC	CACGACGATT	TCCGCGAGTT	TCTACACATA	TATTCCGAAG
901	ATGTGGCGTG	TTACGGTGAA	AACCTGGCCT	ATTTCCCTAA	AGGGTTTATT	GAGAAATATGT
961	TTTTTCGTCTC	AGCCCAATCCC	TGGGTGAGTT	TCACCAAGTT	TGATTTAAAC	GTGGCCCAAT
1021	TGGACAACCT	CTTCGCCCCC	GTTTTACCA	TGGGCAATA	TTATACGCAA	GGCGACAAGG
1081	TGCTGATGCC	GCTGGCGATT	CAGGTTTCATC	ATGCCGCTCTG	TGATGGCTTC	CATGTCGGCA
1141	GAATGCTTAA	TGAATTACAA	CAGTACTGCG	ATGAGTGGCA	GGCGGGGCG	TAAACGCGTG
1201	GATCCGGCTT	ACTAAAAGCC	AGATAACAGT	ATGCGTATTT	GCGCGCTGAT	TTTTGCGGTA
1261	TAAGAATATA	TACTGATATG	TATACCCGAA	GTATGTCAAA	AAGAGGTGTG	CTATGAAGCA
1321	GCGTATTACA	GTGACAGTTG	ACAGCGACAG	CTATCAGTTG	CTCAAGGCAT	ATATGATGTC
1381	AATATCTCCG	GTCTGGTAAG	CACAACCATG	CAGAATGAAG	CCCGTCGTCT	GCGTGGCCGAA
1441	CGCTGGAAAG	CGGAAAATCA	GGAAGGGATG	GCTGAGGTCG	CCCGGTTTAT	TGAAATGAAC
1501	GGCTCTTTTG	CTGACGAGAA	CAGGGACTGG	TGAAATGCAG	TTTAAGGTTT	ACACCTATAA
1561	AAGAGAGAGC	CGTTATCGTC	TGTTTGTTGA	TGTACAGAGT	GATATTATTG	ACACGCCCGG
1621	GCGACGGATG	GTGATCCCCC	TGGCCAGTGC	ACGTCTGCTG	TCAGATAAAG	TCTCCCGTGA
1681	ACTTTACCCG	GTGGTGCATA	TCGGGGATGA	AAGCTGGCGC	ATGATGACCA	CCGATATGGC
1741	CAGTGTGCCG	GTCTCCGTTA	TCGGGGAAGA	AGTGGCTGAT	CTCAGCCACC	GCGAAAATGA
1801	CATCAAAAAC	GCCATTAAAC	TGATGTTCTG	GGGAATATAA	ATGTCAGGCT	CCCTTATACA
1861	CAGCCAGTCT	GCAGGTCGAC	CATAGTGACT	GGATATGTTG	TGTTTTACAG	TATTATGTAG
1921	TCTGTTTTTT	ATGCAAAATC	TAATTTAATA	TATTGATATT	TATATCATTT	TACGTTTCTC
1981	GTTTCAGCTTT	CTTTGACAAA	GTGGTGATAG	CTTGTCGAGA	AGTACTAGAG	GATCATAATC
2041	AGCCATACCA	CATTTGTAGA	GGTTTTACTT	GCTTTAAAAA	ACCTCCCAAC	CCTCCCCCTG
2101	AACCTGAAAC	ATAAAATGAA	TGCAATTGTT	GTTGTTAACT	TGTTTATTGC	AGCTTATAAT
2161	GGTTACAAAT	AAAGCAATAG	CATCACAAT	TTCACAATAA	AAGCATTTTT	TCACTGCAT
2221	TCTAGTTGTG	GTTTGTCCAA	ACTCATCAAT	GTATCTTATC	ATGTCTGGAT	CTGATCACTG
2281	CTTGAGCCTA	GGAGATCCGA	ACCAGATAAG	TGAAATCTAG	TTCCAAACTA	TTTTGTCAAT
2341	TTTAATTTTC	GTATTAGCTT	ACGACGCTAC	ACCCAGTTCC	CATCTATTTT	STCACTCTTC
2401	CCTAAATAT	CCTTAAAAAC	TCCATTTCCA	CCCCCTCCAG	TTCCCAACTA	TTTTGTCCCG
2461	CCACAGCGGG	GCATTTTTCT	TCCTGTTATG	TTTTTAATCA	AACATCCTCG	CAACTCCATG
2521	TGACAAACCG	TCATCTTCGG	CTACTTTTTT	TCTGTCACAG	AATGAAAATT	TTTCTGTCAT

FIGURE 28B

2581 CTCTTCGTTA TTAATGTTTG TAATTGACTG AATATCAACG CTTATTGCA SCCTGAATGG
2641 CGAATGGACG CGCCCTGTAG CGGCGCATTAG AGCGCGGGG GTGTGGTGGT TACGCGCAGC
2701 GTGACCGCTA CACTTGCCAG CGCCCTAGCG CCCGCTCCTT TCGCTTCTTT CCCTTCCTTT
2761 CTCGCCACGT TCGCCGGCTT TCCCGCTCAA GCTCTAAATC GGGGGCTCCC TTAGGGTTC
2821 CGATTTAGTG CTTTACGGCA CCTCGACCCC AAAAACTTG ATTAGGGTGA TGGTTCACGT
2881 AGTGGGCCAT CGCCCTGATA GACGGTTTTT CGCCCTTGA CGTTGGAGTC CACGTTCTTT
2941 AATAGTGGAC TCITGTTCCA AACTGGAACA ACACCAACC CTATCTCGGT CTATTCTTTT
3001 GATTTATAAG GGATTTTGCC GATTTTCGGC TATTGGTTAA AAAATGAGCT GATTTAACAA
3061 AAATTTAACG CGAATTTTAA CAAAATATTA ACGTTTACAA TTTCAGTGG CACTTTTCGG
3121 GGAAATGTGC GCGGAACCCC TAITTGTITA TTTTCTAAA TACATTCAA TATGTATCCG
3181 CTCATGAGAC AATAACCCCTG ATAAATGCTT CAATAATATT GAAAAAGGAA GAGTATGAGT
3241 ATTCAACATT TCCGTGTCGC CCTTATTCCC TTTTTCGGG CATTTTGCTT TCCTGTTTTT
3301 GCTCACCCAG AAACGCTGGT GAAAGTAAAA GATGCTGAG ATCAGTTGGG TGCACGAGTG
3361 GGTACATCG AACTGGATCT CAACAGCGGT AAGATCCTTG AGAGTTTTCG CCCCAGAGAA
3421 CGTTTTCCAA TGATGAGCAC TTTTAAAGTT CTGCTATGTG GCGCGGTATT ATCCCGTATT
3481 GACGCGGGG AAGAGCAACT CGGTCGCGC ATACACTATT CTCAGAATGA CTTGGTTGAG
3541 TACTACCCAG TCACAGAAAA GCATCTTACG GATGGCATGA CAGTAAGAGA ATTATGAGT
3601 GCTGCCATAA CCATGAGTGA TAACACTGCG GCCAACTTAC TTCTGACAA GATCGGAGGA
3661 CCGAAGGAGC TAACCGCTTT TTTGCACAAC ATGGGGGATC ATGTAACCTG CCTTGATCGT
3721 TGGGAACCGG AGCTGAAATGA AGCCATACCA AACGACGAGC GTGACACCAC GATGCCCTGA
3781 GCAATGGCAA CAACGTTGCG CAACTATTA ACTGGCGAAC TACTTACTCT AGCTTCCCGG
3841 CAACAATTA TAGACTGGAT GGAGGCGGAT AAAGTTGCG GACCACTTCT GCGCTCGGCC
3901 CTTCGGCTG GCTGGTTTAT TGCTGATAAA TCTGGAGCCG GTGAGCGTGG GTCTCCGGGT
3961 ATCATGTCAG CACTGGGGCC AGATGGTAAG CCCTCCCGTA TCGTAGTTAT CTACACGACG
4021 GGGAGTCAGG CAACTATGGA TGAACGAAAT AGACAGATCG CTGAGATAGG TGCCTCACTG
4081 ATTAAGCATT GGTAACTGTC AGACCAAGTT TACTCATATA TACTTTAGAT TGATTTAAAA
4141 CTTCAATTTT AATTTAAAAG GATCTAGGTG AAGATCCTTT TTGATAATCT CATGACCAAA
4201 ATCCCTTAAC GTGAGTTTTC GTTCCACTGA GCGTCAGACC CCGTAGAAAA GATCAAGGA
4261 TCTTCTTGAG ATCCTTTTTT TCTGCGCGTA ATCTGCTGCT TGCAAAACAA AAAACCCCG
4321 CTACCGCGG TGGTTTGTG GCCGGATCAA GAGCTACCAA CTCTTTTTC GAAGGTAAC
4381 GGCCTCAGCA GAGCGCAGAT ACCAATACT GTCCTTCTAG TGTAGCCGTA GTTAGGCCAC
4441 CACTTCAAGA ACTCTGTAGC ACCGCTTACA TACCTCGCTC TGCTAATCCT GTTACCAGTG
4501 GCTGCTGCCA GTGGCGATAA GTCGTGCTT ACCGGGTTGG ACTCAAGAG ATAGTTACCG
4561 GATAAGGCGC AGCGGTGCGG CTGAACGGGG GGTTCGTGCA CACAGCCAG CTTGGAGCGA
4621 ACGACCTACA CCGAAGTGA ATACCTACAG CGTGAGCATT GAGAAAGCGC CACGCTTCCC
4681 GAAGGGAGAA AGGCGGACAG GTATCCGGTA AGCGGCAGGG TCGGAACAGG AGAGCGCAG
4741 AGGGAGCTTC CAGGGGGAAG CGCCTGGTAT CTTTATAGTC CTGTGCGGTT TCGCCACCTC
4801 TGACTTGAGC GTCGATTTTT GTGATGCTCG TCAGGGGGG GAGCCTATG GAAAAACGCC
4861 AGCAACCGCG CCTTTTACG GTTCTGGGCC TTTTGCTGGC CTTTGTCTCA CATGTTCTTT
4921 CCTGCGTTAT CCCCTGATTC TGTGGATAAC CGTATTACCG CCTTTGAGTG AGCTGATACC
4981 GCTCGCGCA GCCGAACGAC CGAGCGCAGC GAGTCAGTGA GCGAGGAAGC GGAAGAGCGC
5041 CTGATGCGGT ATTTTCTCCT TACGCATCTG TCGGTATTT CACACCCGAG ACCAGCCCGG
5101 TAACCTGGCA AAATCGGTTA CGGTTGAGTA ATAAATGGAT GCCCTGCGTA AGCGGGTGTG
5161 GGCGGACAAT AAAGTCTTAA ACTGAACAA ATAGATCTAA ACTATGACAA TAAAGTCTTA
5221 AACTAGACAG AATAGTTGTA AACTGAAATC AGTCCAGTTA TGCTGTGAAA AAGCATACTG
5281 GACTTTTGTG ATGGCTAAAG CAACTCTTC ATTTTCTGAA GTGCAAAATG CCGTCTGAT
5341 TAAAGAGGGG CGTGGCCAAG GGCATGGTAA AGACTATATT CGCGCGGTTG TGACAATTTA
5401 CCGAACAACT CCGCGGCCGG GAAGCCGATC TCGGCTTGAA CGAATTGTTA GGTGGCGGTA
5461 CTGGGTGCGA TATCAAAAGT CATCACTTCT TCCCGTATGC CCAACTTTGT ATAGAGAGCC
5521 ACTGCGGGAT CGTCACCGTA ATCTGCTTGC ACGTAGATCA CATAAGCACC AAGCGCGTTG
5581 GCCTCATGCT TGAGGAGATT GATGAGCGCG GTGGCAATGC CCTGCCTCCG GTGCTCGCCG
5641 GAGACTGCGA GATCATAGAT ATAGATCTCA CTACGCGGCT GCTCAAACCT GGGCAGAACG
5701 TAAGCCGCGA GAGCGCCAAC AACCGCTTCT TGGTCGAAGG CAGCAAGCGC GATGAATGTC
5761 TTACTACGGA GCAAGTTCCC GAGGTAATCG GAGTCCGGCT GATGTTGGGA GTAGGTGGCT
5821 ACGTCTCCGA ACTCAGGACC GAAAAGATCA AGAGCAGCCC GCATGGATTT GACTTGGTCA
5881 GGGCCGAGCC TACATGTGCG AATGATGCCC ATACTTGAGC CACCTAATT TGTTTTAGGG
5941 CGACTGCCCT GCTGCGTAAC ATCGTTGCTG CTGCGTAACA TCGTTGCTGC TCCATAACAT
6001 CAAACATCGA CCCACGGCGT AACCGCCTTG CTGCTGGAT GCGCGAGGA TAGACTGTAC

FIGURE 28C

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6061 AAAAAACAG TCATAACAAG CCATGAAAAC CGCCACTGCG CCGTTACCAC CGCTGCGTTC
6121 GGTCAAGGTT CTGGACCAGT TGCCTGAGCG CATACGCTAC TTGCATTACA GTTTACGAAC
6181 CGAACAGGCT TATGTCAACT GGGTTCGTGC CTTTCATCCGT TTCCACGGTG TGCCTCACCC
6241 GGCAACCTTG GGCAGCAGCG AAGTCGAGGC ATTTCTGTCC TGGCTGGCGA ACGAGCGCAA
6301 GGTTCGGTTC TCCACGCATC GTCAGGCATT GCGGCGCTTG CTGTTCTTCT ACGGCAAGGT
6361 GCTGTGCACG GATCTGCCCT GGCTTCAGGA GATCGGAAGA CCTCGGCCGT CGCGGCGCTT
6421 GCGGTGGTGT CTGACCCCGG ATGAAGTGGT TCGCATCCTC GGTTCCTCTG AAGGCGAGCA
6481 TCGTTGTTC GCCCAGGACT CTAGCTATAG TTCTAGTGGT TGGCTA
```

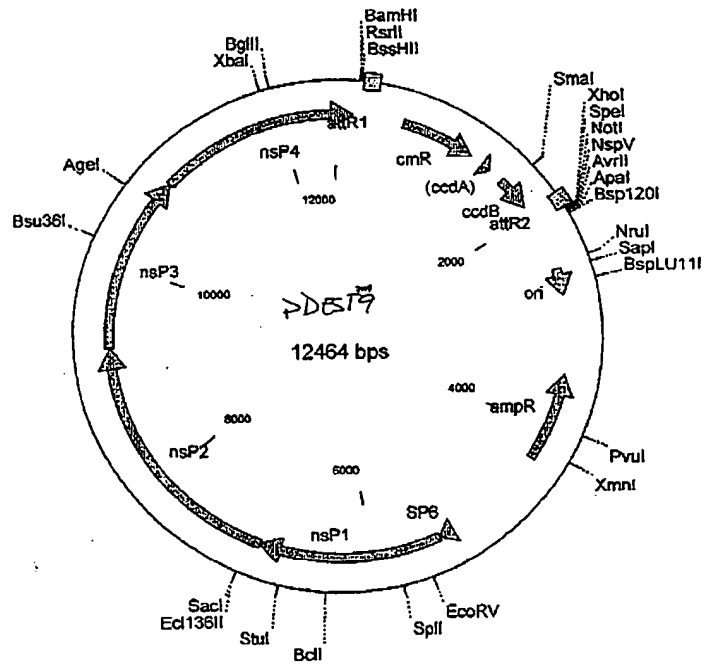
FIGURE 28D

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Figure 29A: pDEST9

Semliki Forest Virus vector

103 ttg gcg agg gac att aag gcg ttt aag aaa ttg aga gga cct gtt ata ~~cat~~
 aac cgc tcc ctg taa ttc cgc aaa ttc ttt aac tct cct gga caa tat ~~ctg~~
 154 ~~ctc tac ggc ggt cct agt ttg gtc~~ cgt taa tac aca gaa ttc tga ttg ~~gat~~
 gag atg ~~ccg cca gga tct aac cgc~~ gca att atg tgt ctt aag act aac cta
 205 ~~ccc ggt ccg aag cgc gct ttc cca tca~~ ~~aca agt tgg tac aac aag gct gga~~
 ggg cca ~~gac ttc gcg cga aag ggt agt~~ ~~tgt tca aac atg ttt tct cga ctc~~



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pDEST9 12464 bp

Location (Base Nos.)	Gene Encoded
355..232	attR1
605..1264	CmR
1384..1468	inactivated ccdA
1606..1911	ccdB
1952..2078	attR2
2532..2782	ori
3482..4282	ampR
5232..5365	SP6 promoter
5365..6965	nsP1:non-structural protein 1
6965..9265	nsP2:non-structural protein 2
9265..10865	nsP3:non-structural protein 3
10865..161	nsP4:non-structural protein 4

1	AGCAAGTGGT	TCCGGACAGG	CTTGGGGGCC	GAACGTGAGG	TGGCACTAAC	ATCTAGGTAT
61	GAGGTAGAGG	GCTGCAAAAG	TATCCTCATA	GCCATGGCCA	CCTTGGCGAG	GGACATTAAAG
121	GCGTTTAAGA	AATTGAGAGG	ACCTGTTATA	CACCTCTACG	GCGGTCCCTAG	ATTGGTGCCT
181	TAATACACAG	AATTCTGATT	GGATCCCGGT	CCGAAGCGCG	CTTTCCCATC	ACAAGTTTGT
241	ACAAAAAAGC	TGAACGAGAA	ACGTAAAATG	ATATAAATAT	CAATATATTA	AATTAGATTT
301	TGCATAAAAA	ACAGACTACA	TAATACTGTA	AAACACAACA	TATCCAGTCA	CTATGGCGGC
361	CGCTAAGTTG	GCAGCATCAC	CCGACGCACT	TTGCGCCGAA	TAAATACCTG	TGACGGAAGA
421	TCACTTCGCA	GAATAAATA	ATCCTGGTGT	CCCTGTTGAT	ACCGGGAAGC	CCTGGGCCAA
481	CTTTTGGCGA	AAATGAGACG	TTGATCGGCA	CGTAAGAGGT	TCCAACCTTC	ACCATAATGA
541	AATAAGATCA	CTACCGGGCG	TATTTTGTGA	GTTATCGAGA	TTTTCAGGAG	CTAAGGAAGC
601	TAAAAATGGAG	AAAAAATCA	CTGGATATAC	CACCGTTGAT	ATATCCCAAT	GGCATCGTAA
661	AGAACATTTT	GAGGCATTTT	AGTCAGTTGC	TCAATGTACC	TATAACCAGA	CCGTTTCAGCT
721	GGATATTACG	GCCTTTTAA	AGACCGTAAA	GAAAAATAAG	CACAAGTTTT	ATCCGGCCTT
781	TATTCACATT	CTTGCCCGCC	TGATGAATGC	TCATCCGGA	TTCCGTATGG	CAATGAAAGA
841	CGGTGAGCTG	GTGATATGGG	ATAGTGTTC	CCCTGTTTAC	ACCGTTTTCC	ATGAGCAAAAC
901	TGAAACGTTT	TCATCGCTCT	GGAGTGAATA	CCACGACGAT	TTCCGGCAGT	TTCTACACAT
961	ATATTTCGCA	GATGTGGCGT	GTTACGGTGA	AAACCTGGCC	TATTTCCCTA	AAGGGTTTTAT
1021	TGAGAATATG	TTTTTCGTCT	CAGCCAATCC	CTGGGTGAGT	TTCAACAGTT	TTGATTTAAA
1081	CGTGGCCAAT	ATGGACAAC	TCCTCGCCCC	CGTTTTTACC	ATGGGCAAT	ATTATACGCA
1141	AGGCGACAAG	GTGCTGATGC	CGCTGGCGAT	TCAGGTTTAT	CATGCCGTCT	GTGATGGCTT
1201	CCATGTCGGC	AGAATGCTTA	ATGAATTACA	ACAGTACTGC	GATGAGTGGC	AGGGCGGGGC
1261	GTAAAGATCT	GGATCCGGCT	TACTAAAAGC	CAGATAACAG	TATGCGTATT	TGCGCGCTGA
1321	TTTTTTCGGT	ATAAGAATAT	ATACTGATAT	GTATACCCGA	AGTATGTCAA	AAAGAGGTGT
1381	GCTATGAAGC	AGCGTATTAC	AGTGACAGTT	GACAGCGACA	GCTATCAGTT	GCTCAAGGCA
1441	TATATGATGT	CAATATCTCC	GGTCTGGTAA	GCACAACCAT	GCAGAATGAA	GCCCGTCGTC
1501	TGCGTGCCGA	ACGCTGGAAA	GCGGAAAATC	AGGAAGGGAT	GGCTGAGGTC	GCCCGTTTTA
1561	TTGAAATGAA	CGGCTCTTTT	GCTGACGAGA	ACAGGGACTG	GTGAAATGCA	GTTTAAGGTT
1621	TACACCTATA	AAAGAGAGAG	CCGTATTCGT	CTGTTTGTGG	ATGTACAGAG	TGATATTATT
1681	GACACGCCCG	GGCGACGGAT	GGTGATCCCC	CTGGCCAGTG	CACGTCTGCT	GTCAGATAAA
1741	GTCTCCCGTG	AACTTTACCC	GGTGGTGAT	ATCGGGGATG	AAAGCTGGCG	CATGATGACC
1801	ACCGATATGG	CCAGTGTGCC	GGTCTCCGTT	ATCGGGGAAG	AAGTGGCTGA	TCTCAGCCAC
1861	CGCGAAAATG	ACATCAAAAA	CGCCATTAA	CTGATGTTCT	GGGGAATATA	AATGTCAGGC
1921	TCCCTTATAC	ACAGCCAGTC	TGCAGGTCGA	CCATAGTGAC	TGGATATGTT	GTGTTTTACA
1981	GTATTATGTA	GTCTGTTTTT	TATGCAAAAG	TGCTAATTTA	ATATATTGAT	ATTATATATCA
2041	TTTTACGTTT	CTCGTTCAGC	TTTCTGTAC	AAAGTGGTGA	TGGGAACTCG	AGTTCACTAG
2101	TCGATCCCGC	GGCCGCTTTC	GAACCTAGGC	AAGCATGCGG	GCCCAAGTGG	TAATTAATTG
2161	AATTACATCC	CTACGCAAA	GTTTTACGGC	CGCCGGTGGC	GCCCGCGCCC	GGCGGCCCGT
2221	CCTTGCCCGT	TGCAGGCCAC	TCCGGTGGCT	CCCGTCGTCC	CCGACTTCCA	GGCCAGCAGC
2281	ATGCAGCAAC	TCATCAGCGC	CGTAAATGCG	CTGACAATGA	GACAGAACGC	AATTGCTCCT
2341	GCTAGGAGCT	TAATTGACG	AATAATTGGA	TTTTTATTIT	ATTTTGCAAT	TGGTTTTTTAA
2401	TATTTCCAAA	AAAAAAAAAA	AAAAAAAAAA	AAAAAAAAAA	AAAAAAAAAA	AAAAAAAAAA

FIGURE 29B

2461 AAAAAAAAAA AAAAAAATA GAAATCGCGA TTTCTAGTCT GCATTAATGA ATCGGCCAAC
 2521 GCGCGGGGAG AGGCGGTTTG CGTATTGGGC GCTCTTCCGC TTCCTCGCTC ACTGACTCGC
 2581 TGCGCTCGGT CGTTCGGCTG CGGCGAGCGG TATCAGCTCA CTCAAAGCGG GTAATACGGT
 2641 TATCCACAGA ATCAGGGGAT AACGCGAGAA AGAACATGTG AGCAAAAGGC CAGCAAAAGG
 2701 CCAGGAACCG TAAAAAGGCC GCGTTGCTGG CGTTTTCCA TAGGCTCCGC CCCCCTGACG
 2761 AGCATCACAA AAATCGACGC TCAAGTCAGA GGTGGCGAAA CCGCAGAGGA CTATAAGAT
 2821 ACCAGGCGTT TCCCCTTGGG AGCTCCCTCG TCGCTCTCC TGTTCGACC CTGCCGCTTA
 2881 CCGGATACCT GTCGCGCTTT CTCCCTTCGG GAAGCGTGGC GCTTTCTCAA TGCTCGCGCT
 2941 GTAGGTATCT CAGTTCGGTG TAGGTCTTC GCTCCAAGCT GGGCTGTGTG CACGAACCCC
 3001 CCGTTCAGCC CGACCGCTGC GCCTTATCCG GTAACATCG TCTTGAGTCC AACCCGGTAA
 3061 GACACGACTT ATCGCCACTG GCAGCAGCCA CTGGTAACAG GATTAGCAGA GCGAGGTATG
 3121 TAGGCGGTGC TACAGAGTTC TTGAAGTGGT GGCCTAACTA CGGCTACACT AGAAGGACAG
 3181 TATTGGGTAT CTGCGCTCTG CTGAAGCCAG TTACCTTCGG AAAAAGAGTT GGTAGCTCTT
 3241 GATCCGGCAA ACAAAACCACC GCTGGTAGCG GTGGTTTTTT TGTTCGAAG CAGCAGATTA
 3301 CGCGCAGAAA AAAAGGATCT CAAGAAGATC CTTTGATCTT TTCTACGGGG TCTGACGCTC
 3361 AGTGAACGA AAACCTACGT TAAGGGATTT TGGTCATGAG ATTATCAAAA AGGATCTTCA
 3421 CCTAGATCCT TTTAAATTA AAATGAAGTT TTAATCAAT CTAAAGTATA TATGAGTAAA
 3481 CTTGGTCTGA CAGTTACCAA TGCTTAATCA GTGAGGCACC TATCTCAGCG ATCTGTCTAT
 3541 TTCGTTTCATC CATAGTTGCC TGACTCCCCG TCGTGTAGAT AACTACGATA CGGGAGGGCT
 3601 TACCATCTGG CCCCAGTCTC GCAATGATAC CGCGAGACCC ACGCTCACC GCTCCAGATT
 3661 TATCAGCAAT AAACCAGCCA GCCGGAAGGG CCGAGCGCAG AAGTGGTCTT GCAACTTTAT
 3721 CCGCCTCCAT CCAGTCTATT AATTGTTGCC GGGAAAGTAG AGTAAGTAGT TCGCCAGTTA
 3781 ATAGTTTGGC CAACGTTGTT GCCATTGCTA CAGGCATCGT GGTGTACGC TCGTCTGTTG
 3841 GTATGGCTTC ATTCAGCTCC GGTTCCTAAC GATCAAGCGC AGTTACATGA TCCCCATGT
 3901 TGTGCAAAA AGCGGTIAGC TCCTTCGGTC CTCCGATCGT TGTGAGAAGT AAGTTGGCCG
 3961 CAGTGTATC ACTCATGTT ATGGCAGCAC TGCATAATTC TCTTACTGTC ATGCCATCCG
 4021 TAAGATGCTT TTCTGTGACT GGTGAGTACT CAACCAAGTC ATTCTGAGAA TAGTGTATGC
 4081 GCGGACCGAG TTGCTCTTGC CCGGCGTCAA TACGGGATAA TACCGCGCCA CATAGCAGAA
 4141 CTTTAAAGT GCTCATCATT GGAAAACGTT CTTCCGGGCG AAAACTCTCA AGGATCTTAC
 4201 CGCTGTTGAG ATCCAGTTCG ATGTAACCCA CTGTCGACCC CAACTGATCT TCAGCATCTT
 4261 TTACTTTTAC CAGCGTTTCT GGGTGAGCAA AAACAGGAAG GCAAAATGCC GCAAAAAGG
 4321 GAATAAGGGC GACACGGA AAATGTAATAC TCATACTCTT CTTTTTCAA TATTATTGAA
 4381 GCATTATCA GGGTATTGTT CTCATGAGCG GATACATATT TGAATGTATT TAGAAAAATA
 4441 AACAAATAGG GGTTCGCGC ACATTTCCCC GAAAAGTGCC ACCTGACGTC TAAGAAACCA
 4501 TTATTATCAT GACATTAACC TATAAAAATA GCGGTATCAC GAGGCCCTTT GCTCTCGCGC
 4561 GTTTCGGTGA TGACGGTGAA AACCTCTGAC ACATGCAGCT CCCGAGACG GTACAGCTT
 4621 CTGTCTAAGC GGATGCCGGG AGCAGACAAG CCCGTCAGGG CCGCTCAGCG GGTGTGGCG
 4681 GGTGTGGGG CTGGCTTAAC TATGCGGCAT CAGAGCAGAT TGTACTGAGA GTGCACCATA
 4741 TCGACGCTCT CCCTTATGCG ACTCCTGCAT TAGGAAGCAG CCCAGTACTA GGTGAGGCC
 4801 GTTAGCACC GCCCGCGCAA GGAATGGTGC ATGCAAGGAG ATGGCGCCA ACAGTCCCCC
 4861 GGCCACGGGG CCTGCCACCA TACCACGCC GAAACAAGCG CTCATGAGCC CGAAGTGGCG
 4921 AGCCCGATCT TCCCCTCGG TGATGTCGGC GATATAGGCG CCAGCAACCG CACCTGTGGC
 4981 GCGGTTGATG CCGGCCACGA TGCGTCCGGC GTAGAGGATC TGGCTAGCGA TGACCCTGCT
 5041 GATTGGTTCG CTGACCATTT CCGGGGTGCG GAACGGCGTT ACCAGAACT CAGAAGGTT
 5101 GTCCAACCAA ACCGACTCTG ACGGCAGTTT ACGAGAGAGA TGATAGGGTC TGCTTCAGTA
 5161 AGCCAGATGC TACACAATTA GGCCTGTACA TATTGTCTGT AGAACGCGGC TACAATTAAT
 5221 ACATAACCTT ATGTATCATA CACATACGAT TTAGGTGACA CTATAGATGG CGGATGTGTG
 5281 ACATACACGA CGCCAAAAGA TTTTGTCCA GCTCTGCCA CCTCCGCTAC GCGAGAGATT
 5341 AACCACCCAC GATGGCCGCC AAAGTGCATG TTGATATTGA GGCTGACAGC CCATTTCATCA
 5401 AGTCTTTGCA GAAGGCATT CCGTCGTTG AGGTGGAGTC ATTGCAGGTC ACACCAATG
 5461 ACCATGCAAA TGCCAGAGCA TTTTCGCACC TGGCTACCAA ATTGATCGAG CAGGAGACTG
 5521 ACAAAGACAC ACTCATCTTG GATATCGGCA GTGCGCTTTC CAGGAGAATG ATGTCTACGC
 5581 ACAAATACCA CTGCGTATGC CCTATGCGCA GCGCAGAAGA CCCCAGAAAG CTCGATAGCT
 5641 ACGCAAAGAA ACTGGCAGCG GCCTCCGGGA AGGTGCTGGA TAGAGAGATC GCAGGAAAAA
 5701 TCACCGACCT GCAGACCGTC ATGGCTACGC CAGACGCTGA ATCTCCTACC TTTTGCCTGC
 5761 ATACAGACGT CAGGTGCTGT ACGGCAGCCG AAGTGGCCGT ATACCAGGAC GTGTATGCTG
 5821 TACATGCACC AACATCGCTG TACCATCAGG CGATGAAAGG TGTGAGAACG GCGTATTGGA
 5881 TTGGGTTTGA CACCACCCG TTTATGTTTG ACGCGCTAGC AGGCGCGTAT CCAACCTACG-

FIGURE 29C

5941 CCACAACTG GCGGACGAG CAGGTGTTAC AGGCCAGGAA CATAGGACTG TGTGCAACAT
6001 CCTTGACTGA GGAAGACTC GGCAAACTGT CCATTCTCCG CAAGAAGCAA TTGAAAACCTT
6061 GCGACACAGT CATGTCTCG GTAGGATCTA CATTTGACAC TGAGAGCAGA AAGCTACTGA
6121 GGAGCTGGCA CTTACCTCC GTATTCCACC TGAAAGGTAA ACAATCCTTT ACCTGTAGGT
6181 GCGATACCAT CGTATCATGT GAAGGGTACG TAGTTAAGAA AATCACTATG TGCCCCSGCC
6241 TGTACGGTAA AACCGTAGGG TACGCCGTGA CGTATCACGC GGAGGGATTC CTAGTGTGCA
6301 AGACCACAGA CACTGTCAAA GGAGAAAGAG TCTCATTCCC TGTATGCACC TACGTCCTCT
6361 CAACCATCTG TGATCAAATG ACTGGCATACT TAGCGACCGA CGTCACACCG GAGGACGAC
6421 AGAAGTTGTT AGTGGGATTG AATCAGAGGA TAGTTGTGAA CGGAAGAACA CAGCGAAACA
6481 CTAACACGAT GAAGAACTAT CTGCTTCCGA TTGTGGCCGT CGCATTTAGC AAGTGGCGGA
6541 GGAATACAA GGCAGACCTT GATGATGAAA AACCTCTGGG TGTCCGAGAG AGGTCACTTA
6601 CTTGCTGCTG CTTGTGGGCA TTAAAAACGA GGAAGATGCA CACCATGTAC AAGAAACAG
6661 ACACCCAGAC AATAGTGAAG GTGCCCTTCAG AGTTTAACTC GTTCGTCATC CCGAGCCTAT
6721 GGTCTACAGG CCTCGCAATC CCAGTCAGAT CACGCATTAA GATGCTTTTG GCCAAGAAGA
6781 CCAAGCGAGA GTTAATACCT GTTCTCGACG CGTCGTCAGC CAGGGATGCT GAACAAGAGG
6841 AGAAGGAGAG GTTGGAGGCC GAGCTGACTA GAGAAGCCTT ACCACCCCTC GTCCCCATCG
6901 CGCCGGCGGA GACGGGAGTC GTGACGTCG ACGTTGAAGA ACTAGAGTAT CACGCAAGTG
6961 CAGGGGTCGT GGAACACCTC CGCAGCGCGT TGAAAGTCAC CGCACAGCCG AACGACSTAC
7021 TACTAGGAAA TTACGTAGTT CTGTCCCGC AGACCGTGCT CAAGAGCTCC AAGTTGSCCC
7081 CCGTGCAACC TCTAGCAGAG CAGGTGAAAA TAATAACACA TAACGGGAGG GCCGGCGGTT
7141 ACCAGGTCGA CGGATATGAC GGCAGGGTCC TACTACCATG TGGATCGGCC ATTCGGSTCC
7201 CTGAGTTTCA GGCCTTGAGC GAGAGCGCCA CTATGGTGTA CAACGAAGAG GAGTTCGTCA
7261 ACAGGAAACT ATACCATATT GCCGTTACG GACCCCTGCT GAACACCGAC GAGGAGAACT
7321 ACAGAAAGT CAGAGCTGAA AGAACTGACG CCGAGTACGT GTTCGACGTA GATAAAATAT
7381 GCTGCGTCAA GAGAGAGGAA GCGTCGGGTT TGGTGTGGT GGGAGAGCTA ACCAACCCCC
7441 CGTTCCATGA ATTCCGCTAC GAAGGGCTGA AGATCAGGCC GTCGGCACCA TATAAGACTA
7501 CAGTAGTAGG AGTCTTTGGG GTTCCGGGAT CAGGCAAGTC TGCTATTATT AAGAGCCTCG
7561 TGACCAAAACA CGATCTGGTC ACCAGCGGCA AGAAGGAGAA CTGCCAGGAA ATAGTTAAGC
7621 ACGTGAAGAA GCACCGCGGG AAGGGGACAA GTAGGGAAAA CAGTGACTCC ATCTGCTTAA
7681 ACGGGTGTG TCGTGCCGTG GACATCCTAT ATGTGACGA GGCTTTGCT TGCCATTCCG
7741 GTACTCTGCT GGCCTTAATT GCTCTGTGA AACCTCGGAG CAAAGTGGTG TTATGCGGAG
7801 ACCCAAAGCA ATGCGGATTC TTCAATATGA TGCAGCTTAA GGTGAACCTC AACACAACA
7861 TCTGCACTGA AGTATGTCAT AAAAGTATAT CCAGACGTTG CACGCGTCCA GTCACGSCCA
7921 TCGTGTCTAC GTTGCACTAC GGAGGCAAGA TGCGCACGAC CAACCCGTGC AACAAACCA
7981 TAATCATAGA CACCACAGGA CAGACCAAGC CCAAGCCAGG AGACATCGTG TTAACATGCT
8041 TCCGAGGCTG GGCAAAGCAG CTGCAGTTGG ACTACCGTGG ACACGAAGTC ATGACAGCAG
8101 CAGCATCTCA GGCCTCACC CGCAAAGGGG TATACGCCGT AAGGCAGAAG GTGAATGAAA
8161 ATCCCTTGTA TGCCCTTGGC TCGGAGCAGC TGAATGTACT GCTGACGCGC ACTGAGSATA
8221 GGCTGGTGTG GAAAACGCTG GCCGGCGATC CTGGATTAA GGTCTATCA AACATTTCCAC
8281 AGGGTAACCT TACGGCCACA TTGGAAGAAT GGCAAGAAGA ACACGACAAA ATAATGAAGG
8341 TGATTGAAGG ACCGGCTGCG CTTGTGGACG CGTTCAGAA CAAAGCGAAC GTGTGTTGGG
8401 CGAAAAGCCT GGTGCTGTG CTGGACACTG CCGGAATCAG ATTGACAGCA GAGGAGTGGA
8461 GCACCATAAT TACAGCATTT AAGGAGGACA GAGCTTACTC TCCAGTGGTG GCCTTGAATG
8521 AAATTTGCAC CAAGTACTAT GGAGTTGACC TGGACAGTGG CCTGTTTTCT GCCCGAAGG
8581 TGTCCCTGTA TTACGAGAAC AACCCTGGG ATAACAGACC TGGTGGAAGG ATGTATGGAT
8641 TCAATGCCGC AACAGCTGCC AGGCTGGAAG CTAGACATAC CTTCTGGAAG GGGCAGTGGC
8701 ATACGGGCAA GCAGGCAGTT ATCGCAGAAA GAAAAATCCA ACCGCTTTCT GTGCTGACCA
8761 ATGTAATTCC TATCAACCGC AGGCTGCCGC ACGCCCTGGT GGCTGAGTAC AAGACGTTA
8821 AAGGCAGTAG GGTTGAGTGG CTGGTCAATA AAGTAAGAGG GTACCAAGTC CTGCTG3TGA
8881 GTGAGTACAA CCTGGCTTTG CTTGACGCA GGGTCACTTG GTTGTACCG CTGAATGTCA
8941 CAGGCGCCGA TAGGTGCTAC GACCTAAGTT TAGGACTGCC GGCTGACGCC GGCAGGTTG
9001 ACTTGGTCTT TGTGAACATT CACACGGAAT TCAGAATCCA CCACTACCAG CAGTGTCTCG
9061 ACCACGCCAT GAAGCTGACG ATGCTTGGGG GAGATGCGCT ACGACTGCTA AAACCCGGCG
9121 GCATCTTGAT GAGAGCTTAC GGATACGCCG ATAAAATCAG CGAAGCCGTT GTTTCTCTCT
9181 TAAGCAGAAA GTTCTCTGCT GCAAGAGTGT TGCGCCCGGA TTGTGTACAC AGCAATACAG
9241 AAGTGTCTCT GCTGTTCTCC AACTTTGACA ACGGAAAGAG ACCCTCTACG CTACACAGGA
9301 TGAATACAA GCTGAGTGCC GTGTATGCCG GAGAAAGCCAT GCACACGGCC GGGTGTGCAC
9361 CATCTACAG AGTTAAGAGA GCAGACATAG CCACGTGCAC AGAAGCGGCT GTGGTTACCG-

Figure 29d

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9421 CAGCTAACGC CCGTGGAACT GTAGGGGATG GCGTATGCAG GCGCGTGGCG AAGAAATGGC
 9481 CGTCAGCCTT TAAGGGAGCA GCAACACCAG TGGGCACAAT TAAACAGTC ATGTGCGGCT
 9541 CGTACCCCGT CATCCACGCT GTAGCGCCTA ATTTCTCTGC CACGACTGAA GCGGAAGGGG
 9601 ACCGCGAATT GCGCGCTGTC TACCGGGCAG TGGCGGCCGA AGTAAACAGA CTGTCACTGA
 9661 GCAGCGTAGC CATCCCGCTG CTGTCCACAG GAGTGTTCAG CGGCGGAAGA GATAGGCTGC
 9721 AGCAATCCCT CAACCATCTA TTCACAGCAA TGGACGCCAC GGACGCTGAC GTGACCATCT
 9781 ACTGCAGAGA CAAAAGTTGG GAGAAGAAAA TCCAGGAAGC CATTGACATG AGGACGGCTG
 9841 TGGAGTTGCT CAATGATGAC GTGGAGCTGA CCACAGACTT GGTGAGAGTG CACCCGAGCA
 9901 GCAGCCTGGT GGGTCGTAAG GGCTACAGTA CCACTGACGG GTCTGCTGAC TCGTACTTTG
 9961 AAGGTACGAA ATTCAACCAG GCTGCTATTG ATATGGCAGA GATACTGACG TTGTGGCCCA
 10021 GACTGCAAGA GGCAACGAA CAGATATGCC TATACGCGCT GGGCGAAACA ATGGACAACA
 10081 TCAGATCCAA ATGTCCGGTG AACGATTCCG ATTCATCAAC ACCTCCAGG ACAGTGCCCT
 10141 GCCTGTGCCG CTACGCAATG ACAGCAGAAC GGATCGCCCG CCTTAGGTCA CACCAAGTTA
 10201 AAAGCATGGT GGTTCGCTCA TCTTTTCCCC TCCCGAAATA CCATGTAGAT GGGGTGACGA
 10261 AGGTAAAGTG CGAGAAGGTT CTCTGTTCG ACCCGACGGT ACCTTCAGTG GTTAGTCCGC
 10321 GGAAGTATGC CGCATCTACG ACGGACCACT CAGATCGGTC GTTACGAGGG TTTGACTTTG
 10381 ACTGGACCAC CGACTCGTCT TCCACTGCCA GCGATACCAT GTCTGCTACCC AGTTTGCAGT
 10441 CGTGTGACAT CGACTCGATC TACGAGCCAA TGGCTCCCAT AGTAGTGACG GCTGACGTAC
 10501 ACCCTGAACC CGCAGGCATC GCGGACCTGG CGGCAGATGT GCACCCTGAA CCCGCAGACC
 10561 ATGTGGACCT GGAGAACCCG ATTCCTCCAC CGCGCCCGAA GAGAGCTGCA TACCTTGCTT
 10621 CCCGCGCGGC GGAGCGACCG GTGCCGGCGC CGAGAAAGCC GACGCCTGCC CCAAGGACTG
 10681 CGTTTAGGAA CAAGCTGCCT TTGACGTTTC GCGACTTTGA CGAGCAGAG GTCCATGCGT
 10741 TGGCCTCCGG GATTACTTTC GGAGACTTCG ACGACGTCTT CCGACTAGGC CGCGCGGGTG
 10801 CATATATTTT CTCTCCGGAC ACTGGCAGCG GACATTTACA ACAAATATCC GTTAGGCAGC
 10861 ACAATCTCCA GTGCGCACAA CTGGATGCGG TCCAGGAGGA GAAATGTAC CCGCCAAAGT
 10921 TGGATACTGA GAGGAGAAAG CTGTTGCTGC TGAATATGCA GATGCACCCA TCGGAGGCTA
 10981 ATAAGAGTCG ATACCAGTCT CGCAAAGTGG AGAATCATGA AGCCACGGTG GTGGACAGGC
 11041 TCACATCGGG GCCCAGATTG TACACGGGAG CGGACGTAGG CCGCATACCA ACATACGCGG
 11101 TTCGGTACCC CCGCCCCGTG TACTCCCTTA CCGTGATCGA AAGATTCTCA AGCCCCGATG
 11161 TAGCAATCGC AGCGTGCAAC GAATACCTAT CCAGAAATTA CCCAACAGTG GCGTCGTACC
 11221 AGATAACAGA TGAATACGAC GCATACCTTG ACATGGTTGA CCGGTCCGAT AGTTGCTTGG
 11281 ACAGAGCGAC ATTCTGCCCC GCGAAGCTCC GGTGCTACCC GAAACATCAT GCGTACCACC
 11341 AGCCGACTGT ACGCAGTGCC GTCCCGTCAC CTTTCAGAA CACACTACAG AACGTGCTAG
 11401 CGGCTGCCAC CAAGAGAAAC TGCAACGTCA CGCAAATGCG AGAATACCC ACCATGGACT
 11461 CGGCAGTGTT CAACGTGGAG TGCTTCAAGC GCTATGCTTG CTCCGGAGAA TATTGGGAAG
 11521 AATATGCTAA ACAACCTATC CGGATAACCA CTGAGAACAT CACTACCTAT GTGACCAAT
 11581 TGAAAGGCCC GAAAGCTGCT GCCTTGTTTC CTAAGACCCA CAACTTGGTT CCGCTGCAGG
 11641 AGTTTCCCAT GGACAGATTG ACGTTCGACA TGAACGAGA TGTCAAAGTC ACTCCAGGGA
 11701 CGAAACACAC AGAGGAAAGA CCCAAAGTCC AGGTAATTCA AGCAGCGGAG CCATTGGCGA
 11761 CCGCTTACCT GTGCGGCATC CACAGGGAAT TAGTAAGGAG ACTAAATGCT GTGTTACGCC
 11821 CTAACGTGCA CACATTGTTT GATATGTCGG CCGAAGACTT TGACGCGATC ATCGCCTCTC
 11881 ACTTCCACCC AGGAGACCCG GTTCTAGAGA CGGACATTGC ATCATTGAC AAAAGCCAGG
 11941 ACGACTCCTT GGCTCTTACA GGTTTAATGA TCCTCGAAGA TCTAGGGGTG GATCAGTACC
 12001 TGCTGGACTT GATCGAGGCA GCCTTTGGGG AAATATCCAG CTGTACCTTA CCAACTGGCA
 12061 CGCGCTTCAA GTTCGGAGCT ATGATGAAAT CGGGCATGTT TCTGACTTTG TTTATTAA
 12121 CTGTTTGA CAATCACCATA GCAAGCAGGG TACTGGAGCA GAGACTCACT GACTCCGCTC
 12181 GTGCGGCCCT CATCGCGGAC GACAACATCG TTCACGGAGT GATCTCCGAC AAGCTGATGG
 12241 CGGAGAGGTG CGCGTCGTGG GTCAACATGG AGGTGAAGAT CATTGACGCT GTCATGGGCG
 12301 AAAAACCCTT ATATTTTGTG GGGGGATTCA TAGTTTTTGA CAGCGTCACA CAGACCGCTT
 12361 GCCGTGTTTC AGACCCACTT AAGCGCTGT TCAAGTTGGG TAAGCCGCTA ACAGCTGAAG
 12421 ACAAGCAGGA CGAAGACAGG CGACGAGCAC TGAGTGACGA GGT

FIGURE 29E

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**Figure 30A: pDEST10 Polyhedron Promoter with N-His₆,
Baculovirus Transfer Plasmid**

→ mRat from polyhedrin promoter

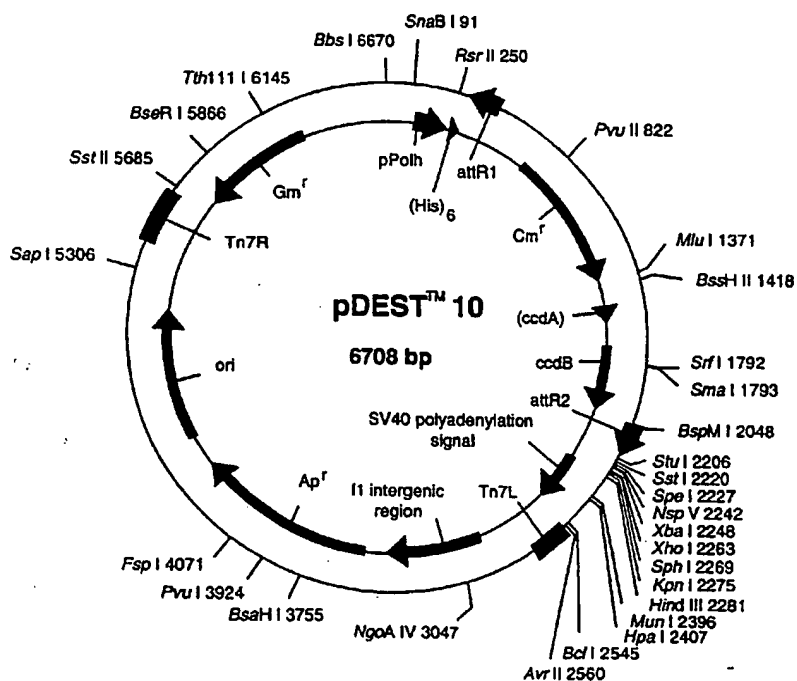
154 aaa taa gta ttt tac tgc ttt cgt aac agt ttt gta ata aaa aaa cct ata
ttt att cat aaa atg aca aaa gca ttg tca aaa cat tat ttt ttt gga tat

205 aat att ccg gat tat tca tac cgt ccc acc atc ggg cgc gga tct cgg tcc
tta taa ggc cta ata agt atg gca ggg tgg tag ccc gcg cct aga gcc agg

256 Met Ser Tyr Tyr His His His His His His Asp Tyr Asp Ile Pro
gaa acc atg tgc tac tac cat cac cat cac cat cac gat tgc gat atc cca
ctt tgg tac agc atg atg gta gtg gta gtg gta gtg cta atg cta tag ggt

TEV protease

307 Thr Thr Glu Asn Leu Tyr Phe Gln Gly Ile Thr Ser Leu Tyr Lys Lys
acg acc gaa aac ctg tat ttt cag ggc atc tgc tca aac atg ttt ttc oga
attR1 Int



pDEST10 6708 bp

Location (Base Nos.)	Gene Encoded
23..152	Ppolh
461..337	attR1
711..1370	CmR
1490..1574	inactivated ccdA
1712..2017	ccdB
2058..2182	attR2
3394..4369	ampR
4510..5164	ori
5658..62	genR

1	CCCCGGATGA	AGTGGTTCGC	ATCCTCGGTT	TTCTGGAAGG	CGAGCATCGT	TGTTTCGCCC
61	AGGACTCTAG	CTATAGTTCT	AGTGGTTGGC	TACGTATACT	CCGGAATATT	AATAGATCAT
121	GGAGATAATT	AAAATGATAA	CCATCTCGCA	AATAAATAAG	TATTTTACTG	TTTTCGTAAC
181	AGTTTTGTAA	TAAAAAACC	TATAAATATT	CCGGATTATT	CATACCGTCC	CACCATCGGG
241	CGCGGATCTC	GGTCCGAAAC	CATGTCGTAC	TACCATCACC	ATCACCATCA	CGATTACGAT
301	ATCCCAACGA	CCGAAAACCT	GTATTTTCAG	GGCATCACAA	GTTTGTACAA	AAAAGCTGAA
361	CGAGAAACGT	AAAATGATAT	AAATATCAAT	ATATTAAATT	AGATTTTGCA	TAAAAACAG
421	ACTACATAAT	ACTGTAAAAC	ACAACATATC	CAGTCACTAT	GGCGGCCGCT	AAGTTGGCAG
481	CATCACCCGA	CGCACTTTGC	GCCGAATAAA	TACCTGTGAC	GGAAGATCAC	TTCCGAGAAT
541	AAATAAATCC	TGGTGTCCCT	GTTGATACCG	GGAAGCCCTG	GGCCAACTTT	TGGCGAAAA
601	GAGACGTTGA	TCGGCACGTA	AGAGGTTCCA	ACTTTCACCA	TAATGAAATA	AGATCACTAC
661	CGGGCGTATT	TTTTGAGTTA	TCGAGATTTT	CAGGAGCTAA	GGAAGCTAAA	ATGGAGAAAA
721	AAATCACTGG	ATATACCACC	GTTGATATAT	CCCAATGGCA	TCGTAAGAGAA	CATTTTGAGG
781	CATTTCACTG	AGTTGCTCAA	TGTACCTATA	ACCAGACCGT	TCAGCTGGAT	ATTACGGCCT
841	TTTTAAAGAC	CGTAAAGAAA	AATAAGCACA	AGTTTATATC	GGCCTTTTAT	CACATTCTTG
901	CCCGCCTGAT	GAATGCTCAT	CCGGAATTCC	GTATGGCAAT	GAAAGACGGT	GAGCTGGTGA
961	TATGGGATAG	TGTTTACCCT	TGTTACACCG	TTTTCCATGA	GCAAACCTGAA	ACGTTTTTCAT
1021	CGCTCTGGAG	TGAATACCAC	GACGATTTC	GGCAGTTTCT	ACACATATAT	TCGCAGAGATG
1081	TGGCGTGTTA	CGGTGAAAAC	CTGGCCTATT	TCCCTAAAGG	GTTTATTGAG	AATATGTTTT
1141	TCGTCTCAGC	CAATCCCTGG	GTGAGTTTCA	CCAGTTTGA	TTTAAACGTG	GCCAATATGG
1201	ACAACCTTCT	CGCCCCCGTT	TTCACCATGG	GCAAATATTA	TACGCAAGGC	GACAAGGTGC
1261	TGATGCCGCT	GGCGATTTCG	GTTTCATCATG	CCGTCTGTGA	TGGCTTCCAT	GTCCGCAGAA
1321	TGCTTAATGA	ATTACAACAG	TACTGCGATG	AGTGGCAGGG	CGGGGCGTAA	ACGCGTGGAT
1381	CCGGCTTACT	AAAAGCCAGA	TAACAGTATG	CGTATTGCGG	CGCTGATTTT	TGCGGTATAA
1441	GAATATATAC	TGATATGTAT	ACCCGAAGTA	TGTCAAAAAG	AGGTGTGCTA	TGAAGCAGCG
1501	TATTACAGTG	ACAGTTGACA	GCGACAGCTA	TCAGTTGCTC	AAGGCATATA	TGATGTCAAT
1561	ATCTCCGGTC	TGGTAAGCAC	AACCATGCAG	AATGAAGCCC	GTCTGTGCGC	TGCCGAACGC
1621	TGGAAAGCGG	AAAATCAGGA	AGGGATGGCT	GAGGTCGCCC	GGTTTATTGA	AATGAACGGC
1681	TCTTTTGCTG	ACGAGAACAG	GGACTGGTGA	AATGCAGTTT	AAGGTTTACA	CCTATAAAAG
1741	AGAGAGCCGT	TATCGTCTGT	TTGTGGATGT	ACAGAGTGAT	ATTATTGACA	CGCCCCGGCG
1801	ACGGATGGTG	ATCCCCCTGG	CCAGTGCACG	TCTGCTGTCA	GATAAAGTCT	CCCGTGAAC
1861	TTACCCGGTG	GTGCATATCG	GGGATGAAAG	CTGGCGCATG	ATGACCACCG	ATATGGCCAG
1921	TGTGCCGGTC	TCCGTTATCG	GGGAAGAAGT	GGCTGATCTC	AGCCACCGCG	AAAATGACAT
1981	CAAAAACGCC	ATTAACCTGA	TGTTCTGGGG	AATATAAATG	TCAGGCTCCC	TTATACACAG
2041	CCAGTCTGCA	GGTCGACCAT	AGTGACTGGA	TATGTTGTGT	TTTACAGTAT	TATGTAGTCT
2101	GTTTTTTATG	CAAAATCTAA	TTTAATATAT	TGATATTTAT	ATCATTTTAC	GTTTCTCGTT
2161	CAGCTTTCTT	GTACAAAGTG	GTGATGCCAT	GGATCCGGAA	TTCAAAGGCC	TACGTCGACG
2221	AGCTCAACTA	GTGCGGCCGC	TTTCGAATCT	AGAGCCTGCA	GTCTCGAGGC	ATGCGGTACC
2281	AAGCTTGTCT	AGAAGTACTA	GAGGATCATA	ATCAGCCATA	CCACATTGTG	AGAGGTTTTA
2341	CTTGCTTTAA	AAAACCTCCC	ACACCTCCCC	CTGAACCTGA	AACATAAAAT	GAATGCAATT
2401	GTTGTGTGTA	ACTTGTTTAT	TGCAGCTTAT	AATGGTTACA	AATAAAGCAA	TAGCATCACA
2461	AATTTACAAA	ATAAAGCATT	TTTTTCACTG	CATTCTAGTT	GTGGTTTGTC	CAAACCTCAT
2521	AATGTATCTT	ATCATGTCTG	GATCTGATCA	CTGCTTGAGC	CTAGGAGATC	CGAACCAGAT
2581	AAGTGAAATC	TAGTTCCAAA	CTATTTTGTC	ATTTTAAATT	TTCTGATTAG	CTTACGACGC

FIGURE 30B

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2641 TACACCCAGT TCCCATCTAT TTTGTCACTC TTCCTAAAT AATCCTTAAA AACTCCATTT
2701 CCACCCCTCC CAGTTCCCAA CTATTTTGTG CGCCACAGC GGGGCATTTT TCTTCCTGTT
2761 ATGTTTTTAA TCAAACATCC TGCCAACTCC ATGTGACAAA CCGTCATCTT CGGCTACTTT
2821 TTCTCTGTCA CAGAATGAAA ATTTTCTGT CATCTCTCG TTATTAATGT TTGTAATTGA
2881 CTGAATATCA ACGCTTATTT GCAGCCTGAA TGGCGAATGG GACGCGCCCT GTAGCGGCGC
2941 ATTAAGCGCG GCGGCTGTGG TGGTACGCG CAGCGTGACC GCTACACTTG CCAGCGCCCT
3001 AGCGCCCGCT CCTTTCGCTT TCTTCCTTC CTTTCTCGCC ACGTTCGCGG GCTTTCGCCG
3061 TCAAGCTCTA AATCGGGGGC TCCTTTAGG GTTCGGAATT AGTGCTTTAC GGCACCTCGA
3121 CCCCCAAAAA CTTGATTAGG GTGATGGTTC ACGTAGTGGG CCATCGCCCT GATAGACGGT
3181 TTTTCGCCCT TTGACGTTGG AGTCCACGTT CTTTAATAGT GGACTCTTGT TCCAAACTGG
3241 AACAACACTC AACCTATCT CGGTCTATTC TTTTGATTTA TAAGGGATTT TGCCGATTTT
3301 GGCCTATTGG TTAATAAATG AGCTGATTTA ACAAATAATT AACGCGAATT TTAACAAAAT
3361 ATTAACGTTT ACAATTTTCA GTGGCACTTT TCGGGGAAAT GTGCGCGGAA CCCCTATTTG
3421 TTTATTTTTC TAAATACATT CAAATATGTA TCCGCTCATG AGACAATAAC CCTGATAAAT
3481 GCTTCAATAA TATTGAAAAA GGAAGAGTAT GAGTATTCAA CATTTCCGTG TCGCCCTTAT
3541 TCCCTTTTTT GCGGCATTTT GCCTTCCTGT TTTTGCTCAC CCAGAAACGC TGGTGAAAGT
3601 AAAAGATGCT GAAGATCAGT TGGGTGCAG AGTGGGTAC ATCGAACTGG ATCTCAACAG
3661 CGGTAAGATC CTTGAGAGTT TTCGCCCGA AGAACGTTTT CCAATGATGA GCACCTTTAA
3721 AGTTCTGCTA TGTGGCGCGG TATTATCCCG TATTGACGCC GGGCAAGAGC AACTCGGTG
3781 CCGCATACAC TATTCTCAGA ATGACTTGGT TGAGTACTCA CCAGTCACAG AAAAGCATCT
3841 TACGGATGGC ATGACAGTAA GAGAATTATG CAGTGCTGCC ATAACCATGA GTGATAACAC
3901 TCGGGCCAAC TTACTTCTGA CAACGATCGG AGGACCGAAG GAGCTAACCG CTTTCTTTCG
3961 CAACATGGGG GATCATGTAA CTCGCCCTGA TCGTTGGGAA CCGGAGCTGA ATGAAGCCAT
4021 ACCAAACGAC GAGCGTGACA CCACGATGCC TGTAGCAATG GCAACAACGT TGCCGAAACT
4081 ATTAACGCGC GAACTACTTA CTCTAGCTTC CCGGCAACAA TTAATAGACT GGATGGAGGC
4141 GGATAAAGTT GCAGGACCAC TTCTGGGCTC GGCCCTTCCG GCTGGCTGGT TTATTGTGTA
4201 TAAATCTGGA GCCGGTGAGC GTGGGTCTCG CGGTATCATT GCAGCACTGG GGCCAGATGG
4261 TAAGCCCTCC CGTATCGTAG TTATCTACAC GACGGGGAGT CAGGCAACTA TCGATGAACG
4321 AAATAGACAG ATCGCTGAGA TAGGTGCCTC ACTGATTAAG CATTGGTAAC TGTCAGACCA
4381 AGTTTACTCA TATATACTTT AGATTGATT AAAACTTCAT TTTTAATTAA AAAGGATCTA
4441 GGTGAAGATC CTTTTTGATA ATCTCATGAC CAAAATCCCT TAACGTGAGT TTTCTGTCCA
4501 CTGAGCGTCA GACCCCGTAG AAAAGATCAA AGGATCTTCT TGAGATCCTT TTTTCTGCG
4561 CGTAATCTGC TGCTTGCAAA CAAAAAACC ACCGCTACCA GCGGTGGTTT GTTTGCCGGA
4621 TCAAGAGCTA CCAACTCTTT TTCCGAAGGT AACTGGCTTC AGCAGAGCGC AGATACCAAA
4681 TACTGTCTTT CTAGTGTAGC CGTAGTTAGG CCACCACTC AAGAACTCTG TAGCACCGCC
4741 TACATACCTC GCTCTGTAA TCCTGTTACC AGTGGCTGCT GCCAGTGGCG ATAAGTCGTG
4801 TCTTACCGGG TTGGACTCAA GACGATAGTT ACCGGATAAG GCGCAGCGGT CGGGCTGAAC
4861 GGGGGGTTTC TGACACAGC CCAGCTTGGA GCGAACGACC TACACCGAAC TGAGATACCT
4921 ACAGCGTGAG CATTGAGAAA GCGCCACGCT TCCCGAAGGG AGAAGGCGG ACAGGTATCC
4981 GGTAAGCGGC AGGGTCGGA CAGGAGAGCG CACGAGGGAG CTTCCAGGGG GAAACGCTG
5041 GTATCTTTAT AGTCCTGTG GGTTCGCCA CCTCTGACTT GAGCGTCGAT TTTGTGATG
5101 CTCGTACGGG GGGCGGAGCC TATGGAAAAA CGCCAGCAAC GCGGCCTTTT TACGGTTCTC
5161 GGCTTTTTC TGCCCTTTTG CTCACATGTT CTTTCTGCG TTATCCCCTG ATTCTGTGGA
5221 TAACCGTATT ACCGCTTTG AGTGAGCTGA TACCGCTCGC GCGAGCCGAA CGACCGAGCG
5281 CAGCGAGTCA GTGAGCGAGG AAGCGGAAGA GCGCCTGATG CGGTATTTTC TCCTTACGCA
5341 TCTGTGCGGT ATTTACACC GCAGACCAGC CGCGTAACCT GGCAAAATCG GTTACGGTTG
5401 AGTAATAAAT GGATGCCCTG CGTAAGCGGG TGTGGCGGA CAATAAAGTC TTAAGTGA
5461 CAAAATAGAT CTAACTATG ACAATAAAGT CTAAACTAG ACAGAATAGT TGTAAGTGA
5521 AATCAGTCCA GTTATGCTGT GAAAAAGCAT ACTGGACTTT TGTTATGGCT AAAGCAAACT
5581 CTTCAITTTT TGAAGTGCAA ATTGCCCGTC GTATTAAAGA GGGGCGTGGC CAAGGGCATG
5641 GTAAAGACTA TATTCGCGG GTTGTGACAA TTTACCGAAC AACTCCGCGG CCGGGAAGCC
5701 GATCTCGGCT TGAACGAATT GTTAGGTGGC GGTACTTGGG TCGATATCAA AGTGATCAC
5761 TTCTTCCCGT ATGCCCAACT TTGTATAGAG AGCCACTGCG GGATCGTCAC CGTAATCTGC
5821 TTGCACGTAG ATCACATAAG CACCAAGCGC GTTGGCTCA TGCTTGAGGA GATTGATGAG
5881 CGCGGTGGCA ATGCCCTGCC TCCGTTGCTC GCCGGAGACT GCGAGATCAT AGATATAGAT
5941 CTCACTACGC GGCTGCTCAA ACCTGGGCG AACGTAAGCC GCGAGAGCGC CAACAACCGC
6001 TTCTTGGTCG AAGGCAGCAA GCGCGATGAA TGTCTTACTA CCGAGCAAGT TCCCGAGGTA
6061 ATCGGAGTCC GGCTGATGTT GGGAGTAGGT GGCTACGCT CCGAACTCAC GACCGAAAAG-

FIGURE 30C

6121 ATCAAGAGCA GCCCGCATGG ATTTGACTTG GTCAGGGCCG AGCCTACATG TCGGAATGAT
6181 GCCCATACIT GAGCCACCTA ACTTTGTITT AGGGCGACTG CCCTGCTGCG TAACATCGTT
6241 GCTGCTGCGT AACATCGTTG CTGCTCCATA ACATCAAACA TCGACCCACG GCGTAACGCG
6301 CTTGCTGCTT GGATGCCCGA GGCATAGACT GTACAAAAAA ACAGTCATAA CAAGCCATGA
6361 AAACCGCCAC TCGCGCGTTA CCACCGCTGC GTTCGGTCAA GGTCTGGAC CAGTTGCGTG
6421 AGCGCATACG CTA CTGTCAT TACAGTTTAC GAACCGAACA GGCTTATGTC AACTGGGTTT
6481 GTGCCCTTCAT CCGTTTCCAC GGTGTGCGTC ACCCGGCAAC CTTGGGCAGC AGCGAAGTCG
6541 AGGCATTTCT GTCTGGCTG GCGAACGAGC GCAAGGTTTC GGTCTCCACG CATCGTCAGG
6601 CATTGGCGGC CTTGCTGTTT TTCTACGGCA AGGTGCTGTG CACGGATCTG CCCTGGCTTC
6661 AGGAGATCGG AAGACCTCGG CCGTCGCGC GCTTGCCGGT GGTGCTGA

FIGURE 30D

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Figure 31A:

PDEST 11

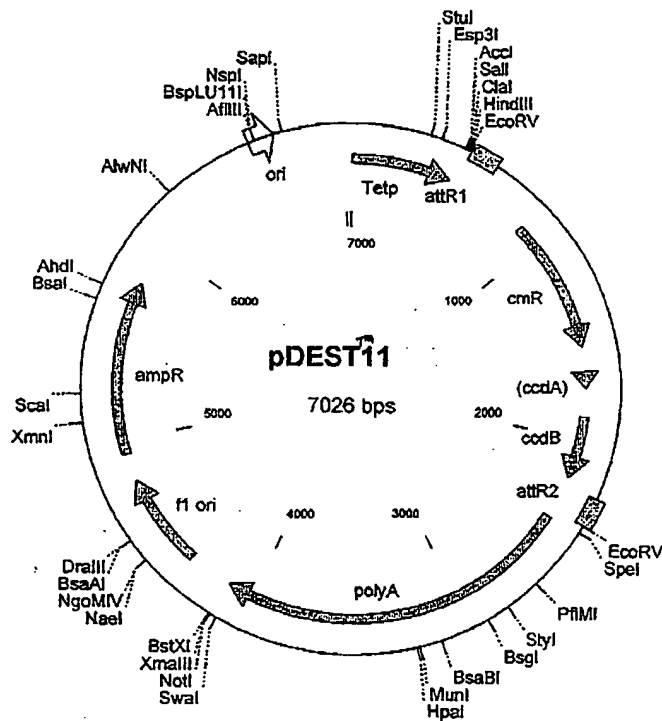
Tet-regulated eukaryotic expression

358 tag tga acc gfc aga tgg cct gga gac gcc atc cac gct gtt tgg acc tcc
atc act tgg cag tct agc gga cct ctg cgg tag gtg cga caa aac tgg agg

409 ata gaa gac acc ggg acc gat cca gcc tcc gcg gcc ccg aat tgg agc tgg
tat ctt ctg tgg ccc tgg cta ggt cgg agg cgc cgg ggc tta agc tgc agc

460 gta ccc ggg gat cct cta gag tgc agg ^{Sal} tcc acc gta ^{Cla} tgg ^{Hind3} atg acc tgg ^{EcoRV} aga
cat ggg ccc cta gga gat ctc agc tcc acc tgc cat agt tat tgg acc tat

511 tca aca agt tgg xap xaa gct gaa cga gaa acg taa tat gaa aga gat
agt tgt tca aac atg ttt ttc cga ctt gct ctt tgc att tta cta tat tta



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pDEST11 7026 bp

Location (Base Nos.)	Gene Encoded
4...479	Tetp ((Tet operator) 7 and min hCMV promoter)
638...514	attR1
888...1547	CmR
1667...1751	inactivated ccdA
1889...2194	ccdB
2235...2359	attR2
2402...4132	polyA
4347...4803	f1 ori
4940...5797	ampR

1	CGAGTTTACC	ACTCCCTATC	AGTGATAGAG	AAAAGTGAAA	GTCGAGTTTA	CCACTCCCCTA
61	TCAGTGATAG	AGAAAAGTGA	AAGTCGAGTT	TACCACTCCC	TATCAGTGAT	AGAGAAAAGT
121	GAAAGTCGAG	TTTACCCTAC	CCTATCAGTG	ATAGAGAAAA	GTGAAAGTCG	AGTTTACCAC
181	TCCCTATCAG	TGATAGAGAA	AAGTGAAAGT	CGAGTTTACC	ACTCCCTATC	AGTGATAGAG
241	AAAAGTGAAA	GTCGAGTTTA	CCACTCCCCTA	TCAGTGATAG	AGAAAAGTGA	AAGTCGAGCT
301	CGGTACCCGG	GTCGAGTAGG	CGGTACCGGT	GGGAGGCCTA	TATAAGCAGA	GCTCGTTTAG
361	TGAACCGTCA	GATCGCCTGG	AGACGCCATC	CACGCTGTTT	TGACCTCCAT	AGAAGACACC
421	GGGACCGATC	CAGCCTCCGC	GGCCCCGAAT	TCGAGCTCGG	TACCCGGGGA	TCCTCTAGAG
481	TCGAGGTCGA	CGGTATCGAT	AAGCTTGATA	TCAACAAGTT	TGTACAAAAA	AGCTGAACGA
541	GAAACGTAAA	ATGATATAAA	TATCAATATA	TTAAATTAGA	TTTTCGATAA	AAAACAGACT
601	ACATAATACT	GTAAAAACACA	ACATATCCAG	TCACTATGGC	GGCCGCTAAG	TTGGCAGCAT
661	CACCCGACGC	ACTTTGGCGC	GAATAAATAC	CTGTGACGGA	AGATCACTTC	GCAGAATAAA
721	TAAATCCTGG	TGTCCCTGTT	GATACCGGGA	AGCCCTGGGC	CAACTTTTGG	CGAAAATGAG
781	ACGTTGATCG	GCACGTAAAG	GGTTCCAACT	TTCAACATAA	TGAAATAAGA	TCACTACCGG
841	GCGTATTTTT	TGAGTTATCG	AGATTTTCAG	GAGCTAAGGA	AGCTAAAATG	GAGAAAAAAA
901	TCACTGGATA	TACCACCGTT	GATATATCCC	AATGGCATCG	TAAAGAACAT	TTTGAGGCAT
961	TTCACTCAGT	TGCTCAATGT	ACCTATAACC	AGACCGTTCA	GCTGGATATT	ACGGCCCTTT
1021	TAAAGACCGT	AAAGAAAAAT	AAGCACAAGT	TTTATCCGGC	CTTTATTTCAC	ATTCTTGCCC
1081	GCCTGATGAA	TGCTCATCCG	GAATTCCGTA	TGGCAATGAA	AGACGGTGAG	CTGGTGATAT
1141	GGGATAGTGT	TCACCCCTGT	TACACCGTTT	TCCATGAGCA	AACTGAAACG	TTTTTCATCG
1201	TCTGGAGTGA	ATACCACGAC	GATTTCCGGC	AGTTTCTACA	CATATATTTC	CAAGATGTGG
1261	CGTGTACCG	TGAAAACCTG	GCCTATTTCC	CTAAAGGGTT	TATTGAGAAT	ATGTTTTTCG
1321	TCTCAGCCAA	TCCCTGGGTG	AGTTTACCCA	GTTTTGATTT	AAACGTGGCC	AATATGGACA
1381	ACTTCTTCGC	CCCCGTTTTT	ACCATGGGCA	AATATTATAC	GCAAGCGCAC	AAGGTGCTGA
1441	TGCCGCTGGC	GATTCAGGTT	CATCATGCCG	TCTGTGATGG	CTTCCATGTC	GGCAGAATGC
1501	TTAATGAATT	ACAACAGTAC	TGCGATGAGT	GGCAGGGCGG	GGCGTAAAGA	TCTGGATCCG
1561	GCTTACTAAA	AGCCAGATAA	CAGTATGCGT	ATTGCGCGCG	TGATTTTTCG	GGTATAAGAA
1621	TATATACTGA	TATGTATACC	CGAAGTATGT	CAAAAAGAGG	TGTGCTATGA	AGCAGCGTAT
1681	TACAGTGACA	GTTGACAGCG	ACAGCTATCA	GTTGCTCAAG	GCATATATGA	TGTCAATATC
1741	TCCGGTCTGG	TAAGCACAAAC	CATGCAGAA	GAAGCCCGTC	GTCTGCGTGC	CGAACGCTGG
1801	AAAGCGGAAA	ATCAGGAAGG	GATGGCTGAG	GTCGCGCGGT	TTATTGAAAT	GAACGGCTCT
1861	TTTGCTGACG	AGAACAGGGA	CTGGTGAAAT	GCAGTTTAAG	GTTTACACCT	ATAAAAAGAGA
1921	GAGCCGTTAT	CGTCTGTTTG	TGGATGTACA	GAGTGATATT	ATTGACACGC	CCGGGCGACG
1981	GATGGTGATC	CCCCTGGCCA	GTGCACGTCT	GCTGTGAGAT	AAAGTCTCCC	GTGAACCTTA
2041	CCCCTGGGTG	CATATCGGGG	ATGAAAGCTG	GCGCATGATG	ACCACCGATA	TGGCCAGTGT
2101	GCCCGTCTCC	GTTATCGGGG	AAGAAGTGGC	TGATCTCAGC	CACCGCGAAA	ATGACATCAA
2161	AAACGCCATT	AACCTGATGT	TCTGGGGAAT	ATAAATGTCA	GGCTCCCTTA	TACACAGCCA
2221	GTCTGCAGGT	CGACCATAGT	GACTGGATAT	GTTGTGTTTT	ACAGTATTAT	GTAGTCTGTT
2281	TTTTATGCAA	AATCTAATTT	AATATATTGA	TATTTATATC	ATTTTACGTT	TCTCGTTTAC
2341	CTTTCTTGTA	CAAAGTGGTT	GATATCGAAT	TCCTGCAGCC	CGGGGGATCC	ACTAGTTCTA
2401	GAGCACTGCG	ATGAGTGGCA	GGGCGGGGCG	TAATTTTTTT	AAGGCAGTTA	TTGGTGCCCT
2461	TAAACGCCCTG	GTGCTACGCC	TGAATAAGTG	ATAATAAGCG	GATGAATGGC	AGAAATTCGC
2521	CGGATCTTTG	TGAAGGAACC	TTACTTCTGT	GGTGTGACAT	AATTGGACAA	ACTACCTACA-

FIGURE 31B

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2581 GAGATTTAAA GCTCTAAGGT AAATATAAAA TTTTAAAGTG TATAATGTGT TAAACTACTG
2641 ATTCTAATTG TTTGTGTATT TTAGATTCCA ACCTATGGAA CTGATGAATG GGAGCAGTGG
2701 TGGAAATGCTT TTAATGAGGA AAACCTGTTT TGCTCAGAAG AAATGCCATC TAGTGATGAT
2761 GAGGCTACTG CTGACTCTCA ACATTCTACT CCTCCAAAAA AGAAGAGAAA GGTAGAAGAC
2821 CCCAAGGACT TTCCTTCAGA AITGCTAAGT TTTTGTAGTC ATGCTGTGTT TAGTAATAGA
2881 ACTCTTGCTT GCTTTGCTAT TTACACCACA AAGGAAAAAG CTGCACTGCT ATACAAGAAA
2941 ATTATGGAAG AATATTCTGT AACCTTTATA AGTAGGCATA ACAGTTATAA TCATAACATA
3001 CTGTTTTTTC TTACTCCACA CAGGCATAGA GTGTCGTCTA TTAATACTA TGCTCAAAAA
3061 TTGTGTACCT TTAGCTTTTT AATTTGTAAA GGGGTTAATA AGGAATATTT GAGTATAGT
3121 GCCTTGACTA GAGATCATAA TCAGCCATAC CACATTTGTA GAGGTTTTAC TTGCTTTAAA
3181 AAACCTCCCA CACCTCCCCC TGAACCTGAA ACATAAAATG AATGCAATTG TTGTTGTAAA
3241 CTTGTTTTATT GCAGCTTATA ATGGTTACAA ATAAAGCAAT AGCATCACAA ATTTACAAAA
3301 TAAAGCATTT TTTTCACTGC ATTCTAGTTG TGGTTTGTCC AAACATCATCA ATGTATCTTA
3361 TCATGTCTGG ATCCCCAGGA AGCTCCTCTG TGCTCCTATA AACCTAAACC TCCTCTACTT
3421 GAGAGGACAT TCCAATCATA GGCTGCCCAT CCACCTCTG TGCTCCTCTG TTAATTAGST
3481 CACTTAACAA AAAGGAAATT GGGTAGGGGT TTTTCACAGA CCGCTTTCTA AGGGTAATTT
3541 TAAAATATCT GGGAGTCCC TTCCACTGCT GTGTTCCAGA AGTGTGTGTA AACAGCCAC
3601 AAATGTCAAC AGCAGAAACA TACAAGCTGT CAGCTTTGCA CAAGGGCCCA ACACCCTGCT
3661 CATCAAGAAG CACTGTGGTT GCTGTGTTAG TAATGTGCAA AACAGGAGGC ACATTTTCCC
3721 CACCTGTGTA GGTTCAAAA TATCTAGTGT TTTCAITTTT ACTTGATCA GGAACCCAGC
3781 ACTCCACTGG ATAAGCATT TCTTATCCA AAACAGCCTT GTGGTCAGTG TTTCTCTGCT
3841 CACTGTCAAC TTAGCATTT TTTGGGGTTA CAGTTTGAGC AGGATATTTG GTCCTGTAGT
3901 TTGCTAACAC ACCCTGCAGC TCCAAAGGTT CCCCACCAAC AGCAAAAAAA TGAAAATTTG
3961 ACCCTTGAAT GGGTTTTCCA GCACCATTTT CATGAGTTT TTGTGTCCCT GAATGCAAGT
4021 TTAACATAGC AGTTACCCCA ATAACCTCAG TTTTAAAGT AACAGCTTCC CACATCAAAA
4081 TATTTCCACA GGTAAAGTCC TCATTTAAAT TAGGCAAGG AATTGCTCTA GAGCGGCCGC
4141 CACCGCGGTG GAGCTCCAAT TCGCCCTATA GTGAGTCGTA TTACGCGCGC TCACCTGGCCG
4201 TCGTTTTACA ACCTCGTGAC TGGGAAAACC CTGGCGTTAC CCAACTTAAT CGCCTTGAC
4261 CACATCCCAT TTTCCGCCAG TGGCGTAATA GCGAAGAGGC CCGCACCGAT CGCCTTCCC
4321 AACAGTTGCG CAGCCTGAAT GCGGAATGGG ACGCGCCCTG TAGCGCGCGA TTAAGCGCGG
4381 CGGGTGTGGT GGTACGCGC AGCGTGACCG CTACACTTGC CAGCGCCCTA GCGCCCGCTC
4441 CTTTCGCTTT CTTCCCTTCC TTTCTCGCCA CGTTCCCGCG CTTTCCCGCT CAAGCTCTAA
4501 ATCGGGGCT CTTTTAGGG TTCCGATTTA GTGCTTTACG GCACCTCGAC CCAAAAAAC
4561 TTGATTAGGG TGATGGTTCA CGTAGTGGGC CATCGCCCTG ATAGACGGTT TTTCCCGCTT
4621 TGACGTTGGA GTCCACGTTT TTTAATAGTG GACTCTTGT CCAACTGGA ACAACACTCA
4681 ACCCTATCTC GGTCTATTCT TTTGATTAT AAGGGATTTT GCCGATTTCG GCCTATTGTT
4741 TAAAAAATGA GCTGATTAA CAAAAATTA ACGCGAATTT TAACAAAATA TTAACGCTTA
4801 CAATTTAGGT GGCATTTTC GGGGAAATGT GCGCGGAACC CCTATTGTT TATTTTCTA
4861 AATACATTCA AATATGTATC CGCTCATGAG ACAATAACCC TGATAAATGC TTCAATAATA
4921 TTGAAAAAGG AAGAGTATGA GTATTCAACA TTTCCGTGTC GCCCTTATTC CCTTTTTTGC
4981 GGCATTTTGC CTTCTGTGTT TTGCTCACCC AGAAACGCTG GTGAAAGTAA AAGATGCTGA
5041 AGATCAGTTG GGTGCACGAG TGGGTTACAT CGAACTGGAT CTCAACAGCG GTAAGATCCT
5101 TGAGAGTTTT CGCCCCGAAG AACGTTTTCC AATGATGAGC ACTTTTAAAG TTCTGCTATG
5161 TGGCGCGGTA TTATCCCGTA TTGACGCGCG GCAAGAGCAA CTCGGTCGCC GCATACACTA
5221 TTCTCAGAAAT GACTTGGTTG AGTACTCACC AGTCACAGAA AAGCATCTTA CGGATGGCAT
5281 GACAGTAAGA GAATTATGCA GTGCTGCCAT AACCATGAGT GATAAAGCTG CGGCCAATCT
5341 ACTTCTGACA ACGATCGGAG GACCGAAGGA GCTAACCGCT TTTTGCACA ACATGGGGGA
5401 TCATGTAATC CGCCTTGATC GTTGGGAACC GGAGCTGAAT GAAGCCATAC CAAACGACGA
5461 GCGTGACACC ACGATGCCCTG TAGCAATGGC AACAACTGTT CGCAAACTAT TAACTGGCGA
5521 ACTACTTACT CTAGCTTCCC GGCAACAATT AATAGACTGG ATGGAGGCGG ATAAAGTTGC
5581 AGGACCACTT CTGCGCTCGG CCTTCCGGC TGGCTGGTTT ATTGCTGATA AATCTGGAGC
5641 CGGTGAGCGT GGGTCTCGCG GTATCATTTG AGCACTGGGG CCAGATGGTA AGCCCTCCCG
5701 TATCGTAGTT ATCTACAGCA CGGGGAGTCA GGCAACTATG GATGAACGAA ATAGACAGAT
5761 CGCTGAGATA GGTGCCTCAC TGATTAAGCA TTGGTAACTG TCAGACCAAG TTTACTCATA
5821 TATACTTTAG ATTGATTTAA AACTTCATTT TTAATTTAAA AGGATCTAGG TGAAGATCCT
5881 TTTTGATAAT CTCATGACCA AAATCCCTTA ACGTGAGTTT TCGTTCCACT GAGCGTCAGA
5941 CCCCCTAGAA AAGATCAAAG GATCTTCTTG AGATCCTTTT TTTCTGCGCG TAATCTGCTG
6001 CTGCAAAACA AAAAAACCA CGCTACCAGC GGTGGTTTGT TTGCGGATC AAGAGCTACC-

FIGURE 31C

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6061 AACTCTTTTT CCGAAGGTAA CTGGCTTCAG CAGAGCGCAG ATACCAAATA CTGTCCTTCT
6121 AGTGTAGCCG TAGTTAGGCC ACCACTTCAA GAACTCTGTA GCACCGCCTA CATACTCGC
6181 TCTGCTAATC CTGTTACCAG TGGCTGCTGC CAGTGGCGAT AAGTCGTGTC TTACCGGGTT
6241 GGAACAAGA CGATAGTTAC CGGATAAGGC GCAGCGGTCG GGCTGAACGG GGGGTTCTG
6301 CACACAGCCC AGCTTGGAGC GAACGACCTA CACCGAAGT AGTACCTAC AGCGTGAGCT
6361 ATGAGAAAGC GCCACGCTTC CCGAAGGGAG AAAGGCGGAC AGGTATCCGG TAAGCGGCAG
6421 GGTCCGAACA GGAGAGCGCA CGAGGGAGCT TCCAGGGGGA AACGCCTGGT ATCTTTATAG
6481 TCCTGTCGGG TTTCGCCACC TCTGACTTGA GCGTCGATTT TTGTGATGCT CGTCAGGGGG
6541 GCGGAGCCTA TGGAAAAACG CCAGCAACGC GGCCTTTTTA CGGTTCTCTG CTTTTGCTG
6601 GCCTTTTGCT CACATGTTCT TTCCTGCGTT ATCCCCTGAT TCTGTGGATA ACCGTATTAC
6661 CGCCTTTGAG TGAGCTGATA CCGCTCGCCG CAGCCGAACG ACCGAGCGCA GCGAGTCAGT
6721 GAGCGAGGAA GCGGAAGAGC GCCCAATACG CAAACCGCCT CTCCCCGCGC GTTGGCCGAT
6781 TCATTAATGC AGCTGGCAGC ACAGGTTTCC CGACTGGAAA GCGGGCAGTG AGCGCAACGC
6841 AATTAATGTG AGTTAGTCA CTCATTAGGC ACCCCAGGCT TTACACTTTA TGCTTCCGGC
6901 TCGTATGTTG TGTGGAATTG TGAGCGGATA ACAATTTTAC ACAGGAAACA GCTATGACCA
6961 TGATTACGCC AAGCGCGCAA TTAACCTCA CTAAAGGGAA CAAAAGCTGG GTACCGGGCC
7021 CCCCCCT
```

FIGURE 31D

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Figure 32A: pDEST12.2 CMV Promoter for Eukaryotic Expression, SV40 Promoter/ori for G418 Resistance

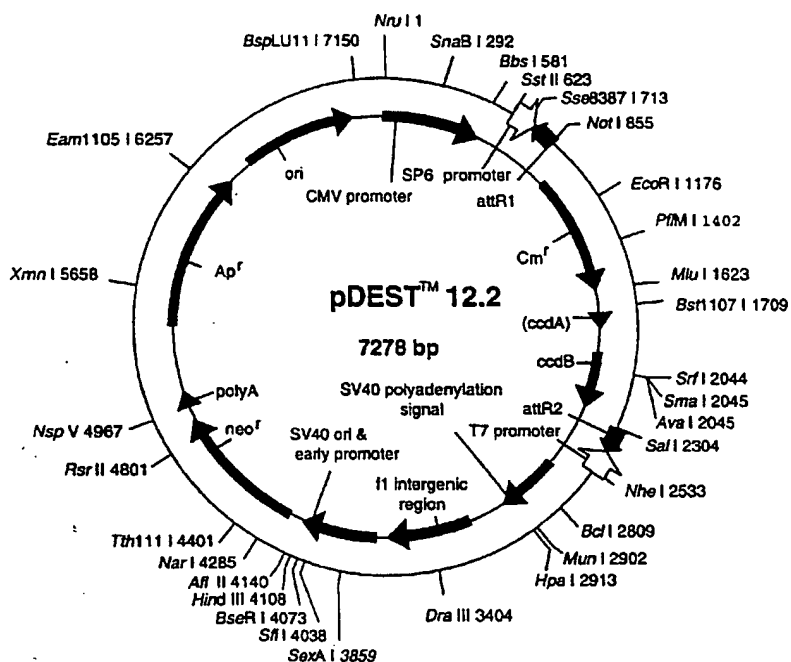
307 *mRNA from CMV promoter*
acc gtc aga tgc cct gga gac gcc atc cac gct gtt ttg acc tcc ata gaa
tgg cag tct agc gga cct ctg cgg tag gtg cga caa aac tgg agg tat ctt

358 gac acc ggg acc gat cea gcc tcc gga ctc tag cct agg ccg cgg agc gga
ctg tgg ccc tgg cta ggt cgg agg cct gag atc gga tcc ggc gcc tgc cct

409 taa caa ttt cac aca gga aac agc tat gac cat tag gcc ttt gca aaa agc
att gtt aaa gtg tgt cct ttg tgc ata ctg gta atc cgg aaa cgt ttt tgc

460 tat tta ggt gac act ata gaa ggt acg cct gca ggt ^{Aga} ^{EcoRI} ~~acc ggt ccg gaa ttc~~
ata aat cca ctg tga tat ctt cca tgc gga cgt cca tgg cca ggc ctt aag

511 ~~cca tca aca agt tgg taa ada gaa gct gaa cga gaa acg taa aat gat ata~~
~~ggt agt tgt tca aac atg ttt tuc cga cgt gct ctt tgc att tca cca tat~~



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pDEST12.2 7278 bp (rotated to position 3900)

Location (Base Nos.)	Gene Encoded
86..136	ori
220..742	CMV promoter
1059..935	attR1
1168..1827	CmR
1947..2031	inactivated ccdA
2169..2474	ccdB
2515..2639	attR2
2824..3186	small t & polyA
3310..3378	lac
4363..5157	neo
5680..6540	ampR

1	GGGGGGCGGA	GCCTATGGAA	AAACGCCAGC	AACGCGGCCT	TTTTACGGTT	CCTGGCCTTT
61	TGCTGGCCTT	TTGCTCACAT	GTTCTTTCCT	GCGTTATCCC	CTGATTCTGT	GGATAACCGT
121	ATTACCGCCT	TTGAGTGAGC	TGATACCGCT	CGCCGCAGCC	GAACGACCGA	GCGCAGCGAG
181	TCAGTGAGCG	AGGAAGCGGA	AGAGCTCGCG	AATGCATGTC	GTTACATAAC	TTACGGTAAA
241	TGGCCCCGCT	GGCTGACCGC	CCAACGACCC	CCGCCCAITG	ACGTCAATAA	TGACGTATGT
301	TCCCATAGTA	ACGCCAATAG	GGACTTTCCA	TTGACGTCAA	TGGGTGGAGT	ATTTACGGTA
361	AACTGCCCAC	TTGGCAGTAC	ATCAAGTGTA	TCATATGCCA	AGTACGCCCC	CTATTGACGT
421	CAATGACCGT	AAATGGCCCC	CCTGGCATT	TGCCCAGTAC	ATGACCTTAT	GGGACTTTCC
481	TACTTGGCAG	TACATCTACG	TATTAGTCAT	CGCTATTACC	ATGGTGAATG	GGTTTGGCA
541	GTACATCAAT	GGGCGTGGAT	AGCGGTTTGA	CTCACGGGGA	TTTCCAAGTC	TCCACCCCAT
601	TGACGTCAAT	GGGAGTTTGT	TTTGGCACCA	AAATCAACGG	GACTTTCCAA	AATGTCGTAA
661	CAACTCCGCC	CCATTGACGC	AAATGGGCGG	TAGGCGTGTA	CGGTGGGAGG	TCTATATAAG
721	CAGAGCTCGT	TTAGTGAACC	GTCAGATCGC	CTGGAGACGC	CATCCACGCT	GTTTGTACCT
781	CCATAGAAGA	CACCGGGACC	GATCCAGCCT	CCGGACTCTA	GCCTAGGCCG	CGGGACGGAT
841	AACAATTTCA	CACAGGAAC	AGCTATGACC	ATTAGGCCTT	TGCAAAAAGC	TATTTAGGTG
901	ACACTATAGA	AGGTACGCCT	GCAGGTACCG	GATCACAAGT	TTGTACAAAA	AAGCTGAACG
961	AGAAACGTAA	AATGATATAA	ATATCAATAT	ATTAAATTAG	ATTTTGATAA	AAAAACAGAC
1021	TACATAATAC	TGTAACACAC	AACATATCCA	GTCATATGG	CGGCCGCATT	AGGCACCCCA
1081	GGCTTTACAC	TTTATGCTTC	CGGCTCGTAT	AATGTGTGGA	TTTGTAGTTA	GGATCCGTCG
1141	AGATTTTCAG	GAGCTAAGGA	AGCTAAATG	GAGAAAAAAA	TCACTGGATA	TACCACCGTT
1201	GATATATCCC	AATGGCATCG	TAAAGAACAT	TTTGAGGCAT	TTCACTCAGT	TGCTCAATGT
1261	ACCTATAACC	AGACCGTTCA	GCTGGATATT	ACGGCCTTTT	TAAAGACCGT	AAAGAAAAAT
1321	AAGCACAAGT	TTTATCCGGC	CTTATTTCAC	ATTCTTGCCC	GCCTGATGAA	TGCTCATCCG
1381	GAATTCCTGT	TGGCAATGAA	AGACGGTGAG	CTGGTGATAT	GGGATAGTGT	TCACCCCTGT
1441	TACACCGTTT	TCCATGAGCA	AACTGAAACG	TTTTCATCGC	TCTGGAGTGA	ATACCACGAC
1501	GATTTCCGGC	AGTTTCTACA	CATATATTCC	CAAGATGTGG	CGTGTTACGG	TGAAACCTTG
1561	GCCTATTTCC	CTAAAGGGTT	TATTGAGAAT	ATGTTTTTCG	TCTCAGCCAA	TCCCTGGGTG
1621	AGTTTCACCA	GTTTGTGATT	AAACGTGGCC	AAATATGGACA	ACTTCTTCGC	CCCCGTTTTT
1681	ACCATGGGCA	AATATTATAC	GCAAGGCGAC	AAGGTGCTGA	TGCCGCTGGC	GATTGAGGTT
1741	CATCATGCCG	TCTGTGATGG	CTTCCATGTC	GGCAGAATGC	TTAATGAATT	ACAACAGTAC
1801	TGCGATGAGT	GGCAGGGCGG	GGCGTAAACG	CGTGGATCCG	GCTTACTAAA	AGCCAGATAA
1861	CAGTATGCGT	ATTTGCGCGC	TGATTTTTCG	GGTATAAGAA	TATATACTGA	TATGTATACC
1921	CGAAGTATGT	CAAAAAGAGG	TGTGCTATGA	AGCAGCGTAT	TACAGTGACA	GTTGACAGCG
1981	ACAGCTATCA	GTTGCTCAAG	GCATATATGA	TGTCAATATC	TCCGGTCTGG	TAAGCACAAC
2041	CATGCAGAA	GAGGCCCGTC	GTCTGCGTGC	CGAACGCTGG	AAAGCGGAAA	ATCAGGAAGG
2101	GATGGCTGAG	GTCGCCCGGT	TTATTGAAAT	GAACGGCTCT	TTTGCTGACG	AGAACAGGGA
2161	CTGGTGAAAT	GCAAGTTTAA	GTTTACACCT	ATAAAGAGA	GAGCCGTTAT	CGTCTGTTTG
2221	TGGATGTACA	GAGTGATATT	ATTGACACGC	CCGGGCGACG	GATGGTGATC	CCCCGGGCA
2281	GTGCACGTCT	GCTGTCAGAT	AAAGTCTCCC	GTGAACCTTA	CCCGGTGGTG	CATATCGGGG
2341	ATGAAAGCTG	GCGCATGATG	ACCACCGATA	TGGCCAGTGT	GCCGGTCTCC	GTTATCGGGG
2401	AAGAAGTGCC	TGATCTCAGC	CACCGCGAAA	ATGACATCAA	AAACGCCATT	AACCTGATGT

FIGURE 32B

2461 TCTGGGGAAT ATAAATGTCA GGCTCCCTTA TACACAGCCA GTCTGCAGGT CGACCATAGT
2521 GACTGGATAT GTTGTGTTTT ACAGTATTAT GTAGTCTGTT TTTTATGCAA AATCTAATTT
2581 AATATATTGA TATTATATC ATTTTACGTT TCTCGTTCAG CTTTCTTGTA CAAAGTGGTG
2641 ATCGCGTGCA TGGCAGCTCA TAGCTCTCTC CCTATAGTGA GTCGTATTAT AAGCTAGGCA
2701 CTGGCCGTG TTTTACAACG TCGTGACTGG GAAAACGTCT AGCTTGGGAT CTTTGTGAAG
2761 GAACCTTACT TCTGTGGTGT GACATAATTG GACAACTAC CTACAGAGAT TTAAAGCTCT
2821 AAGGTAAATA TAAAATTTTT AAGTGTATAA TGTGTTAAAC TAGCTGCATA TGCTTGCTGC
2881 TTGAGAGTTT TGCTTACTGA GTATGATTIA TGAATAATT ATACACAGGA GCTAGTGATT
2941 CTAATTGTTT GTGTATTTTA GATTACAGT CCCAAGGCTC AITTCAGGCC CTTAGTCCCT
3001 CACAGTCTGT TCATGATCAT AATCAGCCAT ACCACATTTG TAGAGGTTTT ACTTGCTTTA
3061 AAAAACCTCC CACACCTCCC CCTGAACCTG AAACATAAAA TGAATGCAAT TGTGTGTGT
3121 AACTTGTTTA TTGCAGCTTA TAATGGTTAC AAATAAAGCA ATAGCATCAC AAATTTTACA
3181 AATAAAGCAT TTTTTTCACT GCATTCTAGT TGTGTTTGT CCAAACCTCAT CAATGTATCT
3241 TATCATGTCT GGATCGATCC TGCATTAATG AATCGGCCAA CGCGCGGGGA GAGGCGGTTT
3301 GCGTATTGGC TGGCGTAATA GCGAAGAGGC CCGCACCAGT CGCCCTTCCC AACAGTTGCG
3361 CAGCCTGAAT GCGGAATGGG ACGCGCCCTG TAGCGCGCGA TTAAGCGCGG CGGCTGTGTT
3421 GGTACGCGC AGCGTGACCG CTACACTTGC CAGCGCCCTA GCGCCCGCTC CTTTCGCTTT
3481 CTTCCCTTCC TTTCTCGCCA CGTTCGCGG CTTTCCCGGT CAAGCTCTAA ATCGGGGGGT
3541 CCCTTTAGGG TTCCGATTTA GTGCTTTACG GCACCTCGAC CCCAAAAAAC TTGATTAGGG
3601 TGATGGTTCA CGTAGTGGC CATCGCCCTG ATAGACGGTT TTTCCGCTTT TGACGTTGGA
3661 GTCCACGTTT TTTAATAGTG GACTCTTGT CCAAACCTGA ACAACACTCA ACCCTATCTC
3721 GGTCTATTCT TTTGATTAT AAGGGAATTT GCCGATTTCG GCCTATTGGT TAAAAATGA
3781 GCTGATTTAA CAAATATTTA ACGCGAATTT TAACAAAATA TTAACGTTTA CAATTTTCGCC
3841 TGATGCGGTA TTTTCTCCTT ACGCATCTGT GCGGTATTTC ACACCGCATA CGCGGATCTG
3901 CGCAGCACA TGGCCTGAAA TAACCTCTGA AAGAGGAAC TGGTTAGGTA CCTTCTGAGG
3961 CGAAAGAAC CAGCTGTGGA ATGTGTGTCA GTTAGGGTGT GGAAGTCCC CCGCTTCCC
4021 AGCAGGCAGA AGTATGCAAA GCATGCATCT CAATTAGTCA GCAACCGAGT GTGGAAAGTC
4081 CCCAGGCTCC CCAGCAGGCA GAAGTATGCA AAGCATGCAT CTCATTTAGT CAGCAACCAT
4141 AGTCCCGCCC CTAACCTCCG CCATCCCGCC CTAACCTCCG CCCAGTTCCG CCCTATTCTC
4201 GCGCCATGGC TGACTAATTT TTTTATTTA TGCAGAGGCC GAGGCCGCT CGGCCTCTGA
4261 GCTATTCCAG AAGTAGTGAG GAGGCTTTT TGGAGGCCCTA GGCTTTTGCA AAAAGCTTGA
4321 TTCTTCTGAC ACAACAGTCT CGAACTTAAG GCTAGAGCCA CATGATTGA ACAAGATGGA
4381 TTGCACGCAG GTTCTCCGGC CGCTTGGGTG GAGAGGCTAT TCGGCTATGA TCGGGCACAA
4441 CAGACAATCG GCTGCTCTGA TGCCGCGGTG TTCCGGCTGT CAGCGCAGGG GCGCCCGGTT
4501 CTTTTTGTCA AGACCGACCT GTCCGGTGCC CTGAATGAAC TGCAGGACGA GGCAGCGCGG
4561 CTATCGTGGC TGGCCACGAC GGGCGTTCC TGCAGCTGTG TGCTCGACGT GTTCACTGAA
4621 GCGGAAGGG ACTGGCTGCT ATTGGGCGAA GTGCCGGGGC AGGATCTCCT GTCATCTCAC
4681 CTGTCTCCTG CCGAGAAAGT ATCCATCATG GCTGATGCAA TGCGGCGGCT GCATACGCTT
4741 GATCCGGCTA CCTGCCCAT CGAACACCAA GCGAAACATC GCATCGAGCG AGCACGTACT
4801 CGGATGGAAG CCGGTCTTGT CGATCAGGAT GATCTGGACG AAGAGCATCA GGGGCTCGCG
4861 CCAGCCGAAC TGTTCCGCCAG GCTCAAGGCG CGCATGCCCG ACGGCGAGGA TCTCGTCTG
4921 ACCCATGGCG ATGCTGCTT GCCGAATATC ATGGTGGAAA ATGGCCGCTT TTCTGGATTG
4981 ATCGACTGTG GCGGCTGGG TGTGGCGGAC CGCTATCAGG ACATAGCGTT GGCTACCCGT
5041 GATATTGCTG AAGAGCTTGG CCGCGAATGG GCTGACCCTT TCCTCGTCTT TACGGTATC
5101 GCGCTCCCG ATTCCGAGCG CATCGCCTTC TATCGCCTTC TTGACGAGTT CTCTGAGCG
5161 GGACTCTGGG GTTCGAAATG ACCGACCAAG CGACGCCCAA CCTGCCATCA CGATGGCCCG
5221 AATAAAATAT CTTTATTTT ATTACATCTG TGTGTTGGTT TTTTGTGTGA ATCGATAGCG
5281 ATAAGGATCC GCGTATGGTG CACTCTCAGT ACAATCTGCT CTGATGCCCG ATAGTTAAGC
5341 CAGCCCCGAC ACCCGCCAAC ACCCGCTGAC GCGCCCTGAC GGGCTTGTCT GTCGCCGCA
5401 TCGCTTACA GACAAGCTGT GACCGTCTCC GGGAGCTGCA TGTGTGAGAG GTTTTACCG
5461 TCATCACCGA AACGCGCGAG ACGAAAGGGC CTCGTGATAC GCCTATTTT ATAGGTTAAT
5521 GTCATGATAA TAATGGTTT TTAGACGTCA GGTGGCACTT TTCGGGAAA TGTGCGCGGA
5581 ACCCTATTT GTTTATTTT CTAAATACAT TCAATATGT ATCCGCTCAT GAGACAATAA
5641 CCCTGATAAA TGCTTCAATA ATATTGAAA AGGAAGAGTA TGAGTATTCA ACATTTCCGT
5701 GTGCGCCTTA TTCCCTTTT TGCGGCATTT TGCTTCTCTG TTTTGTCTCA CCCAGAAACG
5761 CTGGTGAAAG TAAAAGATGC TGAAGATCAG TTGGGTGCAC GAGTGGGTTA CATCGAACTG
5821 GATCTCAACA GCGGTAAAGT CCTTGAGAGT TTTCCGCCCG AAGAACGTTT TCCAATGATG
5881 AGCACTTTTA AAGTCTGCT ATGTGGCGCG GTATTATCCC GTATTGACGC CGGGCAAGAG-

FIGURE 32C

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5941 CAACTCGGTC GCCGCATACA CTATTCTCAG AATGACTTGG TTGAGTACTC ACCAGTCACA
6001 GAAAAGCATC TTACGGATGG CATGACAGTA AGAGAATTAT GCAGTGCTGC CATAACCATG
6061 AGTGATAACA CTGCGGCCAA CTTACTTCTG ACAACGATCG GAGGACCGAA GGAGCTAACC
6121 GCTTTTTTGC ACAACATGGG GGATCATGTA ACTCGCCTTG ATCGTTGGGA ACCGGAGCTG
6181 AATGAAGCCA TACCAAAACGA CGAGCGTGAC ACCACGATGC CTGTAGCAAT GGCAACAACG
6241 TTGCGCAAAC TATTAACCTGG CGAACTACTT ACTCTAGCTT CCCGGCAACA ATTAATAGAC
6301 TGGATGGAGG CGGATAAAGT TGCAGGACCA CTTCTGCGCT CGGCCCTTCC GGCTGGCTGG
6361 TTTATTGCTG ATAAATCTGG AGCCGGTGAG CGTGGGTCTC GCGGTATCAT TGCAGCACTG
6421 GGGCCAGATG GTAAGCCCTC CCGTATCGTA GTTATCTACA CGACGGGGAG TCAGGCAACT
6481 ATGGATGAAC GAAATAGACA GATCGCTGAG ATAGGTGCCT CACTGATTAA GCATTGGTAA
6541 CTGTCAGACC AAGTTTACTC ATATATACTT TAGATTGATT TAAAACTTCA TTTTAAATTT
6601 AAAAGGATCT AGGTGAAGAT CCTTTTTGAT AATCTCATGA CCAAAATCCC TTAACGTGAG
6661 TTTTCGTTCC ACTGAGCGTC AGACCCCGTA GAAAAGATCA AAGGATCTTC TTGAGATCCT
6721 TTTTCTCTGC GCGTAATCTG CTGCTTGCAA ACAAAAAAAC CACCGCTACC AGCGGTGGTT
6781 TGTTTGCCGG ATCAAGAGCT ACCAACTCTT TTTCCGAAGG TAACTGGCTT CAGCAGAGCG
6841 CAGATACCAA ATACTGTCCT TCTAGTGTAG CCGTAGTTAG GCCACCACTT CAAGAACTCT
6901 GTAGCACCCG CTACATACCT CGCTCTGCTA ATCCTGTTAC CAGTGGCTGC TGCCAGTGGC
6961 GATAAGTCGT GTCCTACCGG GTTGGACTCA AGACGATAGT TACCGGATAA GGCGCAGCGG
7021 TCGGGCTGAA CGGGGGGTTT GTGCACACAG CCCAGCTTGG AGCGAACGAC CTACACCGAA
7081 CTGAGATACC TACAGCGTGA GCATTGAGAA AGCGCCACGC TTCCCGAAGG GAGAAAGGCG
7141 GACAGGTATC CGGTAAGCGG CAGGGTCGGA ACAGGAGAGC GCACGAGGGA GCTTCCAGGG
7201 GGAAACGCCT GGTATCTTTA TAGTCCTGTC GGGTTTCGCC ACCTCTGACT TGAGCCTCGA
7261 TTTTGTGAT GCTCGTCA

FIGURE 32D

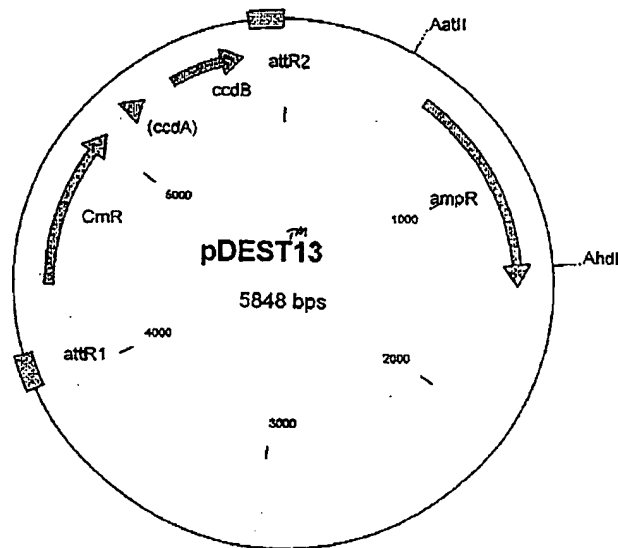
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Figure 33A:

pDEST13

Native protein in *E. coli*: λ PL promoter

3721 ^{BglII}
 tgggcaaacc aagacagcta aagatctctc acctaccaaa caatgcccc ctgcaaaaaa
 acccgtttgg ttctgtcgat ttctagagag tggatgggtt gttacggggg gacgtttttt
 3781 taaattcata taaaaaacat acagataacc atctgcggtg ataaattatc tctggcggtg
 atttaagtat attttttgta tgtctattgg tagacgccac tatttaatag agaccgccac
 3841 ⁻³⁵ λ PL Promoter ⁻¹⁰ ^{mRNA}
 ttgacataaa taccactggc ggtgatactg agcacatcag caggacgcac tgaccaccat
 aactgtattt atggtgaccg ccactatgac tctgtagtc gtctgcgtg actggtggta
 3901 ^{EcoNI}
 gaaggtgacg ctcttaaaaa ttaagecctg aagaaggcca gcattcaag cagaaggctt
 ctccactgc gagaattttt aattcgggac tcttcccg cgttaagttc gtcttccgaa
 3961 ^{att P1}
 tggggtgtgt gatacgaaac gaagcattgg gatcatcaca agttgtaca aaaaagctga
 accccacaca ctatgctttg cttcgttaacc ctagtagtgt tcaaacaatgt ttttcgact



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pDEST13 5848 bp

<u>Location (Base Nos.)</u>		<u>Gene Encoded</u>
599..1458		amp ^R
4123..3998		attR1
4372..5031		Cm ^R
5151..5235		inactivated ccdA
5373..5678		ccdB
5719..5843		attR2

1	TTC	ACT	GGCC	GTC	GT	TTT	TAC	AAC	GT	CGT	GA	CTG	GG	AAA	AC	CCT	GG	CGT	TA	CCCA	ACT	TAA		
61	TCG	CC	TTG	CA	GC	AT	CCCC	CTT	TCG	CC	CAG	CTG	GC	TAAT	AG	CGA	AG	AGG	CCG	CA	CCG	GA		
121	TCG	CC	CTT	CC	CA	AC	AGT	TGC	GC	AG	CC	TGA	TGG	CGA	AT	GG	CG	CT	GAT	GC	GGT	ATTTT	CT	
181	CCT	TAC	GC	AT	CTG	TG	CGG	TAA	TTT	CAC	AC	CG	CAT	AT	GGT	GC	ACT	CT	CAG	TA	CAAT	CTG	CTC	
241	TG	AT	G	CCG	CA	TAG	TTA	AG	CC	CCG	GAC	ACC	GC	CA	ACA	CCG	CTG	AC	G	CG	CC	CTG	ACG	
301	GG	CT	TGT	CTG	CT	CC	CGG	CA	CCG	CT	TAC	ACA	AG	CT	GTG	ACC	GT	CT	CCG	GG	AG	CTG	CA	
361	GT	G	T	CAG	AGG	TTT	T	CAC	CGT	CAT	CAC	CGA	ACG	CG	CGA	GAG	CGA	AGG	GCC	TC	GT	GAT	ACG	
421	CCT	AT	TTTT	TAG	GT	TAA	TG	TC	AT	GATA	AT	AAT	GGT	TTT	CT	TAG	AC	GT	CAG	GTG	GC	ACT	TT	
481	TCG	GG	GAA	AT	GTG	CG	CGG	AA	CCC	CT	ATT	TG	TTT	AT	TTTT	TAA	AT	AC	ATT	CAA	AT	TAT	GTA	
541	TCC	GT	CT	CA	TG	AC	ATA	AT	CCT	GATA	AA	GCT	TT	CA	TAA	TAT	TG	AAAA	GGA	AG	AG	TAT		
601	GAG	TAT	TCAA	CATT	T	CCG	TG	TCG	CC	CT	TAT	TCC	CT	TTTT	TG	CGG	CA	TTTT	GC	CT	T	CT	GT	
661	TTT	T	GCT	CAC	CC	AG	AA	ACG	TGG	TAA	AGT	AAA	AG	AT	GCT	GA	AG	AT	CAG	TGG	GT	GCA	CG	
721	AGT	GGG	TTAC	AT	CGA	ACT	TG	AT	CT	CA	AC	CG	GT	A	AG	AT	C	CT	G	AG	AG	T	T	
781	AGA	AC	GTTTT	CCA	AT	GAT	GA	GCA	CT	TTTT	TAA	AGT	TT	CT	GCTA	TG	TG	CG	CGCG	TAT	TAT	CCCG		
841	TAT	T	GAC	GCC	GGG	CA	AG	AGC	AAC	T	CGG	TG	CC	G	TAC	AT	C	AG	TAT	T	CT	GAT	TG	
901	TG	AG	TACT	CA	CC	AG	T	CAC	AG	AAA	AG	CA	TCT	TAC	GG	AT	G	GC	AT	AA	GAG	AT	TATG	
961	CAG	T	GT	GCC	ATA	ACC	AT	GA	GT	GATA	AC	AC	TG	CG	G	CA	AC	TT	ACT	T	CT	G	AT	
1021	AGG	AC	CGA	AG	GAG	CTA	AC	CG	CT	TTTT	TG	CA	CA	AC	T	GGG	GAT	CA	T	GT	AA	CTC	G	
1081	TCG	TT	GG	AA	CCG	GAG	CT	GA	AT	GA	AC	CA	AC	CA	CG	AG	GAG	CG	T	G	CA	CC	AG	
1141	TGT	AG	CAAT	G	CA	CA	AC	GT	TG	CG	CA	AA	CT	AT	TAA	CT	GG	CA	ACT	T	CT	AG	CT	
1201	CCG	G	CA	CA	AA	TAA	TAG	ACT	GG	AT	GG	AG	GC	GA	TAA	AG	TT	GC	AG	CA	CC	AG	CT	
1261	GG	CC	CTT	CCG	GCT	GG	CT	GGT	TT	ATT	TG	CT	GA	TAA	AT	CT	GG	GC	AT	T	CT	AG	CT	
1321	CGG	TAT	CATT	GC	AG	CA	CT	TG	GG	CC	AG	TG	TAA	AT	CT	GG	GC	GT	GAG	GTG	GG	T	CT	
1381	GAC	GGG	GAGT	CAGG	CA	ACTA	TG	GA	TGA	ACG	AA	TAG	AC	AG	AT	CG	CT	GAG	A	TAG	GT	GC	CT	
1441	ACT	GAT	TAA	G	CAT	TG	TAAC	TG	T	CAG	ACCA	AG	TTT	ACT	CA	TAT	AT	CT	T	AG	AT	T	GATT	
1501	AAA	ACT	TCAT	TTTT	AA	TTTA	AA	AG	GAT	CTA	GG	TGA	AG	AT	C	TTTT	T	GAT	A	AT	CT	CA	TG	
1561	CAA	AA	TCCCT	TAA	CG	TG	AGT	TTT	CG	T	CCA	CTG	AG	CG	T	CA	CC	CG	TAG	AAA	AG	AT	CAA	
1621	AGG	AT	CTTCT	TG	AG	AT	CTT	TTTT	T	CTG	CG	CG	TAA	T	CTG	TG	CA	AA	CA	AAAA	AA	AC	C	
1681	ACC	G	CTACCA	GCG	GT	GGT	TTT	GTT	TG	CCG	GA	TCA	AG	AG	CTA	CCA	ACT	CTT	TT	CCG	AG	AG	T	
1741	AAC	TG	GCTT	AG	CA	GAG	CGC	AG	AT	ACCA	AA	TAC	TG	TTCTT	CT	CTAG	T	GAG	C	CGT	AG	T	TAG	
1801	CC	AC	CACTT	C	AAG	AA	CTG	TAG	CAC	CGC	TAC	AT	AC	CTC	GCT	CT	G	CTA	A	TC	CT	G	T	
1861	AGT	GG	CTGCT	GCC	AG	TGG	CG	ATA	AG	T	CGT	G	TCT	T	AC	CGG	TG	G	ACT	CA	G	AG	T	
1921	ACC	G	GATAAG	GCG	CAG	CGGT	CGG	G	CT	GAAC	GGG	G	G	G	G	TT	G	C	A	C	A	G	C	
1981	GCG	AAC	GACC	TAC	ACC	GAA	C	TG	AG	AT	ACCT	AC	AG	CG	TG	AG	CAT	T	G	A	A	A	A	
2041	TCC	CGA	AGGG	AG	AA	AG	CGG	AC	AG	G	TAT	CC	GG	T	AAG	CGG	AGG	G	T	C	G	A	A	
2101	CAC	GAG	GAG	CTT	C	AGGG	G	GAA	AC	G	CTG	G	GT	AT	CTT	TAT	AG	T	C	T	G	C	C	
2161	CCT	CT	GACTT	GAG	CG	T	CGAT	TTTT	T	G	TATG	G	CT	CG	T	CAGG	G	G	CGG	AG	GC	T	GA	
2221	CGC	CA	GCAAC	GCG	G	CTTTT	TAC	CG	T	T	CTCT	TG	G	C	TTTTG	TG	TG	G	C	T	A	T	T	
2281	CTT	T	CTGCG	TTAT	CCCC	CTG	ATT	CT	G	TGGA	TAA	CCG	TAT	TAC	CG	TAT	T	GG	C	T	T	T	TG	
2341	TAC	CG	CTCGC	CGC	AG	CGG	AA	CG	AC	CGAG	CG	CAG	CG	AG	TCA	GTG	AG	CG	AGG	AAG	CG	GA	GA	
2401	GCG	CCCA	ATA	CG	CA	ACCG	C	TCT	CCCC	CG	GCG	T	GG	CCG	ATT	CAT	TAA	TG	AG	CT	GG	CA	A	
2461	CGA	CAG	GTTT	CCG	AG	CT	GGA	AAG	CGG	CAG	TG	CG	CG	CA	AC	GCA	AT	TAA	TG	AG	T	AG	CT	
2521	CAC	T	CA	TAT	GC	AC	CC	CAGG	CTT	T	CACTT	TAT	G	C	T	TCCG	GCT	CG	TAT	G	TG	TG	GAAT	
2581	TGT	GAG	CGGA	TAA	CA	ATTT	C	AC	AC	AG	AAAA	CAG	CT	AT	G	AC	CAT	G	AT	TACG	CCA	AG	CTT	GG
2641	CTG	CAG	GTGA	TG	ATT	TAC	G	CC	AG	CAG	A	T	TAA	AG	AAAA	CAG	AC	AG	G	T	TAT	T	G	CGC
2701	TTAT	CT	TTTCC	CTTT	AT	TTTTT	GCT	G	CGG	TAA	GTC	G	CAT	AAA	AAC	CA	T	T	CTT	CAT	AAT	T	CAA	

FIGURE 33B

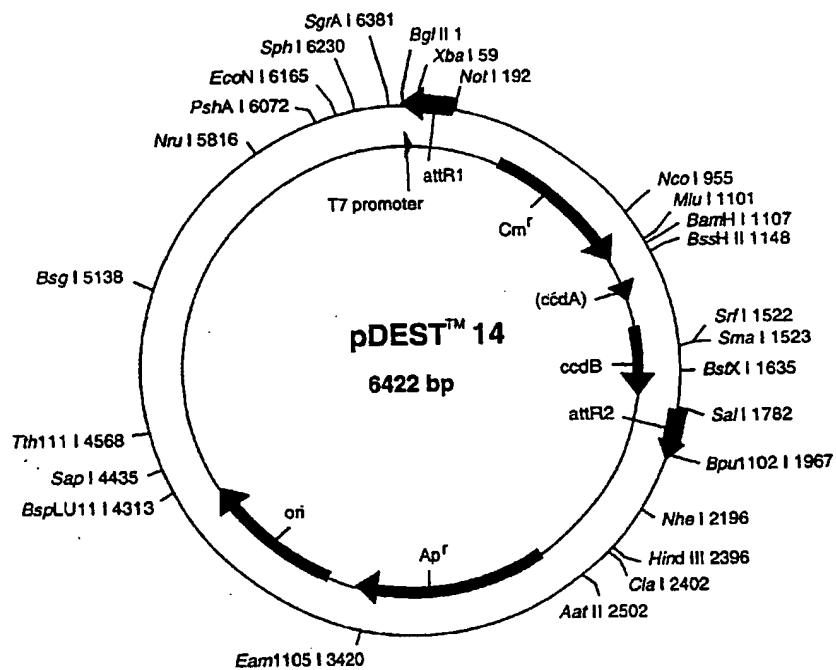
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2761 TCCATTTACT ATGTTATGTT CTGAGGGGAG TGAAAATTCC CCTAATTCCA TGAAGATTCT
2821 TGCTCAATTG TTATCAGCTA TGCGCCGACC AGAACACCTT GCCGATCAGC CAAACGTCTC
2881 TTCAGGCCAC TGACTAGCGA TAACTTTCCC CACAACGGAA CAACCTCAT TGCATGGGAT
2941 CATTGGGTAC TGTGGGTTTA GTGGTTGTAA AAACACCTGA CCGCTATCCC TGATCAGTTT
3001 CTTGAAGGTA AACTCATCAC CCCCAAGTCT GGCTATGCAG AAATCACCTG GCTCAACAGC
3061 CTGCTCAGGG TCAACGAGAA TTAACATTCC GTCAGGAAAG CTTGGCTTGG AGCCTGTTGG
3121 TGCGGTCAATG GAATTACCTT CAACCTCAAG CCAGAAATGCA GAATCACTGG CTTTTTGGT
3181 TGTGCTTACC CATCTCTCCG CATCACCTTT GGTAAAGGTT CTAAGCTTAG GTGAGAACAT
3241 CCCTGCCCTGA ACATGAGAAA AAACAGGGTA CTCATACTCA CTTCTAAGTG ACGGCTGCAT
3301 ACTAACCGCT TCATACATCT CGTAGATTTC TCTGGCGATT GAAGGGCTAA ATTCTTCAAC
3361 GCTAACTTTG AGAATTTTTG CAAGCAATGC GGCCTTATAA GCATTTAATG CATTGATGCC
3421 ATTAAATAAA GCACCAACGC CTGACTGCCC CATCCCATC TTGCTGCGA CAGATTCTCG
3481 GGATAAGCCA AGTTCAATTT TCTTTTTC ATAAATGCT TTAAGGCGAC GTGCGTCTCT
3541 AAGCTGCTCT TGTGTTAATG GTTCTTTTT TGTGCTCATA CGTTAAATCT ATCACCAGCA
3601 GGGATAAATA TCTAACACCG TGCGTGTGA CTATTTTACC TCTGGCGGTG ATAATGGTTG
3661 CATGTACTAA GGAGGTTGTA TGGAAACAACG CATAACCTTG AAAGATTATG CAATGCGCTT
3721 TGGGCAAAAC AAGACAGCTA AAGATCTCTC ACCTACCAAA CAATGCCCCC CTGCAAAAAA
3781 TAAATTCATA TAAAAACAT ACAGATAACC ATCTGCGGTG ATAAATTATC TCTGGCGGTG
3841 TTGACATAAA TACCACTGGC GGTGATACTG AGCACATCAG CAGGACGCAC TGACCACCAT
3901 GAAGGTGACG CTCCTAAAAA TTAAGCCCTG AAGAAGGGCA GCATTCAAAG CAGAAGGCTT
3961 TGGGGTGTGT GATACGAAAC GAAGCATTGG GATCATCACA AGTTTGTAAG AAAAGCTGA
4021 ACGAGAAACG TAAATGATA TAAATATCAA TATATTAAAT TAGATTTTGC ATAAAAACA
4081 GACTACATAA TACTGTAAAA CACAACATAT CCAGTCACTA TGGCGGCCGC TAAAGTTGGA
4141 GCATCACCCG ACGCACTTTG CGCCGAATAA ATACCTGTGA CGGAAGATCA CTTCCGAGAA
4201 TAAATAAATC CTGGTGTCCC TGTGTATACC GGGAAAGCCCT GGGCCAACTT TTGGCGAATA
4261 TGAGACGTTG ATCGGCACGT AAGAGGTTCC AACTTTTACC ATAATGAAAT AAGATCACTA
4321 CCGGGCGTAT TTTTGTAGTT ATCGAGATT TCAGGAGCTA AGGAAGCTAA AATGGAGAAA
4381 AAAATCACTG GATATACCAC CGTTGATATA TCCCAATGGC ATCGTAAAGA ACATTTTGAG
4441 GCATTTCACT CAGTTGTCTA ATGTACCTAT AACCAGACCG TTCAGCTGGA TATTACGGCC
4501 TTTTAAAGA CCGTAAAGAA AAATAAGCAC AAGTTTATC CGGCCTTTAT TCACATTCTT
4561 GCCCGCCTGA TGAATGCTCA TCCGGAATTC CGTATGGCAA TGAAGACGG TGAGCTGGTG
4621 ATATGGGATA GTGTTACACC TTGTTACACC GTTTTCCATG AGCAAACTGA AACGTTTCA
4681 TCGCTCTGGA GTGAATACCA CGACGATTTC CGGCAGTTTC TACACATATA TTCGCAAGAT
4741 GTGGCGTGT ACGGTGAAAA CTTGGCCTAT TTCCCTAAAG GGTTTATTGA GAATATGTTT
4801 TTCGTCTCAG CCAATCCCTG GGTGAGTTTC ACCAGTTTTC ATTTAAACGT GGCCAATATG
4861 GACAACCTCT TCGCCCCCGT TTTCACCATG GGCAATATT ATACGCAAGG CGACAAGGTG
4921 CTGATGCCGC TGGCGATTCA GGTTCATCAT GCCGTCTGTG ATGGCTTCCA TGTCCGCAGA
4981 ATGCTTAATG AATTACAACA GTACTGCGAT GAGTGGCAGG GCGGGGCGTA AACGCGTGA
5041 TCCGGCTTAC TAAAGCCAG ATAACAGTAT GCGTATTTGC GCGCTGATT TTGCGGTATA
5101 AGAATATATA CTGATATGTA TACCCGAAGT ATGTCAAAAA GAGGTGTGCT ATGAAGCAGC
5161 GTATTACAGT GACAGTTGAC AGCGACAGCT ATCAGTTGCT CAAGGCATAT ATGATGTCAA
5221 TATCTCCGGT CTGGTAAGCA CAACCATGCA GAATGAAGCC CGTCTCTGCG GTGCCGAACG
5281 CTGGAAAGCG GAAAATCAGG AAGGGATGGC TGAGGTGCGC CGGTTTATTG AAATGAACGG
5341 CTCTTTTGTG GACGAGAACA GGGACTGGTG AAATGCAATT TAAGGTTTAC ACCTATAAAA
5401 GAGAGAGCCG TTATCGTCTG TTTGTGGATG TACAGAGTGA TATTATTGAC ACGCCCGGGC
5461 GACGGATGGT GATCCCCCTG GCCAGTGCAC GTCTGCTGTC AGATAAAGTC TCCCGTGAAC
5521 TTTACCCGGT GGTGCATATC GGGGATGAAA GCTGGCGCAT GATGACCACC GATATGGCCA
5581 GTGTGCCGGT CTCCGTTATC GGGGAAGAAG TGGCTGATCT CAGCCACCGC GAAAATGACA
5641 TCAAAAACGC CATTAACTG ATGTTCTGGG GAATATAAAT GTCAGGCTCC GTTATACACA
5701 GCCAGTCTGC AGGTCGACCA TAGTGACTGG ATATGTTGTG TTTTACAGTA TTATGTAGTC
5761 TGTTTTTTAT GCAAAATCTA ATTTAATATA TTGATATTTA TATCATTTTA CGTTTCTCGT
5821 TCAGCTTCTT TGTACAAAGT GGTGATAA

FIGURE 33C

Figure 34A: pDEST14 Native Protein Expression in *E. coli*, T7 Promoter

3961 tgccggccac gatcggtccg gcgtagagga tcgagatctc gatcccgcca aatttaatacg
 acggccggtg ctaccgaggc cgcattctct agctctagag ctagggcgct ttaattatgc
 4021 // actcactata gggagaccac aacggtttcc ctttagatca caagtttga caaaaaagct
 // tgagtgatat ccctctggtg ttgccaaagg gagatctagt gttcaaacat gtttttcga //



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pDEST14 6422 bp (rotated to position 4000)

<u>Location (Base Nos.)</u>		<u>Gene Encoded</u>	
185..61		attR1	
435..1094		CmR	
1214..1298		inactivated ccdA	
1436..1741		ccdB	
1782..1906		attR2	
2632..3489		ampR	
1	CGATCCCGCG AAATTAATAC GACTCACTAT AGGGAGACCA CAACGGTTTC CCTCTAGATC		
61	ACAAGTTTGT AAAAAAAGC TGAACGAGAA ACGTAAATG ATATAAATAT CAATATATTA		
121	AATTAGATT TGCATAAAAA ACAGACTACA TAATACTGTA AAACACAACA TATCCAGTCA		
181	CTATGGCGGC CGCTAAGTTG GCAGCATCAC CCGACGCACT TTGCGCCGAA TAAATACCTG		
241	TGACGGAAGA TCACCTTCGCA GAATAAATAA ATCCTGGTGT CCCTGTTGAT ACCGGGAAGC		
301	CCTGGGCCAA CTTTGGCCGA AAATGAGACG TTGATCGGCA CGTAAGAGGT TCCAACTTTC		
361	ACCATAATGA AATAAGATCA CTACCGGGCG TATTTTTTGA GTTATCGAGA TTTTCAGGAG		
421	CTAAGGAAGC TAAAATGGAG AAAAAAATCA CTGGATATAC CACCGTTGAT ATATCCCAAT		
481	GGCATCGTAA AGAACATTTT GAGGCATTTT AGTCAGTTGC TCAATGTACC TATAACCAGA		
541	CGGTTTCAGT GGATATTACG GCCTTTTTAA AGACCGTAAA GAAAAATAAG CACAAGTTTT		
601	ATCCGGCCTT TATTCACATT CTTGCCCGCC TGATGAATGC TCATCCGGA TCCCGTATGG		
661	CAATGAAAGA CGGTGAGCTG GTGATATGGG ATAGTGTTC CCGTTGTTAC ACCGTTTTCC		
721	ATGAGCAAAC TGAAACGTTT TCATCGCTCT GGAGTGAATA CCACGACGAT TTCCGGCAGT		
781	TTCTACACAT ATATTGCAAA GATGTGGCGT GTTACGGTGA AAACCTGGCC TATTTCCCTA		
841	AAGGGTTTAT TGAGAATATG TTTTTCGTCT CAGCCAATCC CTGGGTGAGT TTCACCAGTT		
901	TTGATTTAAA CGTGGCCAAT ATGGAACACT TCCTCGCCCC CGTTTTACC ATGGGCAAAAT		
961	ATTATACGCA AGGCGACAAG GTGCTGATGC CGCTGGCGAT TCAGGTTTCA CATGCCGTCT		
1021	GTGATGGCTT CCATGTCCGC AGAATGCTTA ATGAATTACA ACAGTACTGC GATGAGTGGC		
1081	AGGGCGGGGC GTAAACGCGT GGATCCGGCT TACTAAAAGC CAGATAACAG TATGCGTATT		
1141	TGCGCGCTGA TTTTGGCGGT ATAAGAATAT ATACTGATAT GTATACCCGA AGTATGTCAG		
1201	AAAGAGGTGT GCTATGAAGC AGCGTATTAC AGTGACAGTT GACAGCGACA GCTATCAGTT		
1261	GCTCAAGGCA TATATGATGT CAATATCTCC GGTCTGGTAA GCACAACCAT GCAGAATGAA		
1321	GCCCGTCGTC TGCGTGCCGA ACGCTGGAAA GCGGAAAATC AGGAAGGGAT GGCTGAGGTC		
1381	GCCCGGTTTA TTGAAATGAA CGGCTCTTTT GCTGACGAGA ACAGGGACTG GTGAAATGCA		
1441	GTTTAAGGTT TACACCTATA AAAGAGAGAG CCGTTATCGT CTGTTTGTGG ATGTACAGAG		
1501	TGATATTATT GACACGCCCG GCGACGGAT GGTGATCCCC CTGGCCAGTG CACGTCTGCT		
1561	GTGAGATAAA GTCTCCCGTG AACTTTACCC GGTGGTGATC ATCGGGGATG AAAGCTGGCG		
1621	CATGATGACC ACCGATATGG CCAGTGTGCC GGTCTCCGTT ATCGGGGAAG AAGTGGCTGA		
1681	TCTCAGCCAC GCGGAAAATG ACATCAAAAA CGCCATTAACT CTGATGTTCT GGGGAATATA		
1741	AATGTGAGGC TCCCTTATAC ACAGCCAGTC TGCAGGTCGA CCATAGTGAC TGGATATGTT		
1801	GTGTTTACAA GTATTATGTA GTCTGTTTTT TATGCAAAAT CTAATTTAAT ATATTGATAT		
1861	TTATATCAAT TTACGTTTCT CGTTCAGCTT TCTTGACAAA AGTGGTGATG ATCCGGCTGC		
1921	TAACAAAGCC CGAAAGGAAG CTGAGTTGGC TGCTGCCACC GCTGAGCAAT AACTAGCATA		
1981	ACCCCTTGGG GCCTCTAAAC GGGTCTTGAG GGGTTTTTTG CTGAAAGGAG GAACTATATC		
2041	CGGATATCCA CAGGACGGGT GTGGTCGCCA TGATCGCGTA GTCGATAGTG GCTCCAAGTA		
2101	GCGAAGCGAG CAGGACTGGG CGGCGGCCAA AGCGGTCGGA CAGTGCTCCG AGAACGGGTG		
2161	CGCATAGAAA TTGCATCAAC GCATATAGCG CTAGCAGCAC GCCATAGTGA CTGGCGATGC		
2221	TGTCGGAATG GACGATATCC CGCAAGAGGC CCGGCAGTAC CGGCATAACC AAGCCTATGC		
2281	CTACAGCATC CAGGGTGACG GTGCCGAGGA TGACGATGAG CGCATTGTTA GATTTTCATC		
2341	ACGGTGCCTG ACTGCGTTAG CAATTAACT GTGATAAACT ACCGCATTAA AGCTTATCGA		
2401	TGATAAGCTG TCAAACATGA GAATTCTTGA AGACGAAAGG GCCTCGTGAT ACGCCTATTT		
2461	TTATAGGTTA ATGTCATGAT AATAATGGTT TCTTAGACGT CAGGTGGCAC TTTTCGGGGA		
2521	AATGTGCGCG GAACCCCTAT TTGTTTATTT TTCTAAATAC ATTCAAATAT GTATCCGCTC		
2581	ATGAGACAAT AACCCGTGTA AATGCTTCAA TAATATTGAA AAAGGAAGAG TATGAGTATT		
2641	CAACATTTCG GTGTCGCCCT TATTCCTTT TTTGCGGCAT TTTGCCCTTC TGTTTTGTCT		
2701	CACCCAGAAA CGCTGGTGAA AGTAAAAGAT GCTGAAGATC AGTTGGGTGC ACCAGTGGGT-		

FIGURE 34B

2761 TACATCGAAC TGGATCTCAA CAGCGGTAAG ATCCTTGAGA GTTTTCGCCC CGAAGAACGT
2821 TTTCCAATGA TGAGCACTTT TAAAGTTCTG CTATGTGGCG CGGTATTATC CCGTGTGAC
2881 GCCGGGCAAG AGCAACTCGG TCGCCGCATA CACTATTCTC AGAATGACTT GGTGAGTAC
2941 TCACCAGTCA CAGAAAAGCA TCTTACGGAT GGCATGACAG TAAGAGAATT ATGCAGTGCT
3001 GCCATAACCA TGAGTGATAA CACTGCGGCC AACTTACTTC TGACAACGAT CGGAGGACCG
3061 AAGGAGCTAA CCGCTTTTTT GCACAACATG GGGGATCATG TAACTCGCCT TGATCGTTGG
3121 GAACCGGAGC TGAATGAAGC CATACCAAAC GACGAGCGTG ACACCAGAT GCCTGCAGCA
3181 ATGGCAACAA CGTTGCGCAA ACTATTAAC TACCTCTAGC TTCCCGGCAA
3241 CAATTAATAG ACTGGATGGA GCGGATAAAA GTTGACGAG CACTTCTGCG CTCGGCCCTT
3301 CCGGCTGGCT GGTTTATTGC TGATAAATCT GGAGCCGGTG AGCGTGGGTC TCGCGGTATC
3361 ATTGCAGCAC TGGGGCCAGA TGGTAAGCCC TCCCGTATCG TAGTATCTA CACGACGGGG
3421 AGTCAGGCAA CTATGGATGA ACGAAATAGA CAGATCGCTG AGATAGGTGC CTCAGTGATT
3481 AAGCATTGGT AACTGTGAGA CCAAGTTTAC TCATATATAC TTTAGATTGA TTTAAAACCT
3541 CATTTTTAAAT TTAAGAGGAT CTAGGTGAAG ATCCTTTTTG ATAATCTCAT GACCAAAATC
3601 CCTTAACGTG AGTTTTCGTT CCACTGAGCG TCAGACCCCG TAGAAAAGAT CAAAGGATCT
3661 TCTTGAGATC CTTTTTTTCT GCGGTAATC TGCTGCTTGC AAACAAAAAA ACCACCGCTA
3721 CCAGCGGTGG TTTGTTTGCC GGATCAAGAG CTACCAACTC TTTPTCCGAA GGTAACTGGC
3781 TTCAGCAGAG CGCAGATACC AAATACTGTC CTTCTAGTGT AGCCGTAGTT AGGCCAACAC
3841 TTCAAGAACT CTGTAGCACC GCCTACATAC CTCGCTCTGC TAATCCTGTT ACCAGTGGCT
3901 GCTGCCAGTG GCGATAAGTC GTGCTTACC GGGTTGGACT CAAGACGATA GTTACCGGAT
3961 AAGGCGCAGC GGTCCGGCTG AACGGGGGGT TCGTGACAC AGCCAGCTT GGAGCGAACC
4021 ACCTACACCG AACTGAGATA CCTACAGCGT GAGCTATGAG AAAGCGCCAC GCTTCCCGAA
4081 GGGAGAAAGG CGGACAGGTA TCCGGTAAGC GGCAGGGTCG GAACAGGAGA GCGCACGAGG
4141 GAGCTTCCAG GGGGAAACGC CTGGTATCTT TATAGTCTCG TCGGGTTTCG CCACCTCTGA
4201 CTTGAGCGTC GATTTTTGTG ATGCTCGTCA GGGGGGCGGA GCCTATGGAA AAACGCCAGC
4261 AACGCGGCTT TTTTACGGTT CCTGGCCTTT TGCTGGCCTT TTGCTCAGAT GTTCTTTCTT
4321 CGCTTATCCC CTGATTCTGT GGATAACCGT ATTACCGCCT TTGAGTGAGC TGATACCGCT
4381 CGCCGACGCG GAACGACCGA GCGCAGCGAG TCAGTGAGCG AGGAAGCGGA AGAGCGCCTG
4441 ATGCGGTATT TTCTCCTTAC GCATCTGTGC GGTATTTCAC ACCGCATATA TGGTGCACTC
4501 TCAGTACAAT CTGCTCTGAT GCCGCATAGT TAAGCCAGTA TACACTCCGC TATCGCTACG
4561 TGAAGGGTGC ATGGCTGCGC CCCGACACCC GCCAACACCC GCTGACGCGC CTGACGGGC
4621 TTGTCTGCTC CCGCATCCCG CTTACAGACA AGCTGTGACC GTCTCCGGGA GCTGCATGTG
4681 TCAGAGGTTT TCACCGTCAT CACCGAAACG CGCGAGGCGA CTGCGGTAAA GCTCATCAGC
4741 GTGGTCTGTA AGCGATTAC AGATGTCTGC CTGTTTATCC GCGTCCAGCT CGTTGAGTTT
4801 CTCAGAAGC GTTAATGTCT GGCTTCTGAT AAAGCGGGCC ATGTTAAGGG CGGTTTTTTC
4861 CTGTTTGGTC ACTGATGCCT CCGTGTAAAG GGGATTTCGT TTCATGGGG TAATGATACC
4921 GATGAAACGA GAGAGGATGC TCACGATACG GGTACTGTAT GATGAACATG CCCGTTTACT
4981 GGAACGTTGT GAGGGTAAAC AACTGGCGGT ATGGATGCGG CGGGACCAGA GAAAATCAC
5041 TCAGGGTCAA TGCCAGCGCT TCGTTAATAC AGATGTAGGT GTTCCACAGG GTAGCCAGCA
5101 GCATCTGCG ATGCAGATCC GGAACATAAT GGTGCAGGGC GCTGACTTCC GCGTTTCCAG
5161 ACTTTACGAA ACACGGAAAC CGAAGACCAT TCATGTTGTT GCTCAGGTCG CAGACGTTTT
5221 GCAGCAGCAG TCGCTTCAAG TTGCTGCGG TATCGGTGAT TCATTCTGCT AACCAGTAAG
5281 GCAACCCCGC CAGCCTAGCC GGGTCTCAA CGACAGGAGC ACGATCATGC GCACCCGTGG
5341 CCAGGACCCA ACGCTGCCCG AGATGCGCGC CGTGCGGCTG CTGGAGATGG CGGACCGGAT
5401 GGATATGTTT TGCCAAGGGT TGGTTTGC GC ATTACAGTT CTCCGCAAGA ATTGATTGGC
5461 TCCAATTCTT GGAGTGGTGA ATCCGTTAGC GAGGTGCCGC CGGCTTCCAT TCAGGTCCAG
5521 GTGGCCCGGC TCCATGCACC GCGACGCAAC GCGGGGAGGC AGACAAGGTA TAGGGCGGCG
5581 CCTACAATCC ATGCCAACCC GTTCCATGTG CTCGCGAGG CGGCATAAAT CGCCGTGACG
5641 ATCAGCGGTC CAGTGATCGA AGTTAGGCTG GTAAGAGCCG CGAGCGATCC TTGAAGCTGT
5701 CCCTGATGGT CGTCACTTAC CTGCCTGGAC AGCATGGCCT GCAACGCGGG CATCCCGATG
5761 CCGCCGGAAG CGAGAAGAA CATAATGGGG AAGGCCATCC AGCCTCGCGT CGGAAACGCC
5821 AGCAAGACGT AGCCGAGCGC GTCCGCCGCC ATGCCGCGA TAATGGCCTG CTTCGCGCG
5881 AAACGTTTGG TGGCCGGACC AGTGACGAAG GCTTGAGCGA GGGCGTGCAA GATTCCGAAT
5941 ACCGCAAGCG ACAGGCCGAT CATCGTCGCG CTCCAGCGAA AGCGGTCCTC GCCGAAATG
6001 ACCGAGAGCG CTGCCGCGAC CTGTCCTACG AGTTGCATGA TAAAGAAGAC AGTCATAAGT
6061 GCGCGGACGA TAGTCATGCC CCGCGCCAC CGGAAGGAGC TGAAGGCTTC GAAGGCTCTC
6121 AAGGGCATCG GTCGATCGAC GCTCTCCCTT ATGCGACTCC TGCAATTAGGA AGCAGCCAG
6181 TAGTAGGTTG AGGCCGTTGA GCACCGCCGC CGCAAGGAAT GTGTCATGCA AGGAGATGGC-

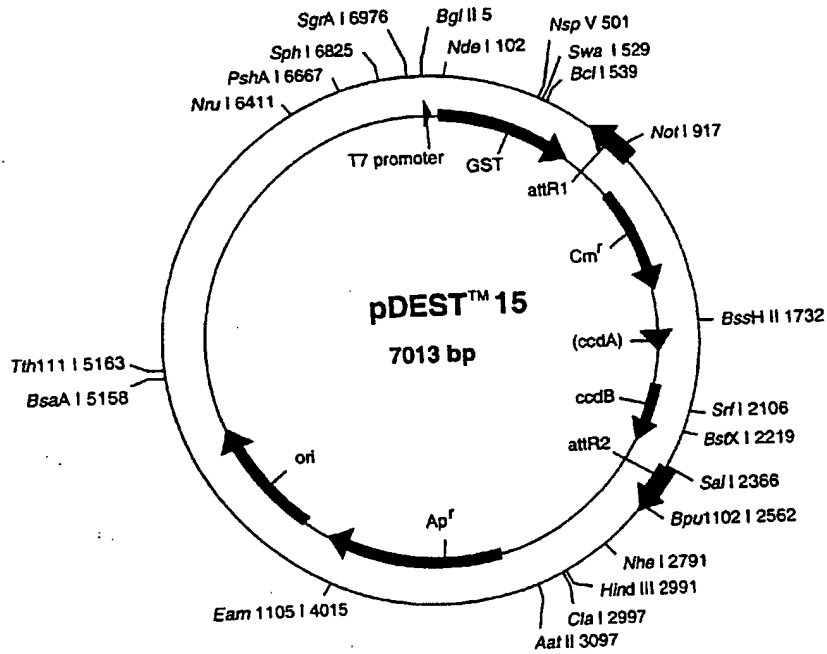
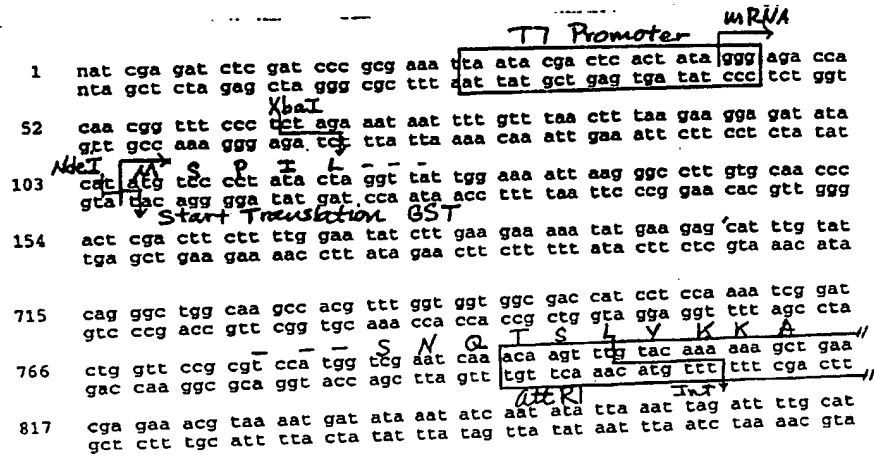
FIGURE 34C

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6241 GCCCAACAGT CCCCCGGCCA CGGGGCCTGC CACCATACCC ACGCCGAAAC AAGCGTCAT
6301 GAGCCCGAAG TGGCGAGCCC GATCTTCCCC ATCGGTGATG TCGGCGATAT AGGCGCCAGC
6361 AACC GCACCT GTGGCGCCGG TGATGCCGGC CACGATGCGT CCGGCGTAGA GGATCGAGAT
6421 CT

FIGURE 34D

Figure 35A: pDEST15 Glutathione-S-transferase Fusion in *E. coli*, T7 Promoter



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pDEST15 7013 bp

<u>Location (Base Nos.)</u>		<u>Gene Encoded</u>
108..776		GST
916..792		attR1
1025..1537		CmR
1804..1888		inactivated ccdA
2026..2331		ccdB
2372..2496		attR2
3233..4093		ampR
1	ATCGAGATCT CGATCCCGCG AAATTAATAC GACTCACTAT AGGGAGACCA CAACGGTTTC	
61	CCTCTAGAAA TAATTTTGTT TAACTTTAAG AAGGAGATAT ACATATGTCC CCTATACTAG	
121	GTTATTGGAA AATTAAAGGC CTTGTGCAAC CCACCTGACT TCTTTTGGAA TATCTTGAAG	
181	AAAAATATGA AGAGCATTTG TATGAGCGCG ATGAAGGTGA TAAATGGCGA AACAAAAAGT	
241	TTGAATTGGG TTGGAGTTT CCCAATCTTC CTTATTATAT TGATGGTGAT GTTAAATTAA	
301	CACAGTCTAT GGCCATCATA CGTTATATAG CTGACAAGCA CAACATGTTG GGTGGTTGTC	
361	CAAAAGAGCG TGCAGAGATT TCAATGCTTG AAGGAGCGGT TTTGGATATT AGATACGGTG	
421	TTTCGAGAAT TGCATATAGT AAAGACTTTG AAACCTCTCA AGTTGATTTT CTTAGCAAGC	
481	TACCTGAAAT GCTGAAAATG TTCGAAGATC GTTTATGTCA TAAACATAT TTAATGGTG	
541	ATCATGTAAC CCATCCTGAC TTCATGTTGT ATGACGCTCT TGATGTGTT TTATACATGG	
601	ACCCAATGTG CCTGGATGCG TTCCCAAAAT TAGTTTGT TTAAAAACGT ATTGAAGCTA	
661	TCCCACAAAT TGATAAGTAC TTGAAATCCA GCAAGTATAT AGCATGGCCT TTGCAGGGCT	
721	GGCAAGCCAC GTTTGGTGGT GGCGACCATC CTCCAAATC GGATCTGGTT CCGCGTCCAT	
781	GGTCGAATCA AACAAGTTTG TACAAAAAG CTGAACGAGA AACGTAAAT GATATAAATA	
841	TCAATATATT AAATTAGATT TTGCATAAAA AACAGACTAC ATAATACTGT AAAACACAAC	
901	ATATCCAGTC ACTATGGCGG CCGCATTAGG CACCCAGGC TTTACACTTT ATGCTTCCGG	
961	CTCGTATAAT GTGTGGATT TTGAGTTAGGA TCCGTGAGGA TTTTCAGGAG CTAAGGAAGC	
1021	TAAAATGGAG AAAAAATCA CTGGATATAC CACCGTTGAT ATATCCCAAT GGCATCGTAA	
1081	AGAACATTTT GAGGCATTTC AGTCAGTTGC TCAATGTACC TATAACCAGA CCGTTCAGTC	
1141	GGATATTACG GCCTTTTAA AGACCGTAAA GAAAAATAAG CACAAGTTT ATCCGGCCTT	
1201	TATTCACATT CTGCCCCGCC TGATGAATGC TCATCCGGAA TTCCGTATGG CAATGAAAGA	
1261	CGGTGAGCTG GTGATATGGG ATAGTGTTC CCCITGTTAC ACCGTTTTC ATGAGCAAAC	
1321	TGAAACGTTT TCATCGCTCT GGAGTGAATA CCACGACGAT TTCCGGCAGT TTCTACACAT	
1381	ATATTGCGAA GATGTGGCGT GTTACGGTGA AAACCTGGCC TATTTCCTA AAGGGTTTAT	
1441	TGAGAATATG TTTTTCGTCT CAGCCAATCC CTGGGTGAGT TTCACCAGTT TTGATTTAAA	
1501	CGTGGCCAAT ATGGACAAC TCTTCGCCCC CGTTTTCACC ATGGGCAAAAT ATTATACGCA	
1561	AGGCGACAAG GTGCTGATGC CGCTGGCGAT TCAGGTTTAT CATGCCGTCT GTGATGGCTT	
1621	CCATGTCGGC AGAATGCTTA ATGAATTACA ACAGTACTGC GATGAGTGGC AGGGCGGGGC	
1681	GTAATCTAGA GGATCCGGCT TACTAAAAGC CAGATAACAG TATGCGTATT TGCGCGCTGA	
1741	TTTTTGGCGT ATAAGAATAT ATACTGATAT GTATACCGA AGTATGTCAA AAAGAGGTGT	
1801	GCTATGAAGC AGCGTATTAC AGTGACAGTT GACAGCGACA GCTATCAGTT GCTCAAGGCA	
1861	TATATGATGT CAATATCTCC GGTCTGGTAA GCACAACCAT GCAGAATGAA GCCCGTCGTC	
1921	TGCGTGCCGA ACGCTGGAAA GCGGAAAATC AGGAAGGGAT GGCTGAGGTC GCCCGGTTTA	
1981	TTGAAATGAA CGGCTCTTTT GCTGACGAGA ACAGGGACTG GTGAAATGCA GTTTAAGGTT	
2041	TACACCTATA AAAGAGAGAG CCGTTATCGT CTGTTTGTGG ATGTACAGAG TGATATTATT	
2101	GACACGCCCG GCGGACGGAT GGTGATCCCC CTGGCCAGTG CACGTCTGCT GTCAGATAAA	
2161	GTCTCCCGTG AACTTTACCC GGTGGTGCAT ATCGGGGATG AAAGCTGGCG CATGATGACC	
2221	ACCGATATGG CCAGTGTGCC GGTCTCCGTT ATCGGGGAAG AAGTGGCTGA TCTCAGCCAC	
2281	CGCGAAAATG ACATCAAAAA CGCCATTAACT CTGATGTTCT GGGGAATATA AATGTCAGGC	
2341	TCCCTTATAC ACAGCCAGTC TGCAGGTCGA CCATAGTGAC TGGATATGTT GTGTTTACA	
2401	GTATTATGTA GTCTGTTTTT TATGCAAAAT CTAATTTAAT ATATTGATAT TTATATCATT	
2461	TIACGTTTCT CGTTCAGCTT TCTTGACAA AGTGGTTTGA TTCCGACCCG GATCCGGCTG	
2521	CTAACAAAGC CCGAAAGGAA GCTGAGTTGG CTGCTGCCAC CGCTGAGCAA TAACTAGCAT	
2581	AACCCCTTGG GGCCTCTAAA CGGCTCTTGA GGGGTTTTTT GCTGAAAGGA GGAACATAT	
2641	CCGATATACC ACAGGACGGG TGTGGTCGCC ATGATCGCGT AGTCGATAGT GGCTCAAAGT	

Figure 35B

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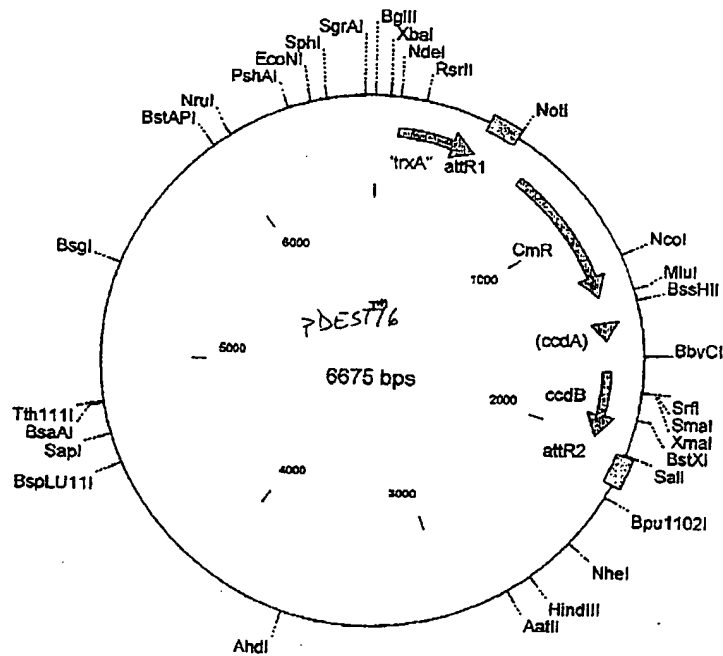
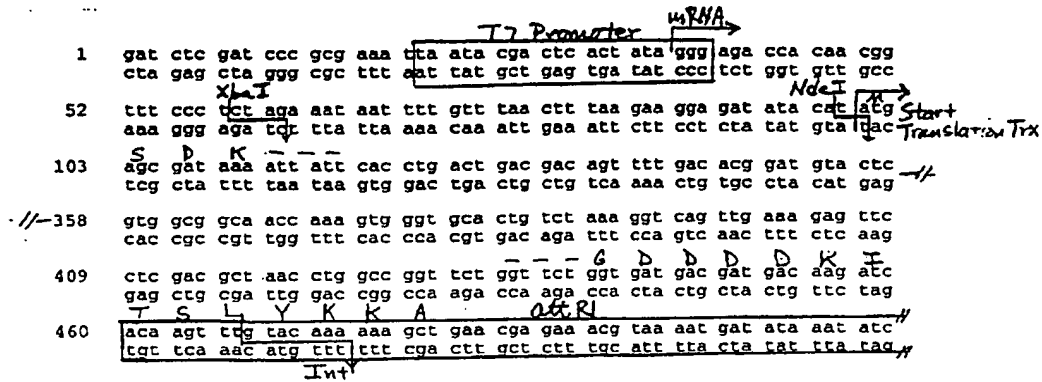
2701 AGCGAAGCGA GCAGGACTGG GCGGCGGCCA AAGCGGTGCG ACAGTGCTCC GAGAACGGGT
 2761 GCGCATAGAA ATTGCATCAA CGCATATAGC GCTAGCAGCA CGCCATAGTG ACTGGCGATG
 2821 CTGTCCGAAT GGACGATATC CCGCAAGAGG CCCGGCAGTA CCGGCATAAC CAAGCCTATG
 2881 CCTACAGCAT CCAGGGTGAC GGTGCCGAGG ATGACGATGA GCGCAITGTT AGATTTTCATA
 2941 CACGGTGCCT GACTGCGTTA GCAATTTAAC TGTGATAAAC TACCGCATT AAGCTTATCG
 3001 ATGATAAGCT GTCAAACATG AGAATTCCTG AAGACGAAAG GGCCTCGTGA TACGCCCTATT
 3061 TTTATAGGTT AATGTCATGA TAATAATGGT TTCTTAGACG TCAGGTGGCA CTTTTCGGGG
 3121 AAATGTGCGC GGAACCCCTA TTTGTTTATT TTTCTAAATA CATTCAAATA TGTATCCGCT
 3181 CATGAGACAA TAACCCCTGAT AAATGCTTCA ATAATATTGA AAAAGGAAGA GTATGAGTAT
 3241 TCAACATTTT CGTGTGCGCC TTATTCCTTT TTTTGGCGCA TTTTGCCTTC CTGTTTTTGC
 3301 TCACCCAGAA ACGCTGGTGA AAGTAAAAGA TGCTGAAGAT CAGTTGGGTG CACGAGTGGG
 3361 TTACATCGAA CTGGATCTCA ACAGCGGTAA GATCCTTGAG AGTITTCGCC CCGAAGAACG
 3421 TTTTCCCAATG ATGAGCACTT TTAAGATTCT GCTATGTGGC GCGGTATTAT CCCGTGTTGA
 3481 CGCCGGGCAA GAGCAACTCG GTCGCCGAT ACACCTATTCT CAGAATGACT TGGTTGAGTA
 3541 CTCACCAATC ACAGAAAAGC ATCTTACGGA TGGCATGACA GTAAGAGAAT TATGCAATGC
 3601 TGCCATAACC ATGAGTGATA ACATGCGGCG CAACCTACTT CTGACAAACA TCGGAGGACC
 3661 GAAGGAGCTA ACCGCTTTTT TGCACAACAT GGGGGATCAT GTAACTCGCC TTGATCGTTG
 3721 GGAACCGGAG CTGAATGAAG CCATACCAA CGACGAGCGT GACACCAAGA TGCTGACAGC
 3781 AATGGCAACA ACGTTGCGCA AACTATTAAC TGGCGAACA CTTACTCTAG CTTCCCGGCA
 3841 ACAATTAATA GACTGGATGG AGCGGGATAA AGTTGCAGGA CCACTTCTGC GCTCGGCCCT
 3901 TCCGGCTGGC TGGTTTATTG CTGATAAATC TGGAGCCGGT GAGCGTGGGT CTCGCGGTAT
 3961 CATTGCAGCA CTGGGGCCAG ATGGTAAGCC CTCGCGTATC GTAGTTATCT ACACGACGGG
 4021 GAGTCAGGCA ACTATGGATG AACGAAATAG ACAGATCGCT GAGATAGGTG CCTCACTGAT
 4081 TAAGCATTGG TAACTGTGAG ACCAAGTTTA CTCATATATA CTTTAGATTG ATTTAAACTG
 4141 TCATTTTAA TTTAAAAGGA TCTAGGTGAA GATCCTTTTT GATAATCTCA TGACCAAAAT
 4201 CCCTTAAGCT GAGTTTTCGT TCCACTGAGC GTCAGACCCC GTAGAAAAGA TCAAAGGATC
 4261 TTCTTGAGAT CCTTTTTTTC TGCGCGTAAT CTGCTGCTTG CAAACAAAAA AACCACCGCT
 4321 ACCAGCGGTG GTTTGTTTGC CGGATCAAGA GCTACCAACT CTTTTTCCGA AGGTAACCTG
 4381 CTTCAGCAGA GCGCAGATAC CAAATACTGT CCTTCTAGTG TAGCCGTAGT TAGGCCACCA
 4441 CTTCAAGAAC TCTGTAGCAC CGCCTACATA CCTCGCTCTG CTAATCCTGT TACCAGTGGC
 4501 TGCTGCCAGT GCGGATAAGT CGTGTCTTAC CGGTTGGAG TCAAGACGAT AGTTACCGGA
 4561 TAAGGCGCAG CGGTCCGGCT GAACGGGGGG TTCGTGCACA CAGCCAGCTG TGGAGCGAAC
 4621 GACCTACACC GAAGTGAAGT ACCTACAGCG TGAGCTATGA GAAAGCGCCA CGCTTCCCGA
 4681 AGGGAGAAAG GCGGACAGGT ATCCGGTAAG CGGCGGGTGC GGAACAGGAG AGCGCACGAG
 4741 GGAGCTTCCA GGGGGAACG CCTGGTATCT TTATAGTCTC GTCCGGTTTC GCCACCTCTG
 4801 ACTTGAGCGT CGATTTTGTG GATGCTCGTC AGGGGGGCGG AGCCTATGGA AAAACGCCAG
 4861 CAACGCGGCC TTTTACGGT TCCTGGCCTT TTGCTGGCCT TTGTCTACA TGTTCCTTCC
 4921 TGCGTTATCC CCTGATTCTG TGGATAACCG TATTACGCC TTTGAGTGAG CTGATACCGC
 4981 TCGCCGCAGC CGAACGACCG AGCGCAGCGA GTCAGTGAGC GAGGAAGCGG AAGAGCGCCT
 5041 GATGCGGTAT TTTCTCCTTA CGCATCTGTG CGGTATTTC CACCGCATAT ATGGTGCACT
 5101 CTCAGTACAA TCTGCTCTGA TGCCGCATAG TTAAGCCAGT ATACACTCCG CTATCGCTAC
 5161 GTGACTGGGT CATGGCTGCG CCCCAGACAC CGCCAACACC CGCTGACGCG CCCTGACGGG
 5221 CTGTCTGCT CCCGGCATCC GCTTACAGAC AAGCTGTGAC CGTCTCCGG AGCTGCATGT
 5281 GTCAGAGGTT TTCACCGTCA TCACCGAAAC GCGCGAGGCA GCTGCGGTAA AGCTCATCAG
 5341 CGTGGTCTG AAGCGATTCA CAGATGTCTG CCTGTTTATC CGCGTCCAGC TCGTTGAGTT
 5401 TCTCCAGAAG CGTTAATGTC TGGCTTCTGA TAAAGCGGGC CATGTTAAGG GCGGTTTTTT
 5461 CCTGTTTGGT CACTGATGCC TCCGTGTAAG GGGGATTTCT GTTCATGGGG GTAATGATAC
 5521 CGATGAAACG AGAGAGGATG CTCACGATAC GGGTTACTGA TGATGAACAT GCCCGGTTAC
 5581 TGGAACGTTG TGAGGGTAAA CAACTGGCGG TATGGATGCG GCGGGACCAG AGAAAAATCA
 5641 CTCAGGGTCA ATGCCAGCGC TTCGTTAATA CAGATGTAGG TGTTCCACAG GGTAGCCAGC
 5701 AGCATCCTGC GATGCAGATC CGGAACATAA TGGTGACGGG CGCTGACTTC CGCGTTTCCA
 5761 GACTTTACGA AACACGGAAA CCGAAGACCA TTATGTTTGT TGCTCAGGTC GCAGACGTTT
 5821 TGCAGCAGCA GTCGCTTCAC GTTCGCTCGC GTATCGGTGA TTCATTCTGC TAACCAAGTAA
 5881 GGCAACCCCG CCAGCCTAGC CGGGTCTTCA ACGACAGGAG CACGATCATG CGCACCCGTG
 5941 GCCAGGACCC AACGCTGCCC GAGATGCGCC GCGTGGGCT GCTGGAGATG GCGGACGCGA
 6001 TGGATATGTT CTGCCAAGGG TTGGTTTGGC CATTACAGT TCTCCGCAAG AATTGATTGG
 6061 CTCCAATTCT TGGAGTGGTG AATCCGTTAG CGAGGTGCCG CCGGCTTCCA TTCAGGTCCA
 6121 GGTGGCCCGG CTCCATGCAC CGCGACGCAA CGCGGGGAGG CAGACAAGGT ATAGGGCGGC-

FIGURE 35C

6181 GCCTACAATC CATGCCAACC CGTTCCATGT GCTCGCCGAG GCGGCATAAA TCGCCGTGAC
6241 GATCAGCGGT CCAGTGATCG AAGTTAGGCT GGTAAGAGCC GCGAGCGATC CTTGAAGCTG
6301 TCCCTGATGG TCGTCATCTA CCTGCCTGGA CAGCATGGCC TGC AACGCGG GCATCCCGAT
6361 GCCGCCGGA GCGAGAAGAA TCATAATGGG GAAGGCCATC CAGCCTCGCG TCGCGAACGC
6421 CAGCAAGACG TAGCCCAGCG CGTCGGCCGC CATGCCGGCG ATAATGGCCT GCTTCTCGCC
6481 GAAACGTTTG GTGGCGGGAC CAGTGACGAA GGCTTGAGCG AGGGCGTGCA AGATTCCGAA
6541 TACCGCAAGC GACAGGCCGA TCATCGTCGC GCTCCAGCGA AAGCGGTCCT CGCCGAAAA
6601 GACCCAGAGC GCTGCCGGCA CCGTCTCTAC GAGTTGCATG ATAAAGAAGA CAGTCATAAG
6661 TGCGGCGACG ATAGTCATGC CCGCGCCCA CCGGAAGGAG CTGACTGGGT TGAAGGCTCT
6721 CAAGGGCATC GGTGATCGA CGCTCTCCCT TATGCCACTC CTGCATTAGG AAGCAGCCCA
6781 GTAGTAGGTT GAGGCCGTTG AGCACCAGCG CCGCAAGGAA TGGTGCATGC AAGGAGATGG
6841 CGCCCAACAG TCCCCCGGCC ACGGGGCGTG CCACCATACC CACGCCGAAA CAAGCGCTCA
6901 TGAGCCCGAA GTGGCGAGCC CGATCTTCCC CATCGGTGAT GTCGGCGATA TAGGCGCCAG
6961 CAACCGCACC TGTGGCGCCG GTGATGCCG CCACGATGCG TCCGGCGTAG AGG

FIGURE 351)

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Figure 36A: γ DEST16Thioredoxin N-Fusion Protein
in E. coli with T7 Promoter

pDEST16 6675 bp

<u>Location (Base Nos.)</u>		<u>Gene Encoded</u>
104..457		trxA
585..461		attR1
694..1353		CmR
1473..1557		inactivated ccdA
1695..2000		ccdB
2041..2165		attR2

1	AGATCTCGAT	CCCGCGAAAT	TAATACGACT	CACTATAGGG	AGACCACAAC	GGTTTCCCTC
61	TAGAAATAAT	TTTGTTTAAC	TTTAAGAAGG	AGATATACAT	ATGAGCGATA	AAATTATTC
121	CCTGACTGAC	GACAGTTTGG	ACACGGATGT	ACTCAAAGCG	GACGGGGCGA	TCCTCGTCGA
181	TTTCTGGGCA	GAGTGGTGCG	GTCCGTGCAA	AATGATCGCC	CCGATTCTGG	ATGAAATCGC
241	TGACGAATAT	CAGGGCAAAAC	TGACCGTTGC	AAAACCTGAAC	ATCGATCAAA	ACCCTGGCAC
301	TGCGCCGAAA	TATGGCATCC	GTGGTATCCC	GACTCTGCTG	CTGTTCAAAA	ACGGTGAAGT
361	GGCGGCAACC	AAAGTGGGTG	CACTGTCTAA	AGGTGAGTTG	AAAGAGTTCC	TCGACGCTAA
421	CCTGGCCGGT	TCTGGTTCCT	GTGATGACGA	TGACAAGATC	ACAAGTTTGT	ACAAAAAAGC
481	TGAACGAGAA	ACGTAAAATG	ATATAAATAT	CAATATATTA	AATTAGATTT	TGCATAAAAA
541	ACAGACTACA	TAATACTGTA	AAACACAACA	TATCCAGTCA	CTATGGCGGC	CGCATTAGGC
601	ACCCGAGGCT	TTACACTTTA	TGCTTCCGGC	TCGTATAATG	TGTGGATTTT	GAGTTAGGAT
661	CCGGCGAGAT	TTTCAGGAGC	TAAGGAAGCT	AAAATGGAGA	AAAAAATCAC	TGGATATACC
721	ACCGTTGATA	TATCCCAATG	GCATCGTAAA	GAACATTTTG	AGGCATTTC	GAGTTAGGCT
781	CAATGTACCT	ATAACAGAGC	CGTTCAGCTG	GATATTACGG	CCTTTTTAAA	GACCGTAAAG
841	AAAAATAAGC	ACAAGTTTAA	TCCGGCCTTT	ATTCACATTC	TGCCCCGCTT	GATGAATGCT
901	CATCCGGAAT	TCCGTATGGC	AATGAAAGAC	GGTGAGCTGG	TGATATGGGA	TAGTGTTCAC
961	CCTTGTAC	CCGTTTTCCA	TGAGCAAAC	GAAACGTTTT	CATCGCTCTG	GAGTGAATAC
1021	CACGACGATT	TCCGGCAGTT	TCTACACATA	TATTCGCAAG	ATGTGGCGTG	TTACGGTGAA
1081	AACCTGGCCT	ATTTCCCTAA	AGGGTTTATT	GAGAATATGT	TTTTCGTCTC	AGCCAATCCC
1141	TGGGTGAGTT	TCACCACTTT	TGATTTAAAC	GTGGCCAATA	TGGACAACCT	CTTCGCCCCC
1201	GTITTCACCA	TGGGCAAATA	TTATACGCAA	GGCGACAAGG	TGCTGATGCC	GCTGGCGATT
1261	CAGGTTATC	ATGCCGCTCT	TGATGGCTTC	CATGTCGGCA	GAATGCTTAA	TGAATTACAA
1321	CAGTACTGCG	ATGAGTGCCA	GGCGGGGGCG	TAAACGCGTG	GATCCGGCTT	ACTAAAAGCC
1381	AGATAACAGT	ATGCGTATTT	GCGCGCTGAT	TTTTGCGGTA	TAAGAAATATA	TACTGATATG
1441	TATACCCGAA	GTATGTCAAA	AAGAGGTGTG	CTATGAAGCA	GCGTATTACA	GTGACAGTTG
1501	ACAGCGACAG	CTATCAGTTG	CTCAAGGCAT	ATATGATGTC	AATATCTCCG	GTCGTGTAAG
1561	CACAACCATG	CAGAATGAAG	CCCCTGCTCT	GCGTGGCGAA	CGCTGGAAAG	CGGAAATCA
1621	GGAAGGGATG	GCTGAGGTCT	CCCGGTTTAT	TGAAATGAAC	GGCTCTTTTG	CTGACGAGAA
1681	CAGGGACTGG	TGAAATGCAG	TTTAAGGTTT	ACACCTATAA	AAGAGAGAGC	CGTTATCGTC
1741	TGTTTGTGGA	TGTACAGAGT	GATATTATTG	ACACGCCCGG	GCGACGGATG	GTGATCCCCC
1801	TGGCCAGTGC	ACGTCTGCTG	TCAGATAAAG	TCTCCCGTGA	ACTTTACCCG	GTGGTGCATA
1861	TCGGGGATGA	AAGCTGGCGC	ATGATGACCA	CCGATATGGC	CAGTGTGCCG	GTCTCCGTTA
1921	TCGGGGGAAGA	AGTGGCTGAT	CTCAGCCACC	GCGAAAATGA	CATCAAAAAC	GCCATTAAAC
1981	TGATGTTCTG	GGGAATATAA	ATGTCAGGCT	CCCTTATACA	CAGCCAGTCT	GCAGGTCGAC
2041	CATAGTGACT	GGATATGTTG	TGTTTTACAG	TATTATGTAG	TCTGTTTTTT	ATGCAAAATC
2101	TAATTTAATA	TATTGATATT	TATATCAITTT	TACGTTTCTC	GTTTCAGCTTT	CTTGATACAA
2161	GTGGTGATGA	TCCGGCTGCT	AACAAAGCCC	GAAAGGAAGC	TGAGTTGGCT	GCTGCCACCG
2221	CTGAGCAATA	ACTAGCATAA	CCCCTTGGGG	CCTCTAAACG	GGTCTTGAGG	GGTTTTTTTG
2281	TGAAAGGAGG	AACTATATCC	GGATATCCAC	AGGACGGGTG	TGGTCGCCAT	GATCGCGTAG
2341	TCGATAGTGG	CTCCAAGTAG	CGAAGCGAGC	AGGACTGGGC	GGCGGCCAAA	GCGGTCCGAC
2401	AGTGCTCCGA	GAACGGGTGC	GCATAGAAAT	TGCATCAACG	CATATAGCGC	TAGCAGCACG
2461	CCATAGTGAC	TGGCGATGCT	GTCCGAATGG	ACGATATCCC	GCAAGAGGCC	CGGCAGTACC
2521	GGCATAACCA	AGCCTATGCC	TACAGCATCC	AGGGTGACGG	TGCCGAGGAT	GACGATGAGC
2581	GCATTGTTAG	ATTTCAATACA	CGGTGCCTGA	CTGCGTTAGC	AATTTAACTG	TGATAAACTA
2641	CCGCATTAAA	GCTTATCGAT	GATAAGCTGT	CAAAACATGAG	AATTCITGAA	GACGAAAGGG
2701	CCTCGTGATA	CGCCTATTTT	TATAGGTTAA	TGTCATGATA	ATAATGGTTT	CTTAGACGTC
2761	AGGTGGCACT	TTTCGGGGAA	ATGTGCGCGG	AACCCCTATT	TGTTTATTTT	TCTAAATACA

Figure 36B

2821 TTCAAATATG TATCCGCTCA TGAGACAATA ACCCTGATAA ATGCTTCAAT AATATTGAAA
 2881 AAGGAAGAGT ATGAGTATTC AACATTTCCTG TGTGCGCCCTT ATTCCCTTTT TTGCGGCATT
 2941 TTGCTTCTCT GTTTTGTGCTC ACCCAGAAAC GCTGGTGAAA GTAAAGATG CTGAAGATCA
 3001 GTTGGGTGCA CGAGTGGGTT ACATCGAACT GGATCTCAAC AGCGGTAAGA TCCTTGAGAG
 3061 TTTTCGCCCC GAAGAACGTT TTCCAATGAT GAGCACTTTT AAAGTTCTGC TATGTGGCGC
 3121 GGTATTATCC CGTGTGACG CCGGGCAAGA GCAACTCGGT CGCCGCATAC ACTATTCTCA
 3181 GAATGACTTG GTTGAGTACT CACCAGTCAC AGAAAAGCAT CTTACGGATG GCATGACAGT
 3241 AAGAGAATTA TGCAGTGTG CCATAACCAT GAGTGATAAC ACTGCGGCCA ACTTACTTCT
 3301 GACAACGATC GGAGGACCGA AGGAGCTAAC CGCTTTTTTG CACAACATGG GGGATCATGT
 3361 AACTCGCCTT GATCGTTGGG AACCGAGCT GAATGAAGCC ATACCAAACG ACGAGCGTGA
 3421 CACCACGATG CCTGCAGCAA TGGCAACAAC GTTGCGCAAA CTATTAACTG GCGAACTACT
 3481 TACTCTAGCT TCCCGGCAAC AATTAATAGA CTGGATGGAG GCGGATAAAG TTGCAGGACC
 3541 ACTTCTGCGC TCGGCCCTTC CGGCTGGCTG GTTTATTGCT GATAAATCTG GAGCCGGTGA
 3601 GCGTGGGTCT CGCGGTATCA TTGCAGCACT GGGGCCAGAT GGTAAGCCCT CCCGTATCGT
 3661 AGTTATCTAC ACGACGGGA GTCAGGCAAC TATGGATGAA CGAAATAGAC AGATGCTGTA
 3721 GATAGGTGCC TCACTGATTA AGCATTGGTA ACTGTCAGAC CAAGTTTACT CATATATACT
 3781 TTAGATTGAT TTAACACTTC ATTTTAAATT TAAAGGATC TAGGTGAAGA TCCTTTTGA
 3841 TAATCTCATG ACCAAAATCC CTTAACGTGA GTTTTCGTTT CACTGAGCGT CAGACCCCGT
 3901 AGAAAAGATC AAAGGATCTT CTTGAGATCC TTTTTCGTC CGCGTAATCT GCTGCTTGCA
 3961 AACAAAAAAA CCACCGCTAC CAGCGGTGGT TTGTTGCGG GATCAAGAGC TACCAACTCT
 4021 TTTTCCGAAG GTAACCTGGT TCAGCAGAGC GCAGATACCA AATACTGTCC TTCTAGTGTA
 4081 GCCGTAGTTA GGCCACCACT TCAAGAACTC TGTAGCACCG CCTACATACC TCGCTCTGCT
 4141 AATCCTGTTA CCAGTGGCTG CTGCCAGTGG CGATAAGTCG TGTCTTACCG GGTGGACTC
 4201 AAGACGATAG TTACCGGATA AGGCGCAGCG GTCCGGCTGA ACGGGGGGT CGTGACACA
 4261 GCCAGCTTG GAGCGAACGA CCTACACCGA ACTGAGATAC CTACAGCGTG AGCTATGAGA
 4321 AAGCGCCACG CTTCCCGAAG GGAGAAAGGC GGACAGGTAT CCGGTAAGCG GCAGGGTCGG
 4381 AACAGGAGAG CGCACGAGGG AGCTTCCAGG GGGAAACGCC TGGTATCTTT ATAGTCCTGT
 4441 CGGGTTTCGC CACCTCTGAC TTGAGCGTCTG ATTTTGTGTA TGCTCGTCAG GGGGGCGGAG
 4501 CCTATGGAAG AACCGCAGCA ACGCGGCTTT TTTACGGTTC CTGGCCTTTT GCTGCGCTTT
 4561 TGCTCACATG TTTCTTCTG CTGTATCCCC TGATTCTGTG GATAACCGTA TTACCGCTTT
 4621 TGAGTGAGCT GATACCGCTC GCCCGAGCCG AACGACCGAG CGCAGCGAGT CAGTGAGCGA
 4681 GGAAGCGGAA GAGCGCCTGA TCGGTATTTT TCTCCTTACG CATCTGTGCG GTATTTCACA
 4741 CCGCATATAT GGTGCACTCT CAGTACAATC TGCTCTGATG CCGCATAGTT AAGCCAGTAT
 4801 AACTCCGCT ATCGCTACGT GACTGGGTCA TGGCTGCGCC CCGACACCCG CCAACACCCG
 4861 CTGACCGGCC CTGACGGGCT TGTCTGCTCC CGGCATCCGC TTACAGACAA GCTGTGACCG
 4921 TCTCCGGGAG CTGCATGTGT CAGAGGTTTT CACCGTCATC ACCGAAACGC GCGAGGCGAGC
 4981 TGCGGTAAAG CTCATCAGCG TGGTCGTGAA GCGATTACCA GATGTCTGCC TGTTCATCCG
 5041 CGTCCAGCTC GTTGAGTTTC TCCAGAAGCG TTAATGTCTG GCTTCTGATA AAGCGGGCCA
 5101 TGTTAAGGCG GGTTTTTTTC TGTGTTGTTA CTGATGCCTC CGTGTAAAGG GGAATTTCTGT
 5161 TCATGGGGGT AATGATACCG ATGAAACGAG AGAGGATGCT CACGATACCG GTTACTGATG
 5221 ATGAACATGC CCGGTTACTG GAACGTTGTG AGGGTAAACA ACTGGCGGTA TGGATGCGGC
 5281 GGGACACAGG AAAAATCACT CAGGGTCAAT GCCAGCGCTT CGTTAATACA GATGTAGGTG
 5341 TTCCACAGGG TAGCCAGCAG CATCCTGCGA TGCAGATCCG GAACATAATG GTGCAGGGCG
 5401 CTGACTTCCG CGTTTCCAGA CTTTACGAAA CACGGAACCC GAAGACCATT CATGTTGTGT
 5461 CTCAGGTCCG AGACGTTTTG CAGCAGCAGT CGCTTCACGT TCGCTCGCGT ATCGGTGATT
 5521 CATTCGTGTA ACCAGTAAGG CAACCCCGCC AGCCTAGCCG GGTCTCTAAC GACAGGAGCA
 5581 CGATCATGCG CACCGTGGC CAGGACCCAA CGCTGCCCGA GATGCGCCGC GTGCGGCTGC
 5641 TGGAGATGGC GGACGCGATG GATATGTTCT GCCAAGGGTT GGTTTGCGCA TTCACAGTTT
 5701 TCCGCAAGAA TTGATTGGCT CCAATTCTTG GAGTGGTGAA TCCGTTAGCG AGGTGCCGCC
 5761 GGCTTCCATT CAGGTGAGG TGGCCCGGCT CCATGCACCG CGACGCAACG CGGGGAGGCA
 5821 GACAAGGTAT AGGGCGGCGC CTACAATCCA TGCCAACCCG TTCCATGTGC TCGCCGAGCG
 5881 GGCATAAATC GCCGTGACGA TCAGCGGTCC AGTGAATGAA GTTAGGCTGG TAAGAGCCGC
 5941 GAGCGATCCT TGAAGCTGTC CCTGATGGTC GTCATCTACC TGCCCTGGCA GCATGGCCTG
 6001 CAACGCGGCG ATCCCGATGC CGCCGGAAGC GAGAAGAATC ATAATGGGGA AGGCCATCCA
 6061 GCCTCGCGTC GCGAACGCCA GCAAGACGTA GCCCAGCGCG TCGGCCGCCA TGCCCGCGAT
 6121 AATGGCCTGC TTCTGCGCGA AACGTTTGGT GCGGGGACCA GTGACGAAGG CTTGAGCGAG
 6181 GCGGTGCAAG ATTCCGAATA CCGCAAGCGA CAGGCCGATC ATCGTCGCGC TCCAGCGAAA
 6241 GCGGTCTCTG CCGAAAATGA CCCAGAGCGC TGCCGCGACC TGTCTACGA GTTGATGAT-

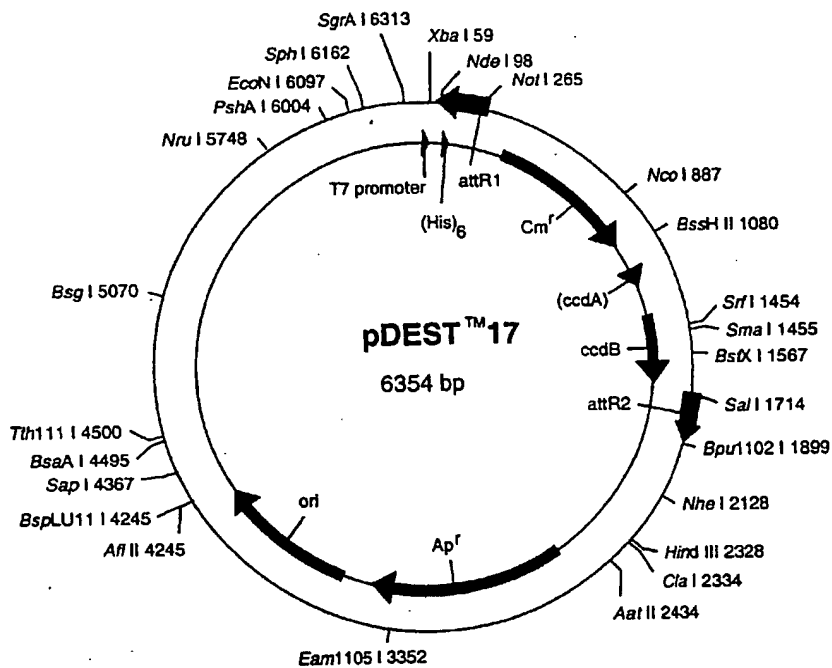
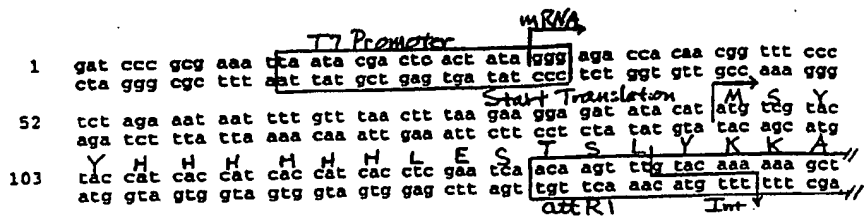
FIGURE 36C

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6301 AAAGAAGACA GTCATAAGTG CGGCGACGAT AGTCATGCC CCGCGCCACC GGAAGGAGCT
6361 GACTGGGTTG AAGGCTCTCA AGGGCATCGG TCGATCGACG CTCTCCCTTA TGC GACTCCT
6421 GCATTAGGAA GCAGCCCA GTAGGTTGA GGCCGTTGAG CACCGCCGCC GCAAGGAATG
6481 GTGCATGCAA GGAGATGGCG CCCAACAGTC CCGCGCCAC GGGGCTGCC ACCATACCCA
6541 CGCCGAAACA AGCGCTCATG AGCCCGAAGT GGCGAGCCCG ATCTTCCCA TCGGTGATGT
6601 CGGCGATATA GCGCCAGCA ACCGCACCTG TGGCGCCGGT GATGCCGCC ACGATGCGTC
6661 CGGCGTAGAG GATCG

FIGURE 36D

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pDEST17 6354 bp

<u>Location (Base Nos.)</u>	<u>Gene Encoded</u>
258..134	attR1
367..1026	CmR
1146..1230	inactivated ccdA
1368..1673	ccdB
1714..1838	attR2
2564..3421	ampR

1	CGATCCGCG	AAATTAATAC	GACTCACTAT	AGGGAGACCA	CAACGGTTTC	CCTCTAGAAA
61	TAATTTTGT	TAACTTTAAG	AAGGAGATAT	ACATATGTCG	TACTACCATC	ACCATCACCA
121	TCACCTCGAA	TCAACAAGTT	TGTACAAAAA	AGCTGAACGA	GAAACGTAAA	ATGATATAAA
181	TATCAATATA	TTAAATTAGA	TTTTCATATA	AAAACAGACT	ACATAATACT	GTAACACACA
241	ACATATCCAG	TCACTATGGC	GGCCGCATTA	GGCACCCAG	GCTTTACACT	TTATGCTTCC
301	GGCTCGTATA	ATGTGTGGAT	TTTGAGTTAG	GATCCGTCGA	GATTTTCAGG	AGCTAAGGAA
361	GCTAAAATGC	AGAAAAAAT	CACCTGGATAT	ACCACCGTTG	ATATATCCCA	ATGGCATCGT
421	AAAGAACATT	TTGAGGCATT	TCAGTCAGTT	GCTCAATGTA	CCTATAACCA	GACCGTTCAG
481	CTGGATATTA	CGGCCITTTT	AAAGACCGTA	AAGAAAAATA	AGCACAAAGT	TTATCCGGCC
541	TTTATTACACA	TTCTTGCCCG	CCTGATGAAT	GCTCATCCCG	AATTCCGTAT	GGCAATGAAA
601	GACGGTGAGC	TGGTGATATG	GGATAGTGT	CACCCITGTT	ACACCGTTT	CCATGAGCAA
661	ACTGAAACGT	TTTCATCGCT	CTGGAGTGAA	TACCACGACG	ATTTCCGGCA	GTTTCTACAC
721	ATATATTTCG	AAGATGTGGC	GTGTTACGGT	GAAAACCTGG	CCTATTTCCT	TAAAGGGTTT
781	ATTGAGAATA	TGTTTTTCGT	CTCAGCCAAT	CCTGGGGTGA	GTTTCACCAG	TTTTGATTTA
841	AACGTGGCCA	ATATGGACAA	CTTCTTCGCC	CCCGTTTTCA	CCATGGGCAA	ATATTATACG
901	CAAGGCGACA	AGGTGCTGAT	GCCGCTGGCG	ATTCAGGTTT	ATCATGCCGT	CTGTGATGGC
961	TTCCATGTG	GCAGAATGCT	TAATGAATTA	CAACAGTACT	GCGATGAGTG	GCAGGCGGG
1021	GCGTAAAGAT	CTGGATCCGG	CTTACTAAAA	GCCAGATAAC	AGTATGCGTA	TTTGGCGGCT
1081	GATTTTTCG	GTATAAGAAT	ATATACTGAT	ATGTATACCC	GAAGTATGTC	AAAAAGAGGT
1141	GTGCTATGAA	GCAGCGTATT	ACAGTGACAG	TTGACAGCGA	CAGCTATCAG	TTGCTCAAGG
1201	CATATATGAT	GTCAATATCT	CCGGTCTGGT	AAGCACAACC	ATGCAGAATG	AAGCCCGTCG
1261	TCTGCGTGCC	GAACGCTGGA	AAGCGGAAAA	TCAGGAAGGG	ATGGCTGAGG	TGCCCCGGTT
1321	TATTGAAATG	AACGGCTCTT	TTGCTGACGA	GAACAGGGAC	TGGTGAAATG	CAGTTTAAGG
1381	TTTACACCTA	TAAAGAGAG	AGCCGTTATC	GTCTGTTTGT	GGATGTACAG	AGTGATATTA
1441	TTGACACGCC	CGGGCGACGG	ATGGTGATCC	CCCTGGCCAG	TGCACGTCTG	CTGTCAGATA
1501	AAGTCTCCCG	TGAACTTTAC	CCGGTGGTGC	ATATCGGGGA	TGAAAGCTGG	CGCATGATGA
1561	CCACCGATAT	GGCCAGTGTG	CCGGTCTCCG	TTATCGGGGA	AGAAGTGGCT	GATCTCAGCC
1621	ACCGCGAAAA	TGACATCAAA	AACGCCATTA	ACCTGATGTT	CTGGGGAATA	TAAATGTCAG
1681	GCTCCCTTAT	ACACAGCCAG	TCTGCAGGTC	GACCATAGTG	ACTGGATATG	TTGTGTTTTA
1741	CAGTATTATG	TAGTCTGTTT	TTTATGCAAA	ATCTAATTTA	ATATATTGAT	ATTTATATCA
1801	TTTTACGTTT	CTCGTTCAGC	TTTCTGTGAC	AAAGTGGTTG	ATTTCAGGCT	GCTAACAAAG
1861	CCCGAAAGGA	AGCTGAGTTG	GCTGCTGCCA	CCGCTGAGCA	ATAACTAGCA	TAACCCCTTG
1921	GGGCCTCTAA	ACGGGTCTTG	AGGGGTTTTT	TGCTGAAAGG	AGGAACTATA	TCCGGATATC
1981	CACAGGACGG	GTGTGGTCCG	CATGATCGCG	TAGTCGATAG	TGGCTCCAAG	TAGCGAAGCG
2041	AGCAGGACTG	GGCGGCGGCC	AAAGCGGTCC	GACAGTGCTC	CGAGAACGGG	TGCGCATAGA
2101	AATTGCATCA	ACGCATATAG	CGCTAGCAGC	ACGCCATAGT	GACTGGCGAT	GCTGTCCGAA
2161	TGGACGATAT	CCCGCAAGAG	GCCCGGCAGT	ACCGGCATAA	CCAAGCCTAT	GCCTACAGCA
2221	TCCAGGGTGA	CGGTGCCGAG	GATGACGATG	AGCGCAATTG	TAGATTTTAT	ACACGGTGCC
2281	TGACTGCGTT	AGCAATTTAA	CTGTGATAAA	CTACCGCATT	AAAGCTTATC	GATGATAAGC
2341	TGTCAAACAT	GAGAATTCTT	GAAGACGAAA	GGGCCTCGTG	ATACGCCTAT	TTTTATAGGT
2401	TAATGTCAAT	ATAATAATGG	TTTCTTAGAC	GTCAGGTGGC	ACTTTTCGGG	GAAATGTGCG
2461	CGGAACCCCT	ATTGTGTTAT	TTTTCTAAAT	ACATTCAAAT	ATGTATCCGC	TCATGAGACA
2521	ATAACCTTGA	TAAATGCTTC	AATAATATTG	AAAAAGGAAG	AGTATGAGTA	TTCAACATTT
2581	CCGTGTCGCC	CTTATTCCTT	TTTTTGCGGC	ATTTTGCCCT	CCTGTTTTTG	CTCACCCAGA
2641	AACGCTGGTG	AAAGTAAAAA	ATGCTGAAGA	TCAGTTGGGT	GCACGAGTGG	GTTACATCGA

FIGURE 37B

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2701 ACTGGATCTC AACAGCGGTA AGATCCTTGA GAGTTTTCGC CCCGAAGAAC GTTTTCCAAT
2761 GATGAGCACT TTTAAAGTTC TGCTATGTGG CGCGGTATTA TCCCGTGTG ACGCCGGGCA
2821 AGAGCAACTC GGTCCGCCGA TACACTATTG TCAGAATGAC TTGGTTGAGT ACTCACCAGT
2881 CACAGAAAAG CATCTTACGG ATGGCATGAC AGTAAGAGAA TTATGCAGTG CTGCCATAAC
2941 CATGAGTGAT AACACTGCGG CCAACTTACT TCTGACAACG ATCGGAGGAC CGAAGGAGCT
3001 AACCGCTTTT TTGCACAACA TGGGGGATCA TGTAACCTCG CTGTATCGTT GGGAAACCGGA
3061 GCTGAATGAA GCCATACCAA ACGACGAGCG TGACACCACG ATGCCTCGAG CAATGGCAAC
3121 AACGTTGCCG AAACATTATA CTGGCGAACT ACTTACTCTA GCTTCCCGGC AACAAATTAAT
3181 AGACTGGATG GAGGCGGATA AAGTTGCAGG ACCACTTCTG CGCTCGGCCC TTCCGGCTGG
3241 CTGGTTTATT GCTGATAAAT CTGGAGCCGG TGAGCGTGGG TCTCGCGGTA TCATTGCAGC
3301 ACTGGGGCCA GATGGTAAGC CCTCCCCTAT CGTAGTTATC TACACGACGG GGAGTCAGGC
3361 AACTATGGAT GAACGAAATA GACAGATCGC TGAGATAGGT GCCTCACTGA TTAAGCATTG
3421 GTAACGTGCA GACCAAGTTT ACTCATATAT ACTTTAGATT GATTTAAAC TTCAATTTTTA
3481 ATTTAAAGG ATCTAGGTGA AGATCCTTTT TGATAATCTC ATGACCAAAA TCCCTTAACG
3541 TGAGTTTTTC TTCCACTGAG CGTCAGACCC CGTAGAAAAG ATCAAAGGAT CTCTTTGAGA
3601 TCCTTTTTTT CTGCGCGTAA TCTGCTGCTT GCAAACAAAA AAACCACCGC TACCAGCGGT
3661 GGTTTGTGTT GCGGATCAAG AGCTACCAAC TCTTTTTCCG AAGGTAACGT GCTTCAGCAG
3721 AGCGCAGATA CCAATACTG TCCCTCTAGT GTAGCCGTAG TTAGGCCACC ACTTCAAGAA
3781 CTCTGTAGCA CCGCTACAT ACCTCGCTCT GCTAATCTCG TTACCACTGG CTGCTGCCAG
3841 TGGCGATAAG TCGTGTCTTA CCGGGTTGGA CTCAGACGA TAGTTACCGG ATAAGGCGCA
3901 GCGGTCGGGC TGAACGGGGG GTTCGTGCAC ACAGCCGAGC TTGGAGCGAA CGACCTACAC
3961 CGAACTGAGA TACCTACAGC GTGAGCTATG AGAAAGCGCC ACGCTTCCCG AAGGGAGAAA
4021 GCGCGACAGG TATCCGCTAA CCGGCAGGGT CGGAACAGGA GAGCGCACGA GGGAGCTTCC
4081 AGGGGGAAAC GCCTGTGATC TTTATAGTCC TGTCGGGTTT CGCCACCTCT GACTTGAGCG
4141 TCGATTTTTG TGATGCTCGT CAGGGGGGCG GAGCCTATGG AAAAAACGCA CGAACCGCGC
4201 CTTTTTACGG TTCTTGGCCT TTTGCTGGCC TTTTGCTCAC ATGTTCTTTC CTGCGTTATC
4261 CCCTGATTCT GTGGATAACC GTATTACCGC CTTTGAGTGA GCTGATACCG CTCGCCGAG
4321 CCGAACGACC GAGCGCAGCG AGTCAGTGAG CGAGGAAGCG GAAGAGCGCC TGATGCGGTA
4381 TTTTCTCCTT ACGCATCTGT GCGGTATTTC ACACCGCATA TATGGTGAC TCTCAGTACA
4441 ATCTGCTCTG ATGCCGCATA GTTAAGCCAG TATACACTCC GCTATCGCTA CGTGACTGGG
4501 TCATGGCTGC GCCCGACAC CCGCAACAC CCGCTGACGC GCCCTGACGG GCTTGTCTCG
4561 TCCCGGCATC CGCTTACAGA CAAGCTGTGA CCGTCTCCGG GAGCTGCATG TGTCAGAGGT
4621 TTTCAACGTC ATCACCAGAA CGCGCGAGGC AGCTGCGGTA AAGCTCATCA GCGTGTCTGT
4681 GAAGCGATTG ACAGATGTCT GCCTGTTTAT CCGCGTCCAG CTCGTTGAGT TTCTCCAGAA
4741 GCGTTAATGT CTGGCTTCTG ATAAAGCGGG CCATGTTAAG GCGGTTTGTG TCCGTGTTGG
4801 TCACTGATGC CTCCTGTGTA GGGGATTTC TGTTTATGGG GGTAATGATA CCGATGAAC
4861 GAGAGAGGAT GCTCACGATA CCGGTTACTG ATGATGAACA TGCCCGGTTA CTGGAACGTT
4921 GTGAGGGTAA ACAACTGGCG GTATGGATGC GCGGGGACCA GAGAAAAATC ACTCAGGGTC
4981 AATGCCAGCG CTTCGTTAAT ACAGATGTAG GTGTTCCACA GGGTAGCCAG CAGCATCTCG
5041 CGATGCAGAT CCGGAACATA ATGGTGCAGG GCGCTGACTT CCGCGTTTCC AGACTTTACG
5101 AAACACGAA ACCGAAGACC ATTCATGTTG TTGCTCAGGT CGCAGACGTT TTGCAGCAGC
5161 AGTCGCTTCA CGTTCGCTCG CGTATCGGTG ATTCATTCTG CTAACCACTA AGGCAACCCC
5221 GCCAGCCTAG CCGGTCCTC AACGACAGGA GCACGATCAT GCGCACCCGT GGCCAGGACC
5281 CAACGCTGCC CGAGATGCGC CGCGTGCGGC TGCTGGAGAT GCGGACGCGC ATGGATATGT
5341 TCTGCCAAGG GTTGGTTTGC GCATTACAG TTCTCCGCAA GAATTGATTG GCTCCAATTC
5401 TTGGAGTGGT GAATCCGTTA GCGAGGTGCC GCCGGCTTCC ATTCAGGTG AGGTGGCCCG
5461 GCTCCATGCA CCGCGACGCA ACGCGGGGAG GCAGACAAGG TATAGGGCGG CGCCTACAA
5521 CCATGCCAAC CCGTTCATG TGCTCGCCGA GCGGCGATAA ATCGCCGTGA CGATCAGCGG
5581 TCCAGTGATC GAAGTTAGGC TGGTAAGAGC CGCGAGCGAT CCTTGAAGCT GTCCCTGATG
5641 GTCGTCTATCT ACCTGCCTGG ACAGCATGGC CTGCAACGCG GGCATCCCGA TGCCGCCGGA
5701 AGCGAGAAGA ATCATAATGG GGAAGGCCAT CCAGCTTCGC GTCCGGAACG CCAGCAAGAC
5761 GTAGCCGAGC GCGTCGGCCG CCATGCCGCG GATAATGGCC TGCTTCTGCG CGAAACGTTT
5821 GGTGGCGGGA CCAGTGACGA AGGCTTGAGC GAGGGCGTGC AAGATTCCGA ATACCGCAAG
5881 CGACAGGCCG ATCATCGTCG CGCTCCAGCG AAAGCGGTCC TCGCCGAAAA TGACCCAGAG
5941 CGCTGCCGCG ACCTGTCTTA CGAGTTGCAT GATAAAGAAG ACAGTCATAA GTGCGGCCGAC
6001 GATAGTCATG CCGCGCGCCC ACCGGAAGGA GCTGACTGGG TTGAAGGCTC TCAAGGGCAT
6061 CCGTCCGATG ACGCTCTCCC TTATGCGACT CTGCAATTAG GAAGCAGCCC AGTAGTAGGT
6121 TGAGGCCGTT GAGCACCGCC GCCGCAAGGA ATGGTGATG CAAGGAGATG GCGCCCAACA-

Figure 37C

6181 GTCCCCCGGC CACGGGGCCT GCCACCATAC CCACGCCGAA ACAAGCGCTC ATGAGCCCGA
6241 AGTGGCGAGC CCGATCTTCC CCATCGGTGA TGTCGGCGAT ATAGGCGCCA GCAACCGCAC
6301 CTGTGGCGCC GGTGATGCCG GCCACGATGC GTCCGGCGTA GAGGATCGAG ATCT

FIGURE 37D

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Figure 38A: pDEST18

FastBac Transfer Vector with p10
Baculovirus Promoter

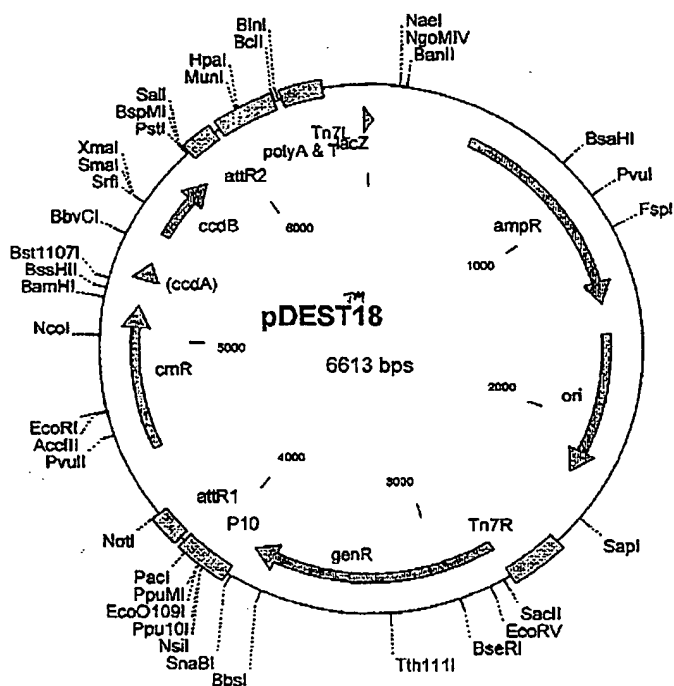
1 gaagacctcg gccgtcgagg cgcttgccgg tgggtgctgac cccggatgaa gtggttcgca
cttctggagc cggcagcgcc gcgaacggcc accacgactg gggcctactt caccaagcgt

61 tccctcggttt tctggaaggg gagcatcggt ttgtcgccca ggactctagc tatagttcta
aggagccaaa agaccttcgg ctctagcaca acaagcgggt cctgagatcg atatcaagat

121 gtggttggtt acgtatcgag caagaactga aaagcgcgaa tgcggtggag tcttctgtac
caccaaccga tgcatagcctc gttcttttat ttctggtttt ggcgaacctc agaaccagac

181 **p10 Promoter**
"tatctttaca agatctcaga ataccgacc attacacaa gggggacta taaatctac"
"ataaaatgt ttctaaatct ttatgcatag tgaatgtgt tccccctgat accttaatac"
241 "cattctgagg atgcggggac ctttaactca accgaacaca atatatata gtaaatatg" mRNA
"gtaaaactcc tacggccctg gaaattgaat tgggtgtgt tatataatat caatttacc"
301 "aattatttat caaatcattt gtatatataa taaataacta tactgtaaat tacattttat
"taataaata gttttagtaa atataaata attttatgat atgacattta atgtaaaata
361 ttacaatgag gatcatcaca agtttgtaca aaaaagctga acgagaacg taaaatgata
aatgttactc ctagtatgt tcaaacatgt tttctcgact tgcctcttgc attttactat

Int. attR1



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pDEST18 6613 bp

Location (Base Nos.)	Gene Encoded
474..1449	ampR
1590..2244	ori
2738..3850	genR
4251..4127	attR1
4501..5160	CmR
5280..5364	inactivated ccdA
5502..5807	ccdB
5848..5972	attR2
6595..25	lacZ

1	GACGCGCCCT	GTAGCGCGC	ATTAAGCGCG	GCGGGTGTGG	TGGTTACGCG	CAGCGTGACC
61	GCTACACTTG	CCAGCGCCCT	AGCGCCCGCT	CCTTTCGCTT	TCTTCCCTTC	CTTTCGCGCC
121	ACGTTCCGCCG	GCTTTCGCCG	TCAAGCTCTA	AATCGGGGGC	TCCCTTTAGG	GTTCCGATTT
181	AGTGCTTTAC	GGCACCTCGA	CCCCAAAAA	CTTGATTAGG	GTGATGGTTC	ACGTAGTGGG
241	CCATCGCCCT	GATAGACGGT	TTTTCGCCCT	TTGACGTTGG	AGTCCACGTT	CTTTAATAGT
301	GGACTCTTGT	TCCAACTGG	AACAACACTC	AACCCCTATCT	CGGTCTATT	TTTGTATT
361	TAAGGGATT	TGCCGATTTC	GGCCTATTGG	TTAAAAAATG	AGCTGATT	ACAAAAATTT
421	AACGCGAATT	TTAACAAAT	ATTAACGTTT	ACAATTTT	GTGGCACTTT	TCGGGGAAAT
481	GTGCGCGGAA	CCCCTATTTC	TTTATTTTTC	TAAATACATT	CAAATATGTA	TCCGCTCATG
541	AGACAATAAC	CCTGATAAAT	GCTTCAATAA	TATTGAAAAA	GGAAGAGTAT	GAGTATTCAA
601	CATTTCCGTG	TCGCCCTTAT	TCCCTTTTTT	GCGGCATTTT	GCCTTCCTGT	TTTTGCTCAC
661	CCAGAAACGC	TGGTGAAAGT	AAAAGATGCT	GAAGATCAGT	TGGGTGCACG	AGTGGGTTAC
721	ATCGAACTGG	ATCTCAACAG	CGGTAAGATC	CTTGAGAGTT	TTGCCCCCGA	AGAACGTTTT
781	CCAATGATGA	GCACCTTTTAA	AGTTCTGCTA	TGTGGCGCGG	TATTATCCCG	TATTGACGCC
841	GGGCAAGAGC	AACTCGGTCG	CCGCATACAC	TATTCTCAGA	ATGACTTGGT	TGAGTACTCA
901	CCAGTCACAG	AAAAGCATCT	TACGGATGGC	ATGACAGTAA	GAGAATTATG	CAGTGTCTGCC
961	ATAACCATGA	GTGATAACAC	TGCGGCCAAC	TACTTCTGTA	CAACGATCGG	AGGACCGAAG
1021	GAGCTAACCG	CTTTTTTGCA	CAACATGGGG	GATCATGTAA	CTCGCCTTGA	TCGTTGGGAA
1081	CCGGAGCTGA	ATGAAGCCAT	ACCAACGAC	GAGCGTGACA	CCACGATGCC	TGTAGCAATG
1141	GCAACAACGT	TGCGCAAACT	ATTAAGTGGC	GAAGTACTTA	CTCTAGCTTC	CCGGCAACAA
1201	TTAATAGACT	GGATGGAGGC	GGATAAAGTT	GCAGGACCAC	TTCTGCGCTC	GGCCCTTCCG
1261	GCTGGCTGGT	TTATTGCTGA	TAAATCTGGA	GCCGGTGAGC	GTGGGTCTCG	CGGTATCATT
1321	GCAGCACTGG	GGCCAGATGG	TAAGCCCTCC	CGTATCGTAG	TTATCTACAC	GACGGGGAGT
1381	CAGGCAACTA	TGGATGAACG	AAATAGACAG	ATCGCTGAGA	TAGGTGCCTC	ACTGATTAA
1441	CATTGGTAAC	TGTCAGACCA	AGTTTACTCA	TATATACTTT	AGATTGATTT	AAAACCTTCAT
1501	TTTTAATTTA	AAAGGATCTA	GGTGAAGATC	CTTTTTGATA	ATCTCATGAC	CAAAATCCCT
1561	TAACGTGAGT	TTTCGTTTCCA	CTGAGCGTCA	GACCCCGTAG	AAAAGATCAA	AGGATCTTCT
1621	TGAGATCCTT	TTTTTCTGCG	CGTAATCTGC	TGCTTGCAAA	CAAAAAAACC	ACCGCTACCA
1681	GCGGTGGTTT	GTTTGCCGGA	TCAAGAGCTA	CCAACCTTTT	TTCCGAAGGT	AACTGGCTTC
1741	AGCAGAGCGC	AGATACCAAA	TACTGTCTTT	CTAGTGTAGC	CGTAGTTAGG	CCACCACTTC
1801	AAGAACTCTG	TAGCACCGCC	TACATACCTC	GCTCTGCTAA	TCCTGTTACC	AGTGGCTGCT
1861	GCCAGTGGCG	ATAAGTCGTG	TCITTACCGG	TTGGACTCAA	GACGATAGTT	ACCGGATAAG
1921	GCGCAGCGGT	CGGGCTGAAC	GGGGGGTTCC	TGCACACAGC	CCAGCTTGGA	GCGAACGACC
1981	TACACCGAAC	TGAGATACCT	ACAGCGTGAG	CATTGAGAAA	GCGCCACGCT	TCCCGAAGGG
2041	AGAAAGGCGG	ACAGGTATCC	GGTAAGCGGC	AGGGTCGGAA	CAGGAGAGCG	CACGAGGGAG
2101	CTTCCAGGGG	GAAACGCCTG	GTATCTTTAT	AGTCCTGTCC	GGTTTCGCCA	CCTCTGACTT
2161	GAGCGTCGAT	TTTTGTGATG	CTCGTCAGGG	GGGCGGAGCC	TATGGAAAAA	CGCCAGCAAC
2221	GCGGCCCTTT	TACGGTTTCT	GGCCTTTTGC	TGGCCTTTTG	CTCACATGTT	CTTTCCTGCG
2281	TTATCCCTCG	ATTCTGTGGA	TAACCGTATT	ACCGCCTTTG	AGTGAGCTGA	TACCGCTCGC
2341	CGCAGCCGAA	CGACCGAGCG	CAGCGAGTCA	GTGAGCGAGG	AAGCGGAAGA	GCGCCTGATG
2401	CGGTATTTTC	TCCTTACGCA	TCTGTGCGGT	ATTTCACACC	GCAGACCAGC	CGCGTAACCT
2461	GGCAAAATCG	GTTACGGTTG	AGTAATAAAT	GGATGCCCTG	CGTAAGCGGG	TGTGGCGGGA-

FIGURE 38B

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2521 CAATAAAGTC TTAAACTGAA CAAAATAGAT CTAAACTATG ACAATAAAGT CTTAAACTAG
2581 ACAGAATAGT TGTAAACTGA AATCAGTCCA GTTATGCTGT GAAAAAGCAT ACTGGACTTT
2641 TGTATGGCT AAAGCAAACCT CTTCAATTTTC TGAAGTGCAA ATTGCCCGTC GTATTAAAGA
2701 GGGGCGTGGC CAAGGGCATG GTAAAGACTA TATTCGCGGC GTTGTGACAA TTTACCCAAC
2761 AACTCCGCGG CCGGGAAGCC GATCTCGGCT TGAACGAATT GTTAGGTGGC GTACTTGGG
2821 TCGATATCAA AGTGCATCAC TTCTTCCCGT ATGCCCAACT TTGTATAGAG AGCCACTGCG
2881 GGATCGTCAC CGTAATCTGC TTGCACGTAG ATCACATAAG CACCAAGCGC GTTGGGCTCA
2941 TGCTTGAGGA GATTGATGAG CGCGGTGGCA ATGCCCTGCC TCCGCTGCTC GCCGGAAGT
3001 GCGAGATCAT AGATATAGAT CTCCTACGCG GGCTGCTCAA ACCTGGGCAG AACGTAAGCC
3061 GCGAGAGCGC CAACAACCGC TTCTTGGTGG AAGGCAGCAA GCGCGATGAA TGTCTTACTA
3121 CGGAGCAAGT TCCCGAGGTA ATCGGAGTCC GGCTGATGTT GGGAGTAGGT GGCTACSTCT
3181 CCGAACTCAC GACCGAAAAG ATCAAGAGCA GCCCGCATGG ATTGACTTGG GTCAAGGCCG
3241 AGCCTACATG TCGCAATGAT GCCCATACTT GAGCCACCTA ACTTTGTTTT AGGGCGACTG
3301 CCTGTGTCGG TAACATCGTT GCTGTGCGT AACATCGTTG CTGCTCCATA ACATCAAACA
3361 TCGACCCACG GCGTAACGCG CTGTCTGCTT GGATGCCCGA GGCATAGACT GTACAAAATA
3421 ACAGTCATAA CAAGCCATGA AAACCGCCAC TCGCGCGTTA CCACCGCTGC GTTCGCTCAA
3481 GGTTCTGGAC CAGTTGCGTG AGCGCATACG CTACTTGAT TACAGTTTAC GAACCGACAA
3541 GGCTTATGTC AACTGGGTTT GTGCCCTTCAT CCGTTTCCAC GGTGTGCGTC ACCCGGCAAC
3601 CTGTGGCGAG AGCGAAGTCG AGGCATTCTT GTCCCTGGCTG GCGAACGAGC GCAAGGTTTC
3661 GGTCTCCACG CATCGTCAGG CATTGGCGGC CTGTCTGTTT TTCTACGGCA AGGTGCTGTG
3721 CACGGATCTG CCTGTGCTTC AGGAGATCGG AAGACCTCGG CCGTCGCGGC GCTTGCCTGG
3781 GGTGCTGACC CCGGATGAAG TGGTTCGCAT CCTCGGTTTT CTGGAAGGCG AGCATCSTTT
3841 GTTCGCCAG GACTCTAGCT ATAGTTCTAG TGGTTGGCTA CGTATCGAGC AAGAAAATAA
3901 AACGCCAAAC GCGTTGGAGT CTGTGTGCTT ATTTTACAA AGATTGAGAA ATACCGATCA
3961 CTTACAAACA GGGGGACTAT GAAATTATGC ATTTTGAGGA TGCCGGGACC TTTAATCAA
4021 CCCAACACAA TATATTATAG TTAAATAAGA ATTATTATC AATCATTTG TATATTAAAT
4081 AAAATACTAT ACTGTAAAT ACATTTTATT TACAATGAGG ATCATCACAA GTTTGTACAA
4141 AAAAGCTGAA CGAGAAACGT AAAATGATAT AAATATCAAT ATATTAAAT AGATTGTGCA
4201 TAAAAACAG ACTACATAAT ACTGTAAAC ACAACATATC CAGTCACTAT GCGGCGCTGT
4261 AAGTTGGCAG CATCACCCGA CGCACTTTGC GCCGAATAAA TACCTGTGAC GGAAGATCAC
4321 TTGCGAGAAAT AAATAAATCC TGGTGTCCCT GTTGATACCG GGAAGCCCTG GGCCAACTTT
4381 TGGCGAAAT GAGACGTTGA TCGGCACGTA AGAGGTTCCA ACTTTCACCA TAATGAAATA
4441 AGATCACTAC CGGCGGTATT TTTTGAGTTA TCGAGATTTT CAGGAGCTAA GGAAGCTAAA
4501 ATGGAGAAAA AAATCACTGG ATATACCACC GTTGATATAT CCCAATGGCA TCGTAAGAA
4561 CATTTTGAGG CATTTCACTG AGTTGCTCAA TGTACCTATA ACCAGACCGT TCAGCTGGAT
4621 ATTACGGCCT TTTTAAAGAC CGTAAAGAAA AATAAGCACA AGTTTATCC GGCCTTTATT
4681 CACATTCTTG CCCGCTGAT GAATGCTCAT CCGGAATTC GTATGGCAAT GAAAGACGGT
4741 GAGCTGGTGA TATGGGATAG TGTTCACCTT TGTACACCG TTTTCCATGA GCAAACTGAA
4801 ACGTTTTCAT CGCTCTGGAG TGAATACCAC GACGATTTC GGCAGTTTC ACACATATAT
4861 TCGCAAGATG TGGCGTGTGA CGGTGAAAAC CTGGCCTATT TCCCTAAAGG GTTTATTGAG
4921 AATATGTTTT TCGTCTCAGC CAATCCCTGG GTGAGTTTCA CCAGTTTGA TTTAAACGTG
4981 GCCAATATGG ACAACTTCTT CGCCCCCGTT TTCACCATGG GCAAATATTA TACGCAAGGC
5041 GACAAGGTGC TGATGCCGCT GCGGATTGAG GTTCATCATG CCGTCTGTGA TGGCTTTCAT
5101 GTCGGCAGAA TGCTTAATGA ATTACAACAG TACTGCGATG AGTGGCAGGG CCGGGCSTAA
5161 ACGCGTGGAT CCGGCTTACT AAAAGCCAGA TAACAGTATG CGTATTTCG CGCTGAATTT
5221 TGCGGTATAA GAATATATAC TGATATGTAT ACCCGAAGTA TGTCAAAAAG AGGTGTCTTA
5281 TGAAGCAGCG TATTACAGTG ACAGTTGACA GCGACAGCTA TCAGTTGCTC AAGGCATATA
5341 TGATGTCAAT ATCTCCGGTC TGGTAAGCAC AACCATGCAG AATGAAGCCC CTCGCTCGCG
5401 TGCCGAACGC TGGAAAGCGG AAAATCAGGA AGGGATGGCT GAGGTGCGCC GGTTTATGTA
5461 AATGAACGGC TCTTTTGTG ACGAGAACAG GGACTGGTGA AATCCAGTTT AAGGTTTACA
5521 CCTATAAAG AGAGAGCCGT TATCGTCTGT TTGTGGATGT ACAGAGTGAT ATTATTGACA
5581 CGCCCCGGCG ACGGATGGTG ATCCCCCTGG CCAAGTCACG TCTGCTGTCA GATAAACTCT
5641 CCCGTGAAC TTAACCGGTG GTGCATATCG GGGATGAAAG CTGGCGCATG ATGACCACCG
5701 ATATGGCCAG TGTGCCGGTC TCCGTTATCG GGAAGAAGT GGCTGATCTC AGCCACCSCG
5761 AAAATGACAT CAAAACGCC ATTAACCTGA TGTCTGTTGG AATATAAATG TCAGGCTCCC
5821 TTATACACAG CCAGTCTGCA GGTGACCAT AGTGAAGTGA TATGTTGTG TTTACAGTAT
5881 TATGTAGTCT GTTTTTTATG CAAAATCTAA TTTAATATAT TGATATTAT ATCATTTTAC
5941 GTTCTCTGTT CAGCTTCTT GTACAAAGTG GTGATAGCTT GTCGAGAAGT ACTAGAGGAT

FIGURE 38C

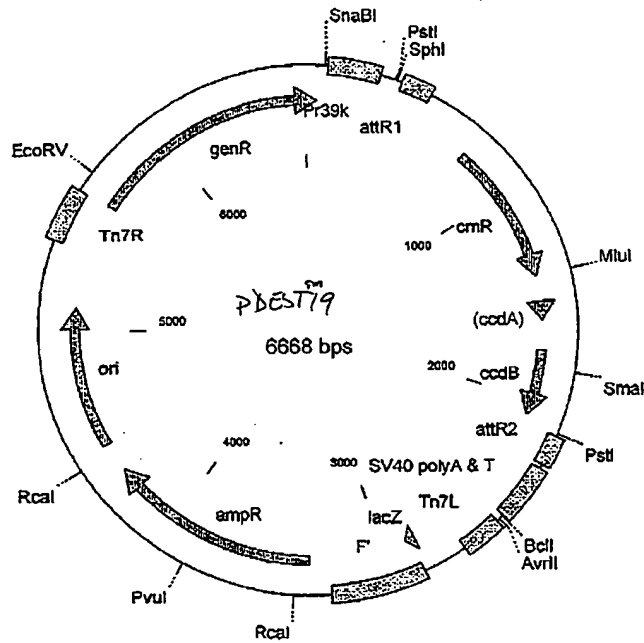
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6001 CATAATCAGC CATAACCACAT TTGTAGAGGT TTTACTTGCT TTAAAAAACC TCCCACACCT
6061 CCCCCTGAAC CTGAAACATA AAATGAATGC AATTGTTGTT GTTAACTTGT TTATTGCAGC
6121 TTATAATGGT TACAAATAAA GCAATAGCAT CACAAATTTT ACAAAATAAG CATTTTTTTC
6181 ACTGCATTCT AGTTGTGGTT TGTCCAAACT CATCAATGTA TCTTATCATG TCTGGATCTG
6241 ATCACTGCTT GAGCCTAGGA GATCCGAACC AGATAAGTGA AATCTAGTTC CAAACTATTT
6301 TGTCAATTTT AATTTTCGTA TTAGCTTAGG ACGCTACACC CAGTTCCCAT CTATTTTGTC
6361 ACTCTTCCCT AAATAATCCT TAAAAACTCC ATTTCACCC CTCCCAGTTC CCAACTATTT
6421 TGTCCGCCCA CAGCGGGGCA TTTTCTTCC TGTATGTTT TTAATCAAAC ATCCTGCCAA
6481 CTCCATGTGA CAAACCGTCA TCTTCGGCTA CTTTTTCTCT GTCACAGAAAT GAAAATTTTT
6541 CTGTCATCTC TTCGTTATTA ATGTTTGTA TTAGCTGAAT ATCAACGCTT ATTTGCAGCC
6601 TGAATGGCGA ATG
```

FIGURE 38D

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1 ggtgacgccc tcattcttcc attgtaacgt aaatggcaac ttgtagatga acgcgctgtc
 ccactgcgcc agtagaaagg taacattgca ttaccggtg aacatctact tgcgagacag
 61 aaaaaaccgg ccagtttctt ccacaaactc gcgcacggct gtctcgtaaa cttttgcgtc
 ttttttgccc ggtcaaagaa ggtgtttgag cgcgtgccga cagagcattt gaaaacgcag
 121 gcaacaatcg cgatgacctc gtggtatgga aattttttct aaaaaagtgt cgttcattgc
cgttgttagc gctactggag caccatacct ttaaaaaaga ttttttcaga gcaagtacag
 181 ggcggcgccg ttgcgcctcc gttacgcgcg acgggcacac agcaggacag ctttgcctcg
ccgcgcgcgc aagcgcgagg ctatgcgcgc tgcccgtgtg tcgtccctgc ggaacaggcc
 241 ctcgattatc ataaacaate ctgcaggcat gcaagctgga tcatacaag ttgtacaaa
gagctaatag tatttgtag gacgtccgta cgttcgacct agtagcttc aaacatgttc
 TAA V



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pDEST19 6668 bp (rotated to position 1000)

<u>Location (Base Nos.)</u>		<u>Gene Encoded</u>
515..391		attR1
765..1424		CmR
1544..1628		inactivated ccdA
1766..2071		ccdB
2112..2236		attR2
2852..2895		lacZ
3344..4319		ampR
4460..5114		ori
5608..52		genR

1	AGTGGTTTCG	ATCCTCGGTT	TTCTGGAAGG	CGAGCATCGT	TGTTTCGCCC	AGGACTCTAG
61	CTATAGTTCT	AGTGGTTGGC	TACGTATATC	AAATACTTGT	AGGTGACGCC	GTCATCTTTC
121	CAITGTAAAC	TAAATGGCAA	CTTGTAGATG	AACGCGCTGT	CAAAAAACCG	GCCAGTTTCT
181	TCCACAAACT	CGCGCACGGC	TGTCTCGTAA	ACTTTTGCCT	CGCAACAATC	GCGATGACCT
241	CGTGGTATGG	AAATTTTTC	TAAAAAAGTG	TCGTTTCATG	CGCGGGCGGG	CQCGTTCCGG
301	CTCCGGTACG	CGCGACGGGC	ACACAGCAGG	ACAGCCTTGT	CCGGCTCGAT	TATCATAAAC
361	AATCTGCAG	GCATGCAAGC	TCGGATCATC	ACAAGTTTGT	ACAAAAAAGC	TGAACGAGAA
421	ACGTAAATG	ATATAAATAT	CAATATATTA	AATTAGATT	TGCATAAAAA	ACAGACTACA
481	TAATACTGTA	AAACACAACA	TATCCAGTCA	CTATGGCGGC	CGCTAAGTTG	GCAGCATCAC
541	CCGACGCACT	TTGCGCCGAA	TAAATACCTG	TGACGGAAGA	TCACCTCGCA	GAATAAATAA
601	ATCCTGGTGT	CCCTGTTGAT	ACCGGGAAGC	CCTGGGCCAA	CTTTTGGCGA	AAATGAGACG
661	TTGATCGGCA	CGTAAGAGGT	TCCAACCTTC	ACCATAATGA	AATAAGATCA	CTACCGGGCG
721	TATTTTGA	GTTATCGAGA	TTTTCAGGAG	CTAAGGAAGC	TAAAAAGGAG	AAAAAATCA
781	CTGGATATAC	CACCGTTGAT	ATATCCCAAT	GGCATCGTAA	AGAACATTTT	GAGGCATTTT
841	AGTCAGTTGC	TCAATGTACC	TATAACCAGA	CCGTTTCAGT	GGATATTACG	GCCTTTTAA
901	AGACCGTAAA	GAAAAATAAG	CACAAGTTT	ATCCGCGCTT	TATTCACATT	CTTGCCCGCC
961	TGATGAATGC	TCATCCGGAA	TTCCGTATGG	CAATGAAAGA	CGGTGAGCTG	GTGATATGGG
1021	ATAGTGTTC	CCCTTGTAC	ACCGTTTTC	ATGAGCAAAC	TGAAACGTTT	TCATCGCTCT
1081	GGAGTGAATA	CCACGACGAT	TTCCGGCAGT	TTCTACACAT	ATATTGCGAA	GATGTGGCGT
1141	GTTACGGTGA	AAACCTGGCC	TATTTCCCTA	AAGGGTTTAT	TGAGAATATG	TTTTTCGTCT
1201	CAGCCAATCC	CTGGGTGAGT	TTCACGAGTT	TTGATTATAA	CGTGGCCAAT	ATGGACAATC
1261	TCITCGCCCC	CGTTTTTACC	ATGGGCAAAT	ATTATACGCA	AGGCGACAAG	GTGCTGATGC
1321	CGCTGGCGAT	TCAGGTTTAT	CATGCCGTCT	GTGATGGCTT	CCATGTCGGC	AGAATGCTTA
1381	ATGAATTACA	ACAGTACTGC	GATGAGTGCC	AGGGCGGGGC	GTAACGCGT	GGATCCGGCT
1441	TACTAAAAGC	CAGATAACAG	TATGCGTATT	TGCGCGCTGA	TTTTTGCGGT	ATAAGAATAT
1501	ATACTGATAT	GTATACCCGA	AGTATGTCAA	AAAGAGGTGT	GCTATGAAGC	AGCGTATTAC
1561	AGTGACAGTT	GACAGCGACA	GCTATCAGTT	GCTCAAGGCA	TATATGATGT	CAATATCTCC
1621	GGTCTGGTAA	GCACAACCAT	GCAGAATGAA	GCCCGTCGTC	TGCGTGCCGA	ACGCTGGAAA
1681	GCGGAAAATC	AGGAAGGGAT	GGCTGAGGTC	GCCCGGTTTA	TTGAAATGAA	CGGCTCTTTT
1741	GCTGACGAGA	ACAGGGACTG	GTGAAATGCA	GTTTAAAGGT	TACACCTATA	AAAGAGAGAG
1801	CCGTTATCGT	CTGTTTGTGG	ATGTACAGAG	TGATATTATT	GACACGCCCG	GGCGACGGAT
1861	GGTGATCCCC	CTGGCCAGTG	CACGTCTGCT	GTCAGATAAA	GTCTCCCGTG	AACTTTACCC
1921	GGTGGTGCAT	ATCGGGGATG	AAAGCTGGCG	CATGATGACC	ACCGATATGG	CCAGTGTGCC
1981	GGTCTCCGTT	ATCGGGGAAG	AAGTGGCTGA	TCTCAGCCAC	CGCGAAAATG	ACATCAAAAA
2041	CGCCATTAA	CTGATGTTCT	GGGGAATATA	AATGTCAGGC	TCCCTTATAC	ACAGCCAGTC
2101	TGCAGGTCGA	CCATAGTGAC	TGGATATGTT	GTGTTTACA	GTATTATGTA	GTCTGTTTTT
2161	TATGCAAAAT	CTAATTTAAT	ATAITGATAT	TTATATCATT	TTACGTTTCT	CGTTCAGCTT
2221	TCTTGTAACA	AGTGGTGATC	GAGAAGTACT	AGAGGATCAT	AATCAGCCAT	ACCACATTTG
2281	TAGAGGTTTT	ACTTGCTTTA	AAAAACCTCC	CACACCTCCC	CCTGAACCTG	AAACATAAAA
2341	TGAATGCAAT	TGTTGTTGTT	AACCTGTTTA	TTGCAGCTTA	TAATGGTTAC	AAATAAAGCA
2401	ATAGCATCAC	AAATTTTACA	AATAAAGCAT	TTTTTTCAC	GCAITCTAGT	TGTGTTTGT
2461	CCAAACTCAT	CAATGTATCT	TATCATGTCT	GGATCTGATC	ACTGCTTGAG	CCTAGGAGAT
2521	CCGAACCAGA	TAAGTGAAAT	CTAGTTCCAA	ACTATTTTGT	CATTTTTAAT	TTTCGTATTA
2581	GCTTACGACG	CTACACCCAG	TTCCCATCTA	TTTTGTCAC	CTTCCTTAA	TAATCCTTAA

FIGURE 39B

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2641 AAACCTCCATT TCCACCCCTC CCAGTTCCCA ACTATTTTGT CCGCCACAG CGGGGCATT
2701 TTCTTCTCTGT TATGTTTTTA ATCAAAACATC CTGCCAATC CATGTGACAA ACCGTCATCT
2761 TCGGCTACTT TTTCTCTGTC ACAGAATGAA AATTTTCTG TCATCTCTTC GTTATTAAATG
2821 TTGTAAATG ACTGAATATC AACGCTTATT TGCAGCCTGA ATGGCGAATG SACGCGCCCT
2881 GTAGCGGCGC ATTAAGCGCG GCGGGTGTGG TGGTTACGCG CAGCGTGACC GCTACACTTG
2941 CCAGCGCCCT AGCGCCGCT CTTTCGCTT TCTTCCCTTC CTTTCTCGCC ACGTTCGCCG
3001 GCTTTCCCG TCAAGCTCTA AATCGGGGCT TCCCTTTAGG GTTCCGATT AGTGCTTTAC
3061 GGCACCTCGA CCCCAAAAAA CTGATTAGG GTGATGGTTC ACGTAGTGGG CCATCGCCCT
3121 GATAGACGGT TTTTCGCCCT TTGACGTGG AGTCCACGTT CTTTAATAGT GGACTCTTGT
3181 TCCAACTGG AACCAACATC AACCTTATCT CGGTCTATTC TTTTGATTTA TAAGGGATTT
3241 TGCCGATTTC GGCCTATTGG TTAATAAATG AGCTGATTTA ACAAAAAATT AACGCGAATT
3301 TTAACAAAAT ATTAACGTTT ACAATTTTCA GTGGCAGTTT TCGGGGAAAT GTGCGCGGAA
3361 CCCCTATTG TTTATTTTTC TAAATACATT CAAATATGTA TCCGCTCATG AGACAATAAC
3421 CCTGATAAAT GCTTCAATAA TATTGAAAAA GGAAGAGTAT GAGTATTCAA CATTTCCGTG
3481 TCGCCCTTAT TCCCTTTTTT GCGGCATTTT GCCTTCCTGT TTTTGCTCAC CCAGAAACGC
3541 TGGTGAAGT AAAAGATGCT GAAGATCAGT TGGGTGCACG AGTGGGTTC ACGCAACTGG
3601 ATCTCAACAG CGGTAAGATC CTTGAGAGTT TTCGCCCCGA AGAAGCTTTT CCAATGATGA
3661 GCACTTTTAA AGTTCTGCTA TGTGGCGCGG TATTATCCCG TATTGACGCC GGGCAAGAGC
3721 AACTCGGTCG CCGCATACAC TATTCTCAGA ATGACTTGGT TGAGTACTCA CCAGTCACAG
3781 AAAAGCATCT TACGGATGCG ATGACAGTAA GAGAATTATG CAGTGCTGCC ATAACCATGA
3841 GTGATAACAC TGCGGCCAAC TTACTTCTGA CAACGATCGG AGGACCGAAG GAGCTAACCG
3901 CTTTTTGTGA CAACATGGGG GATCATGTAA CTCGCCCTGA TCGTTGGGAA CCGGAGCTGA
3961 ATGAAGCCAT ACCAAACGAC GAGCGTGACA CCACGATGCC TGTAGCAATG GCAACAACGT
4021 TGCGCAAAC ATTAAGTGGC GAACTACTTA CTCTAGCTTC CCGGCAACAA TTAATAGACT
4081 GGATGGAGGC GGATAAAGTT GCAGGACCAC TTCTGCGCTC GGCCTTCCG GCTGGCTGGT
4141 TTAITGCTGA TAAATCTGGA GCCGGTGAGC GTGGGTCTCG CGGTATCATT GCAGCATGG
4201 GGCCAGATGG TAAGCCCTCC CGTATCGTAG TTATCTACAC GACGGGGAGT CAGGCAACTA
4261 TGGATGAACG AAATAGACAG ATCGCTGAGA TAGGTGCTTC ACTGATTAG CATTGGTAAC
4321 TGTCAGACCA AGTTTACTCA TATATACTTT AGATTGATT AAAACTTCAT TTTTAATTTA
4381 AAAGGATCTA GGTGAAGATC CTTTGTGATA ATCTCATGAC CAAATCCCT TAACGTGAGT
4441 TTTCTGTCCA CTGAGCGTCA GACCCCGTAG AAAAGATCAA AGGATCTTCT TGAGATCCTT
4501 TTTTCTGCG CGTAATCTGC TGCTTGCAAA CAAAAAACC ACCGCTACCA CCGGTGGTTT
4561 GTTTGCCGGA TCAAGAGCTA CCAACTCTTT TTCCGAAGGT AACTGGCTTC AGCAGAGCGC
4621 AGATACCAA TACTGTCTT CTAGTGTAGC CGTAGTTAGG CCACCCTTC AAGAACTCTG
4681 TAGCACCGCC TACATACCTC GCTCTGCTAA TCCTGTTACC AGTGGCTGCT GCCAGTGGC
4741 ATAAGTCGTG TCTTACCGGG TTGGACTCAA GACGATAGTT ACCGGATAAG SCGCAGCGGT
4801 CGGGCTGAAC GGGGGGTTCG TGCACACAGC CCAGCTTGGG GCGAACGACC TACACCGAAC
4861 TGAGATACCT ACAGCGTAG CATTGAGAAA GCGCCACGCT TCCCGAAGGG AGAAGGCGG
4921 ACAGGTATCC GGTAAAGCGC AGGGTCGGAA CAGGAGAGCG CACGAGGGAG CTTCCAGGGG
4981 GAAACGCCGT GTATCTTTAT AGTCCTGTCT GGTITCGCCA CCTCTGACTT GAGCGTCGAT
5041 TTTGTGTATG CTCGTAGGG GGGCGGAGCC TATGGAAAAA CGCCAGCAAC GCGCCCTTTT
5101 TACGGTTCTT GGCCTTTTGC TGGCCTTTTG CTCACATGTT CTTTCTGCG TTATCCCTG
5161 AITCTGTGGA TAACCGTATT ACCGCCTTTG AGTGAGCTGA TACCGCTCGC CGCAGCCGAA
5221 CGACCGAGCG CAGCGAGTCA GTGAGCGAGG AAGCGGAAGA GCGCTGATG CGGTATTTTC
5281 TCCTTACGCA TCTGTGCGGT ATTTACACCC GCAGACGAGC CGCGTAACCT GGCAAAATCG
5341 GTTACGGTTG AGTAATAAAT GGATGCCCTG CGTAAGCGGG TGTGGGCGGA CAATAAAGTC
5401 TTAAACTGAA CAAAATAGAT CTAAACTATG ACAATAAAGT CTTAAACTAG ACAGAATAGT
5461 TGTAAACTGA AATCAGTCCA GTTATGCTGT GAAAAAGCAT ACTGGACTTT TGTATAGGCT
5521 AAAGCAAAC CTTCATTTTC TGAAGTGCAA ATTGCCGTC GTATTAAAGA GGGCGTGGC
5581 CAAGGGCATG GTAAAGACTA TATTGCGGCG GTTGTGACAA TTTACCGAAC AACTCCGCGG
5641 CCGGGAAGCC GATCTCGGCT TGAACGAAT GTTAGGTGGC GGTACTTGGG TCGATATCAA
5701 AGTGATCAC TTCTTCCCGT ATGCCCAACT TTGTATAGAG AGCCACTGCG GGATCGTCAC
5761 CGTAATCTGC TTGCACGTAG ATCACATAAG CACCAAGCGC GTTGGCTTCA TGCTTGAGGA
5821 GATTGATGAG CGCGGTGGCA ATGCCCTGCC TCCGGTGCTC GCCGAGACT GCGAGATCAT
5881 AGATATAGAT CTCACTACGC GGCTGCTCAA ACCTGGGCGA AACGTAAGCC GCGAGAGCGC
5941 CAACAACCGC TTCTTGGTCG AAGGCAGCAA GCGCGATGAA TGTCTTACTA CCGAGCAAGT
6001 TCCCGAGGTA ATCGGAGTCC GGCTGATGTT GGGAGTAGGT GGCTACGTCT CCGAACTCAC
6061 GACCGAAAAG ATCAAGAGCA GCCCGCATGG ATTTGACTTG GTGAGGGCGG AGCCTACATG-

FIGURE 39C

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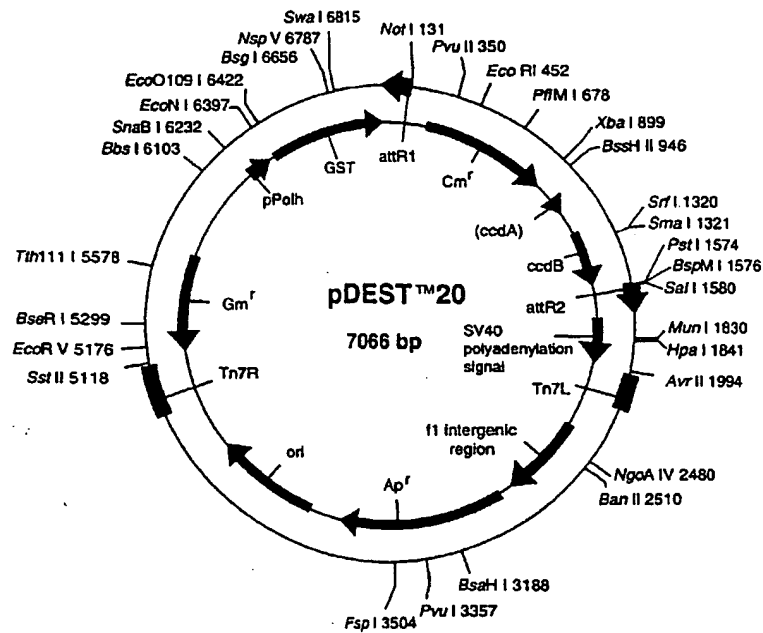
6121 TCGGAATGAT GCCCATACTT GAGCCACCTA ACTTTGTTTT AGGGCGACTG CCCTGCTGCG
6181 TAACATCGTT GCTGCTGCGT AACATCGTTG CTGCTCCATA ACATCAAACA TCGACCCACG
6241 GCGTAACGCG CTGCTGCTT GGATGCCCGA GGCATAGACT GTACAAAAAA ACAGTCATAA
6301 CAAGCCATGA AAACCGCCAC TGCGCCGTTA CCACCGCTGC GTTCGGTCAA GGTTCCTGGAC
6361 CAGTTGCGTG AGCGCATACG CTACTTGCA TACAGTTTAC GAACCGAACA GGCTTATGTC
6421 AACTGGGTTT GTGCCCTTCAT CCGTTTCCAC GGTGTGCGTC ACCCGGCAAC CTTGGGCAGC
6481 AGCGAAGTCG AGGCATTCT GTCTGGCTG GCGAACGAGC GCAAGGTTTC GGTCTCCACG
6541 CATCGTCAGG CATTGGCGGC CTGCTGTTT TTCTACGGCA AGGTGCTGTG CACGGATCTG
6601 CCCTGGCTTC AGGAGATCGG AAGACCTCG CCGTCGCGC GCTTGCCGGT GGTGCTGACC
6661 CCGGATGA

FIGURE 39A

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Figure 40A: pDEST20 Glutathione-S-transferase Fusion with Polyhedron Promoter for Baculovirus Expression

430 ggc tac gta tac tcc gga ata tta ata gat cat gga gat aat taa aat gat
 ccg atg cat atg agg cct tat aat tat cta gta cct cta tta att tta cta
 481 aac cat ctc gca aat aaa taa gta ttt tac tgt ttt cgt aac agt ttt gta
 ttg gta gag cgt tta ttt att cat aaa atg aca aaa gca ttg tca aaa cat
 532 ata aaa aaa cct ata aat att ceg gat tat tca tac cgt ccc acc atc ggy
 tat ttt ttt gga tat tta taa ggc cta ata agt atg gca ggy tag ccc
 Start Transcription → P T - - - GST - -
 583 tgc gga tcc atg gct cct ata cta ggt tat tgg aaa att aag ggc ctt gtg
 gcg cct agg tac cgg gga tat gat cca ata acc ttt taa ttc cgg gaa cac
 1246 S D L V P R H N Q T S L Y K K A
 tgc gat ctg gtt ccg cgt cat aat caa aca agt tgg tac aaa aaa gct gaa
 agc cta gac caa ggc gca gta tta gtt tgt tca aac atg ttt ttt cga ctt
 1297 cga gaa acg taa aat gat ata aat atc aat ata tta aat tag at
 gct ctt tgc att tta cta tat tta tag tta tat aat tta atc ta



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pDEST20 7066 bp (rotated to position 5800)

<u>Location (Base Nos.)</u>		<u>Gene Encoded</u>
592..1263		GST
1397..1273		attR1
1506..2165		CmR
2285..2369		inactivated ccdA
2507..2812		ccdB
2853..2977		attR2
4214..5064		ampR
5263..5843		ori
1	CCACTGCGCC GTTACCACCG CTGCGTTCGG TCAAGGTTCT GGACCAAGTTG CGTGAGCGCA	
61	TACGCTACTT GCATTACAGT TTACGAACCG AACAGGCTTA TGTCAACTGG GTTCGTGCCT	
121	TCATCCGTTT CCACGGTGTG CGTCACCCGG CAACCTTGGG CAGCAGCGAA GTCGAGGCAT	
181	TTCTGTCCTG GCTGGCGAAC GAGCGCAAGG TTTCCGTCTC CACGCATCGT CAGGCATTTGG	
241	CGGCCCTTGT GTTCTTTCTAC GGCAAGGTGC TGTGCACGGA TCTGCCCTGG CTTTCAGGAGA	
301	TCGGAAGACC TCGGCCGTGC CGGCCTTGC CGGTGGTGCT GACCCCGGAT GAAGTGGTTC	
361	GCATCCTCGG TTTTCTGGAA GCGCAGCATC GTTTGTTTCG CCAGGACTCT AGCTATAGTT	
421	CTAGTGGTTG GCTACGTATA CTCCGGAATA TTAATAGATC ATGGAGATAA TTAATAATGAT	
481	AACCATCTCG CAAATAAATA AGTATTTTAC TGTTTTCGTA ACAGTTTGT AATAAAAAA	
541	CCTATAAATA TTCCGGATTA TTCATACCGT CCCACCATCG GCGCGGATC CATGGCCCT	
601	ATACTAGGTT ATTGGAAAT TAAGGCGCTT GTGCAACCCA CTCGACTTCT TTTGGAATAT	
661	CTTGAAGAAA AATATGAAGA GCATTGTAT GAGCGCGATG AAGGTGATAA ATGGCGAAAC	
721	AAAAAGTTTG AATTGGGTTT GGAGTTTCCC AATCTTCCTT ATTATATGA TGGTGATGTT	
781	AAATTAACAC AGTCTATGGC CATCATACGT TATATAGCTG ACAAGCACAA CATGTTGGGT	
841	GGTTGTCCAA AAGAGCGTGC AGAGATTTC AATGCTTGAAG GAGCGGTTT GGATATTAGA	
901	TACGGTGTTC CGAGAATTGC ATATAGTAAA GACTTTGAAA CTCTCAAAGT TGATTTTCTT	
961	AGCAAGCTAC CTGAAATGCT GAAATGTTT GAAGATCGTT TATGTCATAA AACATATTTA	
1021	AATGGTGATC ATGTAACCCA TCCTGACTTC ATGTTGTATG ACGCTCTTGA TGTGTTTGA	
1081	TACATGGACC CAATGTGCCT GGATGCGTTC CCAAAATTAG TTTGTTTAA AAAACGTATT	
1141	GAAGCTATCC CACAAATTGA TAAGTACTTG AAATCCAGCA AGTATATAGC ATGGCCCTTG	
1201	CAGGGCTGGC AAGCCACGTT TGGTGGTGGC GACCATCCTC CAAATCGGA TCTGGTTCCG	
1261	CGTCATAATC AAACAAGTTT GTACAAAAA GCTGAACGAG AAACGTAAAA TGATATAAAT	
1321	ATCAATATAT TAAATTAGAT TTTGCATAAA AAACAGACTA CATAACTCTG TAAACACAA	
1381	CATATCCAGT CACTATGGCG GCCGCATTAG GCACCCAGG CTTTACACTT TATGCTTCCG	
1441	GCTCGTATGT TGTGTGGAAT TTGAGTTAGG ATCCGGCGAG ATTTTCAGGA GCTAAGGAAG	
1501	CTAAAATGGA GAAAAAATC ACTGGATATA CCACCGTTGA TATATCCCAA TGGCATCGTA	
1561	AAGAACATTT TGAGGCATTT CAGTCAGTTG CTCAATGTAC CTATAACCAG ACCGTTCCAGC	
1621	TGGATATTAC GGCCTTTTGA AAGACCGTAA AGAAAAATAA GCACAAGTTT TATCCGGCCT	
1681	TTATTCACAT TCTTGCCCGC CTGATGAATG CTCATCCGGA ATTCGGTATG GCAATGAAAG	
1741	ACGGTGAGCT GGTGATATGG GATAGTTTC ACCCTTGTTA CACCGTTTTC CATGAGCAAA	
1801	CTGAAACGTT TTCATCGCTC TGGAGTGAAT ACCACGACGA TTTCCGGCAG TTTCTACACA	
1861	TATATTCCGA AGATGTGGCG TGTACGGTG AAAACCTGGC CTATTTCCCT AAAGGGTTTA	
1921	TTGAGAATAT GTTTTTCGTC TCAGCCAATC CCTGGGTGAG TTTCAACAGT TTTGATTTAA	
1981	ACGTGGCCAA TATGGACAAC TTCTTCGCCC CCGTTTTCAC CATGGGCAAA TATTATACGC	
2041	AAGGCGACAA GGTGCTGATG CCGCTGGCGA TTCAGGTTCA TCATGCCGTC TGTGATGGCT	
2101	TCCATGTCGG CAGAATGCTT AATGAATTAC AACAGTACTG CGATGAGTGG CAGGCGGGG	
2161	CGTAATCTAG AGGATCCGGC TTAATAAAG CCAGATAACA GTATGCGTAT TTGCGCGCTG	
2221	ATTTTTCGGG TATAAGAATA TATACTGATA TGTATACCCG AAGTATGTCA AAAAGAGGTG	
2281	TGCTATGAAG CAGCGTATTA CAGTGACAGT TGACAGCGAC AGCTATCAGT TGCTCAAGGC	
2341	ATATATGATG TCAATATCTC CGGTCTGGTA AGCACAACCA TGCAGAATGA ACCCCGTCGT	
2401	CTGCGTGCCG AACGCTGGAA AGCGGAAAT CAGGAAGGGA TGGCTGAGGT CGCCGGTTT	
2461	ATTGAAATGA ACGGCTCTTT TGCTGACGAG AACAGGACT GGTGAAATGC AGTTTAAGGT	
2521	TTACACCTAT AAAAGAGAGA GCCGTTATCG TCTGTTTGTG GATGTACAGA GTGATATTAT	
2581	TGACACGCCC GGGCGACGGA TGGTGATCCC CTTGGCCAGT GCACGTCTGC TGTACATAA	
2641	AGTCTCCCGT GAACCTTACC CGGTGGTGCA TATCGGGGAT GAAAGCTGGC GCATGATGAC	

Figure 40B

2701 CACCGATATG GCCAGTGTGC CGGTCTCCGT TATCGGGGAA GAAGTGGCTG ATCTCAGCCA
2761 CCGCGAAAAT GACATCAAAA ACGCCATTAA CCGATGTTT TGGGAATAT AAATGTCAGG
2821 CTCCCTTATA CACAGCCAGT CTGCAGGTCG ACCATAGTGA CTGGATATGT TGTGTTTTAC
2881 AGTATTATGT AGTCTGTTTT TTATGCAAAA TCTAATTAA TATATTGATA TTTATATCAT
2941 TTTACGTTTC TCGTTACAGT TTCTGTGACA AAGTGGTTTT ATAGCTTGTC GAGAAGTACT
3001 AGAGGATCAT AATCAGCCAT ACCACATTG TAGAGGTTTT ACTTGCTTTA AAAAACTCC
3061 CACACCTCCC CCTGAACCTG AAACATAAAA TGAATGCAAT TGTGTTGTT AACCTGTTTA
3121 TTGCAGCTTA TAATGGTTAC AAATAAAGCA ATAGCATCAC AAATTTACACA AATAAAGCAT
3181 TTTTTTCACT GCATTCTAGT TGTGGTTTTG CCAAACTCAT CAATGTATCT TATCATGTCT
3241 GGATCTGATC ACTGCTTGAG CCTAGGAGAT CCGAACCAGA TAAGTGAAT CTAGTTCCAA
3301 ACTATTTTGT CATTTTTAA TTTCTGTTA GCTTACGACG CTACACCCAG TTCCCATCTA
3361 TTTTGTCACT CTTCCTTAAA TAATCCTTAA AAATCCATT TCCACCCCTC CCAGTTCCCA
3421 ACTATTTTGT CCGCCACAG CGGGGCATT TTCTTCCTGT TATGTTTTTA ATCAACATC
3481 CTGCCAATC CATGTGACAA ACCGTCTCT TCGGCTACTT TTTCTCTGTC ACAGAATGAA
3541 AATTTTTCTG TCACTCTTTC GTTATTATG TTTGTAATTG ACTGAATATC AACGCTTATT
3601 TGCAGCCTGA ATGGCGAATG GACGCGCCCT GTAGCGGCGC ATTAAGCGCG GCGGGTGTGG
3661 TGGTTACGCG CAGCGTGACC GCTACACTTG CCAGCGCCCT AGCGCCCGCT CCTTTCGCTT
3721 TCTTCCCTTC CTTCTCGCC ACGTTCGCGG GCTTTCCTCG TCAAGCTCTA AATCGGGGCG
3781 TCCCTTTAGG GTTCCGATTT AGTCTTTAC GGCACCTCGA CCCCCAAAAA CTGATTAGG
3841 GTGATGGTTC ACGTAGTGGG CCATCGCCCT GATAGACGGT TTTTCGCCCT TTGACGTTGG
3901 AGTCCACGTT CTTTAATAGT GGAATCTTGT TCCAACTGG AACAACTC AACCTATCT
3961 CGGTCTATT TTTTGATTTA TAAGGGATTT TGCCGATTT GGCCTATTGG TTAATAAATG
4021 AGCTGATTTA ACAAAAATTT AACCGCAATT TTAACAAAT ATTAACGTTT ACAATTCAG
4081 GTGGCACTTT TCGGGGAAAT GTGCGCGGAA CCGCTATTG TTTATTTTTC TAAATACATT
4141 CAAATATGTA TCCGCTCATG AGACAATAAC CCTGATAAAT GCTTCAATAA TATTGAAAAA
4201 GGAAGAGTAT GAGTATTCAT CATTTCCGTG TCGCCCTTAT TCCCTTTTTT GCGGCATTTT
4261 GCCTTCCTGT TTTTGCTCAC CCAGAAACGC TGGTGAAGT AAAAGATGCT GAAGATCAGT
4321 TGGGTGCACG AGTGGGTTC ATCGAACTGG ATCTCAACAG CGGTAAGATC CTTGAGAGTT
4381 TTCGCCCCGA AGAACGTTTT CCAATGATGA GCATTTTAA AGTTCTGCTA TGTGGCGCGG
4441 TATTATCCCG TATTGACGCC GGGCAAGAGC AACTCGGTG CCGCATACAC TATTCTCAGA
4501 ATGACTTGGT TGAGTACTCA CCAGTCACAG AAAAGCATCT TACGGATGGC ATGACAGTAA
4561 GAGAAATTATG CAGTGCTGCC ATAACCATGA GTGATAACAC TGCGGCCAAC TTACTTCTGA
4621 CAACGATCGG AGGACCGAAG GAGCTAACCG CTTTTTGA CAACATGGGG GATCATGTAA
4681 CTCGCCTTGA TCGTTGGGAA CCGGAGCTGA ATGAAGCCAT ACCAAACGAC GAGCGTGACA
4741 CCACGATGCC TGTAGCAATG GCAACAACGT TGCGCAAACT ATTAACCTGG GAACACTTTA
4801 CTCTAGCTTC CCGGCAACAA TTAATAGACT GGATGGAGGC GGATAAAGTT GCAGGACCAC
4861 TTCTGCGCTC GGCCCTTCCG GCTGGCTGGT TTATTGCTGA TAAATCTGGA GCCGGTGAGC
4921 GTGGGTCTCG CGGTATCAT GCAGCACTGG GGCCAGATGG TAAGCCCTCC CGTATCGTAG
4981 TTATCTACAC GACGGGGAGT CAGGCAACTA TGGATGAACG AAATAGACAG ATCGCTGAGA
5041 TAGGTGCCTC ACTGATTAAG CATTGGTAAC TGTACAGCA AGTTTACTCA TATATACTTT
5101 AGATTGATTT AAAACTTCAT TTTTAATTAA AAAGGATCTA GGTGAAGATC CTTTTTGATA
5161 ATCTCATGAC CAAATCCCT TAACGTGAGT TTTCTGTTCA CTGAGCGTCA GACCCCGTAG
5221 AAAAGATCAA AGGATCTTCT TGAGATCCTT TTTTCTGCG CGTAATCTGC TGCTTGCAAA
5281 CAAAAAACC ACCGCTACCA GCGGTGGTT GTTTGCCGGA TCAAGAGCTA CCAACTCTTT
5341 TTCCGAAGGT AACTGGCTTC AGCAGAGCGC AGATACCAAA TACTGTCTTT CTAGTGTAGC
5401 CGTAGTTAGG CCACCACTTC AAGAACTCTG TAGCACCGCC TACATACCTC GCTCTGCTAA
5461 TCCTGTTACC AGTGGCTGCT GCCAGTGGCG ATAAGTCTGT TCTTACCGGG TTGGACTCAA
5521 GACGATAGTT ACCGGATAAG GCGCAGCGGT CCGGCTGAAC GGGGGGTTCC TGCACACAGC
5581 CCAGCTTGGA GCGAACGACC TACACCGAAC TGAGATACCT ACAGCGTGAG CATTGAGAAA
5641 GCGCCACGCT TCCCGAAGGG AGAAAGGCGG ACAGGTATCC GGTAAGCGGC AGGGTCGGAA
5701 CAGGAGAGCG CACGAGGGAG CTTCAGGGG GAAACGCTG GTATCTTTAT AGTCTGTGCG
5761 GGTTCGCCA CTCTGACTT GAGCGTCGAT TTTTGTGATG CTCGTCAGGG GGGCGGAGCC
5821 TATGGAAAAA CGCCAGCAAC GCGGCTTTT TACGGTTCTT GGCCTTTTGC TGGCCTTTTG
5881 CTCACATGTT CTTTCTGCG TTATCCCTG ATTCTGTGGA TAACCGTATT ACCGCTTTTG
5941 AGTGAGCTGA TACGCTGCG CGCAGCGGAA CGACCGAGCG CAGCGAGTCA GTGACGAGG
6001 AAGCGGAAGA GCGCTGATG CGGTATTTT TCCTTACGCA TCTGTGCGGT ATTTACACCC
6061 GCAGACCAGC CGCGTAACCT GGCAAAATCG GTTACGGTTG AGTAATAAAT GGATGCCCTG
6121 CGTAAGCGGG TGTGGGCGGA CAATAAAGTC TTAACCTGAA CAAAATAGAT CTAACTATG-

FIGURE 40C

6181 ACAATAAAGT CTTAAACTAG ACAGAATAGT TGTAAGTGA AATCAGTCCA GTTATGCTGT
6241 GAAAAAGCAT ACTGGACTTT TGTATGGCT AAAGCAAAC CTTATTTTC TGAAGTGCAA
6301 ATTGCCCCGC GTATTAAAGA GGGGCGTGGC CAAGGGCATG GTAAAGACTA TATTCCGGC
6361 GTTGTGACAA TTTACCGAAC AACTCCGCGG CCGGGAAGCC GATCTCGGCT TGAACGAATT
6421 GTTAGGTGGC GGTACTTGGG TCGATATCAA AGTGCATCAC TTCTTCCCGT ATGCCCAACT
6481 TTGTATAGAG AGCCACTGCG GGATCGTCAC CGTAATCTGC TTGCACGTAG ATCACATAAG
6541 CACCAAGCGC GTTGGCCTCA TGCTTGAGGA GATTGATGAG CGCGGTGGCA ATGCCCTGCC
6601 TCCGGTGCTC GCCGGAGACT GCGAGATCAT AGATATAGAT CTCACTACGC GGCTGCTCAA
6661 ACCTGGGCAG AACGTAAGCC GCGAGAGCGC CAACAACCGC TTCTTGGTCG AAGGCAGCAA
6721 GCGCGATGAA TGTCTTACTA CGGAGCAAAGT TCCCGAGGTA ATCGGAGTCC GGCTGATGTT
6781 GGGAGTAGGT GGCTACGTCT CCGAACTCAC GACCGAAAAG ATCAAGAGCA GCCCGCATGG
6841 ATTTGACTTG GTCAGGGCCG AGCCTACATG TGCGAATGAT GCCCATACTT GAGCCACCTA
6901 ACTTTGTTT AGGGGCGACTG CCCTGCTGCG TAACATCGTT GCTGCTGCGT AACATCGTTG
6961 CTGCTCCATA ACATCAACA TCGACCCACG GCGTAACGCG CTTGCTGCTT GGATGCCCGA
7021 GGCATAGACT GTACAAAAA ACAGTCATAA CAAGCCATGA AAACCG

FIGURE 40D

Figure: 4 (A):

PDt3T21

2-Hybrid Vector with DNA-Binding Domain

ADH PROMOTER

700 ~~ttg ggc ctc tgc tat caa gta taa ata gac ctg caa tca tca atc ttt tgt~~
~~aac ggc gaa acg ata gtt cat att tat ctg ggc att aat aat taa gaa~~

751 ~~ttc ctg gtc att gtt ctg gtt ccc ttt ctt cct tgc ttc ttc tgc aca~~
~~aag gac cag taa caa gaa gaa caa ggc aca aag aag aag acg tgt~~

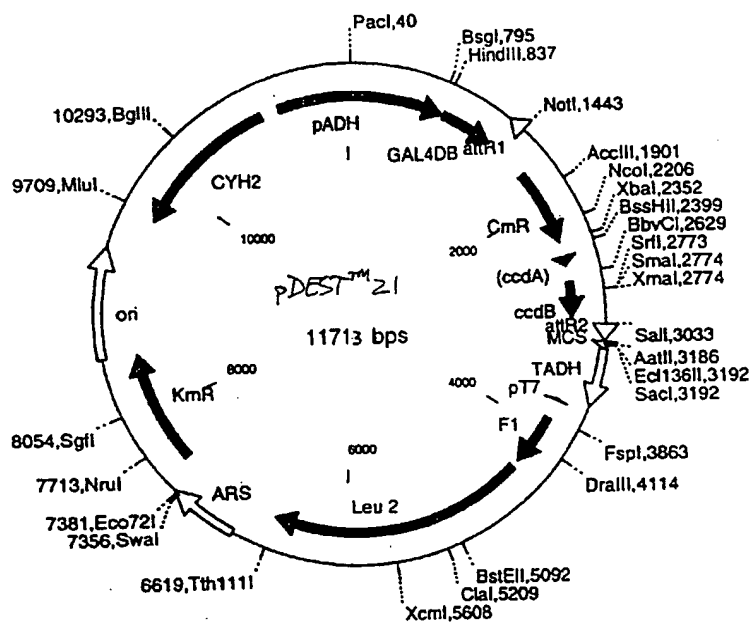
802 ~~ata ttt caa gct ata cca agc ata caa tca act~~ cca agc ttg aag caa gcc
~~tat aag gtt cga tat ggt tgc tat gtt aat tga~~ ggt tgc aac ttc gtt cgg

853 ~~Start Transl~~ ~~M K L L S t t t a c gaa caa gca tgc gat att tgg~~ Gal 4 - DB
~~agg act ttc tac ttc gat gac aga aga tag ctt gtt cgt acg cta taa acg~~

1261 gaa gag agt agt aac aaa ggt caa aga cag ttg act gta tgc tgc agg agt tgc
ctt ctc tca tca ttg ttt cca gtt tct gtc aac tga cat agc agc tcc agc

1312 ~~N Q~~ ~~T S L Y C K K A~~ ~~act RI~~
aat caa tca agt tgc taa aaa aag gct gaa cga gaa acg taa aat gat ata
tta gtt tgc tca aac atg ttt ttt cga ctt gct ctt tgc att tta cta tat

Int ↓



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pDEST21 11713 bp (rotated to position 11000)

<u>Location (Base Nos.)</u>		<u>Gene Encoded</u>
857..1322		GAL4DB
1456..1332		attR1
1706..2365		CmR
2485..2569		inactivated ccdA
2707..3012		ccdB
3053..3177		attR2
3716..3735		pT7 (T7 promoter)
3899..4354		f1 (f1 intergenic region)
4414..6642		Leu2
7541..8515		kanR
9668..10958		CYH2
11118..848		pADH (ADH promoter)

1	TTTATTATGT	TACAATATGG	AAGGGAACCTT	TACACTTCTC	CTATGCACAT	ATATTAATTA
61	AAGTCCAATG	CTAGTAGAGA	AGGGGGGTAA	CACCCCTCCG	CGCTCTTTTC	CGATTTTTTT
121	CTAAACCGTG	GAATATTTTCG	GATATCCTTT	TGTTGTTTCC	GGGTGTACAA	TATGGACTTC
181	CTCTTTTCTG	GCAACCAAAC	CCATACATCG	GGATTCCCTAT	AATACCTTCG	TTGGTCTCCC
241	TAACATGTAG	GTGGCGGAGG	GGAGATATAC	AATAGAACAG	ATACCAGACA	AGACATAATG
301	GGCTAAACAA	GACTACACCA	ATTACACTGC	CTCATTGATG	GTGGTACATA	ACGAACATAAT
361	ACTGTAGCCC	TAGACTTGAT	AGCCATCATC	ATATCGAAGT	TCACTACCC	TTTTTCCATT
421	TGCCATCTAT	TGAAGTAATA	ATAGGCGCAT	GCAACTTCTT	TTCTTTTTTT	TTCTTTTCTC
481	TCTCCCCCGT	TGTTGTCTCA	CCATATCCGC	AATGACAAAA	AAAATGATGG	AAGACACTAA
541	AGGAAAAAAT	TAACGACAAA	GACAGCACCA	ACAGATGTCG	TTGTTCCAGA	GCTGATGAGG
601	GGTATCTTCG	AACACACGAA	ACTTTTTCTT	TCCTTCATTC	ACGCACACTA	CTCTCTAATG
661	AGCAACGGTA	TACGGCCCTT	CTTCCAGTTA	CTTGAATTTG	AAATAAAAAA	AGTTTGCCGC
721	TTTGCTATCA	AGTATAAATA	GACCTGCAAT	TATTAATCTT	TTGTTTCTCT	GTCAATTGTT
781	TCGTTCCCTT	TCTTCTTGT	TTCTTTTTCT	GCACAATATT	TCAAGCTATA	CCAAGCATAC
841	AATCAACTCC	AAGCTTGAAG	CAAGCCTCCT	GAAAGATGAA	GCTACTGTCT	TCTATCGAAC
901	AAGCATGCGA	TATTTGCCGA	CTTAAAAAGC	TCAAGTGCTC	CAAAGAAAAA	CCGAAGTSCG
961	CCAAGTGCT	GAAGAACAAC	TGGGAGTGTC	GCTACTCTCC	CAAAACCAAA	AGGTCTCCGC
1021	TGACTAGGGC	ACATCTGACA	GAACTGGAAT	CAAGGCTAGA	AAGACTGAA	CAGCTATTTT
1081	TACTGATTTT	TCCTCGAGAA	GACCTTGACA	TGATTTTGAA	AATGGATTCT	TTACAGGATA
1141	TAAAAGCATT	GTAAACAGGA	TTATTTGTAC	AAGATAATGT	GAATAAAGAT	CGCCTCACAG
1201	ATAGATTGGC	TTCACTGGAG	ACTGATATGC	CTCTAACATT	GAGACAGCAT	AGAATAAGTG
1261	CGACATCATC	ATCGGAAGAG	AGTAGTAACA	AAGGTCAAAG	ACAGTTGACT	GTATCGTCCA
1321	GGTCGAATCA	AACAAGTTTG	TACAAAAAAG	CTGAACGAGA	AACGTAAAAA	GATATAAATA
1381	TCAATATATT	AAATTAGATT	TTGCATAAAA	AACAGACTAC	ATAATACTGT	AAAACACAAC
1441	ATATCCAGTC	ACTATGGCGG	CCGCTAAGTT	GGCAGCATCA	CCCGACGCAC	TTTGCGCCGA
1501	ATAAATACCT	GTGACGGAAG	ATCACTTCGC	AGAATAAATA	AATCCTGGTG	TCCCTGTTGA
1561	TACCGGGAAG	CCCTGGGCCA	ACTTTTGGCG	AAAATGAGAC	GTTGATCGGC	ACGTAAGAGG
1621	TTCCAACCTT	CACCATAATG	AAATAAGATC	ACTACCGGGC	GTATTTTTTG	AGTTATCGAG
1681	ATTTTCAGGA	GCTAAGGAAG	CTAAAAATGA	GAAAAAATC	ACTGGATATA	CCACCGTTGA
1741	TATATCCCAA	TGGCATCGTA	AAGAACATTT	TGAGGCATTT	CAGTCAGTTG	CTCAATGTAC
1801	CTATAACCAG	ACCGTTTCAGC	TGGATATTAC	GGCCTTTTTA	AAGACCGTAA	AGAAAAATAA
1861	GCACAAGTTT	TATCCGGCCT	TTATTCACAT	TCTTGCCCGC	CTGATGAATG	CTCATCCGGA
1921	ATTCCGTATG	GCAATGAAAG	ACGGTGAGCT	GGTGATATGG	GATAGTGTTT	ACCCTTGTTA
1981	CACCGTTTTT	CATGAGCAAA	CTGAAACGTT	TTTCATCGCTC	TGGAGTGAAT	ACCACGACGA
2041	TTTCCGGCAG	TTTCTACACA	TATATTCGCA	AGATGTGGCG	TGTTACGGTG	AAAACCTGGC
2101	CTATTTCCCT	AAAGGGTTTA	TTGAGAATAT	GTTTTTCGTC	TCAGCCCAATC	CTGGGGTGAG
2161	TTTCACCACT	TTTGATTTAA	ACGTGGCCAA	TATGGACAAC	TTCTTCGCCC	CCGTTTTTAC
2221	CATGGGCAAA	TATTATACCG	AAGGCGACAA	GGTGCTGATG	CCGCTGGCGA	TTCAGGTTCA
2281	TCATGCCGTC	TGTGATGGCT	TCCATGTCCG	CAGAATGCTT	AATGAATTAC	AACAGTACTG
2341	CGATGAGTGG	CAGGGCGGGG	CGTAATCTAG	AGGATCCGGC	TTACTAAAAA	CCAGATAACA
2401	GTATGCGTAT	TTGCGCGCTG	ATTTTTGCGG	TATAAGAATA	TATACTGATA	TGTATACCCG-

FIGURE 41B

2461 AAGTATGTCA AAAAGAGGTG TGCTATGAAG CAGCGTATTA CAGTGACAGT TGACAGCGAC
2521 AGCTATCAGT TGCTCAAGGC ATATATGATG TCAATATCTC CGGTCTGGTA AGCACAACCA
2581 TGCAGAAATGA AGCCCGTCGT CTGCGTGCCG AACGCTGGAA AGCGGAAAT CAGGAAGGGA
2641 TGGCTGAGGT CGCCCGGTTT AITGAAATGA ACGGCTCTTT TGCTGACGAG AACAGGGA
2701 GGTGAAATGC AGTTTAAGGT TTACACCTAT AAAAGAGAGA GCCGTTATCG TCTGTTTGTG
2761 GATGTACAGA GTGATATTAT TGACACGCCC GGGCGACGGA TGGTGATCCC CCTGGCCAGT
2821 GCACGTCTGC TGTCAGATAA AGTCTCCCGT GAACTTTACC CGGTGGTGCA TATCGGGGAT
2881 GAAAGCTGGC GCATGATGAC CACCGATATG GCCAGTGTGC CGGTCTCCGT TATCGGGGAA
2941 GAAGTGGCTG ATCTCAGCCA CCGCGAAAT GACATCAAAA ACGCCATTAA CCTGATGTTT
3001 TGGGGAATAT AAATGTCAGG CTCCCTTATA CACAGCCAGT CTGAGGTGCG ACCATAGTGA
3061 CTGGATATGT TGTGTTTAC AGTATTATGT AGTCTGTTT TTATGCAAAA TCTAATTAA
3121 TATATTGATA TTTATATCAT TTTACGTTTC TCGTTCAGCT TTCTTGATCA AAGTGGTTTG
3181 ATGGCCGCTA AGTAAGTAAG ACGTCGAGCT CTAAGTAAGT AACGGCCGCC ACCGCGGTGG
3241 AGCTTTGGAC TTCTTCGCCA GAGGTTTGGT CAAGTCTCCA ATCAAGGTTG TCGGCTTGTG
3301 TACCTTGCCA GAAATTTACG AAAAGATGGA AAAGGGTCAA ATCGTTGGTA GATACGTTGT
3361 TGACACTTCT AAATAAGCGA ATTTCTTATG ATTTATGATT TTTATTATTA AATAAGTTAT
3421 AAAAAAATA AGTGATACA AATTTTAAAG TGACTCTTAG GTTTTAAAC GAAATTTCTT
3481 ATTCTTGAGT AACTCTTTC TGTAGGTGAG GTTGTCTTCT CAGGTATAGC ATGAGGTGCG
3541 TCTTATTGAC CACACCTCTA CCGCATGCC GAGCAATGC CTGCAATCG CTCCCAATT
3601 CACCCAATTG TAGATATGCT AACTCCAGCA ATGAGTTGAT GAATCTCGGT GTGTATTTTA
3661 TGTCTCAGA GGACAATACC TGTGTAAATC GTTCTTCCAC ACGGATCCCA ATTCGCCCTA
3721 TAGTGAGTCG TATTACAATT CACTGGCCGT CGTTTTACAA CGTCTGACT GGGAAACCC
3781 TGGCGTTACC CAACTTAATC GCCTTGACG ACATCCCTCT TCGCCAGCT GCGTAATAG
3841 CGAAGAGGCC CGCACCGATC GCCCTTCCCA ACAGTTGCGC AGCCTGAATG GCGAATGGAC
3901 GCGCCCTGTA GCGGCGCATT AAGCGCGGCG GGTGTGGTGG TTACGCGCAG CGTGACCGCT
3961 ACACTTGCCA GCGCCCTAGC GCCCGCTCCT TTCGCTTCT TCCCTTCCCT TCTCGCCAGC
4021 TTCGCCGCTT TCCCGCTCA AGCTCTAAAT CCGGGGCTCC CTTAGGGTT CCGATTAGT
4081 GCTTTACGGC ACCTCGACCC CAAAAAATT GATTAGGGTG ATGGTTACAG TAGTGGGCA
4141 TCGCCCTGAT AGACGGTTTT TCGCCCTTTG ACGTTGGAGT CCACGTTCT TAATATGGA
4201 CTCTTGTTCC AAAGTGAAC AACACTCAAC CCTATCTCG TCTATTCTTT TGATTATAA
4261 GGGATTTTGC CGATTTCGGC CTATTGGTTA AAAAATGAGC TGATTTAACA AAAATTAAAC
4321 GCGAATTTTA ACAAATATT AACGTTTACA ATTTCTGAT GCGGTATTT CTCTTACGC
4381 ATCTGTGCGG TATTTCACAC CGCATATCGA CCGTCTGAGG AGAACTTCTA GTATATCCAC
4441 ATACCTAATA TTATTGCTT ATTAATAATG GAATCGGAAC AATTACATCA AAATCCACAT
4501 TCTCTTCAA ATCAATTGTC CTGTACTTCC TTGTTCAATG GTGTTCAAAA ACGTTATATT
4561 TATAGGATAA TTATACTCTA TTTCTCAACA AGTAATTGGT TGTTTGCCG ACGGCTCTAA
4621 GCGCCTGAT TCAAGAAATA TCTTGACCGC AGTTAACTGT GGAATACTC AGGTATCGTA
4681 AGATGAAGA GTTCGAATCT CTAGCAACC ATTATTTTT TCCTCAACAT AACGAGAACA
4741 CACAGGGGCG CTATCGCACA GAATCAAAAT CGATGACTGG AAATTTTTTG TTAATTTTCA
4801 AGGTGCGCTG ACGCATATAC CTTTTTCAAC TGAAAAATTG GGAGAAAAAG GAAAGGTGAG
4861 AGGCCGGAAC CGGCTTTTCA TATAGAATAG AGAAGCGTTC ATGACTAAAT GCTTGCATCA
4921 CAATCTTGA AGTTGACAAT ATTATTTAAG GACCTATTGT TTTTCCAAAT AGGTGGTTAG
4981 CAATCGTCTT ACTTTCTAAC TTTTCTTACC TTTTACATTT CAGCAATATA TATATATATT
5041 TCAAGGATAT ACCATTCTAA TGTCTGCCCC TATGTCTGCC CCTAAGAAGA TCGTCGTTTT
5101 GCCAGGTGAC CACGTTGGTC AAGAAATCAC AGCCGAAGCC ATTAAGGTTT TTAAGGCTAT
5161 TTCTGATGTT CGTTCCAATG TCAAGTTTCA TTTGAAAAT CATTAAATTG GTGGTGCTGC
5221 TATCGATGCT ACAGGTGTC CACTTCCAGA TGAGGCGCTG GAAGCCTCCA AGAAGGTTGA
5281 TGCCGTTTTG TTAGGTGCTG TGGGTGGTCC TAAATGGGGT ACCGGTAGTG TTAGACCTGA
5341 ACAAGGTTTA CTAAAAATCC GTAAAGAACT TCAATTGTAC GCCAACTTAA GACCATGTAA
5401 CTTTGCATCC GACTCTCTTT TAGACTTATC TCCAATCAAG CCACAATTTG CTAAAGGTAC
5461 TGACTTCGTT GTTGTACAGG AATTAGTGGG AGGTATTTAC TTTGGTAAGA GAAAGGAAGA
5521 CGATGGTGAT GGTGTCGCTT GGGATAGTGA ACAATACACC GTTCCAGAAG TGCAAGAAT
5581 CACAAGAATG GCCGCTTTCA TGGCCCTACA ACATGAGCCA CCATTGCCTA TTTGGTCTTT
5641 GGATAAAGCT AATGTTTTGG CTTCTTCAAG ATTATGGAGA AAAACTGTGG AGGAAACCAT
5701 CAAGAACGAA TTCCCTACAT TGAAGGTTCA ACATCAATTG ATTGATTCTG CCGCATGAT
5761 CCTAGTTAAG AACCCAACCC ACCTAAATGG TATTATAATC ACCAGCAACA TGTTTGGTGA
5821 TATCATCTCC GATGAAGCCT CCGTTATCCC AGGTTCCITG GGTGTTGTGC CATCTGCGTC
5881 CTTGGCCTCT TTGCCAGACA AGAACACCGC ATTTGGTTTG TACGAACCAT GCCACGGTTC

FIGURE 41C

5941 TGCTCCAGAT TTGCCAAAGA ATAAGGTTGA CCCTATCGCC ACTATCTTGT CTGCTGCAAT
6001 GATGTTGAAA TTGTCATTGA ACTTGCCTGA AGAAGGTAAG GCCATTGAAG ATGCASITAA
6061 AAAGGTTTTG GATGCAGGTA TCAGAACTGG TGATTTAGGT GGTTCACAAC GTACCACCGA
6121 AGTCGGTGAT GCTGTGCGCG AAGAAGTTAA GAAAATCCTT GCTTAAAAAG ATTCTCTTTT
6181 TTTATGATAT TTGTACATAA ACITTATATA TGAAATTCAT AATAGAAACG ACACGAAATT
6241 ACAAAATGGA ATATGTTTAT AGGGTAGACG AAACATATATA CGCAATCTAC ATACATTTAT
6301 CAAGAAGGAG AAAAAGGAGG ATAGTAAAGG AATACAGGTA AGCAAAATGA TACTAATGGC
6361 TCAACGTGAT AAGGAAAAAG AATTGCACCT TAACATTAAT ATTGACAAGG AGGAGGCGAC
6421 CACACAAAAA GTTAGGTGTA ACAGAAAAATC ATGAAACTAC GATTCTCAAT TTGATATTGG
6481 AGGATTTTCT CTAACAAAAA AAAAATACAA CAAATAAAAA ACACCTCAATG ACCTGACCAT
6541 TTGATGGAGT TTAAGTCAAT ACCTTCTTGA ACCATTTCCTC ATAATGGTGA AAGTTCCCTC
6601 AAGAATTTTA CTCGTGCAGA AACGGCCTTA CGACGTAGTC GATATGGTGC ACTCTCAGTA
6661 CAATCTGCTC TGATGCCGCA TAGTTAAGCC AGCCCCGACA CCCGCCAACA CCCGCTGACG
6721 CGCCCTGACG GGGTTGTCTG CTCCCGGCAT CCGCTTACAG ACAAGCTGTG ACCGCTCTCG
6781 GGAGCTGCAT GTGTCAGAGG TTTTCACCGT CATCACCGAA ACGCGCGAGA CGAAAGGGCC
6841 TCGTGATACG CCTATTTTTA TAGGTTAATG TCATGATAAT AATGGTTTCT TAGGACGGAT
6901 CGCTTGCTG TAACTTACAC GCGCCTCGTA TCTTTTAATG ATGGAATAAT TTGGGAATTT
6961 ACTCTGTGTT TATTTATTTT TATGTTTTGT ATTTGGATTT TAGAAAATAA ATAAAGAAGG
7021 TAGAAGAGTT ACGGAATGAA GAAAAAATAA TAAACAAAGG TTTAAAAAAT TTCAACAAAA
7081 AGCGTACTTT ACATATATAT TTATTAGACA AGAAAAGCAG ATTAATAAGA TATACATTGG
7141 ATTAACGATA AGTAAATGT AAAATCACAG GATTTTCGTG TGTGGTCTTC TACACAGACA
7201 AGATGAAACA ATTCGGCATT AATACCTGAG AGCAGGAAGA GCAAGATAAA AGGTAGTATT
7261 TGTGGCGAT CCCCTAGAG TCTTTTACAT CTTGGGAAAA CAAAACTAT TTTTCTTTTA
7321 ATTTCTTTT TACTTTCTA TTTTAAATTT ATATATTTAT ATTAATAAAT TTAATTTATA
7381 ATTATTTTTA TAGCAGTGA TGAAAAGGAC CCAGGTGGCA CTTTTCGGGG AAATGTGGCG
7441 GGAACCCCTA TTTGTTTTATT TTTCTAAATA CATTCAAATA TGTATCCGCT CATGAGACAA
7501 TAACCCCTGAT AAATGCTTCA ATAATCTGCA GCTCTGGCCC GTGTCTCAAA ATCTCTGATG
7561 TTACATTGCA CAAGATAAAA ATATATCATC ATGAACAATA AAATCTGTCTG CTTACATAAA
7621 CAGTAATACA AGGGGTGTTA TGAGCCATAT TCAACGGGAA ACGTCTTGCT GGAGGCCCGC
7681 ATTAATTTCC AACATGGATG CTGATTATA TGGGTATAAA TGGGCTCGCG ATAAATGTCG
7741 GCAATCAGGT GCGACAATCT TTCGATTGTA TGGGAAGCCC GATGCGCCAG AGTTGTTTCT
7801 GAAACATGGC AAAGGTAGCG TTGCCAATGA TGTATACAGT GAGATGGTCA GACTAAACTG
7861 GCTGACGGAA TTTATGCTC TTCGACCAT CAAGCATTTT ATCCGTACTC CTGATGATGC
7921 ATGGTTACTC ACCACTGCGA TCCGCGGGAA AACAGCATT CAGGTATTAG AAGAATATCC
7981 TGATTCAGGT GAAAATATTG TTGATGCGCT GGCAGTGTTC CTGCGCCGCT TGCATTCTGAT
8041 TCCTGTTTGT AATTGTCTTT TTAACAGCGA TCGCGTATTT CGTCTCGCTC AGGCGCAATC
8101 ACCAATGAAT AACGGTTTGG TTGATGCGAG TGATTTTGAT GACGAGCGTA ATGGCTGGCC
8161 TGTGAAACAA GTCTGGAAG AAATGCATAC GCTTTTGCCA TTCTCACCAG ATTCACTCGT
8221 CACTCATGGT GATTTCCTAC TTGATAACCT TATTTTGTAC GAGGGGAAAT TAATAGGTTG
8281 TATTGATGTT GGACGAGTCG GAATCGCAGA CCGATACCAG GATCTTGCCA TCCTATGGAA
8341 CTGCCTCGGT GAGTTTTTCTC CTTCAITACA GAAACGGCTT TTTCAAAAAT ATGGTATTGA
8401 TAATCCTGAT ATGAATAAAT TGCAGTTTCA TTTGATGCTC GATGAGTTTT TCTAATCAGA
8461 ATTGGTTAAT TGGTTGTAAC ACTGGCAGAG CATTACGCTG ACTTGACGGG ACGGCCCATG
8521 ACCAAATCC CTTAACGTGA GTTTTCGTTT CACTGAGCGT CAGACCCCGT AGAAAAGATC
8581 AAAGGATCTT CTTGAGATCC TTTTTTCTG CCGGTAATCT GCTGCTTGCA AACAAAAAAA
8641 CCACCGCTAC CAGCGGTGGT TTGTTTGCCG GATCAAGAGC TACCAACTCT TTTCCGAAG
8701 GTAACCTGGT TCAGCAGAGC GCAGATACCA AATACTGTCC TTCTAGTGTA GCCGTAGTTA
8761 GGCCACCACT TCAAGAACTC TGTAGCACCG CCTACATACC TCGCTCTGCT AATCCTGTTA
8821 CCAGTGGCTG CTGCCAGTGG CGATAAGTCG TGTCTTACCG GGTGGACTC AAGACCATAG
8881 TTACCGGATA AGGCGCAGCG GTCGGGCTGA ACGGGGGGTT CGTGACACA GCCAGCTTG
8941 GAGCGAACGA CCTACACCGA ACTGAGATAC CTACAGCGTG AGCATTGAGA AAGCGCCACG
9001 CTTCCCGAAG GGAGAAAGGC GGACAGGTAT CCGGTAAGCG GCAGGGTCGG AACAGGAGAG
9061 CGCAGGAGG AGCTTCCAGG GGGGAACGCC TGGTATCTTT ATAGTCTGTG CCGGTTTCGC
9121 CACCTCTGAC TTGAGCGTCG ATTTTGTGTA TGCTCGTCAG GGGGGCCGAG CCTATGGAAA
9181 AACGCCAGCA ACGCGGCCCT TTTACGGTTC CTGGCCTTTT GCTGGCTTTT TGCTCACATG
9241 TTCTTTCTG CGTTATCCCC TGATTCTGTG GATAACCGTA TTACCGCCTT TGAGTGAGCT
9301 GATACCGCTC GCCGCAGCGG AACGACCGAG CGCAGCGAGT CAGTGAGCGA GGAAGCGGAA
9361 GAGCGCCCAA TACGCAAAACC GCCTCTCCCC GCGCSTTGGC CGATTCAATTA ATGCAGCTGG-

FIGURE 4D

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9421 CACGACAGGT TTCCCGACTG GAAAGCGGGC AGTGAGCGCA ACGCAATTAA TGTGAGTTAC
9481 CTCACCTATT AGGCACCCCA GGCTTTACAC TTTATGCTTC CGGCTCCTAT GTTGTGTGGA
9541 ATTGTGAGCG GATAACAATT TCACACAGGA AACAGCTATG ACCATGATT A CGCCAAGCTC
9601 GGAATTAACC CTCACAAAG GGAACAAAAG CTGGTACCGA TCCCGAGCT T TGCAAATTAA
9661 AGCCTTCGAG CGTCCCAAAA CCTTCTCAAG CAAGGTTTT C AGTATAATGT TACATCGGTA
9721 CACGCGTCTG TACAGAAAAA AAAGAAAAAT TTGAAATATA AATAACGTT C TTAATACTAA
9781 CATAACTATA AAAAAATAAA TAGGGACCTA GACTTCAGGT TGTCTAACT C CTTCCTTTTC
9841 GGTTAGAGCG GATGTGGGGG GAGGGCGTGA ATGTAAGCGT GACATAACTA ATTACATGAT
9901 ATCGACAAAG GAAAAGGGGC CTGTTTACTC ACAGGCTTTT TTCAAGTAGG TAATTAAGTC
9961 GTTTCGTCT TTTTCTTCT TCAACCCACC AAAGGCCATC TTGGTACTTT TTTTTTTTTT
10021 TTTTTTTTTT TTTTTTTTTT TTTTTTTTTT TTTTTTTTTT TTTTTTTTTT TTTTTTTTTT
10081 TTTTTTTTTT TTTTTTTTTT TCATAGAAAT AATACAGAAG TAGATGTTGA ATTAGATTAA
10141 ACTGAAGATA TATAATTTAT TGGAAAAATAC ATAGAGCTTT TTGTTGATGC GCTTAAGCGA
10201 TCAATTCAAC AACACCACCA GCAGCTCTGA TTTTCTTTC AGCCAAGCTG GAGACGAATC
10261 TAGCTTTGAC GATACTGGA ACATTGGAA TTCTACCTT ACCCAAGATC TTACCGTAAC
10321 CGGCTGCCAA AGTGTCATA ACTGGAGCAG TTTCTTAGA AGCAGATTC AAGTATGGT
10381 CTCTCTTGTC TTCTGGGATC AATGTCCACA ATTTGTCCAA GTTCAAGACT GGCTTCCAGA
10441 AATGAGCTTG TTGCTTGTG AAGTATCTCA TACCAACCTT ACCGAAATAA CCTGGATGGT
10501 ATTTATCCAT GTTAATTCTG TGGTGATGTT GACCACCGGC CATACCTCTA CCACCGGGT
10561 GCTTCTGTG CTTACCGATA CGACCTTTAC CGGCTGAGAC GTGACCTCTG TGCTTTCTAG
10621 TCTTAGTGAA TCTGGAAGGC ATTCTTGATT AGTTGGATGA TTGTTCTGGG ATTTAATGCA
10681 AAAATCACTT AAGAAGGAAA ATCAACGGAG AAAGCAAACG CCATCTTAAA TATACGGGAT
10741 ACAGATGAAA GGGTTTGAAC CTATCTGGAA AATAGCATTA AACAAAGCGAA AAACCTGCGAG
10801 GAAAATTGTT TCGCTCTCTG CGGGCTATT C ACGCGCCAGA GGAAATAGG AAAAAATAA
10861 GGGCATTAGA AAAATAATTT TGATTTTGGT AATGTGTGGG TCCTGGTGTA CAGATGTTAC
10921 AATTGGTTACA GTACTCTGT TTTTGCTGTG TTTTTCGATG AATCTCCAAA ATGGTTGTTA
10981 GCACATGGAA GAGTCACCGA TGCTAAGTTA TCTCTATGTA AGCTACGTGG CGTGACTTTT
11041 GATGAAGCCG CACAAGAGAT ACAGGATTGG CAACTGCAAA TAGAATCTGG GGATCCCCC
11101 TCGAGATCCG GGATCGAAGA AATGATGGTA AATGAAATAG GAAATCAAGG AGCATGAAGG
11161 CAAAAGACAA ATATAAGGGT CGAACGAAAA ATAAAGTGAA AAGTGTGAT ATGATGTATT
11221 TGGCTTTGCG GCGCCGAAAA AACGAGTTTA CGCAATTGCA CAATCATGCT GACTCTGTGG
11281 CGGACCCGCG CTCTTGCCGG CCCGGCGATA ACGCTGGGCG TGAGGCTGTG CCCGGCGGAG
11341 TTTTTTGCGC CTGCATTTTC CAAGGTTTAC CTGCGCTAA GGGGCGAGAT TGGAGAAGCA
11401 ATAAGAATGC CGGTTGGGGT TCGATGATG ACGACCACGA CAACTGGTGT CATTATTTAA
11461 GTTGCCGAAA GAACCTGAGT GCATTTGCAA CATGAGTATA CTAGAAGAAT GAGCCAAGAC
11521 TTGCGAGACG CGAGTTTGCC GGTGGTGCGA ACAATAGAGC GACCATGACC TTGAAGGTGA
11581 GACGCGCATA ACCGCTAGAG TACTTTGAAG AGGAAACAGC AATAGGGTTG CTACCAGTAT
11641 AAATAGACAG GTACATACAA CACTGGAAAT GGTGTCTGT TTGAGTACGC TTTCAATTCA
11701 TTTGGGTGTG CAC

FIGURE 41E

Figure 42A:

PD3T22

2-Hybrid Vector with Activation Domain

557 acg gac act act ctc taa tga gca acg gta tac ggc ctt cct tcc agt tac
tgc ctg tga tga gag att act cgt tgc cat atg ccg gaa gga agg tca atg

708 ttg aat ttg aaa taa aaa aag ttt gcc gct ttg cta tca agt ata aat aga
aac tta aac ttt att ttt ttc aaa cgg cga aac gat agt tca tat tta tct

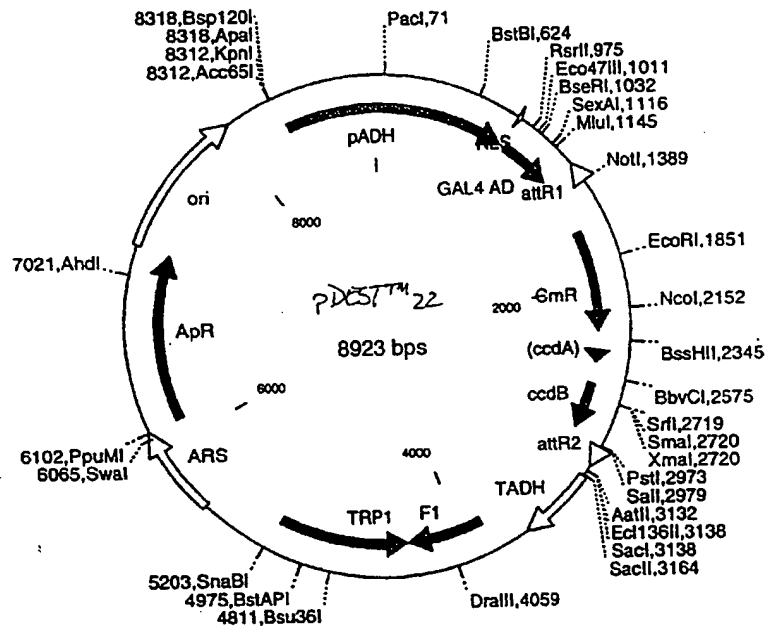
759 cct gca att att aat ctt ttg ttt ctt cgt cat tgt tct cgt tcc ctt tct
gga cgt taa taa tta gaa aac aaa gga gca gta aca aga gca agg gaa aga

810 ~~ccc ttg tct ctt ttt cgt cgt att att tca agt tat acc aag cat acc atc~~
~~ggc aac aag gaa aaa gac atg cta taa agt tgc ata tgg ttt gta tct tag~~

861 ~~aac tcc aag ctt atg ctc aag aag aag cgg aag gtc tgc agc ggc gcc aat~~
~~ttg agg ttc gaa tgc ggg ttc ttc ttc gcc ttc cag agc tgc ccg ggc atg~~

1218 gaa gat acc cca cca aac cca aaa aaa gag gg G G S N S T S
ctt cta tgg ggt ggt ttg ggt ttt ttt ctc cca ccc agc tta gtt tgt tca

1269 ~~ttg tac aaa aaa gct gaa cga gaa acg taa a~~
~~aac atg ttt ttt cga ctt gct ctt tgc att t~~



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pDEST22 8923 bp

Location (Base Nos.)	Gene Encoded
904..1248	GAL4 AD
1388..1264	attR1
1638..2297	CmR
2417..2501	inactivated ccdA
2639..2944	ccdB
2985..3109	attR2
3831..4318	f1 (f1 intergenic region)
4334..5176	TRP1
6110..7194	ampR
8344..866	pADH (yeast ADH promoter)

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1  TTCATTTGGG TGTGCACITT ATTATGTTAC AATATGGAAG GGAACITTTAC ACTTCTCCTA
61  TGCACATATA TTAATTAAAG TCCAATGCTA GTAGAGAAGG GGGGTAACAC CCCTCCGCGC
121 TCTTTTCCGA TTTTITTCCTA AACCGTGGAA TATTTCCGGAT ATCCTTTTGT TGTTTCCGGG
181 TGTACAATAT GGACTTCCTC TTTTCTGGCA ACCAAACCCA TACATCGGGA TTCTTATAAT
241 ACCTTCGTTG GTCTCCCTAA CATGTAGGTG GCGGAGGGGA GATATACAAT AGAACAGATA
301 CCAGACAAGA CATAATGGGC TAAACAAGAC TACACCAATT ACACTGCCTC ATTGATGGTG
361 GTACATAACG AACTAATACT GTAGCCCTAG ACTTGATAGC CATCATCATA TCGAAGTTTC
421 ACTACCCCTT TTCCATTGTC CATCTATTGA AGTAATAATA GGCGCATGCA ACTTCTTTTC
481 TTTTITTTTC TTTTCTCTCT CCCCCTTGTG TGTCTCACCA TATCCGCAAT GACAAAAAAA
541 ATGATGGAAG ACACATAAAGG AAAAAATTAA CGACAAGAC AGCACCAACA GATGTCGTTG
601 TTCCAGAGCT GATGAGGGGT ATCTTCGAAC ACACGAACT TTTTCTTCC TTCATTACG
661 CACACTACTC TCTAATGAGC AACGGTATAC GGCCTTCCTT CCAGTTACTT GAATTGAAA
721 TAAAAAAGT TTGCCGCTTT GCTATCAAGT ATAAATAGAC CTGCAATTAT TAATCTTTTG
781 TTTCTCTGTC ATTGTTCTCG TTCCCTTTCT TCCTTGTTTC TTTTCTGCA CAATATTCTA
841 AGCTATACCA AGCATACAAT CAACTCCAAG CTTATGCCCA AGAAGAAGCG GAAGGTCTCG
901 AGCGGCGCCA ATTTTAATCA AAGTGGGAAT ATTGCTGATA GCTCATGTG CTCCTCTTTC
961 ACTAACAGTA GCAACGGTCC GAACCTCATA ACAACTCAAA CAAATTCTCA AGCGCTTTCA
1021 CAACCAATTG CCTCCTCTAA CGTTCATGAT AACTTCATGA ATAATGAAAT CACGGCTAGT
1081 AAAATTGATG ATGGTAATAA TTCAAAACCA CTGTCACCTG GTTGGACGGA CCAAACTGCG
1141 TATAACGCGT TTGGAATCAC TACAGGGATG TTTAATACCA CTACAATGGA TGATGTATAT
1201 AACTATCTAT TCGATGATGA AGATACCCCA CAAACCCAA AAAAGAGGG TGGGTGCAAT
1261 CAAACAAGTT TGTACAAAAA AGCTGAACGA GAAACGTAAA ATGATATAAA TATCAATATA
1321 TTAATTAGA TTTTGATATA AAAACAGACT ACATAATACT GTAAAAACCA ACATATCCAG
1381 TCACTATGGC GGCCGCTAAG TTGGCAGCAT CACCCGACGC ACTTTGCGCC GAATAAATAC
1441 CTGTGACGGA AGATCACTTC GCAGAATAAA TAAATCCTGG TGTCCCTGTT GATACCGGGA
1501 AGCCCTGGGC CAACTTTTGG CGAAAATGAG ACGTTGATCG GCACGTAAGA GGTTCCAACT
1561 TTCACCATAA TGAAATAAGA TCACTACCGG GCGTATTTT TGAGTTATCG AGATTTTCAG
1621 GAGCTAAGGA AGCTAAAATG GAGAAAAAAA TCACTGGATA TACCACCGTT GATATATCCC
1681 AATGGCATCG TAAAGAACAT TTTGAGGCAT TTCAGTCAGT TGCTCAATGT ACCTATAACC
1741 AGACCGTTCA GCTGGATATT ACGGCCTTTT TAAAGACCGT AAAGAAAAAT AAGCACAAGT
1801 TTTATCCGGC CTTTATTCAC ATTCTTGCCC GCCTGATGAA TGCTCATCCG GAATCCCGTA
1861 TGGCAATGAA AGACGGTGAG CTGGTGATAT GGGATAGTGT TCACCCTTGT TACACCGTTT
1921 TCCATGAGCA AACTGAAACG TTTTCATCGC TCTGGAGTGA ATACCACGAC GATTTCCGGC
1981 AGTTTCTACA CATATATTCT CAAGATGTGG CGTGTACGG TGAACACCTG GCCTATTTC
2041 CTAAGGGTT TATTGAGAAT ATGTTTTTCG TCTCAGCCAA TCCCTGGGTG AGTTTACCA
2101 GTTTTGATTT AAACGTGGCC AATATGGACA ACTTCTTCGC CCCGTTTTC ACCATGGGCA
2161 AATATTATAC GCAAGCGGAC AAGGTGCTGA TGCCGCTGGC GATTCAGGTT CATCATGCCG
2221 TCTGTGATGG CTTCCATGTC GGCAGAATGC TTAATGAATT ACAACAGTAC TGGCATGAGT
2281 GGCAGGGCGG GGCCTAATCT AGAGGATCCG GCTTACTAAA AGCCAGATAA CAGTATGCGT
2341 ATTTGCGCGC TGATTTTTCG GGTATAAGAA TATATACTGA TATGTATACC CGAAGTATGT
2401 CAAAAAGAGG TGTGCTATGA AGCAGCGTAT TACAGTGACA GTTGACAGCG ACAGCTATCA
2461 GTTGCTCAAG GCATATATGA TGTCAATATC TCCGGTCTGG TAAGCACAAAC CATGCAGAAT
2521 GAAGCCCGTC GTCTGCGTGC CGAACGCTGG AAAGCGGAAA ATCAGGAAGG GATGGCTGAG-

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FIGURE 42B

2581 GTCGCCCGGT TTATTGAAAT GAACGGCTCT TTTGCTGACG AGAACAGGGA CTGGTGAAAT
 2641 GCAGTTTAAG GTTTACACCT ATAAAAGAGA GAGCCGTTAT CGTCTGTTT TGGATGTACA
 2701 GAGTGATATT ATTGACACGC CCGGGCGACG GATGGTGATC CCCCTGGCCA GTGCACGTCT
 2761 GCTGTCAGAT AAAGTCTCCC GTGAACTTTA CCCGGTGGTG CATATCGGG ATGAAAGCTG
 2821 GCGCATGATG ACCACCGATA TGGCCAGTGT GCCCGTCTCC GTTATCGGG AAGAAGTGGC
 2881 TGATCTCAGC CACCGCGAAA ATGACATCAA AAACGCCATT AACCTGATGT TCTGGGGAAT
 2941 ATAAATGTCA GGCTCCCTTA TACACAGCCA GTCTGCAGGT CGACCATAGT GACTGSATAT
 3001 GTTGTGTTTT ACAGTATTAT GTAGTCTGTT TTTTATGCAA AATCTAATTT AATATATTGA
 3061 TATTTATATC ATTTTACGTT TCTCGTTCAG CTTTCTTGTA CAAAGTGGTT TGATGGCCGC
 3121 TAAGTAAGTA AGACGTCGAG CTCTAAGTAA GTAACGGCCG CCACCGCGGT GGAGCTTTGG
 3181 ACTTCTTCGC CAGAGGTTTG GTCAAGTCTC CAATCAAGGT TGTCCGCTTG TCTACCTTGC
 3241 CAGAAATTTA CGAAAAGATG GAAAAGGGTC AAATCGTTGG TAGATACGTT GTTGACACTT
 3301 CTAATAAGC GAATTTCTTA TGATTTATGA TTTTATTAT TAAATAAGTT ATAAAAAATA
 3361 TAAGTGATA CAAATTTTAA AGTGACTCTT AGGTTTAAA ACGAAAATTC TTATTTCTGA
 3421 GTAACCTCTT CCTGTAGGTC AGGTTGCTTT CTCAGGTATA GCATGAGGTC GCTCTTATTG
 3481 ACCACACCTC TACCGGCAAT CCGAGCAAAT GCCTGCAAAT CGCTCCCAT TTCACCCAAAT
 3541 TGTAGATATG CTAAGTCCAG CAATGAGTTG ATGAATCTCG GTGTGTATTT TATGTCTCA
 3601 GAGGACAATA CCTGTTGTAA TCGTCTTCC ACACGGATCC CAATTCGCCC TATAGTGAGT
 3661 CGTATTACAA TTCACTGGCC GTCTGTTTAC AACGTCGTGA CTGGGAAAAC CCTGGCGTTA
 3721 CCCAACTTAA TCGCCTTGCA GCACATCCCC CTTTCGCCAG CTGGCGTAAT AGCGAAGAGG
 3781 CCCGCACCGA TCGCCCTTCC CAACAGTTGC GCAGCCTGAA TGGCGAATGG ACGCGCCCTG
 3841 TAGCGCGCA TTAAGCGCGG CCGGTGTGGT GGTACGCGC AGCGTGACCG CTACACTTGC
 3901 CAGCGCCCTA GCGCCCGCTC CTTTCGCTTT CTTCCCTTCC TTTCTCGCCA CGTTCGCCGG
 3961 CTTTCCCGGT CAAGCTCTAA ATCGGGGGCT CCCTTTAGGG TTCCGATTTA GTGCTTTACG
 4021 GCACCTCGAC CCCAAAAAAC TTGATTAGGG TGATGGTTCA CGTAGTGGGC CATCGCCCTG
 4081 ATAGACGGTT TTTCCGCCCT TGACGTTGGA GTCCACGTTT TTTAATAGTG GACTCTTGTT
 4141 CCAAACTGGA ACAACACTCA ACCCTATCTC GGTCTATTCT TTTGATTTAT AAGGGATTTT
 4201 GCGGATTTTG GCCTATTGGT TAAAAAATGA GCTGATTTAA CAAAAATTTA ACGCGAATTT
 4261 TAACAAAATA TTAACGTTTA CAATTTCTCT ATGCGGTATT TTCTCCTTAC GCATCTGTGC
 4321 GGTATTTCAC ACCGCAAGCA AGTGCACAAA CAATACTTAA ATAAATACTA CTCAGTAATA
 4381 ACCTATTTCT TAGCATTTT GACGAAATTT GCTATTTGT TAGAGTCTT TACACCATTT
 4441 GTCTCCACAC CTCCGCTTAC ATCAACACCA ATACGCCAT TTAATCTAAG CGCATCACCA
 4501 ACATTTTCTG GCGTCACTCC ACCAGCTAAC ATAAATGTA AGCTTTCGGG GCTCTCTTGC
 4561 CTTCCAAACC AGTCAGAAAT CGAGTTCCAA TCCAAAAGTT CACCTGTCCC ACCTGCTTCT
 4621 GAATCAACA AGGGAATAAA CGAATGAGGT TTCTGTGAAG CTGCACTGAG TAGTATGTTG
 4681 CAGTCTTTTG GAAATACGAG TCTTTTAATA ACTGGCAAA CAGGGAATC TTGGTATTCT
 4741 TGCCACGACT CATCTCCATG CAGTTGGACG ATATCAATGC CGTAATCATT GACCAGAGCC
 4801 AAAACATCCT CCTTAGGTTG ATTACGAAAC ACGCAACCA AGTATTTCCG AGTGCCTGAA
 4861 CTATTTTAT ATGCTTTTAC AAGACTTGAA ATTTTCTTTC CAATAACCGG GTCAATTGTT
 4921 CTCTTTCTAT TGGGCACACA TATAATACCC AGCAAGTCAG CATCGGAATC TAGAGCACAT
 4981 TCTGCGGCT CTGTGCTCTG CAAGCCGCAA ACTTTCACCA ATGGACCAGA ACTACCTGTG
 5041 AAATTAATAA CAGACATACT CCAAGCTGCC TTTGTGTGCT TAATCAGGTA TACTCAGGTG
 5101 CTCAATAGTC ACCAATGCCC TCCCTCTTGG CCCTCTCCTT TTTCTTTTTC GACCGAATTA
 5161 ATTCTTAATC GGCAAAAAA GAAAAGCTCC GGATCAAGAT TGACGTAAG GTGACAAGCT
 5221 ATTTTCAAT AAGAATATC TTCCACTACT GCCATCTGGC GTCATAACTG CAAAGTACAC
 5281 ATATATTACG ATGCTGTCTA TTAATGCTT CCTATATTAT ATATATAGTA ATGTCGTTTA
 5341 TGGTGCACTC TCAGTACAAT CTGCTCTGAT GCCGCATAGT TAAGCCAGCC CCGACACCCG
 5401 CCAACACCCG CTGACGCGCC CTGACGGGCT TGTCTGTCTC CGGCATCCGC TTACAGACAA
 5461 GCTGTGACCG TCTCCGGGAG CTGCATGTGT CAGAGGTTT CACCGTCATC ACCGAAACGC
 5521 GCGAGACGAA AGGGCCTCGT GATACGCCTA TTTTATAGG TTAATGTCAT GATAATAATG
 5581 GTTCTTAGG ACGGATCGCT TGCTGTAAAC TTACACGCGC CTCGTATCTT TTAATGATGG
 5641 AATAATTGGA GAATTTACTC TGTGTTTAT TATTTTATG TTTTGTATT GGATTTTAGA
 5701 AAGTAATAA AGAAGGTAGA AGAGTTACGG AATGAAGAAA AAAAAATAA CAAAGGTTTA
 5761 AAAAAATTCA AAAAAAGCG TACTTTACAT ATATATTAT TAGACAAGAA AAGCAGATTA
 5821 AATAGATATA CATTCGATTA ACGATAAGTA AAATGTAAAA TCACAGGATT TTCTGTGTG
 5881 GTCTTCTACA CAGACAAGAT GAAACAATTC GGCATTAATA CCTGAGAGCA GGAAGAGCAA
 5941 GATAAAGGT AGTATTGTT GCGCATCCCC CTAGAGTCTT TTACATCTTC GGAAAACAAA
 6001 AACTATTTTT TCTTTAATTT CTTTTTTTAC TTTCTATTTT TAAATTATAT ATTTATATTA-

FIGURE 42C

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6061 AAAAATTAA ATTATAATTA TTTTATAGC ACGTGATGAA AAGGACCCAG GTGGCACTTT
6121 TCGGGGAAAT GTGCGCGGAA CCCCTATTG TTTATTTTC TAAATACATT CAAATATGTA
6181 TCCGCTCATG AGACAATAAC CCTGATAAAT GCTTCAATAA TATTGAAAAA GGAAGAGTAT
6241 GAGTATTCAA CATTTCCTG TCGCCCTTAT TCCCTTTTTT GCGGCATTTT GCCTTCCTGT
6301 TTTTGCTCAC CCAGAAACGC TGGTGAAAGT AAAAGATGCT GAAGATCAGT TGGGTGCACG
6361 AGTGGGTAC ATCGAACTGG ATCTCAACAG CGGTAAGATC CTTGAGAGIT TTCGCCCGA
6421 AGAACGTTTT CCAATGATGA GCACITTTAA AGTTCTGCTA TGTGCGCGG TATTATCCCG
6481 TATTGACGCC GGGCAAGAGC AACTCGGTCG CCGCATACAC TATTCTCAGA ATGACTTGGT
6541 TGAGTACTCA CCAGTCACAG AAAAGCATCT TACGGATGGC ATGACAGTAA GAGAATTATG
6601 CAGTGCTGCC ATAACCATGA GTGATAACAC TCGGCCAAC TTACTTCTGA CAACGATCGG
6661 AGGACCGAAG GAGCTAACCG CTTTTTTTCA CAACATGGGG GATCATGTAA CTCGCCTTGA
6721 TCGTTGGGAA CCGGAGCTGA ATGAAGCCAT ACCAAACGAC GAGCGTGACA CCACGATGCC
6781 TGAGCAATG GCAACAACGT TCGCGAAACT ATTAACCTGG GAACTACTTA CTCTAGCTTC
6841 CCGGCAACAA TTAATAGACT GGTGAGAGG GATAAAGTT GCAGGACCAC TTCTGCGCTC
6901 GGCCTTCCG GCTGGCTGGT TTATGCTGA TAAATCTGGA GCCGGTGAGC GTGGGTCTCG
6961 CGGTATCATT GCAGCACTGG GGCAGATGG TAAGCCCTCC CGTATCGTAG TTATCTACAC
7021 GACGGGCACT CAGGCAACTA TGGATGAACG AAATAGACAG ATCGCTGAGA TAGGTGCTCT
7081 ACTGATTAAG CATTGGTAAC TGTCAGACCA AGTTTACTCA TATATACTTT AGATTGATTT
7141 AAAACTTCAT TTTTAATTAA AAAGGATCTA GGTGAAGATC CTTTTTGATA ATCTCATGAC
7201 CAAAATCCCT TAACGTGAGT TTTCTGTTCA CTGAGCGTCA GACCCCGTAG AAAAGATCAA
7261 AGGATCTTCT TGAGATCCTT TTTTCTGCG CGTAATCTGC TGCTTGCAAA CAAAAAACC
7321 ACCGCTACCA GCGGTGGTTT GTTTGCCGGA TCAAGAGCTA CCAACTCTTT TTCCGAAGGT
7381 AACTGGCTTC AGCAGAGCGC AGATACCAAA TACTGTCTTT CTAGTGTAGC CGTAGTTAGG
7441 CCACCACTTC AAGAACTCTG TAGCACCGCC TACATACCTC GCTCTGTAA TCCTGTTACC
7501 AGTGGCTGCT GCCAGTGGCG ATAAGTCTGT TCTTACCGGG TTGGACTCAA GACGATAGTT
7561 ACCGATAAG GCGCAGCGGT CGGGCTGAAC GGGGGGTTTCG TGCACACAGC CCAGCTTGGA
7621 GCGAACGACC TACACCGAAC TGAGATACCT ACAGCGTGAG CATTGAGAAA CCGCCACGCT
7681 TCCCGAAGGG AGAAAGGCGG ACAGGTATCC GGTAAAGCGG AGGGTCGGAA CAGGAGAGCG
7741 CACGAGGGAG CTTCCAGGGG GGAACGCTTG GTATCTTTAT AGTCCTGTGC GGTTCGCCA
7801 CCTCTGACTT GAGCGTCGAT TTTTGTGATG CTCGTGAGG GGGCCGAGCC TATGAAAAA
7861 CGCCAGCAAC GCGGCTTTT TACGGTTCCT GGCCTTTTGC TGGCCTTTTG CTCACATGTT
7921 CTTTCTGCG TTATCCCTTG ATTCTGTGGA TAACCGTATT ACCGCTTTG AGTGAGCTGA
7981 TACCCTGCG CGCAGCCGAA CGACCGAGCG CAGCGAGTCA GTGAGCGAGG AAGCGGAAGA
8041 GCGCCCAATA CGCAAACCGC CTCTCCCGCG GCGTTGGCCG ATTCATTAAT GCAGCTGGCA
8101 CGACAGGTTT CCCGACTGGA AAGCGGGCAG TGAGCGCAAC GCAATTAATG TGAGTTACCT
8161 CACTCATTAG GCACCCAGG CTTTACACTT TATGCTTCCG GCTCCTATGT TGTGTGGAAT
8221 TGTGAGCGGA TAACAATTC ACACAGGAAA CAGCTATGAC CATGATTACG CCAAGCTCGG
8281 AATTAAACCT CACTAAAGGG AACAAAAGCT GGGTACCGGG CCCCCCTCG AGATCCGGGA
8341 TCGAAGAAAT GATGCTAAT GAAATAGGAA ATCAAGGAGC ATGAAGGCAA AAGACAAATA
8401 TAAGGGTCGA ACGAAAAATA AAGTGAAAAA TGTGATATG ATGTATTGG CTTTGGCGCG
8461 CCGAAAAAAC GAGTTTACGC AATTGCACAA TCATGCTGAC TCTGTGGCGG ACCCGCGCTC
8521 TTGCGGCGCC GCGGATAACG CTGGGCGTGA GGCTGTGCCG GCGGAGTTT TTTGCGCTG
8581 CATTTTCCAA GGTTTACCCT GCGCTAAGGG GCGAGATTGG AGAAGCAATA AGAATGCCGG
8641 TTGGGGTTGC GATGATGACG ACCACGACAA CTGGTGTCAT TATTAAAGTT GCCGAAAGAA
8701 CCTGAGTGCA TTTGCAACAT GAGTATACTA GAAGAATGAG CCAAGACTTG CGAGACGCGA
8761 GTTTGCCCGT GGTGCGAACA ATAGAGCGAC CATGACCTTG AAGGTGAGAC GCGCATAACC
8821 GCTAGAGTAC TTTGAAGAGG AAACAGCAAT AGGGTTGCTA CCAGTATAAA TAGACAGGTA
8881 CATACAACAC TGGAAATGGT TGTCTGTTTG AGTACGCTTT CAA

Figure 42d

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pDEST23

His6 carboxy-fusion vector, T7 promoter

205 atc ccg cga aat taa tac gac tca cta tag gga gat cac aac ggc tto oct att atg ctg agt gat atc cgt ctg gtg ttg cca aag gga cta gat cgc aag ttt gta caa aaa agc tga acg aga aac gta aaa tga tat gat cta ctg ttc aaa cat gtt ttt tca act tgc tot ttg cat ttt act ata

256 cta gat cgc aag ttt gta caa aaa agc tga acg aga aac gta aaa tga tat gat cta ctg ttc aaa cat gtt ttt tca act tgc tot ttg cat ttt act ata

1888 ttt tta tgc aaa atc taa ttt aat ata ttg ata ttt ata tca ttt tac gtt
 aaa aat acg ttt tag att aaa tta tat aac tat aaa tat agt aaa atg caa
 1939 tct cgt tca gct ttg ttg tac aaa gtg gtg att atg tgc tac tac cat cac
aga gca agt cga aag aac atg ttt cac cac taa tac agc atg atg gta gtg
 1990 cat cab cat cac ctc gat gag caa taa cta gca taa ccc ctt ggg gcc tct
gta gtg gta gtg gag cta ctc gtt att gat cgt att ggg gaa ccc cgg aga

// ————— Cm^R ————— ccd B ————— //

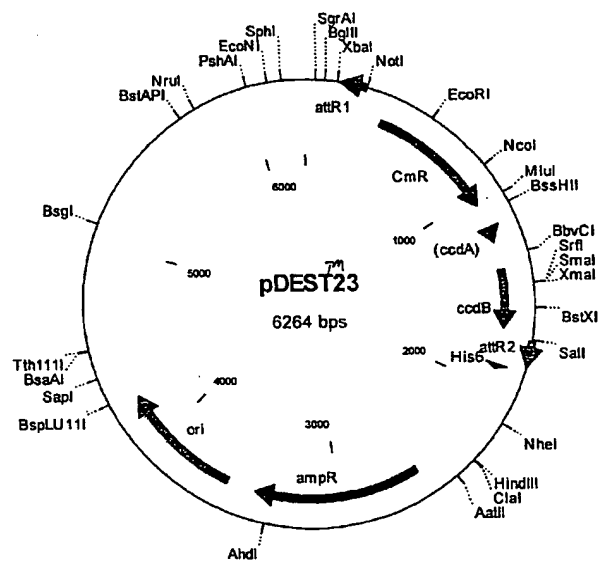


FIGURE 43A

pDEST23 6264 bp

Location (Base Nos.)	Gene Encoded
285..161	attR1
394..1053	CmR
1173..1257	inactivated ccdA
1395..1700	ccdB
1741..1865	attR2
1883..1911	his6
2574..3434	ampR
3583..4222	ori

1	TCTTCCCAT	CGGTGATGTC	GGCGATATAG	GCGCCAGCAA	CCGCACCTGT	GGCGCCGGTG
61	ATGCCCGCCA	CGATGCGTCC	GGCGTAGAGG	ATCGAGATCT	CGATCCCGCG	AAATTAATAC
121	GACTCACTAT	AGGGAGACCA	CAACGGTTTC	CCTCTAGATC	ACAAGTTTGT	ACAAAAAGC
181	TGAACGAGAA	ACGTAAATG	ATATAAATAT	CAATATATTA	AATTAGATTT	TGCATAAAAA
241	ACAGACTACA	TAATACTGTA	AAACACAACA	TATCCAGTCA	CTATGGCGGC	CGCATTAGGC
301	ACCCAGGCT	TTACACTTTA	TGCTCCGGC	TCGTATAATG	TGTGGATTTT	GAGTTAGGAT
361	CCGGCGAGAT	TTTCAGGAGC	TAAGGAAGCT	AAAATGGAGA	AAAAATCAC	TGGATATACC
421	ACCGTTGATA	TATCCCAATG	GCATCGTAAA	GAACATTTTG	AGGCATTTC	GTCAGTTGCT
481	CAATGTACCT	ATAACCAGAC	CGTTCAGCTG	GATATTACGG	CCTTTTAA	GACCGTAAAG
541	AAAAATAAGC	ACAAGTTTFA	TCCGGCCTTT	ATTCACATTC	TTGCCCGCCT	GATGAATGCT
601	CATCCGGAAT	TCCGTATGGC	AATGAAAGAC	GGTGAGCTGG	TGATATGGGA	TAGTGTTCAC
661	CCTTGTTACA	CCGTTTTCCA	TGAGCAAAT	GAAACGTTTT	CATCGCTCTG	GAGTGAATAC
721	CACGACGATT	TCCGGCAGTT	TCTACACATA	TATTCGCAAG	ATGTGGCGTG	TTACGGTGAA
781	AACCTGGCCT	ATTTCCCTAA	AGGGTTTATT	GAGAATATGT	TTTTCTCTC	AGCCAATCCC
841	TGGGTGAGTT	TCACCAGTTT	TGATTTAAAC	GTGGCCAATA	TGGACAACCT	CTTCGCCCCC
901	GTTTTCACCA	TGGGCCAAATA	TTATACGCAA	GGCGACAAGG	TGCTGATGCC	GCTGGCGGAT
961	CAGGTTTCATC	ATGCCGTCTG	TGATGGCTTC	CATGTCGGCA	GAATGCTTAA	TGAATTACAA
1021	CAGTACTGCG	ATGAGTGGCA	GGGCGGGGCG	TAAACGCGTG	GATCCGGCTT	ACTAAAAAGCC
1081	AGATAACAGT	ATGCGTATTT	GCGCGCTGAT	TTTTGCGGTA	TAAGAATATA	TACTGATATG
1141	TATACCCGAA	GTATGTCAAA	AAGAGGTGTG	CTATGAAGCA	GCGTATTACA	GTGACAGTTG
1201	ACAGCGACAG	CTATCAGTTG	CTCAAGGCAT	ATATGATGTC	AATATCTCCG	GTCTGGTAAG
1261	CACAACCATG	CAGAAATGAAG	CCCGTCGTCT	GCGTGCCGAA	CGCTGGAAAG	CGGAAAAATCA
1321	GGAAGGGATG	GCTGAGGTG	CCCGGTTTAT	TGAAATGAAC	GGCTCTTTTG	CTGACGAGAA
1381	CAGGGACTGG	TGAAATGCAG	TTTAAGGTTT	ACACCTATAA	AAGAGAGAGC	CGTTATCGTC
1441	TGTTTGTTGA	TGTACAGAGT	GATATTATTG	ACACGCCCGG	GCGACGGATG	GTGATCCCCC
1501	TGGCCAGTGC	ACGTCTGCTG	TCAGATAAAG	TCTCCCGTGA	ACTTTACCCG	GTGGTGCATA
1561	TCGGGGATGA	AAGCTGGCGC	ATGATGACCA	CCGATATGGC	CAGTGTGCCG	GTCTCCGTTA
1621	TCGGGGAAGA	AGTGGCTGAT	CTCAGCCACC	GCGAAAATGA	CATCAAAAAC	GCCATTAAAC
1681	TGATGTTCTG	GGGAATATAA	ATGTCAGGCT	CCCTTATACA	CAGCCAGTCT	GCAGGTCGAC
1741	CATAGTGACT	GGATATGTTG	TGTTTTACAG	TATTATGTAG	TCTGTTTTTT	ATGCAAAATC
1801	TAATTTAATA	TATTGATATT	TATATCATTT	TACGTTTCTC	GTTTCTGCTT	CTTGTACAAA
1861	GTGGTGATTA	TGTCGTACTA	CCATCACCAT	CACCATCACC	TCGATGAGCA	ATAACTAGCA
1921	TAACCCCTTG	GGGCCCTCTA	ACGGGTCTTG	AGGGGTTTTT	TGCTGAAAGG	AGGAAGTATA
1981	TCCGGATATC	CACAGGACGG	GTGTGCTCGC	CATGATCGCG	TAGTCGATAG	TGGCTCCAAG
2041	TAGCGAAGCG	AGCAGGACTG	GGCGGCGGCC	AAAGCGGTG	GACAGTGCTC	CGAGAACGGG
2101	TGCGCATAGA	AATTGCATCA	ACGCATATAG	CGCTAGCAGC	ACGCCATAGT	GACTGGCGAT
2161	GCTGTCGGAA	TGGACGATAT	CCCGCAAGAG	GCCCGGCAGT	ACCGGCATAA	CCAAGCCTAT
2221	GCCTACAGCA	TCCAGGGTGA	CGGTGCCGAG	GATGACGATG	AGCGCATTTG	TAGATTTCAT
2281	ACACGGTGCC	TGACTGCGTT	AGCAATTTAA	CTGTGATAAA	CTACCGCATT	AAAGCTTTATC
2341	GATGATAAGC	TGTCAAACAT	GAGAATTTCT	GAAGAAGGAA	GGGCCTCGTG	ATACGCCCTAT
2401	TTTTATAGGT	TAATGTCATG	ATAATAATGG	TTTCTTAGAC	GTCAGGTGGC	ACTTTTCGGG
2461	GAAATGTGCG	CGGAACCCCT	ATTTGTTTAT	TTTTCTAAAT	ACATTCAAAT	ATGTATCCGC
2521	TCATGAGACA	ATAACCCCTGA	TAAATGCTTC	AATAATATTG	AAAAAGGAAG	AGTATGAGTA
2581	TTCAACATTT	CCGTGTCGCC	CTTATCCCT	TTTTTGCGGC	ATTTTGCTT	CCTGTTTTCG
2641	CTCACCCAGA	AACGCTGGTG	AAAGTAAAAG	ATGCTGAAGA	TCAGTTGGGT	GCACGAGTGG

FIGURE 43B

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2701 GTTACATCGA ACTGGATCTC AACAGCGGTA AGATCCTTGA GAGTTTTTCG CCCCAGAAGAAC
2761 GTTTTCCAAT GATGAGCACT TTAAAGTTC TGCTATGTGG CGCGGTATTA TCCCGTGTG
2821 ACGCCGGGCA AGAGCAACTC GGTGCGCGCA TACACTATTC TCAGAAATGAC TTGGTTGAGT
2881 ACTCACCAGT CACAGAAAAG CATCTTACGG ATGGCATGAC AGTAAGAGAA TTATGCAAGT
2941 CTGCCATAAC CATGAGTGAT AACACTGCGG CCAACTTACT TCTGACAACG ATCGGAGGAC
3001 CGAAGGAGCT AACCGCTTTT TTGCACAACA TGGGGGATCA GTAACTCGC CTTGATCGTT
3061 GGGAAACCGA GCTGAATGAA GCCATACCAA ACGACGAGCG TGACACCACG ATGCCTGCAG
3121 CAATGGCAAC AACGTTGCGC AAACATTTAA CTGGCGAACT ACTTACTCTA GCTTCCCGGC
3181 AACAAATTAAT AGACTGGATG GAGGCGGATA AAGTTGCAGG ACCACTTCTG CGCTCGGCCC
3241 TTCCGGCTGG CTGGTTTATT GCTGATAAAT CTGGAGCCGG TGAGCGTGGG TCTCGCGGTA
3301 TCATTGCAGC ACTGGGGCCA GATGGTAAGC CCTCCCGTAT CGTAGTTATC TACACGACGG
3361 GGAGTCAGGC AACTATGGAT GAACGAAATA GACAGATCGC TGAGATAGGT GCCTCACTGA
3421 TTAAGCATTG GTAACGTGCA GACCAAGTTT ACTCATATAT ACTTTAGATT GATTTAAAC
3481 TTCATTTTAA ATTTAAAGG ATCTAGGTGA AGATCCTTTT TGATAATCTC ATGACCAAAA
3541 TCCCTTAACG TGAGTTTTCG TTCCACTGAG CGTCAGACCC CGTAGAAAAG ATCAAAAGGAT
3601 CTCTCTGAGA TCCCTTTTTT CTGCGCGTAA TCTGCTGCTT GCAAACAAA AAACCAACCG
3661 TACCAGCGGT GGTGTTGTTG CCGGATCAAG AGCTACCAAC TCTTTTTCCG AAGGTAACGT
3721 GCTTCAGCAG AGCGCAGATA CCAATACTG TCCCTCTAGT GTAGCCGTAG TTAGGCCACC
3781 ACTTCAAGAA CTCTGTAGCA CCGCCTACAT ACCTCGCTCT GCTAATCCTG TTACCACTGG
3841 CTGCTGCCAG TGGCGATAAG TCGTGTCTTA CCGGGTTGGA CTCAAGACGA TAGTTACCGG
3901 ATAAGGCGCA GCGGTCGGGC TGAACGGGGG GTTCGTGCAC ACAGCCACAG TTGGAGCGAA
3961 CGACCTACAC CGAACTGAGA TACCTACAGC GTGAGCTATG AGAAAGCGCC ACGCTTCCCG
4021 AAGGGAGAAA GCGCGACAGG TATCCGGTAA CCGCGAGGGT CGGAACAGGA GAGCGCACGA
4081 GGGAGCTTCC AGGGGAAAC GCCTGGTATC TTTATAGTCC TGTCGGGTTT CGCCACCTCT
4141 GACTTGAGCG TCGATTTTGG TGATGCTCGT CAGGGGGGCG GAGCCTATGG AAAAACGCCA
4201 GCAACGCGGC CTTTTTACGG TTCTTGGCCT TTTGCTGACC TTTTGTCTAC ATGTTCTTTC
4261 CTGCGTTATC CCGTGATTCT GTGATAACC GTATTACCGC CTTTGAGTGA GCTGATACCG
4321 CTCGCCGAG CCGAACGACC GAGCGCAGCG AGTCAGTGAG CGAGGAAGCG GAAGAGCGCC
4381 TGATGCGGTA TTTTCTCCTT ACGCATCTGT GCGGTATTTT ACACCGCATA TATGGTGCAC
4441 TCTCAGTACA ATCTGCTCTG ATGCCGCATA GTTAAGCCAG TATACACTCC GCTATCGCTA
4501 CGTGACTGGG TCATGGCTGC GCCCGACAC CGGCCACAC CCGCTGACGC CCGCTGACGG
4561 GCTTGTCTGC TCCCGGCATC CGCTTACAGA CAAGCTGTGA CCGTCTCCGG GAGCTGCATG
4621 TGTGAGAGGT TTTCACCGTC ATCACCAGAA CGCGCGAGGC AGCTGCGGTA AAGTCAATCA
4681 GCGTGGTCGT GAAGCGATTG ACAGATGTCT GCCTGTTTCT CCGGCTCCAG CTCGTTGAGT
4741 TTCTCCAGAA CGCTTAATGT CTGGCTTCTG ATAAAGCGGG CCAATGTTAA GCGGTTTTTT
4801 TCCTGTTTGG TCACTGATGC CTCCGTGTAA GGGGGATTTC TGTTCAATGG GGTAAATGATA
4861 CCGATGAAAC GAGAGAGGAT GCTCAGATA CCGGTTACTG ATGATGAACA TGCCCGGTTA
4921 CTGGAACGTT GTGAGGGTAA ACAACTGGCG GTATGGATGC GCGGGGACCA GAGAAAAATC
4981 ACTCAGGGTC AATGCCAGCG CTTCGTTAAT ACAGATGTAG GTGTTCCACA GGGTAGCCAG
5041 CAGCATCCTG CGATGCAGAT CCGGAACATA ATGGTGACAG GCGCTGACTT CCGGTTTTCC
5101 AGACTTTACG AAACACGGAA ACCGAAGACC ATTCATGTTG TTGCTCAGGT CGCAGACGTT
5161 TTGCAGCAGC AGTCGCTTCA CGTTCGCTCG CGTATCGGTG ATTCATTCTG CTAACCAATA
5221 AGGCAACCCC GCCAGCCTAG CCGGGTCTCT AACGACAGGA GCACGATCAT GCGCACCCGT
5281 GGCAGGAGAC CAACGCTGCC CGAGATGCGC CGCGTCCGGC TGCTGGAGAT GCGGAGCGCG
5341 ATGGATATGT TCTGCCAAGG GTTGGTTTGC GCATTACAG TTCTCCGCAA GAATTGATTG
5401 GCTCCAATTC TTGGAGTGGT GAATCCGTTA GCGAGGTGCC GCCGGCTTCC ATTCAGGTCT
5461 AGGTGGCCCG GCTCCATGCA CCGCGACGCA ACGCGGGGAG GCAGACAAGG TATAGGGCGG
5521 CGCCTACAAT CCAATGCCAAC CGGTTCCATG TGCTCGCCGA GCGGCATATA ATCCGCGTGA
5581 CGATCAGCGG TCCAGTGATC GAAGTTAGGC TGGTAAGAGC CGCGAGCGAT CCTTGAAGCT
5641 GTCCTGATG GTCGTCTCTT ACCTGCTTGG ACAGCATGGC CTGCAACCGG GGCATCCCGA
5701 TGCCGCCGGA AGCGAGAAGA ATCATAATGG GGAAGGCCAT CCAGCCTCGC GTCGCGAAGC
5761 CCAGCAAGAC GTAGCCGAGC GCGTCGCGCG CCATGCCGCG GATAATGGCC TGCTTCTCGC
5821 CGAAACGTTT GGTGGCGGGA CCAATGACGA AGGCTTGAGC GAGGGCGTGC AAGATTCGGA
5881 ATACCGCAAG CGACAGGCGG ATCATCGTGC CGCTCCAGCG AAAGCGGTCC TCGCCGAAAA
5941 TGACCCAGAG CGCTGCCGCG ACCTGTCCTA CGAGTTGCAT GATAAAGAA ACAGTCATAA
6001 GTGCGCGGAC GATAGTCATG CCCCGCGCCC ACCGGAAGGA GCTGACTGGG TTGAAGGCTC
6061 TCAAGGGCAT CGGTGATCG ACGCTCTCCC TTATGCGACT CTGCTATTAG GAAGCAGCCC
6121 AGTAGTAGGT TGAGGCGGTT GAGCACCGCC GCCGCAAGGA ATGGTGCATG CAAGGAGATG

FIGURE 43C

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6181 GCGCCCAACA GTCCCCCGGC CACGGGGCCT GCCACCATAC CCACGCCGAA ACAAGCGCTC
6241 ATGAGCCCGA AGTGGCGAGC CCGA

FIGURE 43D

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pDEST24
GST carboxy-fusion vector, T7 promoter

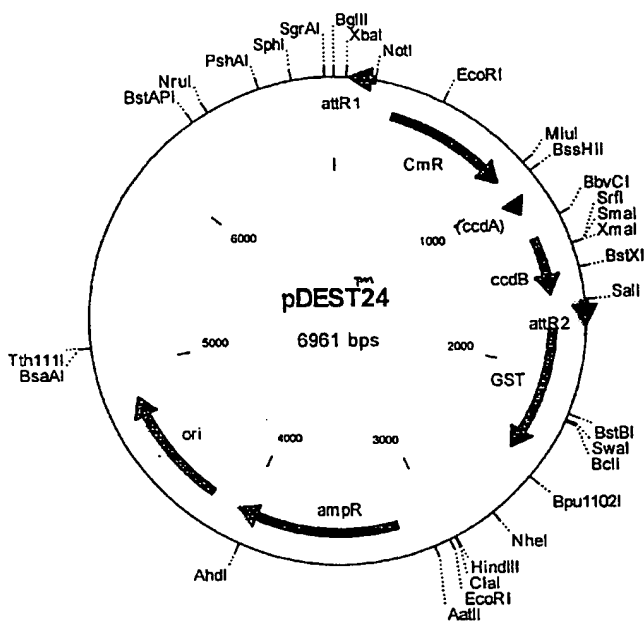
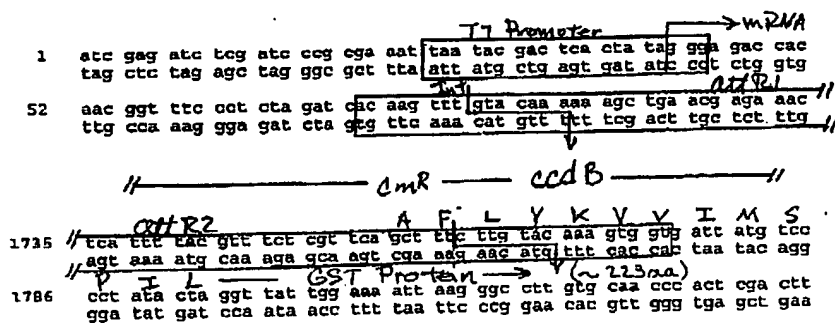


FIGURE 44A

pDEST24 6961 bp

Location (Base Nos.)	Gene Encoded
195..71	attR1
304..963	CmR
1083..1167	inactivated ccdA
1305..1610	ccdB
1651..1775	attR2
1783..2451	GST
3181..4041	ampR
4190..4829	ori

```

1 ATCGAGATCT CGATCCCGCG AAATTAATAC GACTCACTAT AGGGAGACCA CAACGGTTTC
61 CCTCTAGATC ACAAGTTTGT ACAAATAAGC TGAACGAGAA ACGTAAATG ATATAAATAT
121 CAATATATTA AATTAGATTT TGCATAAAAA ACAGACTACA TAATACTGTA AAACACAACA
181 TATCCAGTCA CTATGGCGGC CGCATTAGGC ACCCCAGGCT TTACACTTTA TGCTTCCGGC
241 TCGTATAATG TGTGGATTTT GAGTTAGGAT CCGGCGAGAT TTTCAGGAGC TAAGGAAGCT
301 AAAATGGAGA AAAAAATCAC TGGATATACC ACCGTTGATA TATCCCAATG GCATCGTAAA
361 GAACATTTTG AGGCATTTC A GTCAGTTGCT CAATGTACCT ATAACCAGAC CGTTCAGCTG
421 GATATTACGG CCTTTTTTAA GACCGTAAAG AAAAAAAGC ACAAGTTTAA TCCGGCCTTT
481 ATTCACATTG TTGCCCGCCT GATGAATGCT CATCCGGAAT TCCGTATGGC AATGAAAGAC
541 GGTGAGCTGG TGATATGGGA TAGTGTTTAC CCTTGTTACA CCGTTTCCA TGAGCAAACT
601 GAAACGTTT CATCGCTCTG GAGTGAATAC CACGACGATT TCCGGCAGTT TCTACACATA
661 TATTCGCAAG ATGTGGCGTG TTACGGTGAA AACCTGGCCT ATTTCCCTAA AGGGTTTATT
721 GAGAATATGT TTTTCGTCTC AGCCAATCCC TGGGTGAGTT TCACCAGTTT TGATTTAAAC
781 GTGGCCAATA TGGACAACCT CTTCCGCCCC GTTTTACCA TGGGCAAAATA TTATACGCAA
841 GGGCACAAGG TGCTGATGCC GCTGGCGGAT CAGGTTTATC ATGCCGCTG TGATGGCTTC
901 CATGTCCGCA GAATGCTTAA TGAATTACAA CAGTACTGCG ATGAGTGGCA GGGCGGGGCG
961 TAAACGCGTG GATCCGGCTT ACTAAAAGCC AGATAACAGT ATGCGTATT GCGCGCTGAT
1021 TTTTGCGGTA TAAGAATATA TACTGATATG TATACCCGAA GTATGTCAAA AAGAGGTGTG
1081 CTATGAAGCA CGGTATTACA GTGACAGTTG ACAGCGACAG CTATCAGTTG CTCAAGGCAT
1141 ATATGATGTC AATATCTCCG GTCTGGTAAG CACAACCATG CAGAATGAAG CCCGTCGTCT
1201 GCGTGCCGAA CGCTGGAAAG CGGAAAATCA GGAAGGGATG GCTGAGGTG CCGGTTTAT
1261 TGAAATGAAC GGCTCTTTTG CTGACGAGAA CAGGGACTGG TGAATGCGAG TTTAAGGTTT
1321 ACACCTATAA AAGAGAGAGC CGTTATCGTC TGTTTGTTGA TGACAGAGT GATATTATTG
1381 ACACGCCCCG GCGACGGATG GTGATCCCC TGGCCAGTGC ACGTCTGCTG TCAGATAAAG
1441 TCTCCCGTGA ACTTTACCCG GTGGTGCGTA TCGGGGATGA AAGCTGGCGC ATGATGACCA
1501 CCGATATGGC CAGTGTGCCG GTCTCCGTTA TCGGGGAAGA AGTGGCTGAT CTCAGCCACC
1561 GCGAAAATGA CATCAAAAC GCCATTAAAC TGATGTTCTG GGAATATAA ATGTCAGGCT
1621 CCCTTATACA CAGCCAGTCT GCAGGTGCGC CATAGTGACT GGATATGTTG TGTTTTACAG
1681 TATTATGTAG TCTGTTTTTT ATGCAAAATC TAATTTAATA TATTGATATT TATATCATT
1741 TACGTTTCTC GTTCAGCTTT CTTGTACAAA GTGGTGATTA TGTCCTTAT ACTAGGTTAT
1801 TGGAAAATTA AGGGCCTTGT GCAACCCACT CGACTTCTTT TGGAAATATCT TGAAGAAAAA
1861 TATGAAGAGC ATTTGTATGA GCGCGATGAA GGTGATAAAT GCGGAAACAA AAAGTTTGAA
1921 TTGGGTTTGG AGTTTCCCAA TCTTCCTTAT TATATTGATG GTGATGTTAA ATTAACACAG
1981 TCTATGGCCA TCATACGTTA TATAGCTGAC AAGCACAAAC TGTGGGTGG TTGTCCAAAA
2041 GAGCGTGCAG AGATTTCAAT GCTTGAAGGA GCGGTTTTGG ATATTAGATA CCGGTGTTTCG
2101 AGAATTGCAT ATAGTAAAGA CTTTGAAACT CTCAAAGTTG ATTTTCTTAG CAAGCTACCT
2161 GAAATGCTGA AAATGTTTGA AGATCGTTTA TGTCAATAAA CATATTTAAA TGGTGATCAT
2221 GTAACCCATC CTGACTTCAT GTTGTATGAC GCTCTTGATG TTGTTTTATA CATGGACCCA
2281 ATGTGCCTGG ATGCGTTCCC AAAATTAGTT TGTTTTAAAA AACGTATTGA AGCTATCCCA
2341 CAAATTGATA AGTACTTGAA ATCCAGCAAG TATATAGCAT GGCCTTTGCA GGGCTGGCAA
2401 GCCACGTTTG GTGGTGGCGA CCATCCTCCA AAATCGGATC TGGTTCCGCG TCCATGGGGA
2461 TCCGGCTGCT AACAAAGCCC GAAAGGAAGC TGAGTTGGCT GCTGCCACCG CTGAGCAATA
2521 ACTAGCATAA CCCCTTGGGG CCTCTAAACG GGTCTTGAGG GGTTTTTTGC TGAAAGGAGG
2581 AACTATATCC GGATATCCAC AGGACGGGTG TGGTCGCCAT GATCGCGTAG TCGATAGTGG
2641 CTCCAAGTAG CGAAGCGAGC AGGACTGGGC GCGGGCCAAA CCGGTGCGAC AGTGCTCCGA-

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FIGURE 44B

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2701 GAACGGGTGC GCATAGAAAT TGCATCAACG CATATAGCGC TAGCAGCAGC CCATAGTGCAC
2761 TGGCGATGCT GTCGGAATGG ACGATATCCC GCAAGAGGCC CGGCAGTACC GGCATAACCA
2821 AGCCTATGCC TACAGCATCC AGGGTGACGG TGCCGAGGAT GACGATGAGC GCATTGTTAG
2881 ATTTCATACA CGGTGCTGA CTGCGTTAGC AATTAACTG TGATAAACTA CCGCATTAAA
2941 GCTTATCGAT GATAAGCTGT CAAACATGAG AATTCTTGAA GACGAAAGGG CCTCGTGATA
3001 CGCCTATTTT TATAGGTTAA TGTCATGATA ATAATGGTTT CTTAGACGTC AGGTGGCACT
3061 TTTCCGGGAA ATGTGCGCGG AACCCCTATT TGTTATTTT TCTAAATACA TTCAAATATG
3121 TATCCGCTCA TGAGACATA ACCCTGATAA ATGCTTCAAT AATATTGAAA AAGGAAGAGT
3181 ATGAGTATTG AACATTTCGG TGTCGCCCTT ATTCCCTTTT TTGCGGCATT TTGCCTTCCT
3241 GTTTTTGTCT ACCCAGAAAC GCTGGTGAAG GTAAAGATG CTGAAGATCA GTTGGGTGCA
3301 CGAGTGGGTT ACATCGAACT GGATCTCAAC AGCGGTAAGA TCCTTGAGAG TTTTCGCCCC
3361 GAAGAACGTT TTCCAATGAT GAGCACTTTT AAAGTCTGCG TATGTGGCGC GGTATTATCC
3421 CGTGTGACG CCGGGCAAGA GCAACTCGGT CGCCGCATAC ACTATTCTCA GAATGACTTG
3481 GTTGAGTACT CACCAGTCAC AGAAAAGCAT CTTACGGATG GCATGACAGT AAGAGAATTA
3541 TGCAGTGTG CCATAACCAT GAGTGATAAC ACTGCGGCCA ACTTACTTCT GACAACGATC
3601 GGAGGACCGA AGGAGCTAAC CGCTTTTTTG CACAACATGG GGGATCATGT AACTCGCCTT
3661 GATCGTTGGG AACCGGAGCT GAATGAAGCC ATACCAAACG ACGAGCGTGA CACCAGATG
3721 CCTGCAGCAA TGGCAACAAC GTTGCACAAA CTATTAACCT GCGAACTACT TACTCTAGCT
3781 TCCCGGCAAC AATTAATAGA CTGGATGGAG GCGGATAAAG TTGCAGGACC ACTTCTGCGC
3841 TCGGCCCTTC CGGCTGGCTG GTTTATTGCT GATAAATCTG GAGCCGGTGA GCGTGGGTCT
3901 CGCGGTATCA TTGAGCACT GGGGCCAGAT GGTAAGCCCT CCCGTATCGT AGTTATCTAC
3961 ACGACGGGGA GTCAGGCAAC TATGGATGAA CGAAATAGAC AGATCGCTGA GATAGGTGCC
4021 TCACTGATTA AGCAATTGTA ACTGTCAGAC CAACTTTACT CATATATACT TTAGATTGAT
4081 TTAATACTTC ATTTTAAATT TAAAAGGATC TAGGTGAAGA TCCTTTTGA TAATCTCATG
4141 ACCAAATCC CTTAACGTGA GTTTTCGTT CACTGAGCGT CAGACCCCGT AGAAAAGATC
4201 AAAGGATCTT CTTGAGATCC TTTTCTCTG CGCGTAATCT GCTGCTTGCA AAAAAA
4261 CCACCGCTAC CAGCGGTGGT TGTGTTGCGG GATCAAGAGC TACCAACTCT TTTCCGAAG
4321 GTAACGTGCT TCAGCAGAGC GCAGATACCA AATACTGTCC TTCTAGTGA GCGGTAGTTA
4381 GGCCACCCT TCAAGAACTC TGTAGCACCG CCTACATACC TCGCTCTGCT AATCCTGTTA
4441 CCAGTGGCTG CTGCCAGTGG CGATAAGTCG TGTCTTACCG GGTGGGACTC AAGACGATAG
4501 TTACCGGATA AGGCGCAGCG GTCCGGCTGA ACGGGGGGTT CGTGACACAC GCCAGCTTG
4561 GAGCGAACGA CCTACACCGA ACTGAGATAC CTACAGCGTG AGCTATGAGA AAGCGCCACG
4621 CTTCCCGAAG GGAGAAAGGC GGCAGGGTAT CCGGTAAAGC GCAGGGTCGG AACAGGAGAG
4681 CGCACGAGGG AGCTTCCAGG GGGAAACGCC TGGTATCTTT ATAGTCTGT CCGGTTTCG
4741 CACCTCTGAC TTGAGCGTCG ATTTTGTGA TGCTCGTCAG GGGGGCGGAG CTTTGGAAA
4801 AACGCCAGCA ACGCGGCCTT TTACGGTTC CTGGCCTTTT GCTGGCCTTT TGCTCACATG
4861 TTCTTCTCTG CGTTATCCCC TGATTCTGTG GATAACCGTA TTACCGCCTT TGAGTGAGCT
4921 GATACCGCTC GCCGACGCG AACGACCGAG CGCAGCGAGT CAGTGAGCGA GGAAGCGGAA
4981 GAGCGCCTGA TGGCGTATTT TCTCCTTACG CATCTGTGCG GTATTTCACA CCGCATATAT
5041 GGTGCACTCT CAGTACAATC TGCTCTGATG CCGCATAGTT AAGCCAGTAT AACTCCGCT
5101 ATCGCTACGT GACTGGGTCA TGGCTGCGCC CCGACACCCG CCAACACCCG CTGACGCGCC
5161 CTGACGGGCT TGTCTGTCTC CGGCATCCGC TTACAGACAA GCTGTGACCG TCTCCGGGAG
5221 CTGCATGTGT CAGAGGTTTT CACCGTCATC ACCGAAACGC GCGAGGCAGC TGCGGTAAAG
5281 CTCATCAGCG TGGTCGTGAA GCGATTACA GATGTCTGCC TGTTTCATCCG CGTCCAGCTC
5341 GTTGAGTTTC TCCAGAAAGG TTAATGTCTG GCTTCTGATA AAGCGGGCCA TGTTAAGGGC
5401 GGTTTTTTCC TGTTTGGTCA CTGATGCCTC CGTGTAAAGG GGAATTTCTGT TCATGGGGGT
5461 AATGATACCG ATGAAACGAG AGAGGATGCT CACGATACGG GTTACTGATG ATGAACATGC
5521 CCGGTTACTG GAACGTTGTG AGGGTAAACA ACTGGCGGTA TGGATGCGGC GGGACGAGAG
5581 AAAAATCACT CAGGGTCAAT GCCAGCGCTT CGTTAATACA GATGTAGGTG TTCCACAGGG
5641 TAGCCAGCAG CATCTGCGA TGCAGATCCG GAACATAATG GTGCAGGGCG CTGACTTCCG
5701 CGTTTCCAGA CTTTACGAAA CACGAAACC GAAGACCATT CATGTTGTTG CTCAGGTGCG
5761 AGACGTTTTG CAGCAGCAGT CGCTTCACTG TCGCTCGCGT ATCGGTGATT CATTCTGCTA
5821 ACCAGTAAGG CAACCCCGCC AGCCTAGCCG GGTCTCAAC GACAGGAGCA CGATCATGCG
5881 CACCCGTGGC CAGGACCCAA CGCTGCCCGA GATGCGCCGC GTGCGGCTGC TGGAGATGGC
5941 GGACGCGATG GATATGTTCT GCCAAGGGTT GGTTTGCGCA TTCACAGTTC TCCGCAAGAA
6001 TTGATTGGCT CCAATTCTTG GAGTGTGAA TCCGTTAGCG AGGTGCCGCC GGCCTCCATT
6061 CAGGTCGAGG TGGCCCGGCT CCATGCACCG CAGACGCAAC GCGGGAGGCA GACAAAGGTAT
6121 AGGCGGGCGC CTACAATCCA TGCCAACCCG TTCCATGTGC TCGCCGAGGC GGCATAAATC

FIGURE 44C

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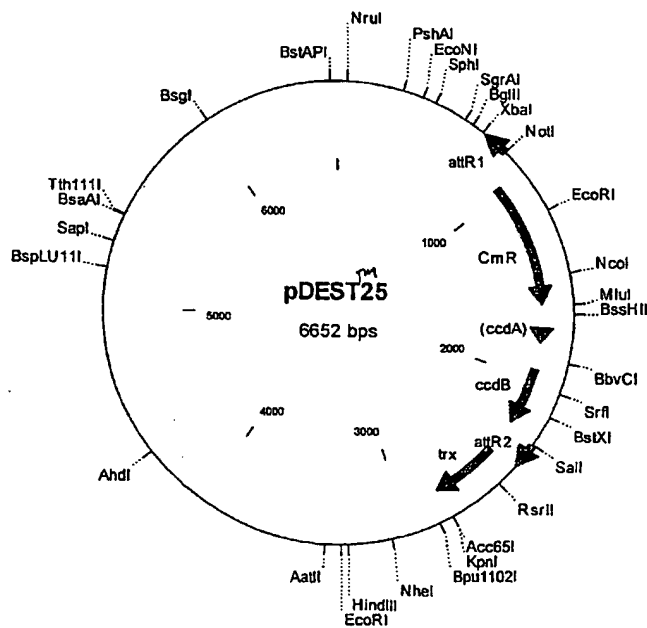
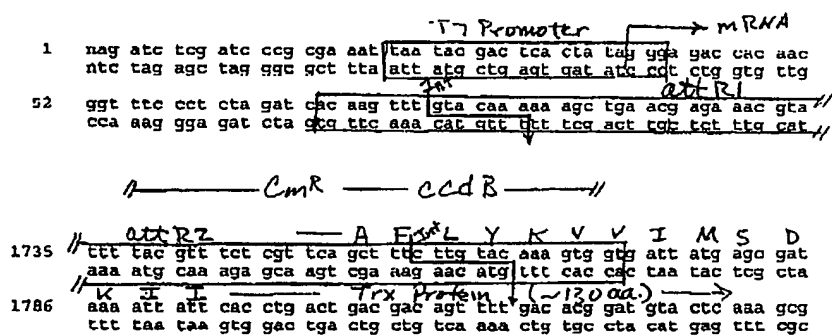
6181 GCCGTGACGA TCAGCGGTCC AGTGATCGAA GTTAGGCTGG TAAGAGCCGC GAGCGATCCT
6241 TGAAGCTGTC CCTGATGGTC GTCATCTACC TGCCTGGACA GCATGGCCTG CAACGCGGGC
6301 ATCCCGATGC CGCCGGAAGC GAGAAGAATC ATAATGGGGA AGGCCATCCA GCCTCGCGTC
6361 GCGAACGCCA GCAAGACGTA GCCCAGCGCG TCGGCCGCCA TGCCGGCGAT AATGGCCTGC
6421 TTCTCGCCGA AACGTTTGGT GGCGGGACCA GTGACGAAGG CTTGAGCGAG GCGGTGCAAG
6481 ATTCCGAATA CCGCAAGCGA CAGGCCGATC ATCGTCGCGC TCCAGCGAAA GCGGTCTCTG
6541 CCGAAAATGA CCCAGAGCGC TGCCGGCACC TGTCTACGA GTTGATGAT AAAGAAGACA
6601 GTCATAAGTG CGGCGACGAT AGTCATGCCC CGCGCCCACC GGAAGGAGCT GACTGGGTTG
6661 AAGGCTCTCA AGGGCATCGG TCGATCGACG CTCTCCCTTA TGCGACTCCT GCATTAGGAA
6721 GCAGCCCACT AGTAGGTTGA GGCCGTTGAG CACCGCCGCC GCAAGGAATG GTGCATGCAA
6781 GGAGATGGCG CCCAACAGTC CCCCGGCCAC GGGGCTGCCC ACCATACCCA CGCCGAAACA
6841 AGCGCTCATG AGCCCGAAGT GGCGAGCCCG ATCTTCCCCA TCGGTGATGT CGGCGATATA
6901 GGCGCCAGCA ACCGCACCTG TGGCGCCGGT GATGCCGCC ACGATGCGTC CGGCGTAGAG
6961 G

FIGURE 44D

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FIGURE 45A

pDEST25
Thioredoxin carboxy-fusion vector, T7 promoter



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pDEST25 6652 bp

Location (Base Nos.)	Gene Encoded
844..720	attR1
953..1612	CmR
1732..1816	inactivated ccdA
1954..2259	ccdB
2300..2424	attR2
2432..2794	trx

1	CCGGAAGCGA	GAAGAATCAT	AATGGGGAAG	GCCATCCAGC	CTCGCGTCGC	GAACGCCAGC
61	AAGACGTAGC	CCAGCGCGTC	GGCCGCCATG	CCGGCGATAA	TGGCCTGCTT	CTCGCCGAAA
121	CGTTTGGTGG	CGGGACCACT	GACGAAGGCT	TGAGCGAGGG	CGTGCAAGAT	TCCGAATACC
181	GCAAGCGACA	GGCCGATCAT	CGTCGCGCTC	CAGCGAAAAGC	GGTCCTCGCC	GAAAATGACC
241	CAGAGCGCTG	CCGGCACCTG	TCCTACGAGT	TGCATGATAA	AGAAGACAGT	CATAAGTGCG
301	GCGACGATAG	TCATGCCCCG	CGCCCCACCG	AAGGAGCTGA	CTGGGTTGAA	GGCTCTCAAG
361	GGCATCGGTC	GATCGACGCT	CTCCCTTATG	CGACTCCTGC	ATTAGGAAAGC	AGCCCCAGTAG
421	TAGGTTGAGG	CCGTGTAGCA	CCGCCGCCCG	AAGGAATGGT	GCATGCAAGG	AGATGGCGCC
481	CAACAGTCCC	CCGGCCACGG	GGCCTGCCAC	CATACCCACG	CCGAAACAAG	CGCTCATGAG
541	CCCGAAGTGG	CGAGCCCGAT	CTTCCCCATC	GGTGATGTCG	GCGATATAGG	CGCCAGCAAC
601	CGCACCCTGT	GCGCCGGTGA	TGCCGGCCAC	GATGCGTCCG	GCGTAGAGGA	TCGAGATCTC
661	GATCCCCGCA	AATTAATACG	ACTCACTATA	GGGAGACCAC	AACGGTTTCC	CTCTAGATCA
721	CAAGTTTGTA	CAAAAAAGCT	GAACGAGAAA	CGTAAATGA	TATAAATATC	AATATATTTAA
781	ATTAGATTIT	GCATAAAAAA	CAGACTACAT	AATACTGTAA	AACACAACAT	ATCCAGTCCAC
841	TATGGCGGCC	GCATTAGGCA	CCCCAGGCTT	TACACTTTAT	GCTTCCGGCT	CGTATAATGT
901	GTGGATTTTG	AGTTAGGATC	CGGCGAGATT	TTCAGGAGCT	AAGGAAGCTA	AAATGGAGAA
961	AAAAATCACT	GGATATACCA	CCGTTGATAT	ATCCCAATGG	CATCGTAAAG	AACATTTTGA
1021	GGCATTTCAG	TCAGTTGCTC	AATGTACCTA	TAACCAGACC	GTTCCAGCTG	ATATTACGGC
1081	CTTTTTAAAG	ACCGTAAAGA	AAAATAAGCA	CAAGTTTAT	CCGGCCTTTA	TTCACATTCT
1141	TGCCCCGCTG	ATGAATGCTC	ATCCGGAATT	CCGTATGGCA	ATGAAAGACG	GTGAGCTGGT
1201	GATATGGGAT	AGTGTTCAAC	CTTGTTACAC	CGTTTTCCAT	GAGCAAACTG	AAACGTTTTT
1261	ATCGCTCTGG	AGTGAATACC	ACGACGATTT	CCGGCAGTTT	CTACACATAT	ATTTCGCAAGA
1321	TGTGGCGTGT	TACGGTGAAA	ACCTGGCCTA	TTTCCCTAAA	GGGTTTATTG	AGAATATGTT
1381	TTTCGTCTCA	GCCAAATCCCT	GGGTGAGTTT	CACCAAGTTT	GATTAAACG	TGGCCAATAT
1441	GGACAACTTC	TTGCCCCCCG	TTTTCAACAT	GGGCAAAATAT	TATACGCAAG	GCGACAAGGT
1501	GCTGATGCCG	CTGGCGATTG	AGGTTTCATCA	TGCCGTCTGT	GATGGCTTCC	ATGTGCGCAG
1561	AATGCTTAAT	GAATTACAAC	AGTACTGCGA	TGAGTGGCAG	GGCGGGGCGT	AAACGCGTGG
1621	ATCCGGCTTA	CTAAAAGCCA	GATAACAGTA	TGCGTATTTG	CGCGCTGATT	TTTGCGGTAT
1681	AAGAATATAT	ACTGATATGT	ATACCCGAAG	TATGTCAAAA	AGAGGTGTGC	TATGAAGCAG
1741	CGTATTACAG	TGACAGTTGA	CAGCGACAGC	TATCAGTTGC	TCAAGGCATA	TATGATGTCA
1801	ATATCTCCGG	TCTGGTAAGC	ACAACCATGC	AGAATGAAAG	CCGTCTGCTG	CGTGCCGAAC
1861	GCTGGAAAGC	GGAAAATCAG	GAAGGGATGG	CTGAGGTCGC	CCGGTTTATT	GAAATGAAAG
1921	GCTCTTTTGC	TGACGAGAAG	AGGGACTGGT	GAAATGCAGT	TTAAGGTTTA	CACCTATAAA
1981	AGAGAGAGCC	GTTATCGTCT	GTTTGTGGAT	GTACAGAGTG	ATATTATTGA	CACGCCCGGG
2041	CGACGGATGG	TGATCCCCCT	GGCCAGTGCA	CGTCTGCTGT	CAGATAAAGT	CTCCCGTGAA
2101	CTTTACCCGG	TGGTGCAATAT	CGGGGATGAA	AGCTGGCGCA	TGATGACCAC	CGATATGGCC
2161	AGTGTGCCGG	TCTCCGTTAT	CGGGGAAGAA	GTGGCTGATC	TCAGCCACCG	CGAAAATGAC
2221	ATCAAAAACG	CCATTAACTT	GATGTTCTGG	GGAATATAAA	TGTGAGGCTC	CCTTATACAC
2281	AGCCAGTCTG	CAGGTCGACC	ATAGTGACTG	GATATGTTGT	GTTTTACAGT	ATTATGTAGT
2341	CTGTTTTTTA	TGCAAAATCT	AATTTAATAT	ATTGATATTT	ATATCATTTT	ACGTTTCTCG
2401	TTCAGCTTTC	TTGTACAAAG	TGGTGATTAT	GAGCGATAAA	ATTATTCAAC	TGACTGACGA
2461	CAGTTTGTAC	ACGGATGTAC	TCAAAGCGGA	CGGGGCGATC	CTCGTCGATT	TCTGGGCAGA
2521	GTGGTGCGGT	CCGTGCAAAA	TGATCGCCCC	GATTCTGGAT	GAAATCGCTG	ACGAATATCA
2581	GGGCAAACTG	ACCGTTGCAA	AACTGAACAT	CGATCAAAAC	CCTGGCACTG	CGCCGAAATA
2641	TGGCATCCGT	GGTATCCCCG	CTCTGCTGCT	GTTCAAAAAC	GGTGAAGTGG	CGGCAACCAA
2701	AGTGGGTGCA	CTGTCTAAAG	GTCAGTTGAA	AGAGTTCTCT	GACGCTAACC	TGGCCGGTTC
2761	TGGTTCTGGT	GATGACGATG	ACAAGGTACC	CGGGGATCGA	TCCGGCTGCT	AACAAAGCCC

Figure 45B

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2821 GAAAGGAAGC TGAGTTGGCT GCTGCCACCG CTGAGCAATA ACTAGCATAA CCCCTTGGGG
2881 CCTCTAAACG GGTCTTGAGG GGTCTTTTGC TGAAAGGAGG AACTATATCC GGATATCCAC
2941 AGGACGGGTG TGGTCGCCAT GATCGCGTAG TCGATAGTGG CTCCAAGTAG CGAAGCGAGC
3001 AGGACTGGGC GCGCGCCAAA GCGGTCGGAC AGTGCTCCGA GAACGGGTGC GCATAGAAAT
3061 TGCATCAACG CATATAGCGC TAGCAGCAGC CCATAGTGAC TGGCGATGCT GTCGGAATGG
3121 ACGATATCCC GCAAGAGGCC CGGCAGTACC GGCATAACCA AGCCTATGCC TACAGCATCC
3181 AGGGTGACGG TGCCGAGGAT GACGATGAGC GCATTGTTAG ATTTTCATACA CGGTGCCTGA
3241 CTGCGTTAGC AATTTAACTG TGATAAACTA CCGCATTAAA GCTTATCGAT GATAAGCTGT
3301 CAAACATGAG AATTCTTGAA GACGAAAGGG CCTCGTGATA CGCCTATTTT TATAGGTTAA
3361 TGTCTAGATA ATAATGGTTT CTTAGACGTC AGGTGGCACT TTTCGGGGAA ATGTGCGCGG
3421 AACCCCTATT TGTATTATTT TCTAAATACA TTCAAAATATG TATCCGCTCA TGAGACAATA
3481 ACCCTGATAA ATGCTTCAAT AATATTGAAA AAGGAAGAGT ATGAGTATTC AACATTTCG
3541 TGTCGCCCTT ATTCCCTTTT TTGCGGCATT TTGCCCTTCT GTTTTTGTCT ACCCAGAAAC
3601 GCTGGTGAAA GTAAAGATG CTGAAGATCA GTTGGGTGCA CGAGTGGGTT ACATCGAAT
3661 GGATCTCAAC AGCGGTAGA TCCTTGAGAG TTTTCGCCCC GAAGAACGTT TTCCAATGAT
3721 GAGCACTTTT AAAGTTCTGC TATGTGGCGC GGTATTATCC CGTGTGACG CCGGGCAAGA
3781 GCAACTCGGT CGCCGCATAC ACTATTCTCA GAATGACTTG GTTGAGTACT CACCACTCAC
3841 AGAAAAGCAT CTTACGGATG GCATGACAGT AAGAGAATTA TGCAGTGCTG CCATAACCAT
3901 GAGTGATAAC ACTGCGGCCA ACTTACTTCT GACAACGATC GGAGGACCGA AGGAGCTAAC
3961 CGCTTTTTTG CACAACATGG GGGATCATGT AACTCGCCTT GATCGTTGGG AACCGGAGCT
4021 GAATGAAGCC ATACCAAACG ACGAGCGTGA CACCACGATG CCTGCAGCAA TGGCAACAAC
4081 GTTGGCGAAA CTATTAACTG GCGAACTACT TACTCTAGCT TCCCGGCAAC AATTAATAGA
4141 CTGGATGGAG GCGGATAAAG TTGCAGGACC ACTTCTGCGC TCGGCCCTTC CGGCTGGCTG
4201 GTTTATTGCT GATAAATCTG GAGCGGTGTA GCGTGGGTCT CGCGGTATCA TTGCAGCACT
4261 GGGGCCAGAT GGTAAAGCCT CCCGTATCGT AGTTATCTAC ACGACGGGGA GTCAAGCAAC
4321 TATGGATGAA CGAAATAGAC AGATCGCTGA GATAGGTGCC TCACTGATTA AGCATTGGTA
4381 ACTGTGAGAC CAAGTTTACT CATATATACT TTAGATTGAT TTAACCTTC ATTTTTAATT
4441 TAAAAGGATC TAGGTGAAGA TCCTTTTGA TAATCTCATG ACCAAAATCC CTTAACGTGA
4501 GTTTTCGTTT CACTGAGCGT CAGACCCCGT AGAAAAGATC AAAGGATCTT CTTGAGATCC
4561 TTTTTTCTG CGCGTAATCT GCTGCTTGCA AACAAAAAAA CCACCGTAC CAGCGGTGGT
4621 TTGTTTGGCG GATCAAGAGC TACCAACTCT TTTTCCGAAG GTAAGTGGCT TCAGCAGAGC
4681 GCAGATACCA AATACTGTCC TTCTAGTGTA GCCGTAGTTA GGCCACCACT TCAAGAACTC
4741 TGTAGCACCG CCTACATACC TCGCTCTGCT AATCCTGTGA CCAGTGGCTG CTGCCAGTGG
4801 CGATAAGTCG TGCTTACC GGTGGGACTC AAGACGATAG TTACCGGATA AGCGCGAGCG
4861 GTCGGGCTGA ACGGGGGGTT CGTGACACA GCCCAGCTTG GAGCGAAGCA CCTACACCGA
4921 ACTGAGATAC CTACAGCGTG AGCTATGAGA AAGCGCCACG CTTCCCGAAG GGAGAAAGGC
4981 GGACAGGTAT CCGGTAAGCG GCAGGGTCGG AACAGGAGAG CGCACGAGGG AGCTTCCAGG
5041 GGGAAACGCC TGGTATCTTT ATAGTCCTGT CGGGTTTCGC CACCTCTGAC TTGAGCGTCG
5101 ATTTTGTGA TGCTCGTCAG GGGGGCGGAG CCTATGGA AAAACGCCAGCA ACGCGGCCTT
5161 TTACCGGTTT CTGGCCTTTT GCTGGCCTTT TGCTCACATG TTCCTTCTG CGTTATCCCC
5221 TGATTCTGTG GATAACCGTA TTACCGCCTT TGAGTGAGCT GATACCGCTC GCCGCGAGCG
5281 AACGACCGAG CGCAGCGAGT CAGTGAGCGA GGAAGCGGAA GAGCGCCTGA TCGGTTATT
5341 TCTCCTTACG CATCTGTGCG GTATTTCACA CCGCATATAT GGTGCACTCT CAGTACAATC
5401 TGCTCTGATG CCGCATAGTT AAGCCAGTAT ACACTCCGCT ATCGCTACGT GACTGGGTCA
5461 TGGCTGGGCC CCGACACCGG CCAACACCGG CTGACGCGCC CTGACGGGCT TGTCTGCTCC
5521 CGGCATCCGC TTACAGACAA GCTGTGACCG TCTCCGGGAG CTGCATGTGT CAGAGGTTTT
5581 CACCGTCATC ACCGAAACGC GCGAGGCAGC TGCGGTAAAG CTCATCAGCG TGGTCTGTGA
5641 CGGATTACCA GATGTCTGCC TGTTCATCCG CGTCCAGCTC GTTGAGTTTC TCCAGAAGCG
5701 TTAATGTCG GCTTCTGATA AAGCGGGCCA TGITAAGGGC GGTTTTTTCC TGTTTGGTCA
5761 CTGATGCCTC CGTGTAAAGG GGATTCTGT TCATGGGGGT AATGATACCG ATGAACGAG
5821 AGAGGATGCT CACGATACGG GTTACTGATG ATGAACATGC CCGGTTACTG GAACGTTGTG
5881 AGGGTAAACA ACTGGCGGTA TGGATGCGGC GGGACCAGAG AAAAATCACT CAGGGTCAAT
5941 GCCAGCGCTT CGTTAATACA GATGTAGGTG TTCCACAGGG TAGCCAGCAG CATCTGCGA
6001 TGCAGATCCG GAACATAATG GTGCAGGGCG CTGACTTCCG CGTTTCCAGA CTTTACGAAA
6061 CACGGAAACC GAAGACCATT CATGTTGTTG CTCAGGTCCG AGACGTTTTG CAGCAGCAGT
6121 CGCTTCACGT TCGCTCGCGT ATCGGTGATT CATTTCTGCTA ACCAGTAAGG CAACCCCGCC
6181 AGCCTAGCCG GGTCTCTAAC GACAGGAGCA CGATCATGCG CACCCGTGGC CAGGACCCAA
6241 CGCTGCCCGA GATGCGCCG GTGCGGCTGC TGGAGATGGC GGACGCGATG GATATGTTCT-

FIGURE 45C

6301 GCCAAGGGTT GGTTCGCGCA TTCACAGTTC TCCGCAAGAA TTGATTGGCT CCAATTCTTG
6361 GAGTGGTGAA TCCGTTAGCG AGGTGCCCGC GGCTTCCATT CAGGTCGAGG TGGCCCGGCT
6421 CCATGCACCG CGACGCAACG CGGGGAGGCA GACAAGGTAT AGGGCGGCGC CTACAATCCA
6481 TGCCAACCCG TTCCATGTGC TCGCCGAGGC GGCATAAATC GCCGTGACGA TCAGCGGTCC
6541 AGTGATCGAA GTTAGGCTGG TAAGAGCCGC GAGCGATCCT TGAAGCTGTC CCTGATGGTC
6601 GTCATCTACC TGCCTGGACA GCATGGCCTG CAACGCGGGC ATCCCGATGC CG

FIGURE 45D

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FIGURE 46A

pDEST26 His6 Amino Fusion in pCMV Sport-neo Vector

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600  ttg acg tca atg gga gtt tgt ttt ggc aac aaa atc aac ggg act ttc caa
    aac tgc agt tac cct caa aca aaa cag tgg ttt tag ttg ccc tga aag gtt

651  aat gtc gta aca act cag ccc oat tga cgc aaa tgg cgc gta ggc gtg tac
    tta cag cat tgt tga ggc ggg gta act ggc ttt acc cgc cat cgc cac atg

702  // ggt ggg agg tct ata taa gca gag ctc gtt tag tga acc gtc aga tgg ttt
    //cca ccc tcc aga tat att cgt ctc gag caa atc act tgg cag tct ago gga

753  gga gac gcc atc cac gct gtt ttg acc tcc ata gaa gac acc ggg acc gat
    cct ctg cgg tag gtg cga caa aac tgg agg tat ctt ctg tgg ccc tgg cta

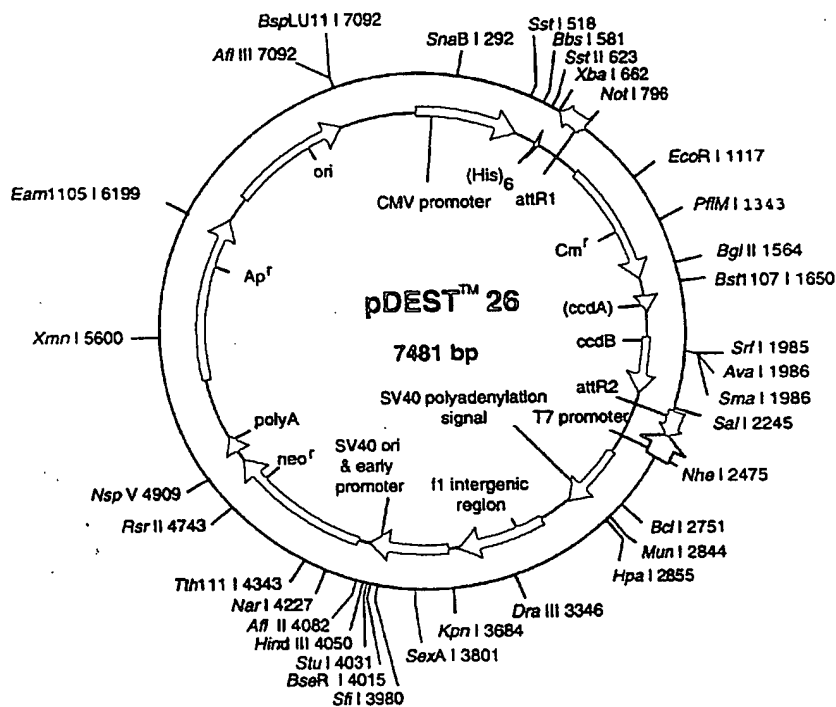
804  cca gcc tcc gga ctc tag cct agg ccg cgg acc latg gcg tac tac cat cac
    ggt cgg agg cct gag atc gga tcc gcc gcc tgg tac cgc atg atg gta gtc

855  H H H H S R S T S H Y K K A
    dat cac cat cac tct aga tca aca agt ttg tac aaa aaa gct gaa cga gaa
    gta gtg gta gtg aga tct agt tgt tca aac atg ttt ttt cga ott gct ctt //
    
```

CMV Promoter → M13

Start Transl → M A Y Y H H

Int



pDEST26 7481 bp

Location (Base Nos.)	Gene Encoded
492..509	his6
619..519	attR1
752..1411	CmR
1531..1615	inactivated ccdA
1753..2058	ccdB
2099..2223	attR2
2409..2771	SV40 polyA
2966..3421	f1 intergenic region
3485..3903	SV40 promoter
3948..4742	neo
4806..4854	polyA
5265..6125	Apr
6274..6913	ori
7344..385	CMV promoter

1	GTAAACTGCC	CACCTGGCAG	TACATCAAGT	GTATCATATG	CCAAGTACGC	CCCCATTGTA
61	CGTCAATGAC	GGTAAATGGC	CCGCCTGGCA	TTATGCCCCAG	TACATGACCT	TATGGGACTT
121	TCCTACTTGG	CAGTACATCT	ACGTATTAGT	CATCGCTATT	ACCATGGTGA	TGCGGTTTGT
181	GCAGTACATC	AATGGGCGTG	GATAGCGGTT	TGACTCACGG	GGATTTCCTAA	GTCTCCACCC
241	CATTGACGTC	AATGGGAGTT	TGTTTGGCA	CCAAATCAA	CGGGACTTTC	CAAAATGTCG
301	TAACAACTCC	GCCCCATTGA	CGCAAAATGGG	CGGTAGGCGT	GTACGGTGGG	AGGTCTATAT
361	AAGCAGAGCT	CGTTTAGTGA	ACCGTCAGAT	CGCCTGGAGA	CGCCATCCAC	GCTGTTTGA
421	CCTCCATAGA	AGACACCGGG	ACCGATCCAG	CCTCCGGACT	CTAGCCTAGG	CCGCGGACCA
481	TGGCGTACTA	CCATCACCAT	CACCATCACT	CTAGATCAAC	AAGTTGTAC	AAAAAAGCTG
541	AACGAGAAAC	GTAATATGAT	ATAAATATCA	ATATATTAAA	TTAGATTGTG	CATAAAAAAC
601	AGACTACATA	ATACTGTAAA	ACACAACATA	TCCAGTCACT	ATGGCGGCCG	CATTAGGCAC
661	CCCAGGCTTT	ACACTTTATG	CTTCCGGCTC	GTATAATGTG	TGGATTTTGA	GTTAGGATCC
721	GGCGAGATT	TCAGGAGCTA	AGGAAGCTAA	AATGGAGAAA	AAAATCACTG	GATATACCAC
781	CGTTGATATA	TCCCAATGGC	ATCGTAAAGA	ACATTTTGAG	GCATTTCAGT	CAGTTGCTCA
841	ATGTACCTAT	AACCAGACCG	TTCAGCTGGA	TATTACGGCC	TTTTTAAAGA	CCGTAAAGAA
901	AAATAAGCAC	AAGTTTTATC	CGGCCCTTAT	TCACATTCTT	GCCCGCCTGA	TGAATGCTCA
961	TCGGGAATTC	CGTATGGCAA	TGAAAGACGG	TGAGCTGGTG	ATATGGGATA	GTGTTACCCC
1021	TTGTTACACC	GTTTTCCATG	AGCAAACTGA	AACGTTTTCA	TCGCTCTGGA	GTGAATACCA
1081	CGACGATTTT	CGGCAGTTTC	TACACATATA	TTCGCAAGAT	GTGGCGTGTT	ACGGTGAAAA
1141	CCTGGCCTAT	TTCCCTAAAG	GGTTTATTGA	GAATATGTTT	TTCGTCTCAG	CCAATCCCTG
1201	GGTGAGTTTC	ACCAGTTTTG	ATTTAAACGT	GGCCAATATG	GACAACTTCT	TCGCCCCCGT
1261	TTTCACCATG	GGCAAATATT	ATACGCAAGG	CGACAAGGTG	CTGATGCCGC	TGGCGATTCA
1321	GGTTCATCAT	GCCGTCTGTG	ATGGCTTCCA	TGTCGGCAGA	ATGCTTAATG	AATTACAACA
1381	GTACTGCGAT	GAGTGGCAGG	GCGGGGCGTA	AAGATCTGGA	TCCGGCTTAC	TAAAGGCCAG
1441	ATAACAGTAT	GCGTATTGTC	GCGCTGATTT	TTGCGGTATA	AGAATATATA	CTGATATGTA
1501	TACCCGAAGT	ATGTCAAAAA	GAGGTGTGCT	ATGAAGCAGC	GTATTACAGT	GACAGTTGAC
1561	AGCGACAGCT	ATCAGTTGCT	CAAGGCATAT	ATGATGTCAA	TATCTCCGGT	CTGGTAAGCA
1621	CAACCATGCA	GAATGAAGCC	CGTCGTCTGC	GTGCCGAACG	CTGGAAAGCG	GAAAAACAGG
1681	AAGGGATGGC	TGAGGTGCGC	CGGTTTATTG	AAATGAACGG	CTCTTTTGCT	GACGAGAACA
1741	GGGACTGGTG	AAATGCAGTT	TAAAGTTTAC	ACCTATAAAA	GAGAGAGCCG	TTATCGTCTG
1801	TTTGTGGATG	TACAGAGTGA	TATTATTGAC	ACGCCCGGGC	GACGGATGGT	GATCCCCCTG
1861	GCCAGTGCAC	GTCGTCTGTC	AGATAAAGTC	TCCCGTGAAC	TTTACCCGGT	GGTGCAATATC
1921	GGGGATGAAA	GCTGGCGCAT	GATGACCACC	GATATGGCCA	GTGTGCCGGT	CTCCGTTATC
1981	GGGGAAGAAG	TGGCTGATCT	CAGCCACCGC	GAAAATGACA	TCAAAAACGC	CATTAACTTG
2041	ATGTTCTGGG	GAATATAAAT	GTCAGGCTCC	CTTATACACA	GCCAGTCTGC	AGGTCGACCA
2101	TAGTGACTGG	ATATGTTGTG	TTTTACAGTA	TTATGTAGTC	TGTTTTTTAT	GCAAAATCTA
2161	ATTTAATATA	TTGATATTTA	TATCATTTTA	CGTTTCTCGT	TCAGCTTTCT	TGTACAAAGT
2221	GGTTGATCGC	GTGCATGCGA	CGTCATAGCT	CTCTCCCTAT	AGTGAGTCGT	ATTATAAGCT
2281	AGGCACTGGC	CGTCGTTTTA	CAACGTCGTG	ACTGGGAAAA	CTGCTAGCTT	GGGATCTTTG -

FIGURE 46B

2341 TGAAGGAACC TTACTTCTGT GGTGTGACAT AATTGGACAA ACTACCTACA GAGATTAAAA
2401 GCTCTAAGGT AAATATAAAA TTTTAAAGTG TATAATGTGT TAAACTAGCT GCATATGCTT
2461 GCTGCTTGAG AGTTTGTCTT ACTGAGTATG ATTTATGAAA ATATTATACA CAGGAGCTAG
2521 TGATTCTAAT TGTTTGTGTA TTTTAGATTC ACAGTCCCAA GGCTCATTC AGGCCCTCA
2581 GTCCTCACAG TCTGTTTCATG ATCATAATCA GCCATACCAC ATTTGTAGAG GTTTTACTTG
2641 CTTTAAAAAA CCTCCACAC CTCCCCCTGA ACCTGAAACA TAAATGAAT GCRAATTGTTG
2701 TTGTTAACTT GTTTATTGCA GCTTATAATG GTTACAAATA AAGCAATAGC ATCACAATT
2761 TCACAAATAA AGCAITTTTT TCACTGCATT CTAGTTGTGG TTTGTCCAAA CTCATCAATG
2821 TATCTTATCA TGTCTGGATC GATCCTGCAT TAATGAATCG GCCAACGCGC GGGGAGAGGC
2881 GGTITGCGTA TTGGCTGGCG TAATAGCGAA GAGGCCCCGA CCGATCGCCC TTCCCAACAG
2941 TTGCGCAGCC TGAATGGCGA ATGGGACGCG CCCTGTAGCG GCGCATTAAAG CGCGCGGGT
3001 GTGGTGGTTA CGCGCAGCGT GACCGCTACA CTTGCCAGCG CCCTAGCGCC CGCTCCTTTC
3061 GCTTCTTCC CTTCCTTCTT CGCCACGTTT GCGCGCTTTC CCCGTCAAAG TCTAAATCGG
3121 GGGCTCCCTT TAGGGTTCCG ATTTAGTGCT TTACGGCACC TCGACCCCAA AAACTTGAT
3181 TAGGGTGATG GTTCACGTAG TGGGCCATCG CCCTGATAGA CGGTTTTTCG CCCTTTGACG
3241 TTGGAGTCCA CGTCTTTAA TAGTGGACTC TTGTTCCAAA CTGGAACAACT ACTCAACCTT
3301 ATCTCGGTCT ATTCTTTTGA TTTATAAGGG ATTTTGCCGA TTTCCGCCCTA TTGGTTAAAA
3361 AATGAGCTGA TTTAACAAAT AITTAACGCG AATTTTAAAC AAATATTAAC GTTTTCAATT
3421 TCGCCTGATG CGGTATTTTC TCCTTACGCA TCTGTGCGGT ATTTACACCC GCATACGCGG
3481 ATCTGCGCAG CACCATGGCC TGAATAAACC TCTGAAAGAG GAACTTGGTT AGGTACCTTC
3541 TGAGGCGGAA AGAACCAGCT GTGGAATGTG TGTCAGTTAG GGTGTGAAA GTCCCGAGGC
3601 TCCCGAGCAG GCAGAAGTAT GCAAAGCATG CATCTCAATT AGTCAGCAAC CAGGTGTGGA
3661 AAGTCCCCAG GCTCCCCAGC AGGCAGAAAT ATGCAAAGCA TGCACTCTAA TTAGTCAGCA
3721 ACCATAGTCC CGCCCTTAAC TCCGCCATC CCGCCCTTAA CTCCGCCAG TTCCGCCCAT
3781 TCTCCGCCCC ATGGCTGACT AATTTTTTTT ATTTATGCG AGGCCGAGGC CGCCTCGGCC
3841 TCTGAGCTAT TCCAGAAGTA GTGAGGAGGC TTTTTTGAG GCCTAGGCTT TTGCAAAAAG
3901 CTTGATTCTT CTGACACAAC AGTCTCGAAC TTAAGGCTAG AGCCACCATG ATTGAACAAG
3961 ATGGATTGCA CGCAGTTCT CCGGCCGCTT GGGTGGAGAG GCTATTCCGG TATGACTGGG
4021 CACAACAGAC AATCGGCTGC TCTGATGCCG CCGTGTCCG GCTGTACGCG CAGGGGCGCC
4081 CGGTTCTTTT TGTCAAGACC GACCTGTCCG GTGCCCTGAA TGAAGTCAG GACGAGGCG
4141 CGCGGTATC GTGGCTGGCC ACGACGGGCG TTCCTTGCAG AGCTGTGCTC GACGTTGTCA
4201 CTGAAGCGGG AAGGGATCG CTGCTATTGG GCGAAGTGCC GGGGAGGAT CTCCTGTCTC
4261 CTCACCTTGC TCCTGCCGAG AAAGTATCCA TCATGGCTGA TGCAATGCGG CGGCTGCATA
4321 CGCTTGATCC GGCTACCTGC CCATTTCGACC ACCAAGCGAA ACATCGCATC GAGCGAGCAC
4381 GTACTCGGAT GGAAGCCGGT CTTGTGATC AGGATGATCT GGACGAAGAG CATCAGGGGC
4441 TCGCGCCAGC CGAAGTGTTC GCCAGGCTCA AGGCGCGCAT GCCCGACGGC GAGGATCTCG
4501 TCGTGACCCA TGGCGATGCC TGCTTGCCGA ATATCATGGT GGAATAATGGC CGCTTTTCTG
4561 GATTTCATCA CTGTGGCCGG CTGGGTGTGG CGGACCGCTA TCAGGACATA GCGTTGGCTA
4621 CCCGTGATAT TGCTGAAGAG CTTGGCGGCG AATGGGCTGA CCGCTTCTC GTGCTTTACG
4681 GTATCGCCGC TCCCGATTCC CAGCGCATCG CCTTCTATCG CCTTCTTGAC GAGTTCTTCT
4741 GAGCGGGACT CTGGGGTTCG AAATGACCGA CCAAGCGAGC CCCAACCTGC CATCACGATG
4801 GCGCAATAA AATATCTTTA TTTTCATTAC ATCTGTGTGT TGGTTTTTGT TGTGAATCGA
4861 TAGCGATAAG GATCCGCGTA TGGTGCACTC TCAGTACAAT CTGCTCTGAT GCCGCATAGT
4921 TAAGCCAGCC CCGACACCCG CCAACACCCG CTGACGCGCC CTGACGGGCT TGTCTGTCTC
4981 CGGCATCCGC TTACAGACAA GCTGTGACCG TCTCCGGGAG CTGCATGTGT CAGAGGTTTT
5041 CACCGTCATC ACCGAAACGC GCGAGACGAA AGGGCCTCGT GATACGCCCTA TTTTATAGG
5101 TTAATGTCTAT GATAATAATG GTTCTTAGA CGTCAGGTGG CACTTTTCGG GGAATGTGC
5161 GCGGAACCCC TATTTGTTTA TTTTCTAAA TACATTCAA TATGTATCCG CTCATGAGAC
5221 AATAACCTCG ATAAATGCTT CAATAATATT GAAAAAGGAA GAGTATGAGT ATTCAACATT
5281 TCCGTGTGCG CCTTATTCCT TTTTGTGCG CATTGTGCTT TCCTGTTTTT GCTCACCAG
5341 AAACGCTGGT GAAAGTAAAA GATGCTGAAG ATCAGTTGGG TGCACGAGTG GGTTACATCG
5401 AACTGGATCT CAACAGCGGT AAGATCCTTG AGAGTTTTCG CCCCAGAGAA CGTTTTCCAA
5461 TGATGAGCAC TTTTAAAGTT CTGCTATGTG GCGCGGTATT ATCCGCTATT GACGCCGGGC
5521 AAGAGCAACT CGGTCGCGCG ATACACTATT CTCAGAATGA CTTGGTTGAG TACTCACCAG
5581 TCACAGAAAA GCATCTTACG GATGGCATGA CAGTAAGAGA ATTATGCACT GCTGCCATAA
5641 CCATGAGTGA TAACACTGCG GCCAATTAC TTCTGACAAC GATCGGAGGA CCGAAGGAGC
5701 TAACCGCTTT TTTGCACAAC ATGGGGGATC ATGTAACCTG CCTTGATCGT TGGGAACCGG
5761 AGCTGAATGA AGCCATACCA AACGACGAGC GTGACACCAC GATGCCTGTA GCAATGGCAA -

FIGURE 46C

5821 CAACGTTGCG CAAACTATTA ACTGGCGAAC TACTTACTCT AGCTTCCCGG CAACAATTAA
5881 TAGACTGGAT GGAGGCGGAT AAAGTTGCAG GACCACTTCT GCGCTCGGCC CTTCGGGCTG
5941 GCTGGTTTAT TGCTGATAAA TCTGGAGCCG GTGAGCGTGG GTCTCGCGGT ATCATTGCAG
6001 CACTGGGGCC AGATGGTAAG CCTCCCGTA TCGTAGTTAT CTACACGACG GGGAGTCAGG
6061 CAACTATGGA TGAACGAAAT AGACAGATCG CTGAGATAGG TGCCTCACTG ATTAAGCATT
6121 GGTAACTGTC AGACCAAGTT TACTCATATA TACTTTAGAT TGATTTAAAA CTTCATTTTT
6181 AATTTAAAAG GATCTAGGTG AAGATCCTTT TTGATAATCT CATGACCAAA ATCCCTTAAC
6241 GTGAGTTTTC GTTCCACTGA GCGTCAGACC CCGTAGAAAA GATCAAAGGA TCTTCTTGAG
6301 ATCCTTTTTT TCTGCGCGTA ATCTGCTGCT TGCAAAACAA AAAACCAACG CTACCAGCGG
6361 TGGTTTGTTC GCCGGATCAA GAGCTACCAA CTCTTTTTC GAAGGTAAC GGCTTCAGCA
6421 GAGCGCAGAT ACCAAATACT GTCCTTCTAG TGTAGCCGTA GTTAGGCCAC CACTTCAAGA
6481 ACTCTGTAGC ACCGCCTACA TACCTCGCTC TGCTAATCCT GTTACCAGTG GCTGTGCCA
6541 GTGGCGATAA GTCGTGTCTT ACCGGGTTGG ACTCAAGACG ATAGTTACCG GATAAGGCGC
6601 AGCGGTCGGG CTGAACGGGG GGTTCGTGCA CACAGCCAG CTGGAGCGA ACGACCTACA
6661 CCGAACTGAG ATACCTACAG CGTGAGCATT GAGAAAGCGC CACGCTTCCC GAAGGGAGAA
6721 AGGCGGACAG GTATCCGGTA AGCGGCAGGG TCGGAACAGG AGAGCGCAGC AGGGAGCTTC
6781 CAGGGGAAAA CGCCTGGTAT CTTTATAGTC CTGTCGGGTT TCGCCACCTC TGACTTGAGC
6841 GTCGATTTTT GTGATGCTCG TCAGGGGGGC GGAGCCTATG GAAAAACGCC AGCAACGCGG
6901 CCTTTTACG GTTCTCGGCC TTTTGCTGGC CTTTTGCTCA CATGTTCTTT CTGCGTTAT
6961 CCCCTGATTC TGTGGATAAC CGTATTACCG CCTTTGAGTG AGCTGATACC GCTCGCCGCA
7021 GCCGAACGAC CGAGCGCAGC GAGTCAGTGA GCGAGGAAGC GGAAGAGCGC CCAATACGCA
7081 AACCGCCTCT CCGCGCGCGT TGGCCGATTC ATTAATGCAG AGCTTGCAAT TCGCGCGTTT
7141 TTCAATATTA TTGAAGCATT TATCAGGGTT ATTGTCTCAT GAGCGGATAC ATATTGAT
7201 GTATTTAGAA AAATAAACAA ATAGGGGTTT CGCGCACATT TCCCCGAAAA GTGCCACCTG
7261 ACGTCTAAGA AACCATTATT ATCATGACAT TAACCTATAA AAATAGGCGT AGTACGAGGC
7321 CCTTTCACTC ATTAGATGCA TGTCGTTACA TAACTTACGG TAAATGGCCC GCCTGGCTGA
7381 CCGCCCAACG ACCCCCGCCC ATTGACGTCA ATAATGACGT ATGTTCCCAT AGTAACGCCA
7441 ATAGGGACTT TCCATTGACG TCAATGGGTG GAGTATTAC G

FIGURE 4b

136/240
FIGURE 47ApDEST 27 GST Amino Fusion in pCMV Sport-neo:
Vector

CMV Promoter

600 // nac ggt ggg agg tct ata taa gca gag ctc gtt tag tga acc gtc aga tcy
ntg cca ccc tcc aga tar att cgt ctc gag caa atc act tgg dag tct agc

651 cct gga gac gcc atc cac gct gtt ttg acc tcc ata gaa gac acc ggg acc
gga cct ctg cgg tag gtg cga caa aac tgg agg tat ctt ctg tgg occ tgg

702 gat cca gcc tcc gga ctc tag cct agg cgg cgg acc atg gcc cct ata cta
cta ggt cgg agg cct gag atc gga tcc gga gcc tgg cag cgg gga tat gat

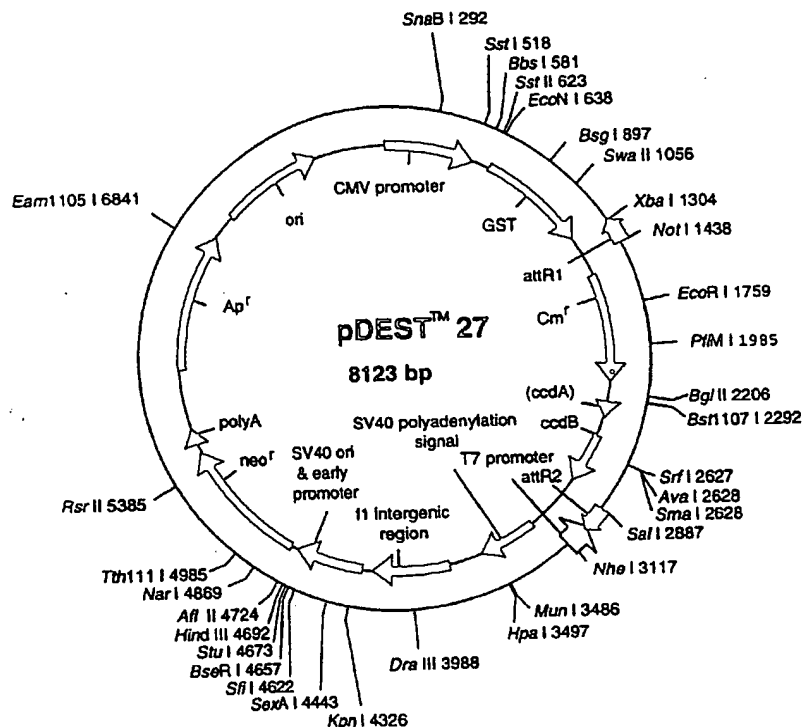
753 ggt tat tgg aaa att aag ggc ctt gtg caa ccc act oga ctt ctt ttg gaa
cca ata acc ttt taa ttc ccg gaa cac gtt ggg tga gct gaa gaa aac ctt

804 tat ctt gaa gaa aaa tat gaa gag cat ttg tat gag cgc gat gaa ggt gat
ata gaa ctt ctt ttt ata ctt ctc gta aac ata ctc gcg cta ctt cca cta

1365 // ttt ggt ggt ggc gac cat cct cca aaa tgg gat ctg gtt ccg cgt tct aga
aaa cca cca ccg ctg gta gga ggt ttt agc cta gac caa gga gca aga tct

1416 tca aca agt ttg tac aaa aaa gct gaa cga gaa acg
agt tgt tca aac atg ttg ttt cga ctt gcc ctt tgc

attR1



pDEST27 8123 bp (rotated to position 7800)

Location (Base Nos.)	Gene Encoded
130..793	GST
803..927	attR1
1036..1695	CmR
1815..1899	inactivated ccdA
2037..2342	ccdB
2383..2507	attR2
2693..3055	SV40 polyA
3250..3705	f1 intergenic region
3769..4187	SV40 promoter
4232..5026	neo
5090..5138	polyA
5549..6409	Apr
6558..7197	ori
7628..27	CMV promoter

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1 ATAAGCAGAG CTCGTTAGT GAACCGTCAG ATCGCCTGGA GACGCCATCC ACGCTGTTTT
61 GACCTCCATA GAAGACACCG GGACCGATCC AGCCTCCGGA CTCTAGCCTA GGCCGCGGAC
121 CATGGCCCTT ATACTAGGTT ATTGGAATAT TAAGGGCCTT GTGCAACCCA CTCGACTTCT
181 TTTGGAATAT CTTGAAGAAA AATATGAAGA GCATTGTAT GAGCGCGATG AAGGTGATAA
241 ATGGCGAAAC AAAAAGTTTG AATTGGGTTT GGAGTTTCCC AATCTTCCTT ATTATATTGA
301 TGGTGATGTT AAATTAACAC AGTCTATGGC CATCATACGT TATATAGCTG ACAAGCACAA
361 CATGTTGGGT GGTGTGCCAA AAGAGCGTGC AGAGATTCA ATGCTTGAAG GAGCGGTTTT
421 GGATATTAGA TACGGTGTTC CGAGAATTGC ATATAGTAAA GACTTTGAAA CTCTCAAAGT
481 TGATTTCTT AGCAAGCTAC CTGAAATGCT GAAAAATGTC GAAGATCGTT TATGTCATAA
541 AACATATTTA AATGGTGATC ATGTAACCCA TCCTGACTTC ATGTTGTATG ACGCTCTTGA
601 TGTTGTTTTA TACATGGACC CAATGTGCCT GGATGCGTTC CCAAAATTAG TTTGTTTTAA
661 AAAACGTATT GAAGCTATCC CACAAATTGA TAAGTACTTG AAATCCAGCA AGTATATAGC
721 ATGGCCTTTG CAGGGCTGGC AAGCCACGTT TGGTGGTGGC GACCATCCTC CAAAATCGGA
781 TCTGGTCCCG CGTTCTAGAT CAACAAGTTT GTACAAAAAA GCTGAACGAG AAACGTAAAA
841 TGATATAAAT ATCAATATAT TAAATTAGAT TTTGCATAAA AAACAGACTA CATAATACTG
901 TAAACACAAA CATATCCAGT CACTATGGCG GCCGCATTAG GCACCCGAGG CTTTACACTT
961 TATGCTTCCG GCTCGTATAA TGTGTGGATT TTGAGTTAGG ATCCGGCGAG ATTTTCAGGA
1021 GCTAAGGAAG CTAAATGGA GAAAAAATC ACTGGATATA CCACCGTTGA TATATCCCAA
1081 TGGCATCGTA AAGAACATTT TGAGGCATTT CAGTCAGTTG CTCATGTAC CTATAACCAAG
1141 ACCGTTCCAG TGGATATTAC GGCCTTTTTA AAGACCGTAA AGAAAAATAA GCACAAGTTT
1201 TATCCGGCCT TTATTCACAT TCTTGCCCGC CTGATGAATG CTCATCCGGA ATTCCGTATG
1261 GCAATGAAAG ACGGTGAGCT GGTGATATGG GATAGTGTTC ACCCTTGTTA CACCGTTTTC
1321 CATGAGCAAA CTGAAACGTT TTCATCGCTC TGGAGTGAAT ACCACGACGA TTTCCGGCAG
1381 TTTCTACACA TATATTCGCA AGATGTGGCG TGTTACGGTG AAAACCTGGC CTATTTCCCT
1441 AAAGGGTTTA TTGAGAATAT GTTTTTCGTC TCAGCCAATC CCTGGGTGAG TTTCAACAGT
1501 TTTGATTATA ACGTGCCCAA TATGGACAAC TTCTTCGCCC CCGTTTTCAC CATGGGCAAA
1561 TATTATACGC AAGGCGACAA GGTGCTGATG CCGCTGGCGA TTCAGGTTCA TCATGCCGTC
1621 TGTGATGGCT TCCATGTCCG CAGAATGCTT AATGAATTAC AACAGTACTG CGATGAGTGG
1681 CAGGGCGGGG CGTAAAGATC TGGATCCGGC TTAATAAAG CCAGATAACA GTATGCGTAT
1741 TTGCGCGCTG ATTTTTCGGG TATAAGAATA TATACTGATA TGTATACCCG AAGTATGTCA
1801 AAAAGAGGTG TGCTATGAAG CAGCGTATTA CAGTGACAGT TGACAGCGAC AGCTATCAGT
1861 TGCTCAAGGC ATATATGATG TCAATATCTC CGGTCTGGTA AGCACAACCA TGCAGAAATGA
1921 AGCCCGTCGT CTGCGTGCCG AACGCTGGAA AGCGGAAAAT CAGGAAGGGA TGGCTGAGGT
1981 CGCCCGGTTT ATTGAAATGA ACGGCTCTTT TGCTGACGAG AACAGGACT GGTGAAATGC
2041 AGTTTAAGGT TTACACCTAT AAAAGAGAGA GCCGTTATCG TCTGTTTGTG GATGTACAGA
2101 GTGATATTAT TGACACGCCC GGGCGACGGA TGGTGATCCC CCTGGCCAGT GCACGTCTGC
2161 TGTGAGATAA AGTCTCCCGT GAACCTTACC CGGTGGTGCA TATCGGGGAT GAAAGCTGGC
2221 GCATGATGAC CACCGATATG GCCAGTGTGC CGGTCTCCGT TATCGGGGAA GAAGTGGCTG
2281 ATCTCAGCCA CCGCGAAAAT GACATCAAAA ACGCCATTAA CTGATGTTC TGGGGAATAT -

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Figure 47B

2341 AAATGTCAGG CTCCCTTATA CACAGCCAGT CTGCAGGTCG ACCATAGTGA CTGGATATGT
2401 TGTGTTTTAC AGTATTATGT AGTCTGTTTT TTATGCAAAA TCTAATTAA TATATTGATA
2461 TTTATATCAT TTTACGTTTC TCGTTCAGCT TTCITGTACA AAGTGGTTGA TCGCGTGCAT
2521 GCGACGTCAT AGCTCTCTCC CTATAGTGAG TCGTATTATA AGCTAGGCAC TGGCCGTCGT
2581 TTTACAACGT CGTGACTGGG AAAACTGCTA GCTTGGGATC TTTGTGAAGG AACCTTACTT
2641 CTGTGGTGTG ACATAATTGG ACAAACTACC TACAGAGATT TAAAGCTCTA AGGTAAATAT
2701 AAAATTTTTA AGTGATATAAT GTGTTAAACT AGCTGCATAT GCTTGCTGCT TGAGAGTTTT
2761 GCTTACTGAG TATGATTTAT GAAAATATTA TACACAGGAG CTAGTGATTCT TAATTGTTTG
2821 TGTATTTTAG ATTCACAGTC CCAAGGCTCA TTTCAGGCCC CTCAGTCTCT ACAGTCTGTT
2881 CATGATCATA ATCAGCCATA CCACATTTGT AGAGGTTTTA CTTGCTTTAA AAAACCTCCC
2941 ACACCTCCCC CTGAACCTGA AACATAAAAT GAATGCAATT GTTGTGTTA ACTTGTTTAT
3001 TGCAGCTTAT AATGGTTACA AATAAAGCAA TAGCATCACA AATTTCACAA ATAAAGCAAT
3061 TTTTTCACGT CATTCTAGTT GTGGTTTGTC CAAATCATC AATGTATCTT ATCATGTCTG
3121 GATCGATCCT GCATTAATGA ATCGGCCAAC GCGCGGGGAG AGGCGGTTTG CGTATTGGCT
3181 GCGGTAATAG CGAAGAGGCC CGCACCGATC GCCCTTCCCA ACAGTTGCGC AGCCTGAATG
3241 GCGAATGGGA CGCGCCCTGT AGCGGCGCAT TAAGCGCGGC GGGTGTGGTG GTTACGCGCA
3301 GCGTGACCGC TACACTTGCC AGCGCCCTAG CGCCCGCTCC TTTGCTTTC TTCCCTTCTT
3361 TTCTCGCCAC GTTCGCGGCG TTTCCCGTCT AAGCTCTAAA TCGGGGGCTC CCTTATAGGT
3421 TCCGATTTAG TGCTTTACGG CACCTCGACC CCAAAAAACT TGATTAGGGT GATGGTTCAC
3481 GTAGTGGGCC ATCGCCCTGA TAGACGGTTT TCGCCCTTT GACGTGGAG TCCACGTCTT
3541 TTAATAGTGG ACTCTTGTT CAAACTGGAA CAACACTCAA CCCTATCTCG GTCTATTCTT
3601 TTGATTTATA AGGGATTTTG CCGATTTCGG CCTATTGGTT AAAAAATGAG CTGATTTAAC
3661 AAATATTATA CGCGAATTTT AACAAAATAT TAACGTTTAC AATTTGCGCT GATCGCGTAT
3721 TTTCTCCTTA CGCATCTGTG CGGTATTTC CACCGCATAC GCGGATCTGC GCAGCACCAT
3781 GGCCTGAAAT AACCTCTGAA AGAGGAACCT GGTAGGTAC CTTCTGAGGC GGAAGGAACC
3841 AGCTGTGGAA TGTGTGTCAG TTAGGGTGTG GAAAGTCCCC AGGCTCCCCA GCAGGCAGAA
3901 GTATGCAAAAG CATGCATCTC AATTAGTCAG CAACCAAGTG TGGAAAGTCC CCAGGCTCCC
3961 CAGCAGGCAG AAGTATGCAA AGCATGCATC TCAATTAGTC AGCAACCATA GTCCCCCCCC
4021 TAACCTCGCC CATCCCGCCC CTAACCTCCG CAGTTCCGC CCATTCTCCG CCCCATGGCT
4081 GACTAATTTT TTTTATTAT GCAGAGGCCG AGGCCGCTCT GGCCTCTGAG CTATTCCAGA
4141 AGTAGTGAGG AGGCTTTTTT GGAGGCTTAG GCTTTTGCAA AAAGCTTGAT TCTTCTGACA
4201 CAACAGTCTC GAACTTAAGG CTAGAGCCAC CATGATTGAA CAAGATGGAT TGCACGCAGG
4261 TTCTCCGGCC GCTTGGGTGG AGAGGCTATT CGGCTATGAC TGGGCACAA AGACAATCGG
4321 CTGCTCTGAT GCGCGCTGT TCCGGCTGTC AGCGCAGGGG CGCCCGGTTT TTTTGTCAA
4381 GACCGACCTG TCCGGTGCCC TGAATGAAC GCAGGACGAG GCAGCGCGGC TATCGTGGCT
4441 GGGCACGACG GGGCTTCCCT GCGCAGCTGT GCTCGACGTT GTCAGTGAAG CGGGAAGGGA
4501 CTGGCTGCTA TTGGGCGAAG TGCCGGGGCA GGATCTCCTG TCATCTCACC TTGCTCTGTC
4561 CGAGAAAGTA TCCATCATGG CTGATGCAAT GCGGCGGCTG CATACGCTTG ATCCGGCTAC
4621 CTGCCCATTG GACCACCAAG CGAAACATCG CATCGAGCGA GCACGTACTC GGATGGAAGC
4681 CGGTCTTGTC GATCAGGATG ATCTGGAAGA AGAGCATCAG GGGCTCGGCG CAGCCGAAC
4741 GTTCGCCAGG CTCAAGGCGC GCATGCCCGA CGCGGAGGAT CTCGTCGTGA CCCATGGCGA
4801 TGCCCTGCTG CCGAATATCA TGGTGGAAAA TGCCCGCTTT TCTGGATTCA TCGACTGTGG
4861 CCGGCTGGGT GTGGCGGACC GCTATCAGGA CATAGCGTTG GCTACCCGTG ATATTGCTGA
4921 AGAGCTTGGC GCGGAATGGG CTGACCGCTT CCTCGTGTG TACGCTATCG CCGCTCCCGA
4981 TTCGACGCGC ATCGCCTTCT ATCGCCTTCT TGACGAGTTC TTCTGAGCGG GACTCTGGGG
5041 TTCGAAATGA CCGACCAAGC GACGCCCAAC CTGCCATCAC GATGGCCGCA ATAAATATC
5101 TTTATTTTCA TTACATCTGT GTGTTGGTTT TTTGTGTGAA TCGATAGCGA TAAGGATCCG
5161 CGTATGGTGC ACTCTCAGTA CAATCTGCTC TGATGCCGCA TAGTTAAGCC AGCCCCGACA
5221 CCGCCCAACA CCGCTGACG CGCCCTGACG GGCTTGTCTG CTCCCGGCT CCGCTTACAG
5281 ACAAGCTGTG ACCGTCTCCG GGAGCTGCAT GTGTCAGAGG TTTTACCCTG CATCACCGAA
5341 ACGCGCGAGA CGAAAGGGCC TCGTGATACG CCTATTTTAA TAGGTTAATG TCATGATAAT
5401 AATGGTTTCT TAGACGTCAG GTGGCACTTT TCGGGGAAAT GTGCGCGGAA CCCCTATTTG
5461 TTTATTTTTT TAAATACATT CAAATATGTA TCCGCTCATG AGACAATAAC CCTGATAAAT
5521 GCTTCAATAA TATTGAAAAA GGAAGAGTAT GAGTATTCAA CATTTCCGTG TCGCCCTTAT
5581 TCCCTTTTTT GCGGCATTTT GCCTTCTGT TTTTGCTCAC CCAGAAACGC TGGTGAAAGT
5641 AAAAGATGCT GAAGATCAGT TGGGTGCACG AGTGGGTTAC ATCGAACTGG ATCTCAACAG
5701 CGGTAAGATC CTTGAGAGTT TCGCCCCGA AGAACGTTTT CCAATGATGA GCACTTTTAA
5761 AGTTCTGCTA TGTGGCGCGG TATTATCCCG TATTGACGCC GGGCAAGAGC AACTCGGTGC

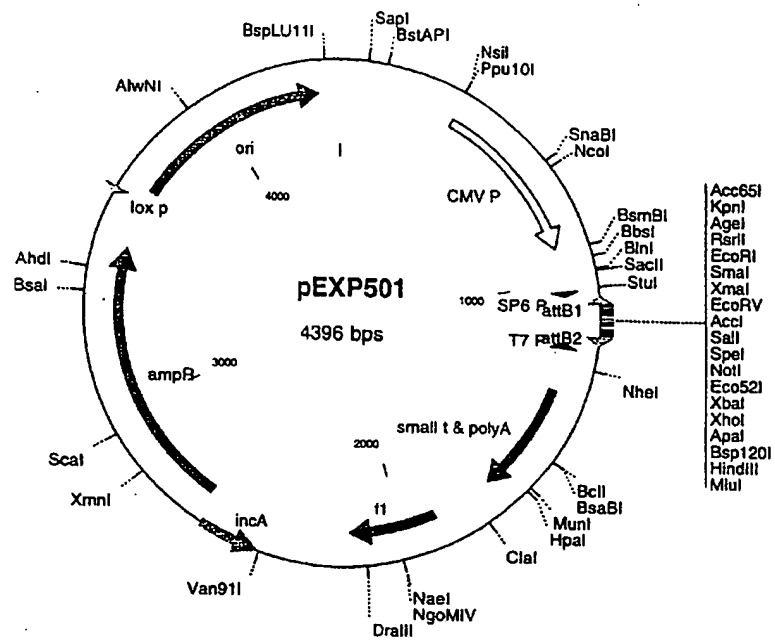
FIGURE 47C

5821 CCGCATACAC TATTCTCAGA ATGACTTGGT TGAGTACTCA CCAGTCACAG AAAAGCATCT
5881 TACGGATGGC ATGACAGTAA GAGAAATTATG CAGTGCTGCC ATAACCATGA GTGATAACAC
5941 TGCGGCCAAC TTACTTCTGA CAACGATCGG AGGACCGAAG GAGCTAACCG CTTTTTTGCA
6001 CAACATGGGG GATCATGTAA CTCGCCCTGA TCGTTGGGAA CCGGAGCTGA ATGAAGCCAT
6061 ACCAAACGAC GAGCGTGACA CCACGATGCC TGTAGCAATG GCAACAACGT TGGCAAACCT
6121 ATTAACCTGC GAACTACTTA CTCTAGCTTC CCGGCAACAA TTAATAGACT GGATGGAGGC
6181 GGATAAAGTT GCAGGACCAC TTCTGCGCTC GGCCCTTCCG GCTGGCTGGT TTATTGCTGA
6241 TAAATCTGGA GCCGGTGAGC GTGGGTCTCG CGGTATCATT GCAGCACTGG GGCAGATGG
6301 TAAGCCCTCC CGTATCGTAG TTATCTACAC GACGGGGAGT CAGGCAACTA TGGATGAACG
6361 AAATAGACAG ATCGCTGAGA TAGGTGCCTC ACTGATTAAG CATTGGTAAC TGTGAGACCA
6421 AGTTTACTCA TATATACTTT AGATTGATTT AAAACTTCAT TTTTAATTTA AAAGGATCTA
6481 GGTGAAGATC CTTTTTGATA ATCTCATGAC CAAATCCCT TAACGTGAGT TTTCTGTTCA
6541 CTGAGCGTCA GACCCCGTAG AAAAGATCAA AGGATCTTCT TGAGATCCTT TTTTTCTGCG
6601 CGTAATCTGC TGTCTGCAA CAAAAAACC ACCGCTACCA CCGGTGGTTT GTTTGCCGGA
6661 TCAAGAGCTA CCAACTCTTT TTCCGAAGGT AACTGGCTTC AGCAGAGCGC AGATACCAAA
6721 TACTGTCCCT CTAGTGTAGC CGTAGTTAGG CCACCACTTC AAGAACTCTG TAGCACCGCC
6781 TACATACCTC GCTCTGCTAA TCCTGTTACC AGTGGCTGCT GCCAGTGGCG ATAAGTCGTG
6841 TCTTACCGGG TTGGACTCAA GACGATAGTT ACCGGATAAG GCGCAGCGGT CGGGCTGAAC
6901 GGGGGGTTCC TGACACAGC CCAGCTTGGA GCGAACGACC TACACCGAAC TGAGATACCT
6961 ACAGCGTGAG CATGAGAAA GCGCCACGCT TCCCGAAGGG AGAAAGGCGG ACAGGTATCC
7021 GGTAAGCGGC AGGGTCGGAA CAGGAGAGCG CACGAGGGAG CTTCAGGGG GAAACGCCTG
7081 GTATCTTTAT AGTCTGTGCG GGTTTCGCCA CCTCTGACTT GAGCGTCGAT TTTTGTGATG
7141 CTCGTACGGG GGGCGGAGCC TATGGAAAA CGCCAGCAAC GCGGCCTTTT TACGGTTCCCT
7201 GGCCTTTTGC TGGCCTTTTG CTCACATGTT CTTTCTGCG TTATCCCTG ATTCTGTGGA
7261 TAACCGTATT ACCGCTTTG AGTGAGCTGA TACCGCTCGC GCGAGCCGAA CGACCGAGCG
7321 CAGCGAGTCA GTGAGCGAGG AAGCGGAAGA GCGCCCAATA CGCAAACCGC CTCTCCCGC
7381 GCGTTGGCCG ATTCATTAAT GCAGAGCTTG CAATTCGCGC GTTTTCAAT ATTATTGAAG
7441 CATTTATCAG GGTATTGTG TCATGAGCGG ATACATATTT GAATGTATTT AGAAAAATAA
7501 ACAAATAGGG GTTCCGCGCA CATTTCCCG AAAAGTGCCA CCTGACGTCT AAGAAACCAT
7561 TATTATCATG ACATTAACCT ATAAAAATAG GCGTAGTACG AGGCCCTTTC ACTCATTAGA
7621 TGCATGTCGT TACATAACTT ACGGTAAATG GCCCGCCTGG CTGACCGCCC AACGACCCCC
7681 GCCCATGAC GTCAATAATG ACGTATGTT CCATAGTAAC GCCAATAGGG ACTTTCATT
7741 GACGTCAATG GGTGGAGTAT TTACGGTAAA CTGCCCACTT GGCAGTACAT CAAGTGTATC
7801 ATATGCCAAG TACGCCCCCT ATTGACGTCA ATGACGGTAA ATGGCCCCG TGGCATTATG
7861 CCCAGTACAT GACCTTATGG GACTTTCCTA CTTGGCAGTA CATCTACGTA TTAGTCATCG
7921 CTATTACCAT GGTGATGCGG TTTTGGCAGT ACATCAATGG GCGTGGATAG CGGTTTGACT
7981 CACGGGGATT TCCAAGTCTC CACCCCATG ACGTCAATGG GAGTTTGTG TGGCACCAAA
8041 ATCAACGGGA CTTTCCAAAA TGTCGTAACA ACTCCGCCCC ATTGACGCAA ATGGGCGGTA
8101 GCGGTGTACG GTGGGAGGTC TAT

FIGURE 471

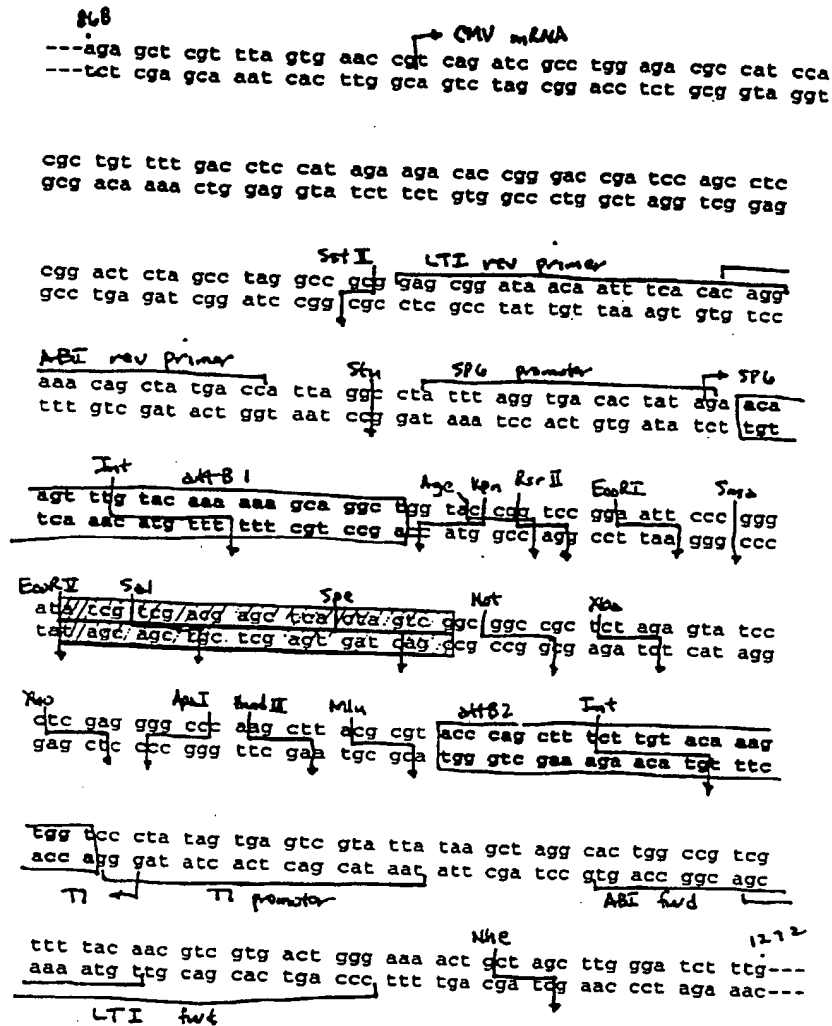
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Figure 4B A: pEXP501: pCMV-SPORT 6 host for attB Libraries



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Figure 48B: pEXP501 (cont'd). **Features of the att B cloning vector, pEXP501.** Bases within hatched area are replaced by cDNA in some LTI cDNA libraries.



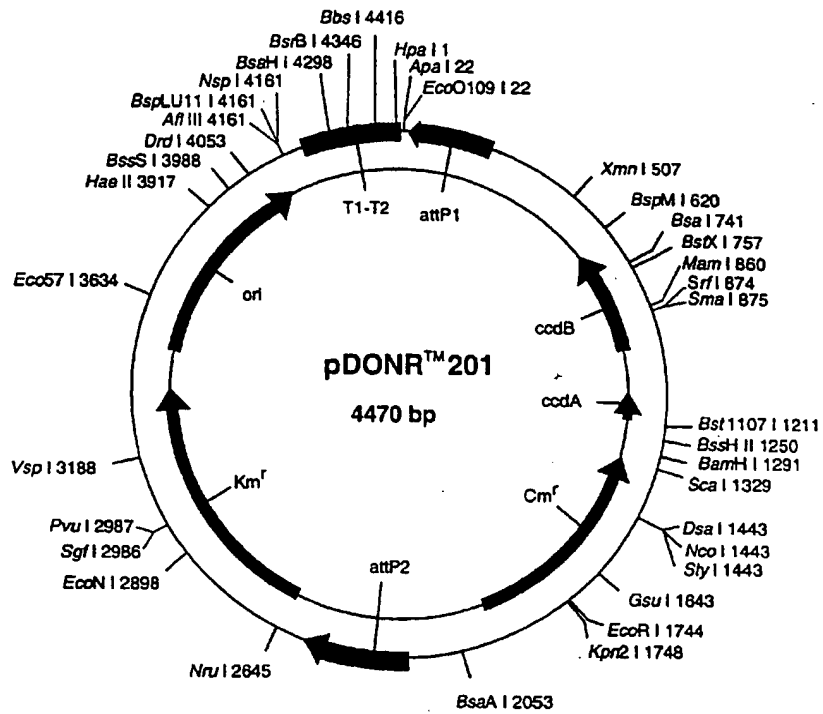
pEXP501 4396 bp

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1 CCATTCGCCA TTCAGGCTGC GCAACTGTTG GGAAGGGCGA TCGGTGCGGG CCTCTTCGCT
61 ATTACGCCAG CCAATACGCA AACCGCCTCT CCCCGCGCGT TGGCCGATTC ATTAATGCAG
121 GATCGATCCA GACATGATAA GATACATTGA TGAGTTTGGA CAAACCACAA CTAGAATGCA
181 GTGAAAAAAA TGCTTTATTT GTGAAATTTG TGATGCTATT GCTTTATTTG TAACCATTAT
241 AAGCTGCAAT AAACAAGTTA ACACAAACAA TTGCATTTCAT TTTATGTTTC AGGTTCCAGGG
301 GGAGGTGTGG GAGGTTTTTT AAAGCAAAGTA AAACCTCTAC AAATGTGGTA TGGCTGATTA
361 TGATCATGAA CAGACTGTGA GGACTGAGGG GCCTGAAATG AGCCTTGGGA CTGTGAATCT
421 AAAATACACA AACAAATTAGA ATCACTAGCT CCTGTGTATA ATATTTTCAT AAATCATACT
481 CAGTAAGCAA AACTCTCAAG CAGCAAGCAT ATGCAGCTAG TTTAACACAT TATACACTTA
541 AAAATTTTAT ATTTACCTTA GAGCTTTAAA TCTCTGTAGG TAGTTTGTCC AATTATGTCA
601 CACCACAGAA GTAAGGTTCC TTCACAAAGA TCCCAAGCTA GCAGTTTTC CAGTCACGAC
661 GTTGTAAGAC GACGGCCAGT GCCTAGCTTA TAATACGACT CACTATAGGG ACCACTTTGT
721 ACAAGAAAGC TGGGTACGCG TAAGCTTGGG CCCCTCGAGG GATCCTCTAG AGCGGCCGCC
781 GACTAGTGAG CTCGTCGACG ATATCCCGGG AATTCCGGAC CGGTACCAGC CTGCTTTTTT
841 GTACAAACTT GTTCTATAGT GTCACCTAAA TAGGCCTAAT GGTCAATAGT GTTCTCTGTG
901 TGAATTTGTT ATCCGCTCCG CGGCTTAGGC TAGAGTCCGG AGGCTGGATC GGTCCCGGTG
961 TCTTCTATGG AGGTCAAAC AGCGTGGATG GCGTCTCCAG GCGATCTGAC GGTTCACATA
1021 ACGAGCTCTG CTTATATAGA CCTCCCACCG TACACGCCTA CCGCCCATTT GCGTCAATGG
1081 GCGCGAGTTG TTACGACATT TTGAAAGTC CCGTTGATTT TGGTGCCAAA ACAAACTCCC
1141 ATTGACGTCA ATGGGGTGGA GACTTGGAAA TCCCGGTGAG TCAAACCGCT ATCCACGCCC
1201 ATTGATGTAC TGCCAAAACC GCATCACCAT GGTAAATAGCG ATGACTAATA CGTAGATGTA
1261 CTGCCAAGTA GGAAAGTCCC ATAAGGTCAT GTACTGGGCA TAATGCCAGG CGGGCCATTT
1321 ACCGTCAATG ACGTCAATAG GGGGCGTACT TGGCATATGA TACACTTGAT GTACTGCCAA
1381 GTGGGCGATT TACCGTAAAT ACTCCACCCA TTGACGTCAA TGGAAAGTCC CTATTGGCGT
1441 TACTATGGGA ACATACGTCA TTATTGACGT CAATGGGCGG GGGTCTGTGG GCGGTACAGC
1501 AGGCGGGCCA TTTACCGTAA GTTATGTAAC GACATGCATC TAATGAGTGA AAGGGCCTCG
1561 TACTACGCCT ATTTTATAGT GTTATGTCA TGATAATAAT GGTTCCTTAG ACGTCAGGTG
1621 GCACCTTTTC GGGAAATGTG CGCGGAACCC CTATTGTGTT ATTTTCTTAA ATACACTCAA
1681 ATATGTATCC GCTCATGAGA CANTAACCCT GATAAATGCT TCAATAATAT TGA AAAACGC
1741 GCGAATTGCA AGCTCTGCAT TAATGAATCG GCCAACGCGC GGGGAGAGGC GGTTCGCGTA
1801 TTGGGCGCTC TTCCGCTTCC TCGCTCACTG ACTCGCTGCG CTCGGTCTGT GCGCTGCGGC
1861 GAGCGGTATC AGCTCACTCA AAGGCGGTAA TACGGTTATC CACAGAATCA GGGGATAACG
1921 CAGGAAAGAA CATGTAGCA AAAGGCCAGC AAAAGGCCAG GAACCGTAAA AAGGCCCGCT
1981 TGCTGGCGTT TTTCCATAGG CTCCGCCCCC CTGACGAGCA TCACAAAAAT CGACGCTCAA
2041 GTCAGAGGTG GCGAAACCCG ACAGGACTAT AAAGATACCA GCGGTTTCCC CTTGGAAGCT
2101 CCCTCGTGCG CTCCTCTGTT CCGACCCTGC CGCTTACCGG ATACCTGTCC GCCTTCTTCC
2161 CTTGCGGAAG CGTGGCGCTT TCTCAATGCT CACGCTGTAG GTATCTCAGT TCGGTGTAGG
2221 TCGTTCGCTC CAAGCTGGGC TGTGTGCACG AACCCCCCGT TCAGCCCGAC CGCTGCGCCT
2281 TATCCGGTAA CTATCGTCTT GAGTCCAACC CGGTAAGACA CGACTTATCG CCACTGGCAG
2341 CAGCCACTGG TAACAGGATT AGCAGAGCGA GGTATGTAGG CCGTGCTACA GAGTCTTGA
2401 AGTGGTGGCC TAACACGGC TACACTAGAA GGACAGTATT TGTATCTGC GCTCTGTGA
2461 AGCCAGTTAC CTTCGGAAAA AGAGTTGGTA GCTCTTGATC CGGCAACAA ACCACCGCTG
2521 GTAGCGGTGG TTTTGTGTT TGCAAGCAGC AGATTACGCG CAGAAAAAAA GGATCTCAAG
2581 AAGATCCTTT GATCTTTTCT ACGGGGTCTG ACGCTCAGTG GAACGAAAAC TCACGTTAAG
2641 GGAATTTGGT CATGCCATAA CTTGCTATAG CATACATTAT ACGAAGTTAT GGCATGAGAT
2701 TATCAAAAAG GATCTTCACC TAGATCCTTT TAAATTAAAA ATGAAGTTT AAATCAATCT
2761 AAAGTATATA TGAGTAAACT TGGTCTGACA GTTACCAATG CTTAATCAGT GAGGCACCTA
2821 TCTCAGCGAT CTGTCTATTT CGTTCATCCA TAGTTGCTG ACTCCCGTC GTGTAGATAA
2881 CTACGATACG GGAGGGCTTA CCATCTGGCC CCAGTGTGTC AATGATACCG CGAGACCCAC
2941 GCTCAGCGGC TCCAGATTTA TCAGCAATAA ACCAGCCAGC CGGAAGGGCC GAGCGCAGAA
3001 GTGGTCCTGC AACTTTATCC GCCTCCATCC AGTCTATTAA TTGTTGCCGG GAAGCTAGAG
3061 TAAGTAGTTC GCCAGTTAAT AGTTTGCACA ACGTTGTTGC CATTGCTACA GGCATCGTGG
3121 TGTACGCTC GTCGTTTGGT ATGCTTCAT TCAGCTCCGG TTCCCAACGA TCAAGCGGAG-
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FIGURE 48C

3181 TTACATGATC CCCCATGTTG TGCAAAAAAG CGGTTAGCTC CTTCCGGTCCT CCGATCGTTG
3241 TCAGAAGTAA GTTGGCCGCA GTGTTATCAC TCATGGTTAT GGCAGCACTG CATAATTCTC
3301 TTACTGTCAT GCCATCCGTA AGATGCTTTT CTGTGACTGG TGAGTACTCA ACCAAGTCAT
3361 TCTGAGAATA GTGTATGCGG CGACCGAGTT GCTCTTGCCC GGCCTCAATA CGGGATAATA
3421 CCGCGCCACA TAGCAGAACT TTAAAAGTGC TCATCATTGG AAAACGTTCT TCGGGGCGAA
3481 AACTCTCAAG GATCTTACCG CTGTTGAGAT CCAGTTCGAT GTAACCCACT CGTGCAACCA
3541 ACTGATCTTC AGCATCTTTT ACTTTCACCA GCGTTTCTGG GTGAGCAAAA ACAGGAAGGC
3601 AAAATGCCGC AAAAAAGGGA ATAAGGGCGA CACGGAAATG TTGAATACTC ATACTCTTCC
3661 TTTTTCATAA TTATTGAAGC ATTTATCAGG GTTATTGTCT CATGCCAGGG GTGGGCACAC
3721 ATATTTGATA CCAGCGATCC CTACACAGCA CATAATTCAA TCGACTTCC CTCTATCGCA
3781 CATCTTAGAC CTTTATTCTC CCTCCAGCAC ACATCGAAGC TGCCGAGCAA GCCGTTCTCA
3841 CCACTCCAAG ACCTGGCATG AGCGGATACA TATTTGAATG TATTTAGAAA AATAAACAAA
3901 TAGGGGTTC GCGCACATTT CCCCAGAAAAG TGCCACCTGA AATTGTAAAC GTTAATATTT
3961 TGTTAAAATT CGCGTTAAAT TTTTGTAAA TCAGCTCATT TTTTAACCA TAGGCCGAAA
4021 TCGGCAAAAT CCCTTATAAA TCAAAAGAAT AGACCGAGAT AGGGTTGAGT GTTGTTCAG
4081 TTTGGAACAA GAGTCCACTA TTAAAGAACG TGGACTCCAA CGTCAAAGGG CGAAAAACCG
4141 TCTATCAGGG CGATGGCCCA CTACGTGAAC CATCACCTTA ATCAAGTTTT TTGGGGTCGA
4201 GGTGCCGTAA AGCACTAAAT CGGAACCCTA AAGGGAGCCC CCGATTAGA GCTTGACGGG
4261 GAAAGCCGGC GAACGTGGCG AGAAAGGAAG GGAAGAAAGC GAAAGGAGCG GCGCTAGGG
4321 CGCTGGCAAG TGTAGCGGTC ACGCTGCGCG TAACCACCAC ACCCGCCGCG CTTAATGCGC
4381 CGCTACAGGG CGCGTC

FIGURE 48D



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pDONR201 4470 bp (rotated to position 3516)

Location (Base Nos.)	Gene Encoded
260..29	attP1
656..961	ccdB
1099..1184	ccdA
1303..1962	Cmr
2210..2442	attP2
2565..3374	Kmr
3495..4134	ori

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1 GTTAAACGCTA GCATGGATCT CGGGCCCCAA ATAATGATTT TATTTTGACT GATAGTGACC
61 TGTTTCGTTGC AACAAATTGA TGAGCAATGC TTTTATATAA TGCCAACTTT GTACAAAAAA
121 GCTGAACGAG AAACGTAAAA TGATATAAAT ATCAATATAT TAAATTAGAT TTGTCATAAA
181 AAACAGACTA CATAATACTG TAAAACACAA CATATCCAGT CACTATGAAT CAACTACTTA
241 GATGGTATTA GTGACCTGTA GTCGACCGAC AGCCTTCCAA ATGTTCTTCG GGTGATGCTG
301 CCAACTTAGT CGACCGACAG CTTTCCAAAT GTTCTTCTCA AACGGAATCG TCGTATCCAG
361 CCTACTCGCT ATTGTCCCTA ATGCCGTATT AAATCATAAA AAGAAATAAG AAAAAGAGGT
421 GCGAGCCTCT TTTTGTGTG ACAAAATAAA AACATCTACC TATTCATATA CGCTAGTGTC
481 ATAGTCCTGA AAATCATCTG CATCAAGAAC AATTCACAA CTCTTATACT TTCTCTTAC
541 AAGTCGTTTC GCTTCATCTG GATTTTCAGC CTCTATACTT ACTAAACGTG ATAAAGTTTC
601 TGTAATTTCT ACTGTATCGA CTGCAGACT GGCTGTGTAT AAGGGAGCCT GACATTATATA
661 TTCCCCAGAA CATCAGGTTA ATGGCGTTT TGATGTCATT TTCGCGGTGG CTGAGATCAG
721 CCACCTTCTC CCCGATAACG GAGACCGGCA CACTGGCCAT ATCGGTGGTC ATCATGCGCC
781 AGCTTTTCAT CCCGATATGC ACCACCGGGT AAAGTTCACG GGAGACTTTA TCTGACAGCA
841 GACGTGCACT GGCCAGGGGG ATCACCATCC GTCGCCCCGG CGTGTCATA ATATCACTCT
901 GTACATCCAC AAACAGACGA TAACGGCTCT CTCTTTTATA GGTGTAAACC TTAACTTGCA
961 TTTCAACAGT CCCTGTTCTC GTCAGCAAAA GAGCCGTTCA TTTCAATAAA CCGGGCGACC
1021 TCAGCCATCC CTTCTGATT TCCGCTTTC CAGCGTTCGG CACGAGACG ACGGGCTTCA
1081 TTCTGCATGG TTGTGCTTAC CAGACCGGAG ATATGACAT CATATATGCC TTGAGCAACT
1141 GATAGCTGTC GCTGTCAACT GTCAGTGTAA TACGCTGCTT CATAGCACAC CTCTTTTGA
1201 CATACTTCGG GTATACATAT CAGTATATAT TCTTATACCG CAAAAATCAG CGCGCAATA
1261 CGCATACTGT TATCTGGCTT TTAGTAAGCC GGATCCACGC GATTACGCCC CGCCCTGCCA
1321 CTCATCGCAG TACTGTTGTA ATTCATTAAG CATTCGCGC ACATGGAAGC CATCACAGAC
1381 GGCATGATGA ACCTGAATCG CCAGCGGCAT CAGCACCTTG TCGCCTTGGG TATAATATTT
1441 GCCCATGGTG AAAACGGGGG CGAAGAAGTT GTCCATATTG GCCACGTTTA AATCAAAACT
1501 GGTGAAACTC ACCCAGGGAT TGGCTGAGAC GAAAAACATA TTCTCAATAA ACCCTTTAGG
1561 GAAATAGGCC AGGTTTTCAC CGTAACACGC CACATCTTGC GAATATATGT GTAGAAACTG
1621 CCGGAAATCG TCGTGGTATT CACTCCAGAG CGATGAAAC GTTTCAGTTT GCTCATGGAA
1681 AACGGTGTA CAAGGGTGAA CACTATCCCA TATCACCAGC TCACCGTCTT TCATTGCCAT
1741 ACGGAATTCC GGTATGAGCAT TCATCAGGCG GGCAAGAATG TGAATAAAGG CCGGATAAAA
1801 CTGTGTCTTA TTTTCTTTA CGGTCTTTAA AAAGGCCGTA ATATCCAGCT GAACGGTCTG
1861 GTTATAGGTA CATTGAGCAA CTGACTGAAA TGCCCTCAAA TGTTCCTTAC GATGCCATTG
1921 GGTATATATCA ACGGTGGTAT ATCCAGTGAT TTTTCTTCC ATTTTAGCTT CCTTAGCTCC
1981 TGAAAATCTC GATAACTCAA AAAATACGCC CGGTAGTGAT CTTATTTTAT TATGGTGAAA
2041 GTTGGAACCT CTTACGTGCC GATCAACGTC TCATTTTCGC CAAAAGTTGG CCCAGGGCTT
2101 CCCGGTATCA ACAGGGACAC CAGGATTTAT TTATCTGCG AAGTGATCTT CCGTCACAGG
2161 TATTTATTCG GCGCAAAGTG CGTCGGGTGA TGCTGCCAAC TTAGTCGACT ACAGGTCACT
2221 AATACCATCT AAGTAGTTGA TTCATAGTGA CTGGATATGT TGTGTTTTAC AGTATTATGT
2281 AGTCTGTTTT TTATGCAAAA TCTAATTTAA TATATTGATA TTTATATCAT TTTACGTTTC
2341 TCGTTCAGCT TTCTTGTA CAAGTTGGCAT TATAAGAAAG CATTGCTTAT CAATTGTTG
2401 CAACGAACAG GTCACATCA GTCAAAATAA AATCATTATT TGCCATCCAG CTGCAGCTCT
2461 GGCCCGTGTC TCAAAATCTC TGATGTTACA TTGCACAAGA TAAAAATATA TCATCATGAA
2521 CAATAAAACT GTCTGCTTAC ATAAACAGTA ATACAGGGG TGTATGAGC CATATTC AAC
2581 GGGAAACGTC GAGGCCGCGA TTAATTTCCA ACATGGATGC TGATTTATAT GGGTATAAAT
2641 GGGCTCGCGA TAATGTGCGG CAATCAGGTG CGACAATCTA TCGCTTGAT GGGAAAGCCG
2701 ATGCGCCAGA GTTGTCTCTG AAACATGGCA AAGGTAGCGT TGCCATGAT GTTACAGATG -

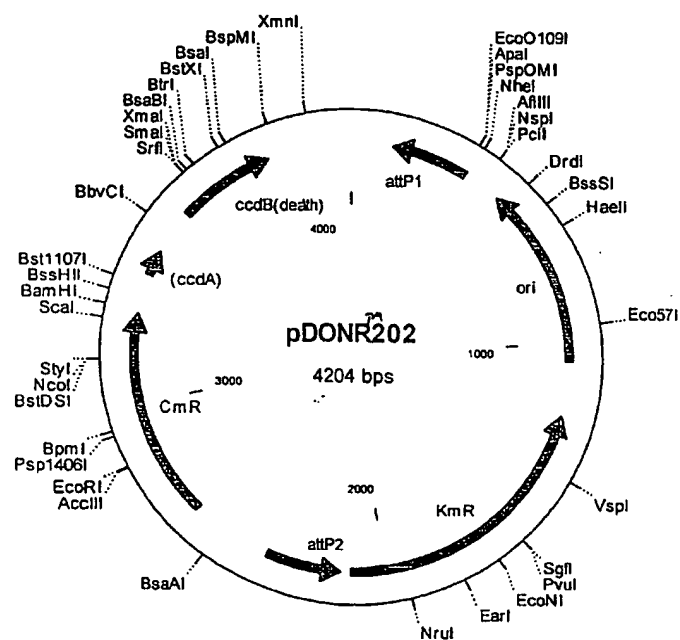
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FIGURE 49B

2761 AGATGGTCAG ACTAAACTGG CTGACGGAAT TTATGCCTCT TCCGACCATC AAGCATTTTA
2821 TCCGTACTCC TGATGATGCA TGGTTACTCA CCACTGCGAT CCCCAGGAAA ACAGCATTTCC
2881 AGGTATTAGA AGAATATCCT GATTTCAGGTG AAAATATTGT TGATGCGCTG GCAGTGTTCC
2941 TGCGCCGGTT GCATTGCAIT CCTGTTTGTG ATTGTCTTTT TAACAGCGAT CGCGTATTTT
3001 GTCTCGCTCA GCGCGCAATCA CGAATGAATA ACGGTTTGGT TGATGCGAGT GATTTTGATG
3061 ACGAGCGTAA TGGCTGGCCT GTTGAAACAAG TCTGGAAGA AATGCATAAA CTTTTCGCAT
3121 TCTCACCGBA TTCAGTCGTC ACTCATGGTG ATTTCTCACT TGATAACCTT ATTTTGTAGC
3181 AGGGGAAATT AATAGGTTGT ATTGATGTTG GACGAGTCGG AATCGCAGAC CGATACCAGG
3241 ATCTTGCCAT CCTATGGAAC TGCCTCGGTG AGTTTTCCTC TTCATTACAG AAACGGCTTT
3301 TTCAAAAATA TGGTATTGAT AATCCTGATA TGAATAAATT GCAGTTTCAT TTGATGCTCG
3361 ATGAGTTTTT CTAATCAGAA TTGGTTAATT GGTGTGAACA CTGGCAGAGC ATTACGCTGA
3421 CTGACGGGA CGGCGCAAGC TCATGACCAA AATCCCTTAA CGTGAGTTTT CGTTCCACTG
3481 AGCGTCAGAC CCCGTAGAAA AGATCAAAGG ATCTTCTTGA GATCCTTTTT TTCTGCGCGT
3541 AATCTGCTGC TTGCAAAACAA AAAAACCACC GCTACCAGCG GTGGTTTGTG TGCCGGATCA
3601 AGAGCTACCA ACTCTTTTTC CGAAGGTAAC TGGCTTCAGC AGAGCGCAGA TACCAAATAC
3661 TGTCCTTCTA GTGTAGCCGT AGTTAGGCCA CCACTTCAAG AACTCTGTAG CACCGCCTAC
3721 ATACCTCGCT CTGCTAATCC TGTTACCAGT GGCTGCTGCC AGTGGCGATA AGTCGTGTCT
3781 TACCGGGTTG GACTCAAGAC GATAGTTACC GGATAAGGCG CAGCGGTCGG GCTGAACGGG
3841 GGGTTCGTGC ACACAGCCCA GCTTGGAGCG AACGACCTAC ACCGAAGTGA GATACCTACA
3901 GCGTGAGCTA TGAGAAAGCG CCACGCTTCC CGAAGGGAGA AAGGCGGACA GGTATCCGGT
3961 AAGCGGCAGG GTCGGAACAG GAGAGCGCAC GAGGGAGCTT CCAGGGGGAA ACGCCTGGTA
4021 TCTTTATAGT CCTGTCGGGT TTCGCCACCT CTGACTTGAG CGTCGATTTT TGTGATGCTC
4081 GTCAGGGGGG CGGAGCCTAT GGAATAACGC CAGCAACGCG GCCTTTTAC GGTTCCTGSC
4141 CTTTTGCTGG CCTTTTGCTC ACATGTTCTT TCCTGCGTTA TCCCTTGATT CTGTGGATAA
4201 CCGTATTACC GCTAGCCAGG AAGAGTTTGT AGAAACGCAA AAAGGCCATC CGTCAGGATG
4261 GCCTTCTGCT TAGTTTGATG CCTGGCAGTT TATGGCGGGC GTCTGCCCCG CCACCTCCG
4321 GGCCGTTGCT TCACAACGTT CAAATCCGCT CCGGCGGAT TTGTCTTACT CAGGAGAGCG
4381 TTCACCGACA AACACAGAT AAAACGAAAG GCCCAGTCTT CCGACTGAGC CTTTCGTTTT
4441 ATTTGATGCC TGGCAGTTCC CTACTCTCGC

FIGURE 49C

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FIGURE 50A: pDONR202 (kan^R)



pDONR202 4204 bp

<u>Location (Base Nos.)</u>		<u>Gene Encoded</u>
369..127		attP1
486..1059		ori
1228..2107		KmR
2381..2140		attP2
2629..3288		CmR
3408..3492		inactivated ccdA
3630..3935		ccdB

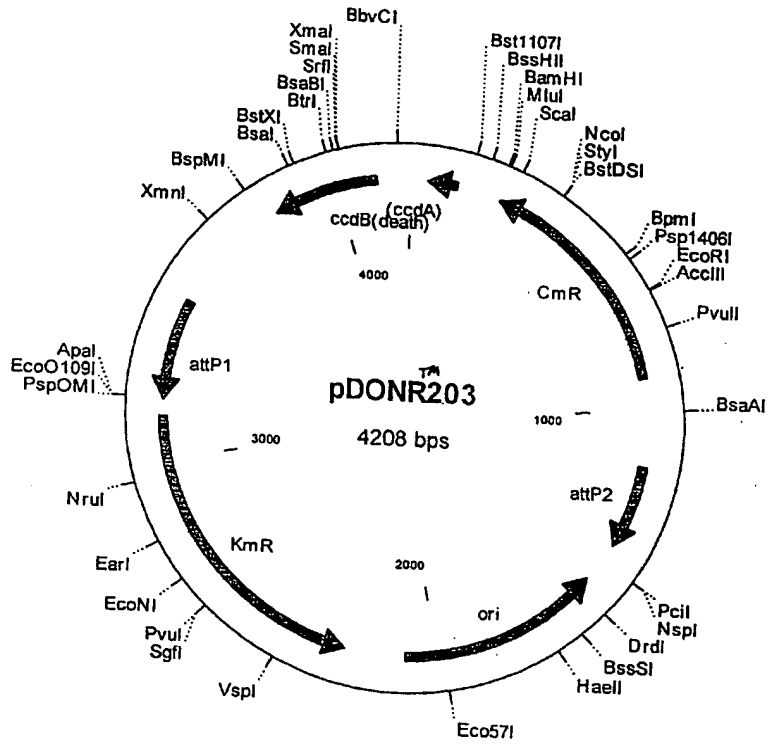
1	CGGCATTGAG	GACAATAGCG	AGTAGGCTGG	ATACGACGAT	TCCGTTTGAG	AAGAACATTT
61	GGAAGGCTGT	CGGTCGACTA	AGTTGGCAGC	ATCACCCGAA	GAACATTTGG	AAGGCTGTCTG
121	GTCGACTACA	GGTCACTAAT	ACCATCTAAG	TAGTTGATTG	ATAGTGACTG	GATATGTTGT
181	GTTTTACAGT	ATTATGTAGT	CTGTTTTTTA	TGCAAAATCT	AATTTAATAT	ATTGATATTT
241	ATATCATTTT	ACGTTTCTCG	TTCAGCTTTT	TTGTACAAAG	TTGGCATTAT	AAAAAAGCAT
301	TGCTCATCAA	TTTGTGCGAA	CGAACAGGTC	ACTATCAGTC	AAAAATAAAT	CATTATTTTGG
361	GGCCCGAGAT	CCATGCTAGC	GGTAATACGG	TTATCCACAG	AATCAGGGGA	TAACGCAGGA
421	AAGAACATGT	GAGCAAAAGG	CCAGCAAAAG	GCCAGGAACC	GTAATAAGGC	CGCGTTGCTG
481	GCGTTTTTCC	ATAGGCTCCG	CCCCCTGAC	GAGCATCACA	AAAATCGACG	CTCAAGTCAG
541	AGGTGGCGAA	ACCCGACAGG	ACTATAAAGA	TACCAGGCGT	TTCCCCCTGG	AAGCTCCCTC
601	GTGCGCTCTC	CTGTTCCGAC	CCTGCCGCTT	ACCGGATACC	TGTCCGCTT	TCTCCCTTCG
661	GGAAGCGTGG	CGCTTTCTCA	TAGCTCACGC	TGTAGGTATC	TCAGTTCCGT	GTAGGTCGTT
721	CGCTCCAAGC	TGGGCTGTGT	GCACGAACCC	CCCGTTCAGC	CCGACCGCTG	CGCCTTATCC
781	GGTAACTATC	GTCTTGAGTC	CAACCCGGTA	AGACACGACT	TATCGCCACT	GCGAGCAGCC
841	ACTGGTAACA	GGATTAGCAG	AGCGAGGTAT	GTAGGCGGTG	CTACAGAGTT	CTTGAAGTGG
901	TGGCCTAACT	ACGGCTACAC	TAGAAGGACA	GTATTTGGTA	TCTGCGCTCT	GCTGAAGCCA
961	GTTACCTTCG	GAAAAAGAGT	TGGTAGCTCT	TGATCCGGCA	AACAAACCC	CGCTGGTAGC
1021	GGTGGTTTTT	TTGTTTGCAA	GCAGCAGATT	ACGCGCAGAA	AAAAAGGATC	TCAAGAAGAT
1081	CCTTTGATCT	TTTCTACGGG	GTCTGACGCT	CAGTGGAAAG	AAAATCAGC	TTAAGGGATT
1141	TTGGTCATGA	GCTTGCGCCG	TCCCGTCAAG	TCAGCGTAAT	GCTCTGCCAG	TGTTACAAACC
1201	AATTAACCAA	TTCTGATTAG	AAAAACTCAT	CGAGCATCAA	ATGAAACTGC	AAITTTATCA
1261	TATCAGGATT	ATCAATACCA	TATTTTGA	AAAGCCGTTT	CTGTAATGAA	GGAGAAACT
1321	CACCGAGGCA	GTTCCATAGG	ATGGCAAGAT	CCTGGTATCG	GTCTGCGATT	CCGACTCGTC
1381	CAACATCAAT	ACAACCTATT	AATTTCCCTT	CGTCAAAAT	AAGGTTATCA	AGTGAGAAAT
1441	CACCATGAGT	GACGACTGAA	TCCGGTGAGA	ATGGCAAAAG	TTTATGCATT	TCTTTCAG
1501	CTTGTTCAAC	AGGCCAGCCA	TTACGCTCGT	CATCAAAATC	ACTCGCATCA	ACCAAACCGT
1561	TATTCATTCT	TGATTGCGCC	TGAGCGAGAC	GAAATACGCG	ATCGCTGTTA	AAAGGACAAT
1621	TACAAACAGG	AATCGAATGC	AACCGGCGCA	GGAACACTGC	CAGCGCATCA	ACAATATTTT
1681	CACCTGAATC	AGGATATTCT	TCTAATACCT	GGAATGCTGT	TTTTCCGGGG	ATCGCAGTGG
1741	TGAGTAACCA	TGCATCATCA	GGAGTACGGA	TAAATGCTT	GATGGTCGGA	AGAGGCATAA
1801	ATTCCGTCAG	CCAGTTTAGT	CTGACCATCT	CATCTGTAAC	ATCATGGCA	ACGCTACCTT
1861	TGCCATGTTT	CAGAAACAC	TCTGGCGCAT	CGGGCTTCCC	ATACAAGCGA	TAGATTGTCTG
1921	CACCTGATTG	CCCGACATTA	TGCGGAGCCC	ATTTATACCC	ATATAAATCA	GCATCCATGT
1981	TGGAATTTAA	TGCGGGCCTC	GACGTTTCCC	GTTGAATATG	GCTCATAACA	CCCCTTGTAT
2041	TACTGTTTTAT	GTAAGCAGAC	AGTTTTATTG	TTCATGATGA	TATATTTTTA	TCTTGTGCAA
2101	TGTAACATCA	GAGATTTTGA	GACACGGGCC	AGAGCTGCAG	CTGGATGGCA	AATAATGATT
2161	TTATTTTGAC	TGATAGTGAC	CTGTTCTGTT	CAACAAATG	ATAAGCAATG	CTTTCTTATA
2221	ATGCCAACTT	TGTACAGAA	AGCTGAACGA	GAAACGTAAA	ATGATATAAA	TATCAATATA
2281	TTAAATTAGA	TTTTGCATAA	AAAACAGACT	ACATAATACT	GTAACACACA	ACATATCCAG
2341	TCATATGAA	TCAACTACTT	AGATGGTATT	AGTGACCTGT	AGTCGACTAA	GTTGGCAGCA
2401	TCACCCGACG	CACCTTGGCG	CGAATAAATA	CCTGTGACGG	AAGATCACTT	CGCAGAATAA
2461	ATAAATCCTG	GTGTCCCTGT	TGATACCGGG	AAGCCCTGGG	CCAACCTTTG	GCGAAAATGA
2521	GACGTTGATC	GGCACGTAAG	AGGTTCCAAC	TTTACCATA	ATGAAATAAG	ATCACTACCG
2581	GGCGTAATTT	TTGAGTTATC	GAGATTTTCA	GGAGCTAAGG	AAGCTAAAT	GGAGAAAAAA
2641	ATCACTGGAT	ATACCAACCGT	TGATATATCC	CAATGGCATC	GTAAGAACA	TTTTGAGGCA
2701	TTTCAGTCAG	TTGCTCAATG	TACCTATAAC	CAGACCGTTC	AGCTGGATAT	TACGGCCTTT

Figure 50B

2761 TTAAAGACCG TAAAGAAAA TAAGCACAAG TTTTATCCGG CCTTTATTCA CATTCTTGCC
2821 CGCCTGATGA ATGCTCATCC GGAATTCGGT ATGGCAATGA AAGACGGTGA GCTGGTGATA
2881 TGGGATAGTG TTCACCCTTG TTACACCGTT TTCCATGAGC AAAGTGAAC GTTTTCATCG
2941 CTCTGGAGTG AATACCACGA CGATTTCGG CAGTTTCTAC ACATATATTC GCAAGATGTG
3001 GCGTGTTACG GTGAAAACCT GGCCCTATTC CTTAAAGGGT TTATTGAGAA TAAGTTTTC
3061 GTCTCAGCCA ATCCCTGGGT GAGTTTCACC AGTTTGTATT TAAACGTGGC CAATATGGAC
3121 AACTTCTTCG CCCCCGTTT CACCATGGGC AAATATTATA CGCAAGGCGA CAAGGTGCTG
3181 ATGCCGCTGG CGATTTCAGT TCATCATGCC GTCTGTGATG GCTTCCATGT CGGCAGAATG
3241 CTTAATGAAT TACAACAGTA CTGCGATGAG TGGCAGGGCG GGGCGTAATC GCGTGGATCC
3301 GGCTTACTAA AAGCCAGATA ACAGTATGCG TATTGCGCG CTGATTTTTC CGGTATAAGA
3361 ATATATACTG ATATGTATAC CCGAAGTATG TCAAAAAGAG GTGTGCTATG AAGCAGCGTA
3421 TTACAGTGAC AGTTGACAGC GACAGCTATC AGTTGCTCAA GGCATATATG ATGTCAATAT
3481 CTCCGGTCTG GTAAGCACAA CCATGCAGAA TGAAGCCCGT CGTCTGCGTG CCGAACGCTG
3541 GAAAGCGGAA AATCAGGAAG GGATGGCTGA GGTGCGCCCG TTTATTGAAA TGAACGGCTC
3601 TTTTGCTGAC GAGAACAGGG ACTGGTGAAA TGCAGTTTAA GGTTTACACC TATAAAGAG
3661 AGAGCCGTTA TCGTCTGTTT GTGGATGTAC AGAGTGATAT TATTGACACG CCCGGGCGAC
3721 GGATGGTGAT CCCCCTGGCC AGTGCACGTC TGCTGTCAGA TAAAGTCTCC CGTGAACTTT
3781 ACCCGGTGGT GCATATCGGG GATGAAAGCT GGCGCATGAT GACCACCGAT ATGGCCAGTG
3841 TGCCGGTCTC CGTTATCGGG GAAGAAGTGG CTGATCTCAG CCACCGCGAA AATGACATCA
3901 AAAACGCCAT TAACCTGATG TTCTGGGGAA TATAATGTC AGGCTCCCTT ATACACAGCC
3961 AGTCTGCAGG TCGATACAGT AGAAATTACA GAACTTTAT CACGTTTAGT AAGTATAGAG
4021 GCTGAAAATC CAGATGAAGC CGAACGACTT GTAAGAGAAA AGTATAAGAG TTGTGAAATT
4081 GTTCTTGATG CAGATGATTT TCAGGACTAT GACACTAGCG TATATGAATA GGTAGATGTT
4141 TTTATTTTGT CACACAAAA AGAGGCTCGC ACCTCTTTT CTTATTTCTT TTTATGATTT
4201 AATA

FIGURE 50C

FIGURE 51A pDONR203 (kan^R)



pDONR203 4208 bp

Location (Base Nos.)	Gene Encoded
47..131	inactivated ccdA
251..910	CmR
1158..1398	attP2
1509..2082	ori
2251..3130	KmR
3464..3174	attP1
3812..4117	ccdB

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1 GCGTTCGGCA CGCAGACGAC GGGCTTCATT CTGCATGGTT GTGCTTACCA GACCCGAGAT
61 ATTGACATCA TATATGCCTT GAGCAACTGA TAGCTGTCGC TGTCAACTGT CACTGTAATA
121 CGCTGCTTCA TAGCACACCT CTTTGTGACA TACTTCGGGT ATACATATCA GTATATATTC
181 TTATACCGCA AAAATCAGCG CGCAAATACG CATACTGTTA TCTGGCTTTT AGTAAGCCGG
241 ATCCACGCGT TTACGCCCGG CCCTGCCACT CATCGCAGTA CTGTTGTAAT TCATTAAAGCA
301 TTCTGCCGAC ATGGAAGCCA TCACAGACGG CATGATGAAC CTGAATCGCC AGCGGCATCA
361 GCACCTTGTC GCCTTGCGTA TAATATTTGC CCATGGTGAA AACGGGGGCG AAGAAGTTGT
421 CCATATTGGC CACGTTTAAA TCAAACTGGG TGAAACTCAC CCAGGGATTG GCTGAGACGA
481 AAAACATATT CTCAATAAAC CCTTTAGGGA AATAGGCCAG GTTTTCACCG TAACACGCCA
541 CATCTTGCGA ATATATGTGT AGAAACTGCC GGAATTCGTC GTGGTATTCA CTCCAGAGCG
601 ATGAAAACGT TTCAGTTTGC TCATGGAAAA CGGTGTAACA AGGGTGAACA CTATCCCATTA
661 TCACCAGCTC ACCGCTTTTC ATTGCCATAC GGAATTCGCG ATGAGCATTC ATCAGGCGGG
721 CAAGAAATGT AATAAAGGCC GATAAAACT TGTGCTTATT TTTCTTTACG GTCTTTAAAA
781 AGGCCGTAAT ATCCAGCTGA ACGGCTCTGG TATAGGTACA TTGAGCAACT GACTGAAATG
841 CCTCAAAATG TTCTTTACGA TGCCATTGGG ATATATCAAC GGTGGTATAT CCAGTGATTT
901 TTTTCTCCAT TTTAGCTTCC TTAGCTCCTG AAAATCTCGA TAACTCAAAA AATACGCCCC
961 GTAGTGATCT TATTTTCATTA TGGTGAAAGT TGGAACCTCT TACGTGCCGA TCAACGTCTC
1021 ATTTTCGCCA AAGTTGGGCC CAGGGCTTCC CGGTATCAAC AGGGACACCA GGATTTATTT
1081 ATTCTGCGAA GTGATCTTCC GTCACAGGTA TTTATTCGGC GCAAAGTCCG TCGGGTGATG
1141 CTGCCAACTT AGTCGACTAC AGGTCACATA TACCATCTAA GTAGTTGATT CATAGTGACT
1201 GGATATGTTG TGTTTTACAG TATTATGTAG TCTGTTTTTT ATGCAAAATC TAATTTAATA
1261 TATTGATATT TATATCATTT TACGTTTCTC GTTCAGCTTT CTGTGACAAA GTTGGCATT
1321 TAAGAAAGCA TTGCTTATCA ATTTGTTGCA ACGAACAGGT CACTATCAGT CAAAATAAAA
1381 TCATTATTTG CCATCCAGCT AGCGGTAATA CGGTTATCCA CAGAATCAGG GGATAACGCA
1441 GGAAGAAGCA TGTGAGCAAA AGGCCAGCAA AAGGCCAGGA ACCGTAAAAA GGCCGCGTTG
1501 CTGGCGTTTT TCCATAGGCT CCGCCCCCTT GACGAGCATC ACAAAAATCG ACGCTCAAGT
1561 CAGAGGTGGC GAAACCCGAC AGGACTATAA AGATACCAGG CGTTTCCCCC TGGAAGCTCC
1621 CTCGTGCGCT CTCCTGTTCC GACCCCTGCC CTTACCGGAT ACCTGTCCGC CTTTCTCCCT
1681 TCGGGAAGCG TGGCGCTTTC TCATAGCTCA CGCTGTAGGT ATCTCAGTTC GGTGTAGGTC
1741 GTTCGCTCCA AGCTGGGCTG TGTGCACGAA CCCCCCGTTC AGCCCCGACC GTGCGCCTTA
1801 TCCGGTAACT ATCGTCTTGA GTCCAACCCG GTAAGACACG ACTTATCGCC ACTGGCAGCA
1861 GCCACTGGTA ACAGGATTAG CAGAGCGAGG TATGTAGGCG GTGCTACAGA GTTCTTGAAG
1921 TGGTGGCCTA ACTACGGCTA CACTAGAAGA ACAGTATTG GTATCTGCGC TCTGCTGAAG
1981 CCAGTTACCT TCGGAAAAAG AGTTGGTAGC TCTTGATCCG GCAAAACAAAC CACCGCTGGT
2041 AGCGGTGGTT TTTTGTGTTG CAAGCAGCAG ATTACGCGCA GAAAAAAGG ATCTCAAGAA
2101 GATCCTTTGA TCTTTTCTAC GGGGTCTGAC GCTCAGTGA ACGAAAATC ACGTTAAGGG
2161 ATTTTGGTCA TGAGCTTGGC CCGTCCCGTC AAGTCAGCGT AATGCTCTGC CAGTGTTACA
2221 ACCAATTAAC CAATTCGTAT TAGAAAAACT CATCGAGCAT CAAATGAAAC TGCAATTTAT
2281 TCATATCAGG ATTATCAATA CCAATTTTTT GAAAAAGCCG TTTCTGTAAT GAAGGAGAAA
2341 ACTCACCGAG GCAGTTCCAT AGGATGGCAA GATCCTGGTA TCGGTCTGCG ATTCCGACTC
2401 GTCCAACATC AATACAACCT ATTAATTTCC CCTCGTCAAA AATAAGGTTA TCAAGTGAGA
2461 AATCACCATG AGTGACGACT GAATCCGGTG AGAATGGCAA AAGTTTATGC ATTTCTTTCC
2521 AGACTTGTTT AACAGGCCAG CCATTACGCT CGTCATCAAA ATCACTCGCA TCAACCAAC
2581 CGTTATTCAAT TCGTGATTGC GCCTGAGCGA GACGAAATAC GCGATCGCTG TTAAGAGGAC
2641 AATTACAAAC AGGAATCGAA TGCAACCGGC GCAGGAACAC TGCCAGCGCA TCAACAATAT
2701 TTTCACCTGA ATCAGGATAT TCTTCTAATA CCTGGAATGC TGTTTTTCCG GGGATCGCAG-

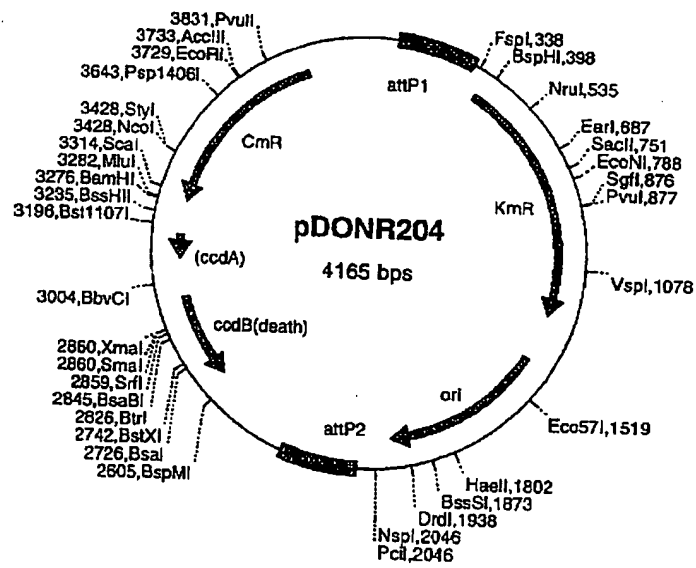
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Figure 51B

2761 TGGTGAGTAA CCATGCATCA TCAGGAGTAC GGATAAAATG CTTGATGGTC GGAAGAGGCA
2821 TAAATTCCGT CAGCCAGTTT AGTCTGACCA TCTCATCTGT AACATCATTG GCAACGCTAC
2881 CTTTGCCATG TTTCAGAAAC AACTCTGGCG CATCGGGCTT CCCATACAAG CGATAGATTG
2941 TCGCACCTGA TTGCCCCGACA TTATCGCGAG CCCATTTATA CCCATATAAA TCAGCATCCA
3001 TGTGGGAATT TAATCGCGGC CTCGACGTTT CCCGTTGAAT ATGGCTCATA ACACCCCTTG
3061 TATTACTGTT TATGTAAGCA GACAGTTTTA TTGTTTCATGA TGATATATTT TTATCTTGTG
3121 CAATGTAACA TCAGAGATTT TGAGACACGG GCCAGAGCTG CAGCTAGCAT GGATCTCGGG
3181 CCCCAAATAA TGATTTTATT TTGACTGATA GTGACCTGTT CGTTGCAACA AATTGATGAG
3241 CAATGCTTTT TTATAATGCC AACTTTGTAC AAAAAAGCTG AACGAGAAAC GTAAAAATGAT
3301 ATAAATATCA ATATATTAAA TTAGATTTTG CATAAAAAAC AGACTACATA ATACTGTAAA
3361 ACACAACATA TCCAGTCACT ATGAATCAAC TACTTAGATG GTATTAGTGA CCTGTAGTCG
3421 ACCGACAGCC TTCCAAATGT TCTTCGGGTG ATGCTGCCAA CTTAGTCGAC CGACAGCCTT
3481 CCAATGTTT TCTCAAACG GAATCGTCGT ATCCAGCCTA CTCGCTATTG TCCTCAATGC
3541 CGTATTAAAT CATAAAAAGA AATAAGAAAA AGAGGTGCGA GCCTCTTTT TGTGTGACAA
3601 AATAAAAAACA TCTACCTATT CATATACGCT AGTGTCATAG TCCTGAAAAAT CATCTGCATC
3661 AAGAACAATT TCACAACCTCT TATACTTTTC TCTTACAAGT CGTTCTGGCTT CATCTGGATT
3721 TTCAGCCTCT ATACTTTACTA AACGTGATAA AGTTTCTGTA ATTTCTACTG TATCGACCTG
3781 CAGACTGGCT GTGTATAAGG GAGCCTGACA TTTATATTCC CCAGAACATC AGGTTAATGG
3841 CGTTTTTGAT GTCATTTTCG CGGTGGCTGA GATCAGCCAC TTCTTCCCG ATAACGGAGA
3901 CCGGCACACT GGCCATATCG GTGGTCATCA TCGCCAGCT TTCATCCCCG ATATGCACCA
3961 CCGGGTAAAG TTCACGGGAG ACTTTATCTG ACAGCAGACG TGCAGTGGCC AGGGGGATCA
4021 CCATCCGTCG CCCGGGCGTG TCAATAATAT CACTCTGTAC ATCCACAAAC AGACGATAAC
4081 GGCTCTCTCT TTTATAGGTG TAAACCTTAA ACTGCATTTT ACCAGTCCCT GTTCTCGTCA
4141 GCAAAAGAGC CGTTCATTTC AATAAACCGG GCGACCTCAG CCATCCCTTC CTGATTTTCC
4201 GCITTTCCA

FIGURE 51C

FIGURE 52A pDONR204 (kanR)



pDONR204 4165 bp

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1 CGGCATTGAG GACAATAGCG AGTAGGCTGG ATACGACGAT TCCGTTTGAG AAGAACATTT
61 GGAAGGCTGT CGGTCGACTA CAGGTCACCTA ATACCATCTA AGTAGTTGAA TCATAGTGAC
121 TGGATATGTT GTGTTTTACA GTATTATGTA GTCTGTTTTT TATGCAAAAT CTAAATTTAAT
181 ATATTGATAT TTATATCATT TTACGTTTCT CGTTCAGCTT TTTGTACAA AGTTGGCATT
241 ATAAAAAAGC ATTGCTTATC AATTTGTTGC AACGAACAGG TCACTATCAG TCAAAATAAA
301 ATCATTATTT GGGGCCCGAG ATCCATGCTA GCTGCAGTGC GCAGGGCCCG TGTCTCAAAA
361 TCTCTGATGT TACATTGCAC AAGATAAAAA TATATCATCA TGAACAATAA AACTGTCTCG
421 TTACATAAAC AGTAATACAA GGGGTGTTAT GAGCCATATT CAACGGGAAA CGTCTTGCTG
481 GAGGCCGCGA TTAATTTCCA ACATGGATGC TGATTTATAT GGGTATAAAT GGGCTCGCGA
541 TAATGTCGGG CAATCAGGTG CGACAATCTT TCGATTGTAT GGGAAAGCCCG ATGCGCCAGA
601 GTTGTTCCTG ARAATGGCA AAGGTAGCGT TGCCAATGAT GTTACAGATG AGATGGTCAG
661 ACTAAACTGG CTGACGGAAT TTATGCCTCT TCCGACCATC AAGCATTTTA TCCGTACTCC
721 TGATGATGCA TGGTTACTCA CCATGCGCAT CCGCGGGAAA ACAGCATTTCC AGGTATTAGA
781 AGAATATCCT GATTCAAGTG AAAATATTGT TGATGCGCTG GCAGTGTTCG TCGCCCGGTT
841 GCATTTCGAT CTGTTTGTGA ATTGTCCTTT TAACAGCGAT CGCGTATTTC GTCTCGCTCA
901 GCGCAATCA CGAATGAATA ACGGTTTGGT TGATGCGAGT GATTTTGATG ACGAGCGTAA
961 TGGCTGGCCT GTTGAACAAG TCTGGAAAGA AATGCATACG CTTTGGCCAT TCTCACCGBA
1021 TTCAGTCGTC ACTCATGGTG ATTTCTCACT TGATAACCTT ATTTTGTACG AGGGGAAATT
1081 AATAGGTTGT ATTGATGTTG GACGAGTCGG AATCGCAGAC CGATACCAGG ATCTTGCCAT
1141 CCTATGGAAC TGCCCTCGGTG AGTTTTCTCC TTCATTACAG AAACGGCTTT TTCAAAAATA
1201 TGGTATTGAT AATCCTGATA TGAATAAAAT GCAGTTTCAT TTGATGCTCG ATGAGTTTTC
1261 CTAATCAGAA TTGGTTAATT GGTGTAACA CTGGCAGAGC ATTACGCTGA CTTGACGGGA
1321 CGGCGNCAAT ACCAAATCC CTTAACGTGA GTTTTCGTTT CACTGAGCGT CAGACCCCGT
1381 AGAAAAGATC AAAGGATCTT CTTGAGATCC TTTTTCCTG CGCGTAATCT GCTGCTTGCA
1441 AACAAAAAAA CCACCGCTAC CAGCGGTGGT TTGTTTCCCG GATCAAGAGC TACCAACTCT
1501 TTTTCCGAAG GTAACGTGCT TCAGCAGAGC GCAGATACCA AATACTGTCC TTCTAGTGTA
1561 CCCGTAGTTA GGCACCACT TCAAGAACTC TGTAAGCAGG CCTACATACC TCGCTCTGCT
1621 AATCCTGTTA CCAGTGGCTG CTGCCAGTGG CGATAAGTCG TGTCTTACCG GGTGGACCTC
1681 AAGACGATAG TTACCGGATA AGGCGCAGCG GTCGGGCTGA ACGGGGGGTT CGTGACACAC
1741 GCCCAGCTTG GAGCGAACGA CCTACACCGA ACTGAGATAC CTACAGCGTG AGCTATGAGA
1801 AAGCGCCACG CTTCCCGAAG GGAGAAAGGC GGAACAGTAT CCGGTAAGCG GCAGGCTCGG
1861 AACAGGAGAG CGCAGCAGGG AGCTTCCAGG GGGAAACGCC TGGTATCTTT ATAGTCTGCT
1921 CGGGTTTCGC CACCTCTGAC TTGAGCGTCG ATTTTGTGA TGCTCGTCAG GGGGCGGAG
1981 CCTATGGAAA AACGCCAGCA ACGCGGCTT TTTACGGTTC CTGGCCTTTT GCTGGCCTTT
2041 TGCTCACATG TTCTTTCCTG CGTTATCCCC TGATTCTGTG GATAACCGTA TTACCCTAG
2101 CTGGATCGGC AAATAATGAT TTTATTTTGA CTGATAGTGA CCTGTTCGTT GCAACAAATT
2161 GATAAGCAAT GCTTTTTTAT AATGCCAACT TTGTACAAGA AAGCTGAACG AGAAACGTAA
2221 AATGATATAA ATATCAATAT ATTAAATTAG ATTTTGCATA AAAAAACAGC TACATAATAC
2281 TGTAACACAC AACATATCCA GTCACATGTA TTCAACTACT TAGATGGTAT TAGTGACCTG
2341 TAGTCGACTA AGTTGGCAGC ATCACCAGAC GCACCTTTCG CCGAATAAAT ACCTGTGACG
2401 GAAGATCACT TCGCAGAAATA AATAAATCCT GGTGTCCCTG TTGATACCGG GAAGCCCTGG
2461 GCCAATTTT GCGGAAAATG AGACGTTGAT CGGCACATT CACAACCTTT ATACTTTTCT
2521 CTTACAAGTC GTTCGGCTTC ATCTGGATTT TCAGCCTCTA TACTTACTAA ACGTGATAAA
2581 GTTTCTGTAA TTTCTACTGT ATCGACCTGC AGACTGGCTG TGTATAACGG AGCCTGACAT
2641 TTATATTCCC CAGAACATCA GGTAAATGGC GTTTTGTATG TCATTTTCGC GGTGGCTGAG
2701 ATCAGCCACT TCTTCCCCGA TAACGGAGAC CGGCACACTG GCCATATCGG TGGTCATCAT
2761 GCGCCAGCTT TCATCCCCGA TATGCACCAC CGGGTAAAGT TCACGGGAGA CTTTATCTGA
2821 CAGCAGACGT GCACTGGCCA GGGGGATCAC CATCCGTCG CCGGGCGTGT CAATAATATC
2881 ACTCTGTACA TCCACAAACA GACGATAACG GCTCTCTCTT TTATAGGTGT AAACCTTAAA
2941 CTGCATTTCA CCAGTCCCTG TTCTCGTCAG CAAAAGAGCC GTTCATTTCA ATAAACCGGG
3001 CGACCTCAGC CATCCCTTCC TGATTTTCCG CTTTCCAGCG TTCCGACCGG AGACGACGGG
3061 CTTCAATCTG CATGGTTGTG CTTACCAGAC CGGAGATATT GACATCATAT ATGCCTTGAG
3121 CAACTGATAG CTGTCGCTGT CAACTGTCAC TGAATACGC TGCTTCATAG CACACCTCTT-
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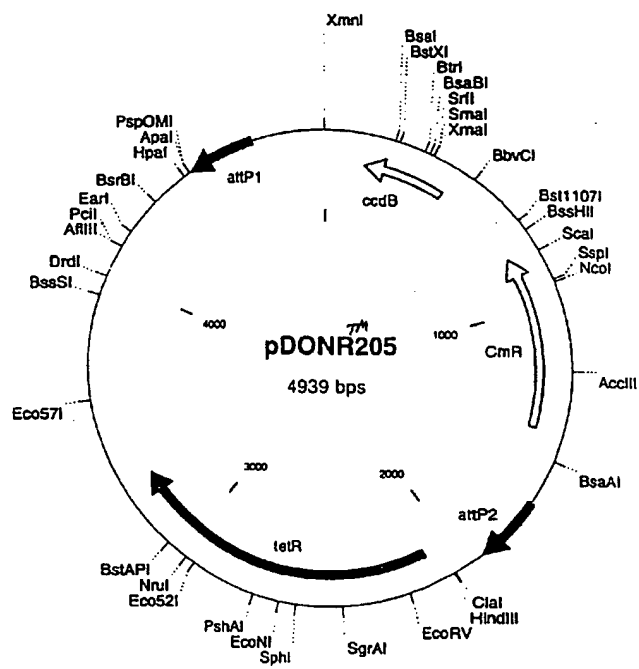
FIGURE 52B

3181 TTGACATAC TTCGGGTATA CATATCAGTA TATATTCTTA TACCGCAAAA ATCAGCGCGC
3241 AAATACGCAT ACTGTTATCT GGCTTTTAGT AAGCCGGATC CACCGGTTTA CGCCCCGCC
3301 TGCCACTCAT CGCAGTACTG TTGTAATTCA TTAAGCATTG TGCCGACATG GAAGCCATCA
3361 CAGACGGCAT GATGAACCTG AATCGCCAGC GGCATCAGCA CCTTGTCGCC TTGCGTATAA
3421 TATTTGCCCA TGGTGAAAAC GGGGGCGAAG AAGTTGTCCA TATTGGCCAC GTTTAAATCA
3481 AAACGTGTGA AACTCACCCA GGGATTGGCT GAGACGAAAA ACATATTCTC AATAAACCTT
3541 TTAGGGAAAT AGGCCAGGTT TTCACCGTAA CACGCCACAT CTGCGAATA TATGTGTAGA
3601 AACTGCCGGA AATCGTCGTG GTATTCACTC CAGAGCGATG AAAACGTTTC AGTTTGCTCA
3661 TGGAAAACGG TGTAACAAGG GTGAACACTA TCCCATATCA CCAGCTCACC GTCTTTTCATT
3721 GCCATACGGA ATTCCGGATG AGCATTTCATC AGCGGGGCAA GAATGTGAAT AAAGGCCGGA
3781 TAAAACTTGT GCTTATTTT CTTTACGGTC TTTAAAAAGG CCGTAATATC CAGCTGAACG
3841 GTCTGGTTAT AGGTACATTG AGCAACTGAC TGAAATGCCT CAAAATGTTT TTTACGATC
3901 CATTGGGATA TATCAACGGT GGTATATCCA GTGATTTTTT TCTCCATTTT AGCTTCCTTA
3961 GCTCCTGAAA ATCTCGATAA CTCAAAAAAT ACGCCCGGTA GTGATCTTAT TTCATTATGG
4021 TGAAAGTTGG AACCTCTTAC TGTCTTGAT GCAGATGAT TTCAGGACTA TGACACTAGC
4081 ATATATGAAT AGGTAGATGT TTTTATTTG TCACACAAA AAGAGGCTCG CACCTCTTTT
4141 TCTTATTTCT TTTTATGATT TAATA

FIGURE 52C

156/240

Figure 53A: pDONR205 (tetR)



pDONR205 4939 bp

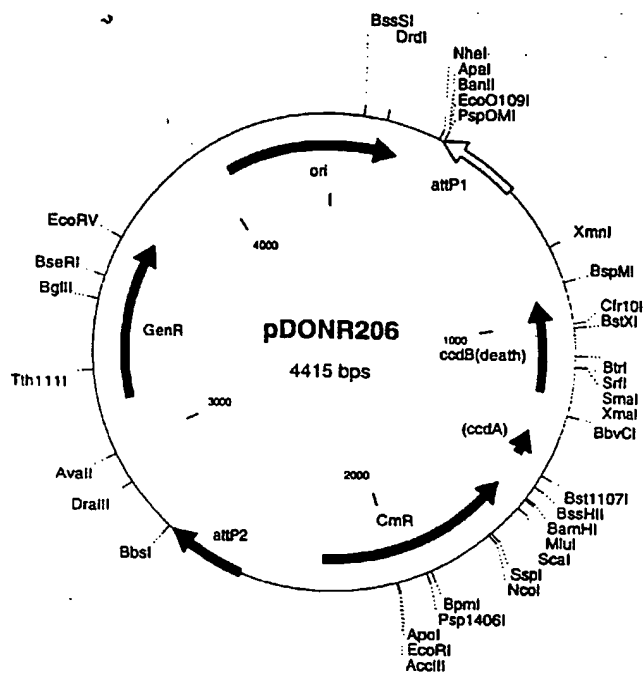
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TTACATTGCACAAGATAAAATATATCATCATGAATTTCTCATGTTTGACAGCTTATCATC
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ATGAAATCTAACAAATGCGCTCATCGTCATCTCGGCACCGTCAACCTGGATGCTGTAGGC
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CCCGTTCTCGGAGCACTGTCGACCGCTTGGCCCGCGCCAGTCTGCTCGCTTCGCTA
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GCCGAGCAGCATCGTGCCCGCATCACCGCGCCACAGGTGCGGTTGCTGGCGCTATATC
GCCGACATCACCAGTGGGAAGATCGGGCTCGCCACTTCGGGCTCATGAGCGCTTGTTC
GGCGTGGGTATGGTGGCAGGCCCGCTGGCCGGGGGACTGTTGGCGGCCATCTCTTGCAT
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AGCTCCTTCGGTGGCGCGGGCATGACTATCGTCGCGCACTTATGACTGTCTTCTTT
ATCATGCAACTCGTAGGACAGGTGCCGGCAGCGCTTGGGTCAATTTTCGGCGAGGACCGC
TTTCGCTGGAGCGCGACGATGATCGGCCTGTGCTGCGGTATTCGGAATCTTGACGCGC
CTCGCTCAAGCCTTCGTCACTGGTCCCGCCACCAACGTTTCGGCGAGAAGCAGGCCATT
ATCGCCGGCATGGCGGCCGACGCGTGGGCTACGCTTGTCTGGCGTTTCGCGACGCGAGGC
TGGATGGCCTTCCCCATTATGATTTCTCTGCTTCGGCGGCATCGGGATGCCCGCGTTG
CAGGCCATGCTGTCAGGCAGGTAGATGACGACCATCAGGGACAGCTTCAAGGATCGCTC
GCGGCTCTTACCAGCCTAACTTCGATCATTGGACCGCTGATCGTCACGCGGATTTATGCC
GCCTCGCGGAGCACATGGAACGGGTTGGCATGGATTGTAGGCGCCGCCCTATACCTTGTG
TGCTTCCCGCGCTTGCCTGCGGTGATGGAGCCGGGCCACCTCGACCTGAATGGAAGCC
GGCGGCACCTCGCTAACGATTCACCACTCCAGAATGGAGCCAATCAATCTTGGCGGA
GAACGTGTAATGCGCAAAACCAACCTTGGCAGAACATATCCATCGCATGACCAAAATCCC
TTAACGTGAGTTTTCGTTCCACTGAGCGTCAGACCCCGTAGAAAAGATCAAAGGATCTTC
TTGAGATCCTTTTTTCTGCGCGTAATCTGCTGCTTGCAAAACAAAAAACACCGCTACC
AGCGGTGGTTTGTGTTGCCGGATCAAGAGCTACCAACTCTTTTTCGGAAGGTAAGTGGCTT
CAGCAGAGCGCAGATACCAATACTGCTCTTCTAGTGTAGCCGTAGTTAGGCCACCACTT
CAAGAACTCTGTAGCACCGCCTACATACCTCGCTCTGCTAATCTGTTACCAAGTGGCTGC
TGCCAGTGGCGATAAGTGTGCTTACCGGGTTGGACTCAAGACGATAGTTACCGGATAA
GGCGCAGCGGTGGGCTGAACGGGGGGTTCGTGCACACAGCCAGCTTGGAGCGAACGAC
CTACACCGAACTGAGATACCTACAGCGTGAGCTATGAGAAAGCGCCACGCTTCCCGAAGG
GAGAAAGGCGGACAGGTATCCGTAAGCGGCAGGGTCGGAACAGGAGAGCGCAGAGGGA
GCTTCCAGGGGAAACGCGCTGTATCTTTATAGTCTGTCGGGTTTCGCCACCTCTGACT
TGAGCGTCGATTTTTGTGATGCTGTCAGGGGGCGGAGCCTATGGAAAAACGCCAGCAA-

FIGURE 53B

CGCGGCCTTTTACGGTTCCTGGCCTTTTGCTGGCCTTTTGCTCACATGTTCTTTCCTGC
GTTATCCCCTGATTCTGTGGATAACCGTATTACCGCTAGCCAGGAAGAGTTGTAGAAAC
GCAAAAAGGCCATCCGTCAGGATGGCCTTCGCTTAGTTTGATGCCTGGCAGTTTATGGC
GGGCGTCTGCCCCGCCACCTCCGGGCGGTGCTTCACAACGTTCAAATCCGCTCCCGGC
GGATTTGTCTACTCAGGAGAGCGTTTACCGACAAACAACAGATAAAACGAAAGGCCAG
TCTTCCGACTGAGCCTTTCGTTTATTGATGCTGGCAGTTCCCTACTCTCGCGTTAAC
GCTAGCATGGATCTCGGGCCCCAAATAATGATTTTATTTTGACTGATAGTGACCTGTTG
TTGCAACAAATTGATGAGCAATGCTTTTATAATGCCAACTTTGTACAAAAAAGCTGAA
CGAGAAACGTAAAATGATATAAATATCAATATATTAAATTAGATTTGCATAAAAAACAG
ACTACATAACTGTAAAAACAACATATCCAGTCACTATGAATCAACTACTTAGATGGT
ATTAGTGACCTGTAGTCGACCGACAGCCTTCCAAATGTTCTTCGGGTGATGCTGCCAACT
TAGTCGACCGACAGCCTTCCAAATGTTCTTCTCAAACGGAATCGTCGTATCCAGCCTACT
CGCTATTGTCCTCAATGCCGTATTAAATCATAAAAAGAAAATAAGAAAAAGAGGTGCGAGC
CTCTTTTGTGTGACAAAATAAAAAACATCTACCTATTCATATACGCTAGTGTATAGTC
CTGAAAATCATCTGCATCAAGAACAAATTCACAACCTTTATACTTTTCTTTACAAGTCG
TTGGGCTTCATCTGGATTTTCAGCCTCTATACTTACTAAACGTGATAAGGTTTCTGTAAT
TTCTACTGTATCGACCTGCAGACTGGCTGTGTATAAGGGAGCCTGACATTTATATCCCC
AGAACATCAGGTTAATGGCGTTTGTATGTCATTTTCGCGGTGGCTGAGATCAGCCACTT
CTTCCCGGATAACGGAGACCGGCACACTGGCCATATCGGTGGTCATCATGCGCCAGCTTT
CATCCCCGATATGCACCACCGGGTAAAGTTTACGGGAGACTTTATCTGACAGCAGACGTG
CACTGGCCAGGGGATCACCATCCGTCGCCCGGGCGTGTCAATAATATCACTCTGTACAT
CCACAAACAGACGATAACGGCTCTCTCTTTTATAGGTGTAAACCTTAAACTGCATTTCAC
CAGTCCCTGTTCTCGTCAGCAAAAGAGCGGTTCAATTCATAAACCGGGCGACCTCAGCC
ATCCCTTCTCGATTTTCCGCTTTCCAGCGTTCGGCACGCAGACGACGGGCTTCATTCTGC
ATGGTTGTGCTTACCAGACCGGAGATATTGACATCATATATGCCTTGAGCAACTGATAGC
TGTCGCTGTCACTGTCACTGTAATACGCTGCTTCATAGCACACCTCTTTTGACATACT
TCGGGTATACATATCAGTATATATTCTTATACCGCAAAATCAGCGCGCAAAATACGCATA
CTGTTATCTGGCTTTAGTAAGCCGGATCCACGCGATTACGCCCCGCCCTGCCACTCATC
GCAGTACTGTTGTAATTCATTAAAGCATTCTGCCGACATGGAAGCCATCACAGACGGCATG
ATGAACCTGAATCGCCAGC

FIGURE 53C

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pDONR206 4415 bp

CGGCATTGAGGACAATAGCGAGTAGGCTGGATACGACGATTCCGTTTGAGAAGAACATT
GGAAGGCTGTCGGTCGACTACAGGTCACTAATACCATCTAAGTAGTTGAATCATAGTGAC
TGGATATGTTGTGTTTTACAGTATTATAGTAGTCTGTTTTTTATGCAAAATCTAATTTAAT
ATATTGATATTTATATCATTTTACGTTTCTCGTTACGCTTTTTTGACAAAGTTGGCATT
ATAAAAAGCATTGCTTATCAATTTGTTGCAACGAACAGGTCACTATCAGTCAAAATAAA
ATCATTATTTGGGGCCCGAGATCCATGCTAGCGGTAATACGGTTATCCACAGAATCAGGG
GATAACGCAGGAAAGAACATGTGAGCAAAAGGCCAGCAAAAGGCCAGAACCGTAAAAAG
GCCGCGTTGCTGGCGTTTTTCCATAGGCTCCGCCCCCTGACGAGCATCACAAAAATCGA
CGCTCAAGTCAGAGGTGGCGAAACCCGACAGGACTATAAGATACCAGGCGTTTCCCCCT
GGAAGCTCCCTCGTGGCTCTCTCTGTTCCGACCCCTGCCGCTTACCGGATACCTGTCCGCC
TTTCTCCCTTCGGGAAGCGTGGCGCTTCTCATAGCTCACGCTGTAGGTATCTCAGTTCG
GTGTAGGTCGTTCTGCTCCAAGCTGGGCTGTGTGCACGAACCCCGTTACGCCGACCGC
TGCGCCTTATCCGGTAACATATCTGTTGAGTCCAAACCCGGTAAGACACGACTTATCGCCA
CTGGCAGCAGCCACTGGTAACAGGATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAGAG
TTCTTGAAAGTGGTGGCTAACTACGGCTACACTAGAAGGACAGTATTGGTATCTCGCT
CTGCTGAAGCCAGTTACCTTCGGAAGAGAGTTGGTAGCTCTTGATCCGGCAACAAACC
ACCGCTGGTAGCGGTGGTTTTTTTGTGTTGCAAGCAGCAGATTACGCGCAGAAAAAGGA
TCTCAAGAAAGATCCTTTGATCTTTTCTACGGGTCTGACGCTCAGTGGAAACGAAAACTCA
CGTTAAGGGATTTTGGTCATGNCGCCGTCCCGTCAAGTCAGCGTAATGCTCTGCCAGTGT
TACAAACCAATTAACCAATCTGATTAGAAAACTCATCGAGCATCAAAAGAACTGCAAT
TTATTATATCAGGATTATCAATACCATATTTTGAAAAAGCCGTTTCTGTAATGAAGGA
GAAAACTCACCGAGGAGTTCATAGGATGGCAAGATCCTGGTATCGGTCTGCGATTCCG
ACTCGTCCAACATCAATACAACTATTAGCCGAGGTCTTCCGATCTCTGAGCCAGGGC
AGATCCGTGCACAGCACCTTGCCGTAGAAGAACAGCAAGGCCGCCAATGCTGACGATGC
GTGGAGACCGAAACCTTGCGCTCGTTCCGACGACAGGACAGAAATGCTCGACTTCGCTG
CTGCCCAAGGTTGCCGGGTACGCACACCGTGGAAACGGATGAAGGCACGAACCCAGTTG
ACATAAGCCTGTTCCGGTTCGTTAACTGTAATGCAAGTAGCGTATGCGCTCACGCACTGG
TCCAGAACCTTGACCGAACGACGCGGTGTTACGGCGCAGTGGCGGTTTTTATGGCTTGT
TATGACTGTTTTTTGTACAGTCTATGCCCTCGGGCATCCAAGCAGCAAGCGGTTACGCC
GTGGGTTCGATGTTTGTATGTTATGGAGCAGCAACGATGTTACGACGACGACGATGTTAC
GCAGCAGGCGAGTCCGCTAAACAAAGTTAGGTGGCTCAAGTATGGGCATCATTGCGAC
ATGTAGGCTCGGCTGACCAAGTCAAACTCATGCGGGCTGCTCTGATCTTTTCGGTCTG
TGAGTTTCGGAGACGTAGCCACCTACTCCCAACATCAGCCGGACTCCGATTACCTCGGGAA
CTTGCTCCGTAGTAAGACATTCATCGCGCTTGCTGCTTCGACCAAGAAGCGGTTGTTGG
CGCTCTCGCGGCTTACGTTCTGCCAGGTTTGAAGCAGCGCGTAGTGAGATCTATATCTA
TGATCTCGCAGTCTCGGCGAGCACCGAGGAGGGCATTGCCACCGCGCTCATCAATCT
CCTCAAGCATGAGGCCAACGCGCTTGGTGCTTATGTGATCTACGTGCAAGCAGATTACGG
TGACGATCCCGCAGTGGCTCTCTATACAAAGTTGGGCATACGGGAAGAAGTGTATGCACTT
TGATATCGACCCAAGTACCGCCACCTAACAAATTCGTTCAAGCCGAGATCGGCTTCCCGGC
CTAATTTCCCTCGTCAAAATAAGGTTATCAAGTGAGAAATCACCATGAGTGACGACTG
AATCCGGTGAGAAATGGCAAAAGCGTATGCATTTCTTTCCAGACTTGTTCAACAGGCCAGC
CATTACGCTCGTCATCAAAATCACTCGCATCAACCAACCGTTATTCAATTCGTGATTGCG
CCTGAGCGAGACGAAATACGCGATCGCTGTTAAAGGACAATTACAAACAGGAATCGAAT
GCAACCGGCGCAGGAACACTGCCAGCGCATCAACAATATTTTACCTGAATCAGGATATT
CTTCTAATACCTGGAATGCTGTTTTCCCGCGGATCGCAGTGGTGAGTAACCATGCATCAT
CAGGAGTACGGATAAAATGCTTGATGGTGGAGAGGCATAAAATCCGTCAGCCAGTTTA
GTCTGACCATCTCATCTGTAACTCATTTGGCAACGCTACCTTTGCCATGTTTCAGAAACA
ACTCTGGCGCATCGGGCTTCCCATACAATCGAAAGATTGTGCGACCTGATTGCCCGACAT
TATCGCGAGCCCATTTATACCCATATAAATCAGCATCCATGTTGGAAATTAATCGCGGCC
TCCAGCAAGACGTTTCCCGTTGAATATGGCTCATAACACCCCTTGATTACTGTTTATGT
AAGCAGACAGTTTTATTGTTTCATGATGATATATTTTTATCTTGTCGAATGTAACATCAGA
GATTTTTGAGACACGGGCCCGCGCACTGCGAGCTGGATCGGCAAAATAATGATTTTTATTG
ACTGATAGTGACCTGTTCTGTTGCAACAAATGATAAGCAATGCTTTTTTATAATGCCAAC -

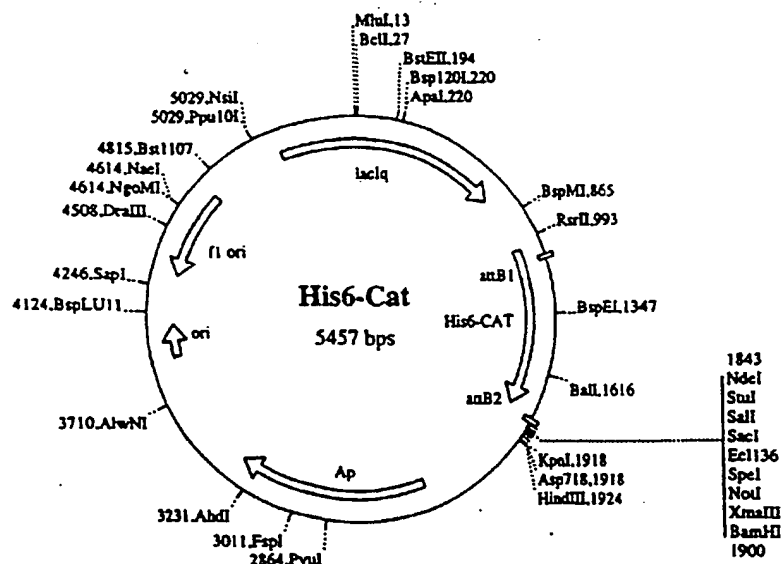
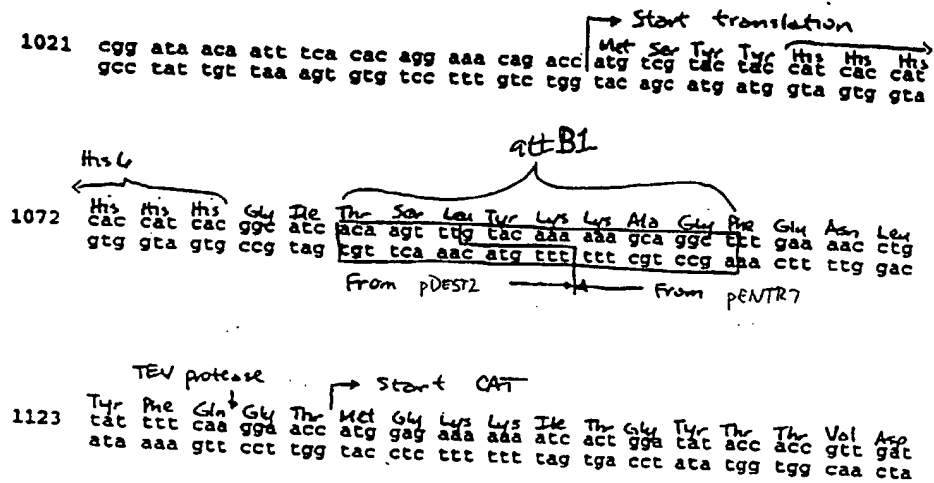
FIGURE 54B

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TTTGTACAAGAAAGCTGAACGAGAAACGTAAAATGATATAAATATCAATATATTAAATTA
GATTTTGCATAAAAAACAGACTACATAAATCTGTAAAACACAACATATCCAGTCACTATG
ATTCAACTACTTAGATGGTATTAGTGACCTGTAGTCGACTAAGTTGGCAGCATCACCCGA
CGCACTTTGCGCCGAATAAATACCTGTGACGGAAGATCACTTCGCAGAATAAATAAATCC
TGGTGTCCCTGTTGATACCGGGAAGCCCTGGGCCAATTTTGGCGAAAATGAGACGTTGA
TCGGCACGTAAGAGGTTCCAACCTTCACCATAATGAAAATAAGATCACTACCGGGCGTATT
TTTTGAGTTATCGAGATTTTCAGGAGCTAAGGAAGCTAAAATGGAGAAAAAATCACTGG
ATATACCACCGTTGATATATCCCAATGGCATCGTAAAGAACATTTTGAGGCATTTTCAGTC
AGTTGCTCAATGTACCTATAACCAGACCGTTCAGCTGGATATTACGGCCTTTTTAAAGAC
CGTAAAGAAAAATAAGCACAAGTTTTATCCGGCCTTTATTCACATTCTTGCCCGCCTGAT
GAATGCTCATCCGGAATTCGGTATGGCAATGAAAGACGGTGAGCTGGTGATATGGGATAG
TGTTCAACCTTGTACACCGTTTTCCATGAGCAAACTGAAACGTTTTTCATCGCTCTGGAG
TGAATACCACGACGATTTCCGGCAGTTTCTACACATATATTGCAAGATGTGGCGTGTTA
CGGTGAAAACCTGGCCTATTCCCTAAAGGGTTTATTGAGAATAAGTTTTTCGTCTCAGC
CAATCCCTGGGTGAGTTTCAACAGTTTTGATTAAACGTGGCCAATATGGACAACTTCTT
CGCCCCCGTTTTCAACCATGGGCAAAATATTATACGCAAGGCGACAAGGTGCTGATGCCGCT
GGCGATTCAAGTTTCATCATGCCGCTCTGTGATGGCTTCCATGTGCGCAGAAATGCTTAATGA
ATTACAACAGTACTGCGATGAGTGGCAGGGCGGGGCGTAAACGCGTGATCCGGCTTACT
AAAAGCCAGATAACAGTATGCGTATTTCGCGCTGATTTTGGCGGTATAAGAATATATAC
TGATATGTATACCCGAAGTATGTCAAAAGAGGTGTGCTATGAAGCAGCGTATTACAGTG
ACAGTTGACAGCGACAGCTATCAGTTGCTCAAGGCATATATGATGTCAATATCTCCGGTC
TGGTAAGCACAACCATGCAGAATGAAGCCCGTCGTCTGCGTGCCGAACGCTGGAAAGCGG
AAAATCAGGAAGGGATGGCTGAGGTGCGCCCGTTTTATTGAAATGAACGGCTCTTTTGCTG
ACGAGAACAGGGACTGGTGAAATGCAGTTTAAAGGTTTACACCTATAAAAAGAGAGCCGT
TATCGTCTGTTTGTGGATGTACAGAGTGATATTATGACACGCCCGGGCGACGGATGGTG
ATCCCCCTGGCCAGTGACGCTCTGCTGTGATGATAAAGTCTCCCGTGAACTTTACCCGGTG
GTGCATATCGGGGATGAAGCTGGCGCATGATGACACCGATATGGCCAGTGTGCCGGTG
TCCGTTATCGGGGAAGAAGTGGCTGATCTCAGCCACCGGAAAATGACATCAAAAACGCC
ATTAACCTGATGTTCTGGGGAATATAAATGTCAGGCTCCGTTATACACAGCCAGTCTGCA
GGTCGATACAGTAGAAAATTACAGAACTTTATCACGTTTAGTAAGTATAGAGGCTGAAAA
TCCAGATGAAGCCGAACGACTTGTAAAGAGAAAAGTATAAGAGTTGTGAAATGTTCTTGA
TGCAGATGATTTTCAGGACTATGACACTAGCATATATGAATAGGTAGATGTTTTATTTT
GTCACACAAAAAGAGGCTCGCACCTCTTTTCTTATTTCTTTTATGATTTAATA

FIGURE 54c

Figure 55 An Entry (pENTR7) Clone of CAT Subcloned into pDEST2



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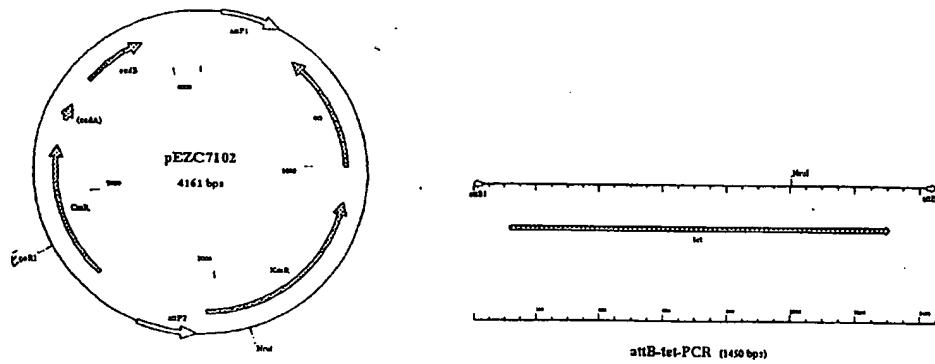


FIGURE 5b

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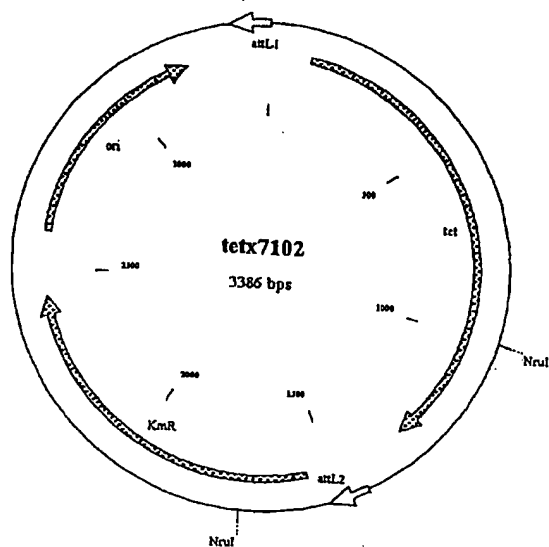


FIGURE 57

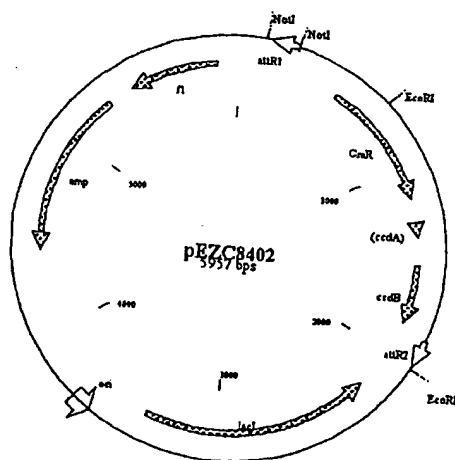


FIGURE 5B

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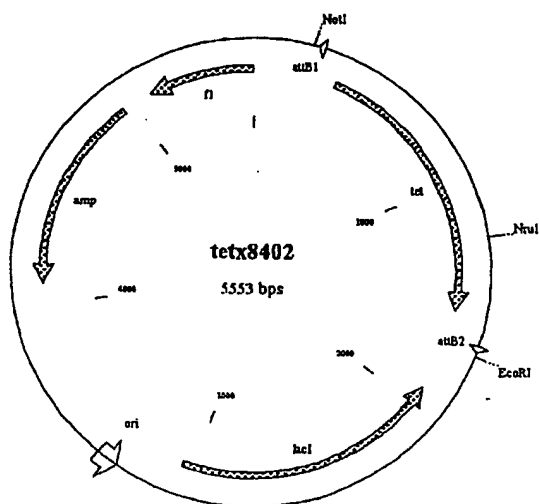


FIGURE 59

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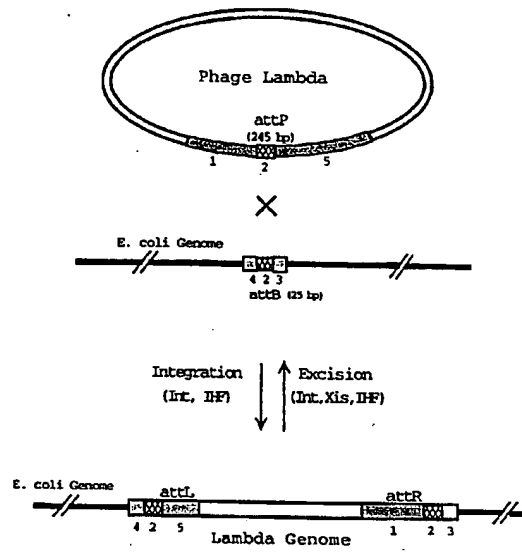


FIGURE 60

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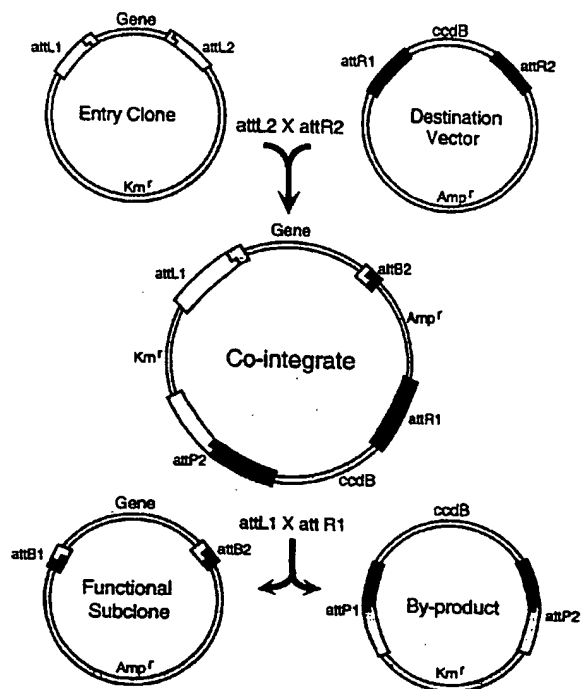


FIGURE 61

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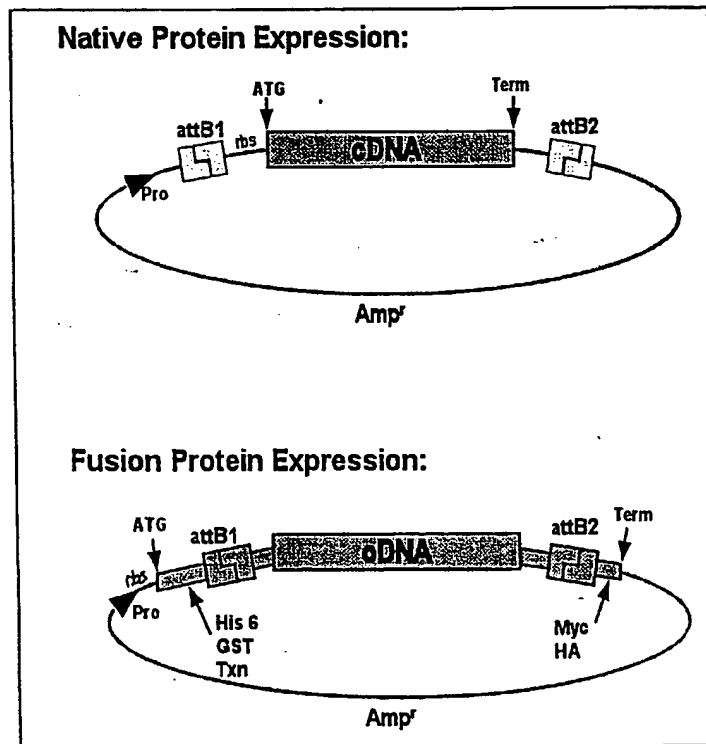


FIGURE 62

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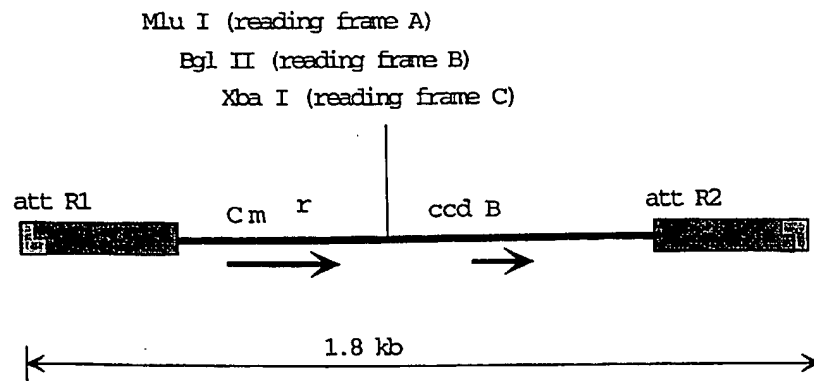


FIGURE 63

FIGURE 64A

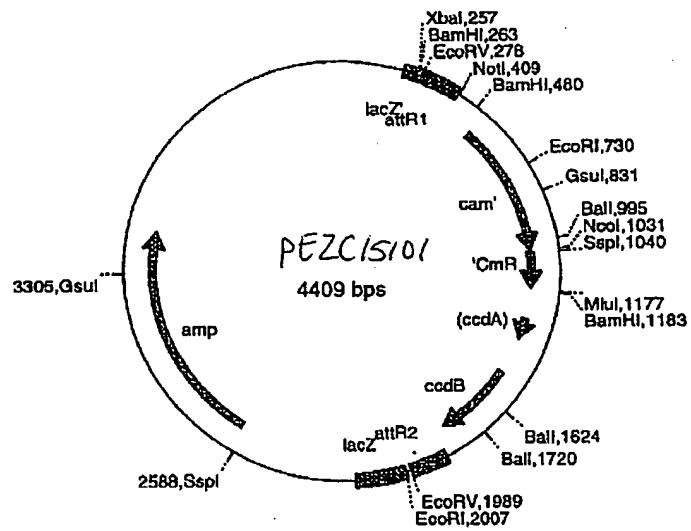
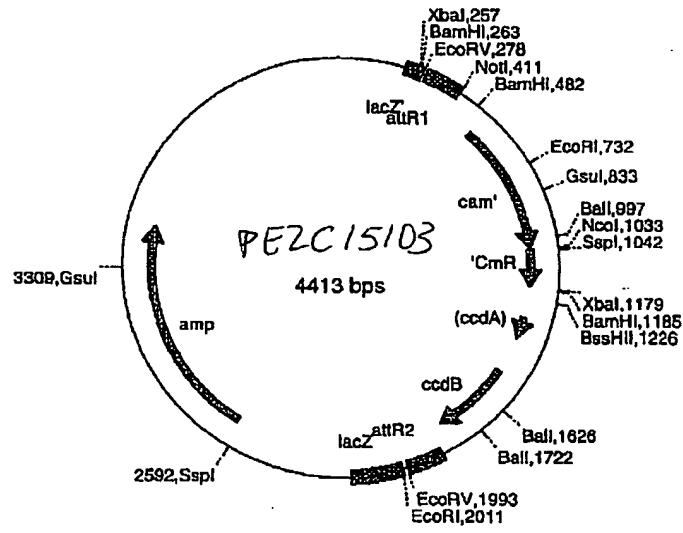


FIGURE 64C



Primers for Amplifying *tetR* and *ampR* for Cloning by Recombination

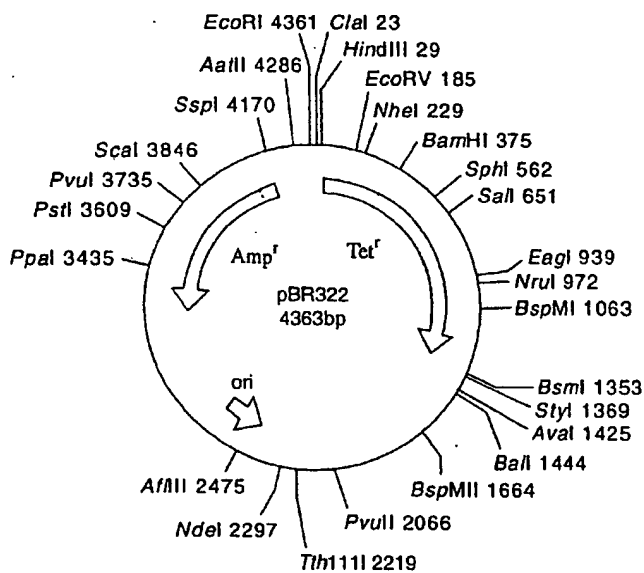
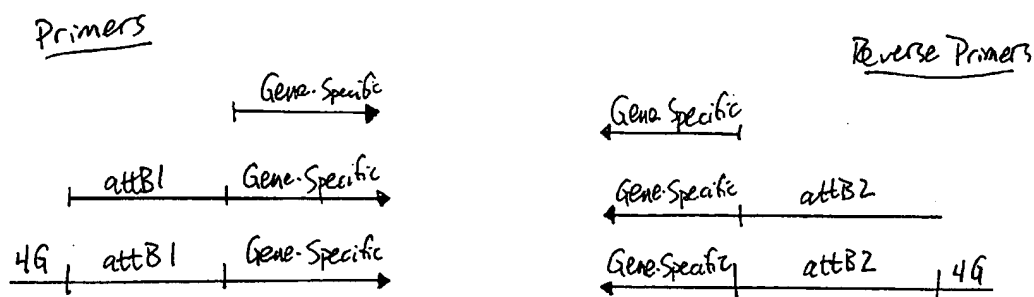


FIGURE 65

**Results of Cloning
tet and amp PCR Products
by Recombination**

PCR Product Used in GCS Reactions	No. Colonies Obtained (100 μ l plated)	Form of DNA Analyzed	Colonies Obtained of Predicted Size
tet	6, 10	SC	0 of 8
attB-tet	9, 6	SC	1 of 8
attB+4G-tet	824, 1064	SC AvaI+Bam	7 of 7 7 of 7
amp	7, 13	SC	0 of 8
attB-amp	18, 22	SC	3 of 8
attB+4G-amp	3020, 3540	SC PstI	8 of 8 8 of 8
attB Plasmid (Pos. Control)	320, 394		

FIGURE 66

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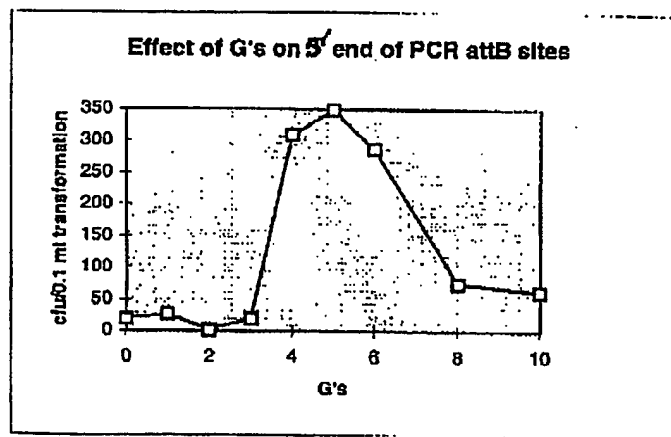


FIGURE 67

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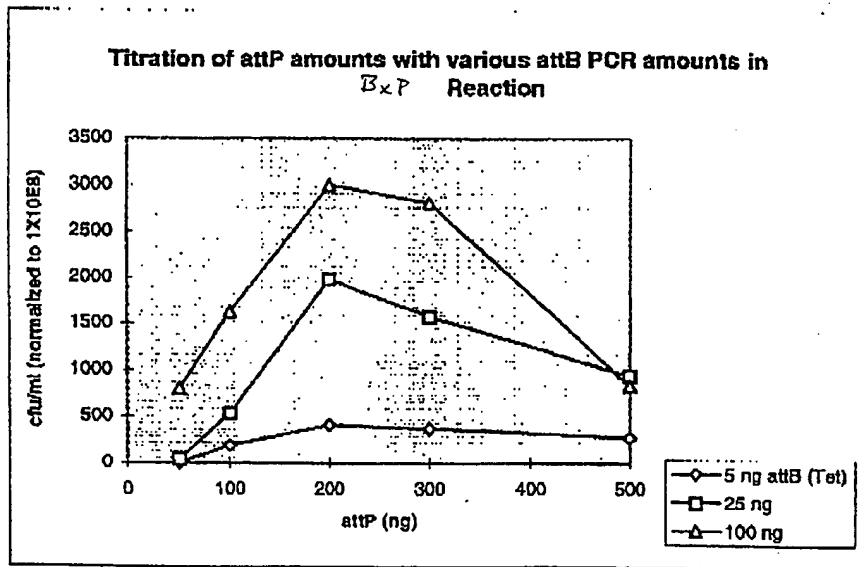
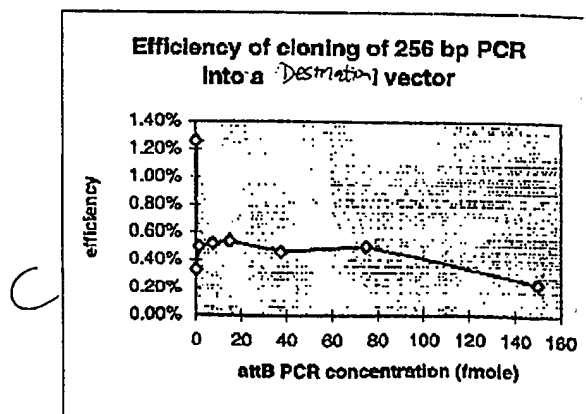
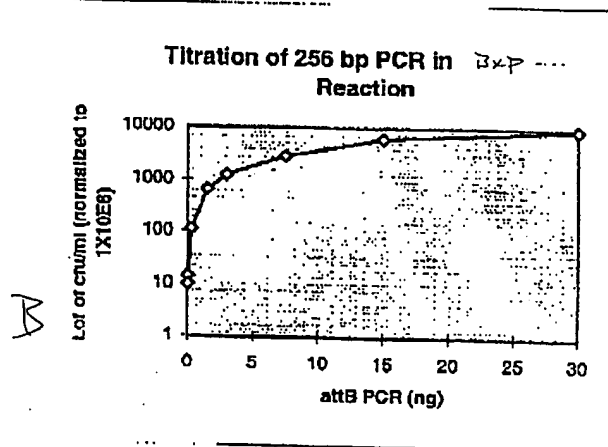
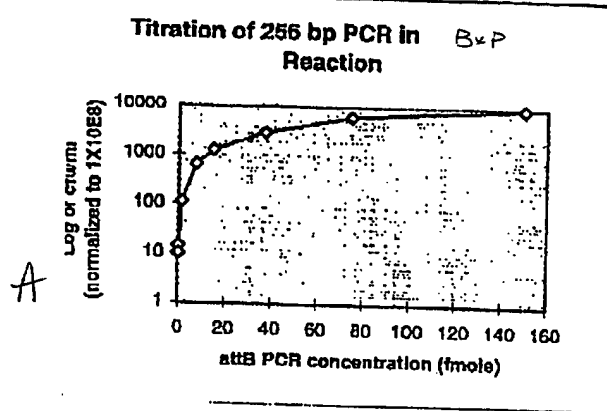


FIGURE 68

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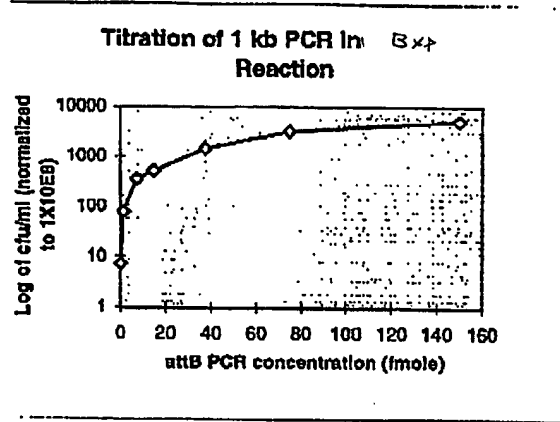
FIGURE
69



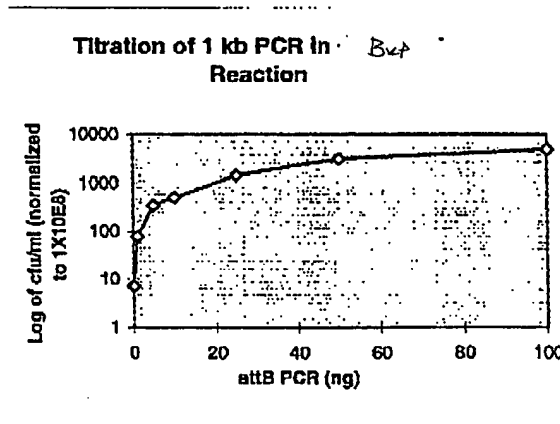
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FIGURE
70

A



B



C

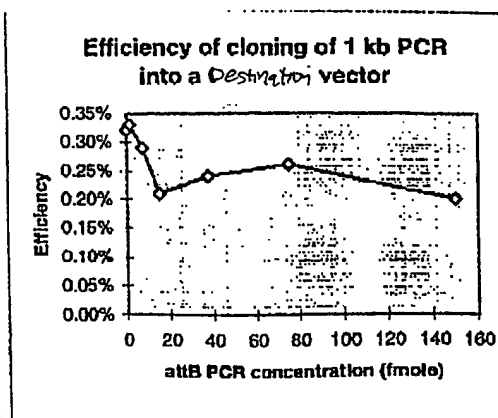
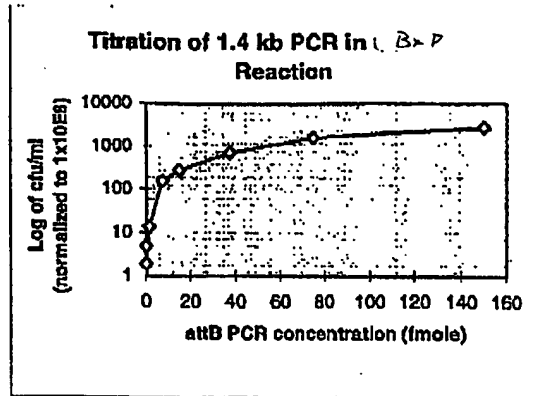
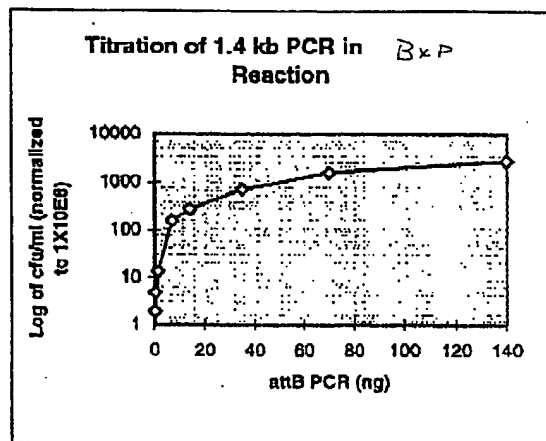


FIGURE 71

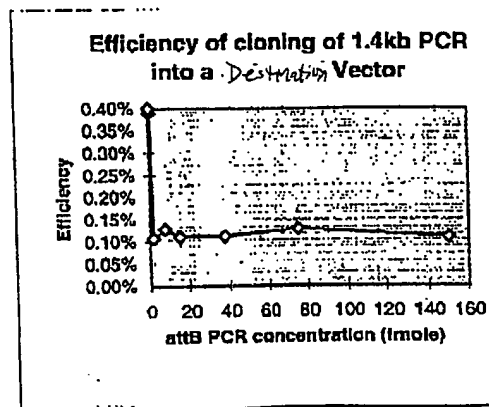
A



B



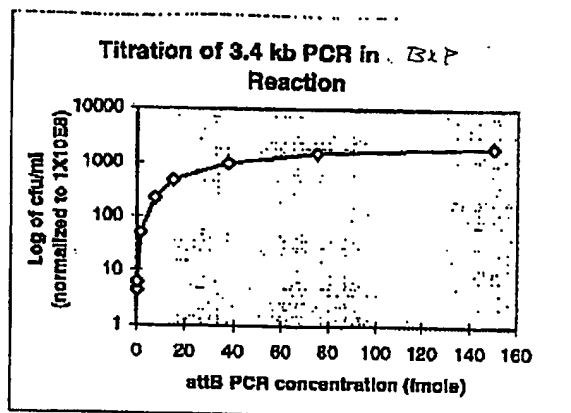
C



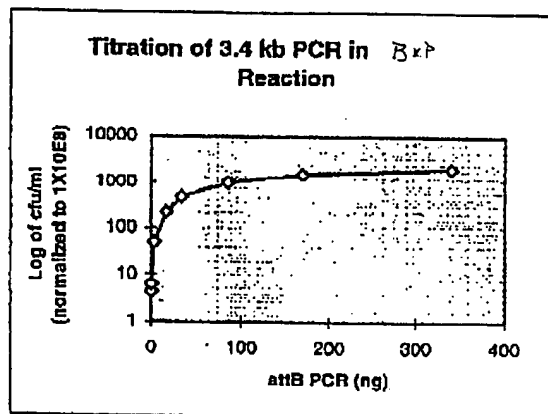
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FIGURE 72

A



B



C

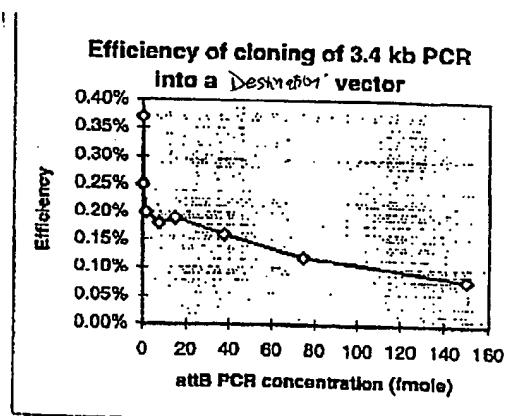
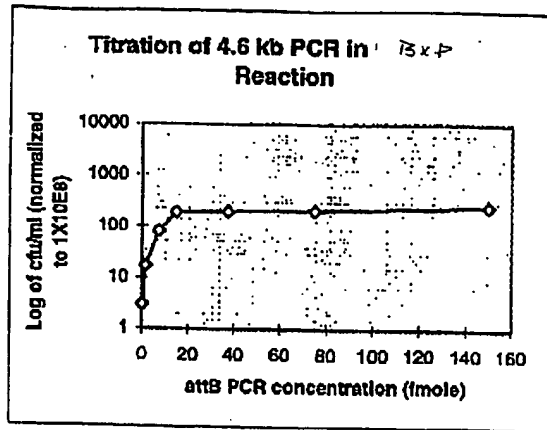
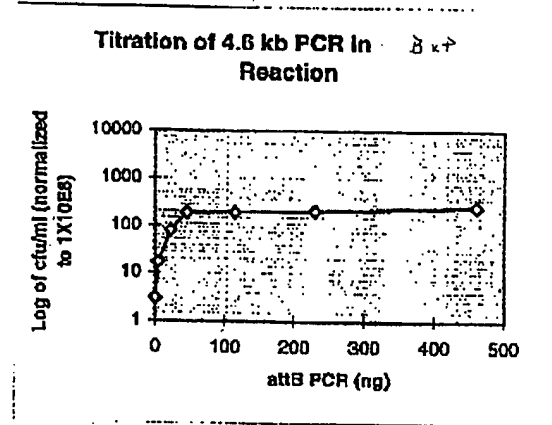


FIGURE 73

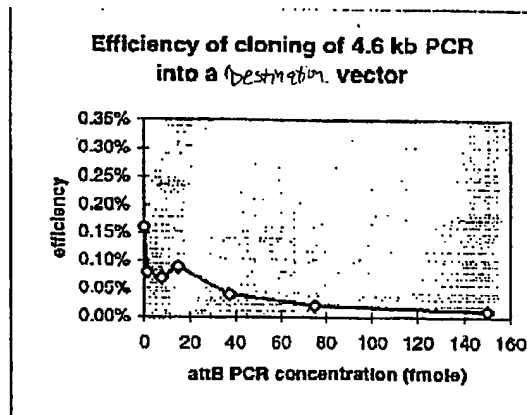
A



B



C



6.9 kb PCR DNA Titration in α B \times P Reaction

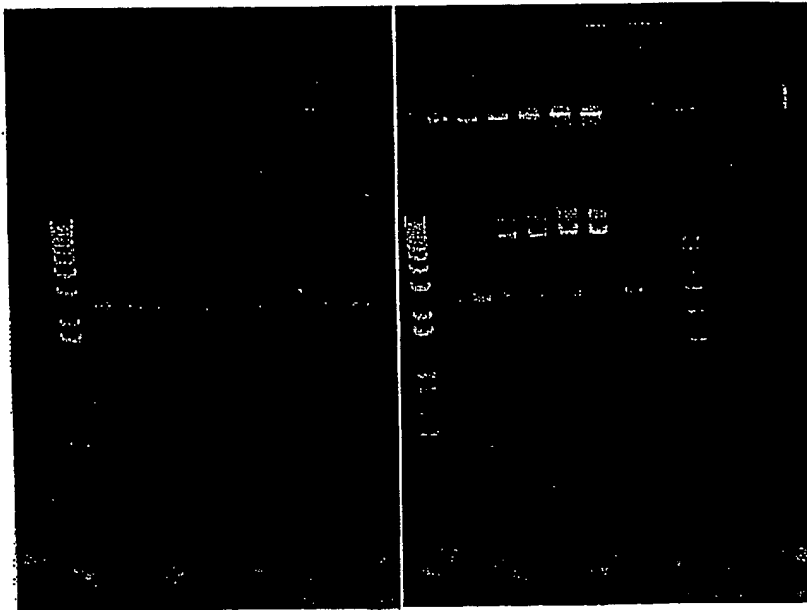


FIGURE 74

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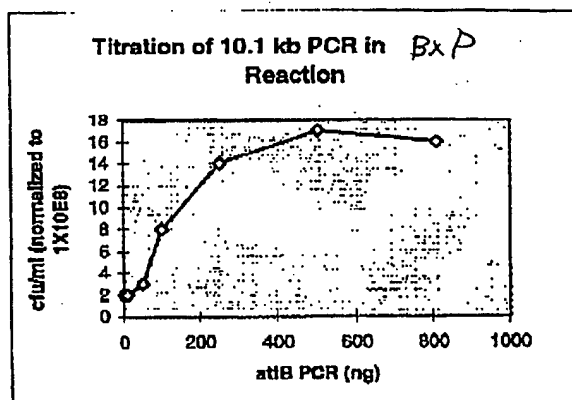


FIGURE 75-

10.1 kb PCR DNA Titration in Bx7 Reaction

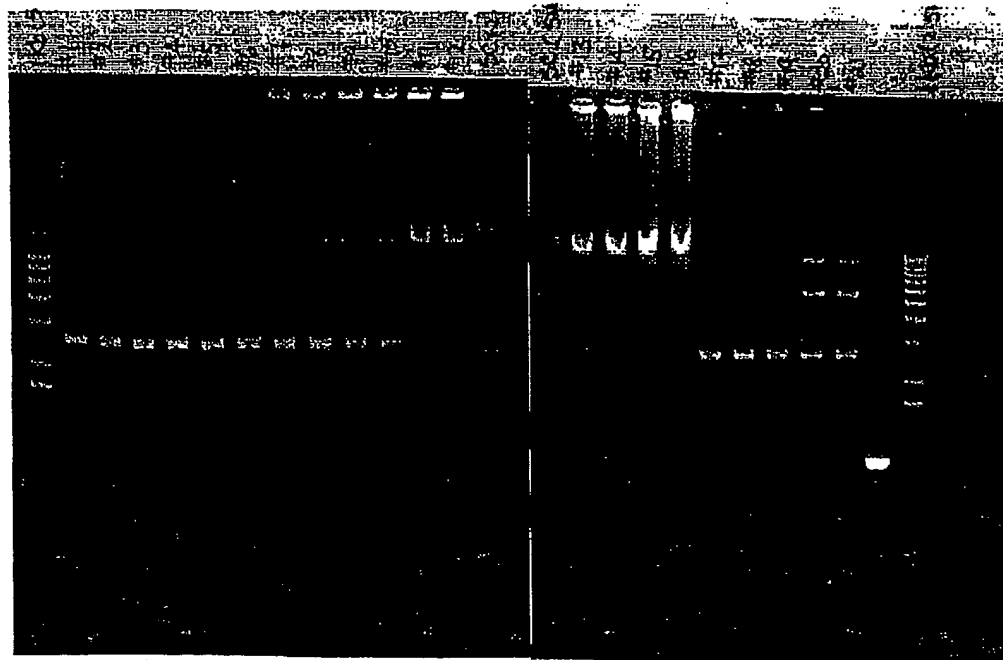


FIGURE 76

**Cloning of PCR Products of Different Sizes with the
GATEWAY™ PCR Cloning System**

Size	fmols PCR DNA	ng PCR DNA	Cols/ml Transformation (pUC=10 ⁸ CFU/ml)	Correct Clones/Total Examined**
0.26 kb*	15	3	1223	10/10 (a)
	37.5	7.5	2815	
1.0 kb	15	10	507	49/50 (b)
	37.5	25	1447	
1.4 kb	15	14	271	48/50 (c)
	37.5	35	683	
3.4 kb	15	34	478	9/10 (a)
	37.5	85	976	
4.6 kb	15	46	190	10/10 (a)
	37.5	115	195	
6.9 kb	15	69	30 (235)**	47/50 (b)
	37.5	173	54 (463)**	

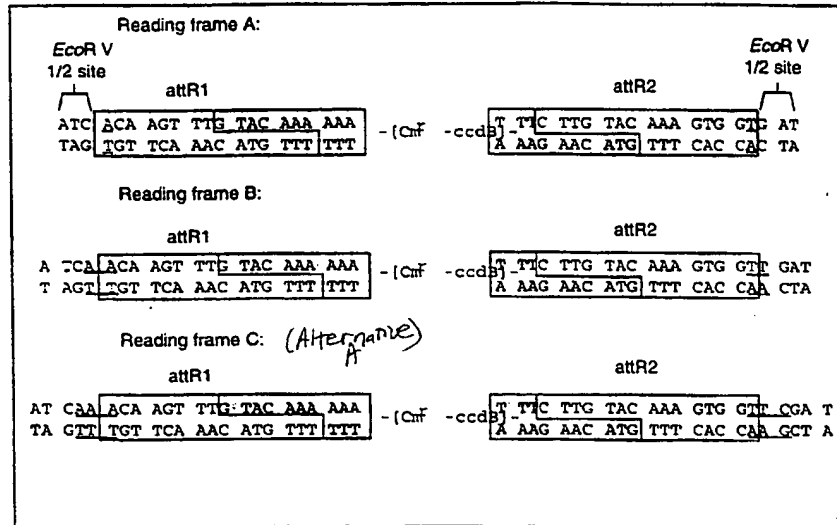
*The 0.26 kb PCR product was used unpurified; all the others were purified by precipitation with PEG/MgCl₂ as described in the text of Example 9, to remove primer dimers potentially present. Standard incubations were for 60 min.

**overnight incubation

- (a) DNA minipreps
- (b) ampR/kanR
- (c) tetR/kanR

Figure 77

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Reading frame C: (Alternative)

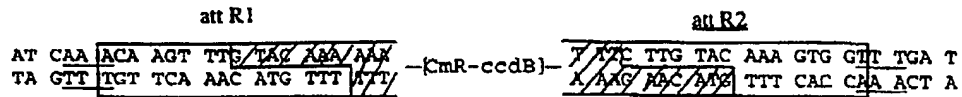


FIGURE 78

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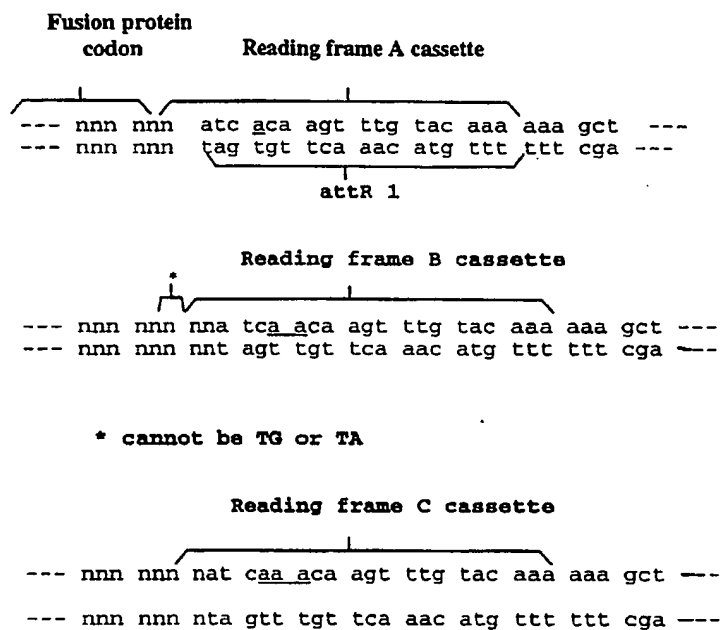


FIGURE 79

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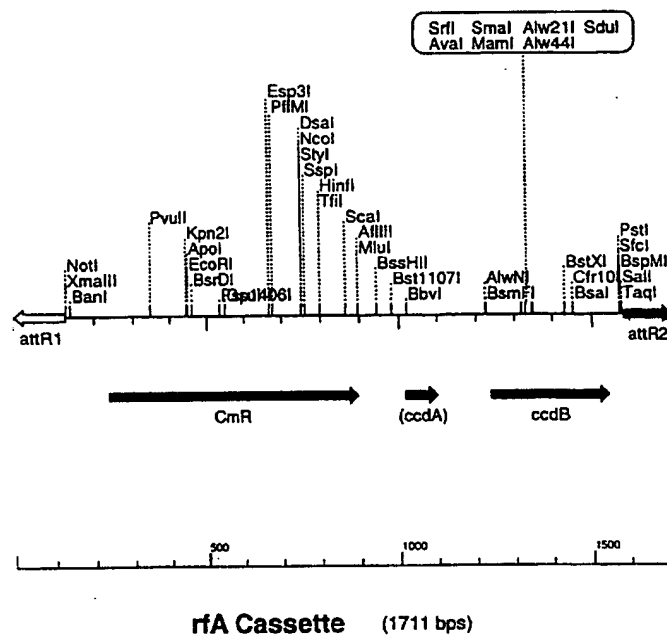


FIGURE 80

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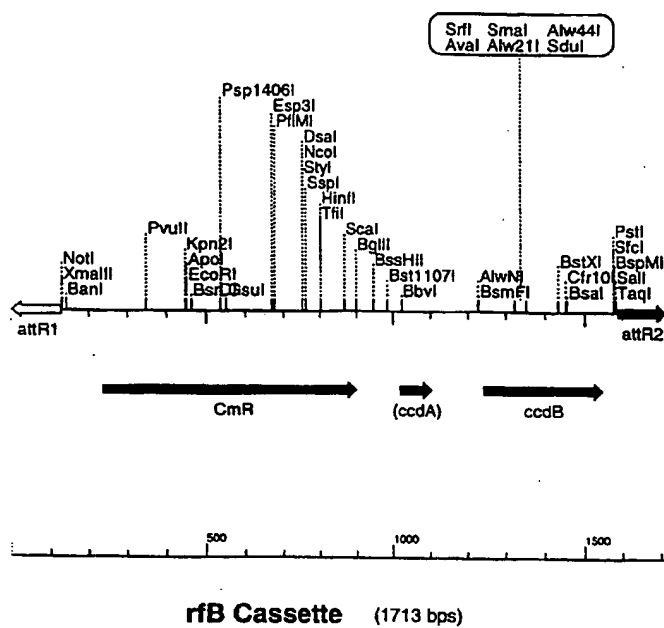
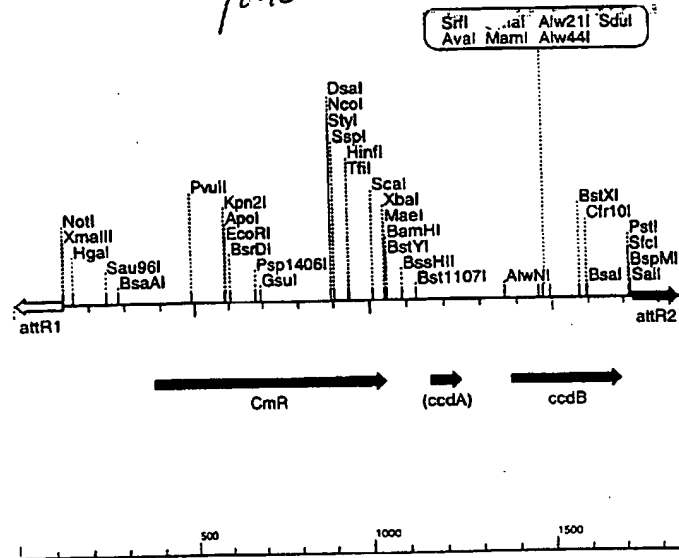


FIGURE 81

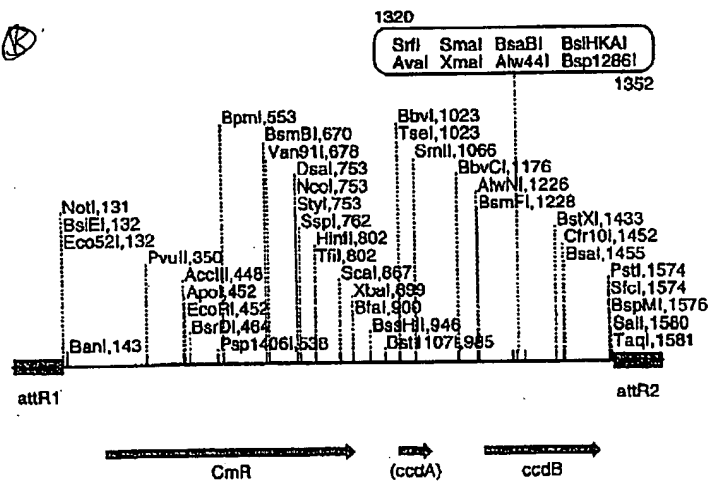
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(A)



rC Cassette (1856 bps)

(B)



rC cassette (1715 bps)

FIGURE 02

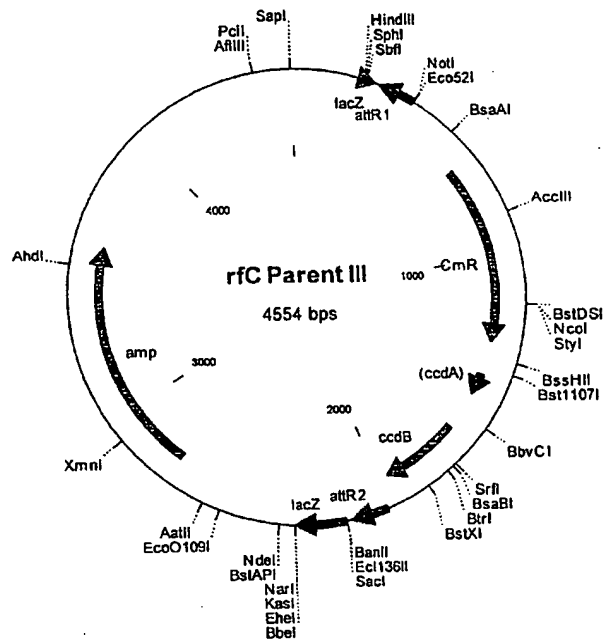


FIGURE 83A

prfC Parent III 4554 bp

<u>Location (Base Nos.)</u>		<u>Gene Encoded</u>
410..286		attR1
660..1319		CmR
1439..1523		inactivated ccdA
1661..1966		ccdB
2007..2131		attR2
2753..3613		amp

1	GCGCCCAATA	CGCAAACCGC	CTCTCCCCGC	GCGTTGGCCG	ATTCATTAAT	GCAGCTGGCA
61	CGACAGGTTT	CCCGACTGGA	AAGCGGGCAG	TGAGCGCAAC	GCAATTAATG	TGAGTTAGCT
121	CACTCATTAG	GCACCCGAGG	CTTTACACTT	TATGCTTCCG	GCTCGTATGT	TGTGTGGAAT
181	TGTGAGCGGA	TAACAATTTT	ACACAGGAAA	CAGCTATGAC	CATGATTACG	CCAAGCTTGC
241	ATGCCCTGAG	GTCGACTCTA	GAGGATCCCC	GGGTACCGAT	ATCAAACAAG	TTTGTACAAA
301	AAAGCTGAAC	GAGAAACGTA	AAATGATATA	AATATCAATA	TATTAAATTA	GATTTTGCAT
361	AAAAAACAGA	CTACATAATA	CTGTAAAAACA	CAACATATCC	AGTCACATATG	GCGGCCCGCTA
421	AGTTGGCAGC	ATCACCCGAC	GCACCTTGGC	CCGAATAAAT	ACCTGTGACG	GAAGATCACT
481	TCGCAGAATA	AATAAATCCT	GGTGTCCTGT	TTGATACCGG	GAAGCCCTGG	GCCAACTTTT
541	GGCGAAAATG	AGACGTTGAT	CGGCACGTAA	GAGGTTCCAA	CTTTCACCAT	AATGAAATAA
601	GATCACTACC	GGGCGTATTT	TTTGAGTTAT	CGAGATTTTC	AGGAGCTAAG	GAAGCTAAAA
661	TGGAGAAAAA	AATCACTGGA	TATACCACCG	TTGATATATC	CCAATGGCAT	CGTAAAGAAC
721	ATTTTGAGGC	ATTTCACTCA	GTTGCTCAAT	GTACCTATAA	CCAGACCGTT	CAGCTGGATA
781	TTACGGCCTT	TTTAAAGACC	GTAAAGAAAA	ATAAGCACAA	GTTTTATCCG	GCCTTTATTC
841	ACATTCCTTG	CCGCGTGATG	AATGCTCATC	CGGAATTTCCG	TATGGCAATG	AAAGACGGTG
901	AGCTGGTGAT	ATGGGATAGT	GTTTACCCTT	GTTACACCGT	TTTCCATGAG	CAAACTGAAA
961	CGTTTTCATC	GCTCTGGAGT	GAATACCACG	ACGATTTCCG	GCAGTTTCTA	CACATATATT
1021	CGCAAGATGT	GGCGTGTAC	GGTGAAGAAC	TGGCCTATTT	CCCTAAAGGG	TTTATTGAGA
1081	ATATGTTTTT	CGTCTCAGCC	AATCCCTGGG	TGAGTTTCAC	CAGTTTGTAT	TTAAACGTGG
1141	CCAATATGGA	CAACTTCTTC	GCCCCCGTTT	TCACCATGGG	CAAAATATTAT	ACGCAAGGCG
1201	ACAAGGTGCT	GATGCCGCTG	GCGATTCAAG	TTTCATCATG	CGTCTGTGAT	GGCTTCCATG
1261	TCGGCAGAAT	GCTTAATGAA	TTACAACAGT	ACTGCGATGA	GTGGCAGGGC	GGGGCGTAAT
1321	CTAGAGGATC	CGGCTTACTA	AAAGCCAGAT	AACAGTATGC	GTATTTGCGC	GCTGATTTTT
1381	GCGGTATAAG	AATATATACT	GATATGTATA	CCCGAAGTAT	GTCAAAAAGA	GGTGTGCTAT
1441	GAAGCAGCGT	ATTACAGTGA	CAGTTGACAG	CGACAGCTAT	CAGTTGCTCA	AGGCATATAT
1501	GATGTCAATA	TCTCCGGTCT	GGTAAGCACA	ACCATGCAGA	ATGAAGCCCC	TCGTCTGCGT
1561	GCCGAACGCT	GGAAGCGGGA	AAATCAGGAA	GGGATGGCTG	AGGTCCGCCC	GTTTATTGAA
1621	ATGAACGGCT	CTTTTGCTGA	CGAGAACAGG	GACTGGTGAA	ATGCAGTTTA	AGGTTTACAC
1681	CTATAAAAAG	GAGAGCCGTT	ATCGTCTGTT	TGTGGATGTA	CAGAGTGATA	TTATTGACAC
1741	GCCCCGGCGA	CGGATGGTGA	TCCCCCTGGC	CAGTGCACGT	CTGCTGTCAG	ATAAAGTCTC
1801	CCGTGAACCT	TACCCGGTGG	TGCATATCGG	GGATGAAAGC	TGGCGCATGA	TGACCACCGA
1861	TATGGCCAGT	GTGCCGGTCT	CCGTTATCGG	GGAAGAAGTG	GCTGATCTCA	GCCACCGCGA
1921	AAATGACATC	AAAAACGCCA	TTAACCTGAT	GTTCTGGGGA	ATATAAATGT	CAGGCTCCGT
1981	TATACACAGC	CAGTCTGCGA	GTCGACCATA	GTGACTGGAT	ATGTTGTGTT	TTACAGTATT
2041	ATGTAGTCTG	TTTTTTATGC	AAAATCTAAT	TTAATATATT	GATATTTATA	TCATTTTACG
2101	TTTCTCGTTC	AGCTTTCTTG	TACAAAAGTG	TTGATATATC	GTACCGAGCT	CGAATTCAC
2161	GGCCGTCGTT	TTACAACGTC	GTGACTGGGA	AAACCCCTGGC	GTTACCCAAC	TTAATCGCCT
2221	TGCAGCACAT	CCCCCTTTTG	CCAGCTGGCG	TAATAGCGAA	GAGGCCCGCA	CCGATCGCCC
2281	TTCCCAACAG	TTGCGCAGCC	TGAATGGCGA	ATGGCGCCTG	ATGCGGTATT	TTCTCCTTAC
2341	GCATCTGTGC	GGTATTTTAC	ACCGCATATG	GTGCACTCTC	AGTACAATCT	GCTCTGATGC
2401	CGCATAGTTA	AGCCAGCCCC	GACACCCGCG	AACACCCGCT	GACGCGCCCT	GACGGGCTTG
2461	CTGTGTCCTG	GCATCCGCTT	ACAGACAAGC	TGTGACCGTC	TCCGGGAGCT	GCATGTGTCA
2521	GAGGTTTTCA	CCGTCATCAC	CGAAACGCGC	GAGACGAAAG	GGCCTCGTGA	TACGCCTATT
2581	TTTATAGGTT	AATGTCATGA	TAATAATGGT	TTCTTAGACG	TCAGGTGGCA	CTTTTCGGGG
2641	AAATGTGCGC	GGAACCCCTA	TTTGTATTAT	TTTCTAAATA	CAITCAAATA	TGTATCCGCT
2701	CATGAGACAA	TAACCCCTGAT	AAATGCTTCA	ATAATATTGA	AAAAGGAAGA	GTATGAGTAT
2761	TCAACATTTT	CGGTGCGCCC	TTATTCCCTT	TTTTGCGGCA	TTTTGCCTTC	CTGTTTTTGC

FIGURE 83B

2821 TCACCCAGAA ACGCTGGTGA AAGTAAAAGA TGCTGAAGAT CAGTTGGGTG CACGAGTGGG
2881 TTACATCGAA CTGGATCTCA ACAGCGGTAA GATCCTTGAG AGTTTTCGCC CCGAAGAACG
2941 TTTTCCAATG ATGAGCACTT TTAAGTTCT GCTATGTGGC GCGGTATTAT CCGTATTGA
3001 CGCCGGGCAA GAGCAACTCG GTCGCCGCAT ACACTATTCT CAGAATGACT TGGTTGAGTA
3061 CTCACCAAGT ACAGAAAAGC ATCTTACGGA TGGCATGACA GTAAGAGAAT TATGCAGTGC
3121 TGCCATAACC ATGAGTGATA ACACCTGCGC CAACCTACTT CTGACAACGA TCGGAGGACC
3181 GAAGGAGCTA ACCGCTTTTT TGCACAACAT GGGGGATCAT GTAACCTGCC TTGATCGTTG
3241 GGAACCGGAG CTGAATGAAG CCATACCAA CGACGAGCGT GACACCACGA TGCCTGTAGC
3301 AATGGCAACA ACGTTGCGCA AACTATTAA TGGCGAACTA CTTACTCTAG CTTCCCGCA
3361 ACAATTAATA GACTGGATGG AGGCGGATAA AGTTGCAGGA CCACTTCTGC GCTCGGCCCT
3421 TCCGGCTGGC TGGTTTATTG CTGATAAATC TGGAGCCGGT GAGCGTGGGT CTCGCGGTAT
3481 CATTCAGCA CTGGGGCCAG ATGGTAAGCC CTCCCGTATC GTAGTTATCT ACACGACGGG
3541 GAGTCAGGCA ACTATGGATG AACGAAATAG ACAGATCGCT GAGATAGGTG CCTCACTGAT
3601 TAAGCATTGG TAACTGTCAG ACCAAGTTTA CTCATATATA CTTTAGATTG ATTTAAAACT
3661 TCATTTTAA TTTAAAAGGA TCTAGGTGAA GATCCTTTTT GATAATCTCA TGACCAAAAT
3721 CCCTTAACGT GAGTTTTCGT TCCACTGAGC GTCAGACCCC GTAGAAAAGA TCAAAGGATC
3781 TTCTTGAGAT CCTTTTTTTC TGC GCGTAAT CTGCTGCTTG CAAACAAAAA AACCACCGCT
3841 ACCAGCGGTG GTTTGTTTGC CGGATCAAGA GCTACCAACT CTTTTCCGA AGGTAAGTGG
3901 CTTCAAGCAG GCGCAGATAC CAAATACTGT CCTTCTAGTG TAGCCGTAGT TAGGCCACCA
3961 CTTCAAGAAC TCTGTAGCAC CGCCTACATA CCTCGCTCTG CTAATCCTGT TACCAGTGGC
4021 TGCTGCCAGT GGCATAAAGT CGTGTCTTAC CGGGTTGGAC TCAAGACGAT AGTTACCGGA
4081 TAAGGCGCAG CGGTGCGGCT GAACGGGGGG TTCGTGCACA CAGCCAGCT TGGAGCGAAC
4141 GACCTACACC GAACTGAGAT ACCTACAGCG TGAGCTATGA GAAAGCGCCA CGCTTCCCGA
4201 AGGGAGAAAG GCGGACAGGT ATCCGGTAAG CGGCAGGGTC GGAACAGGAG AGCGCACGAG
4261 GGAGCTTCCA GGGGAAACG CCTGGTATCT TTATAGTCCT GTCGGGTTTC GCCACCTCTG
4321 ACTTGAGCGT CGATTTTGT GATGCTCGTC AGGGGGGCGG AGCCTATGGA AAAACGCCAG
4381 CAACGCGGCC TTTTACGGT TCCTGGCCTT TTGCTGGCCT TTTGCTCACA TGTTCCTTCC
4441 TCGGTTATCC CCTGATTCTG TGGATAACCG TATTACCGCC TTTGAGTGAG CTGATACCGC
4501 TCGCCGCAGC CGAACGACCG AGCGCAGCGA GTCAGTGAGC GAGGAAGCGG AAGA

FIGURE 83C

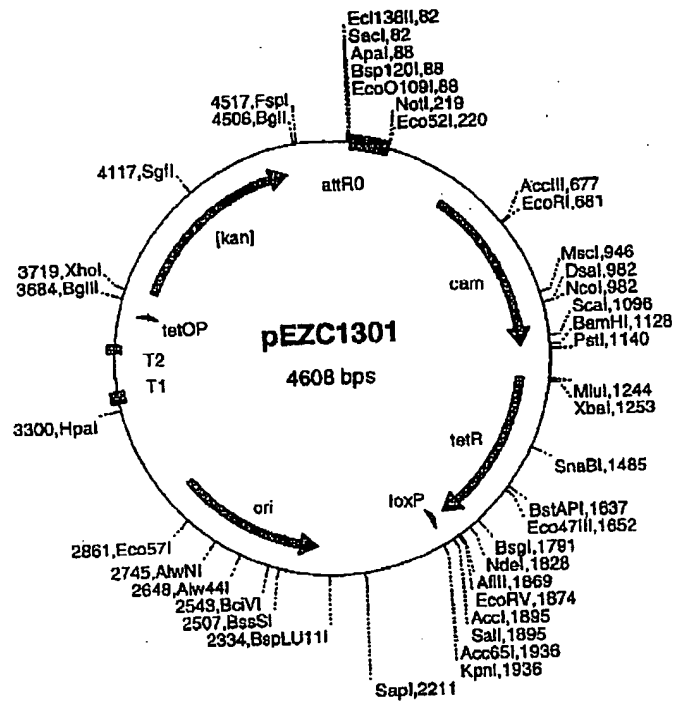


FIGURE 84

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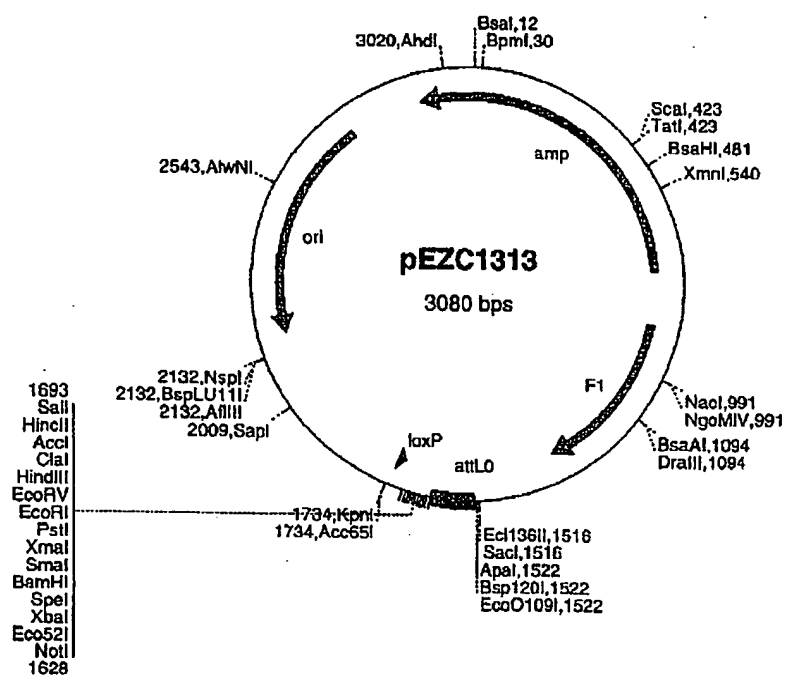


FIGURE 85

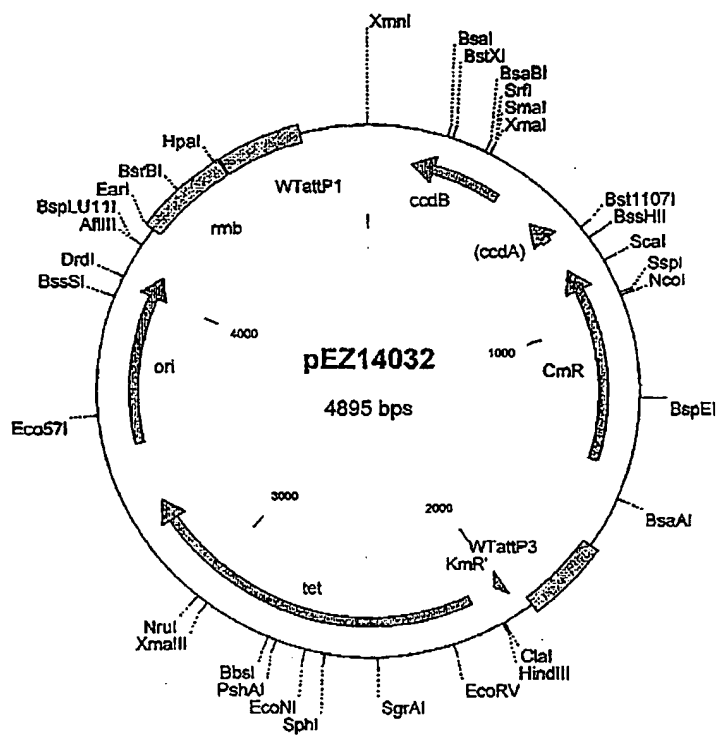


FIGURE 86

FIGURE 87

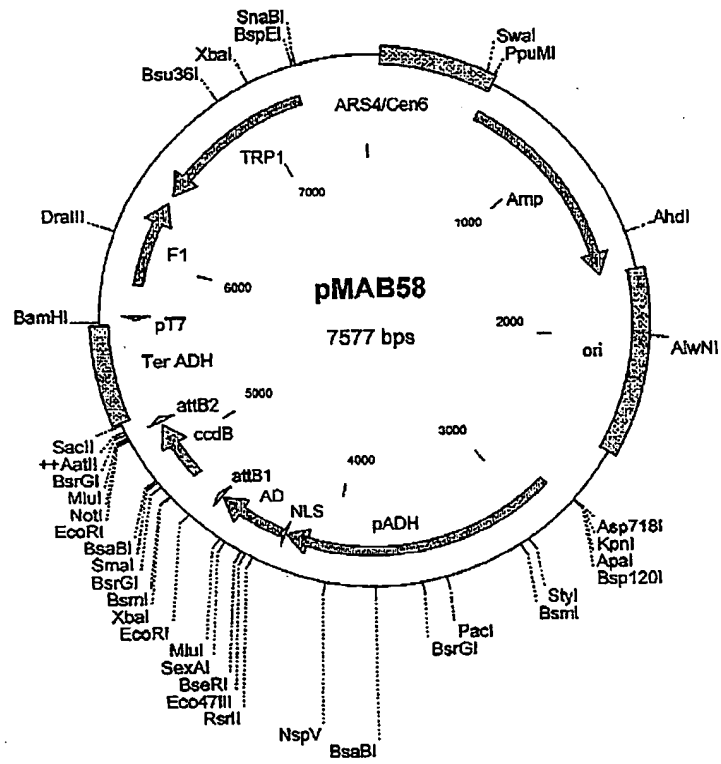
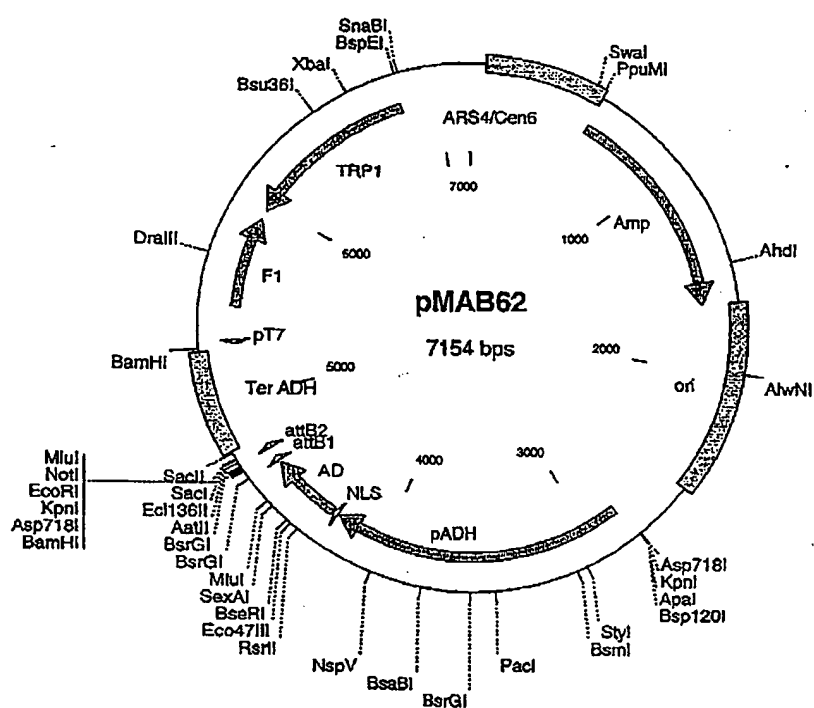
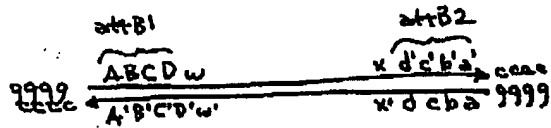
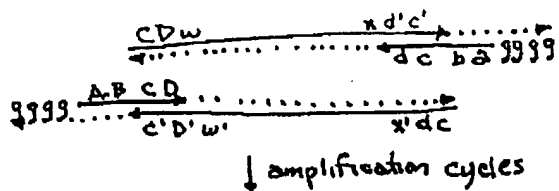
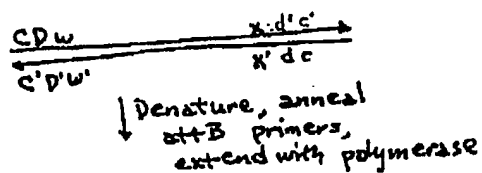
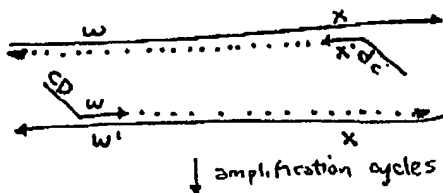
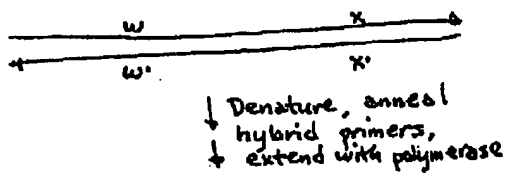


FIGURE 88



DNA to be amplified (5' → 3'):



attB1 primer:
9999 ABCD

attB2 primer:
9999 abcd

Hybrid primers (part
attB, part gene
specific):

CDw

ed x'

FIGURE 89

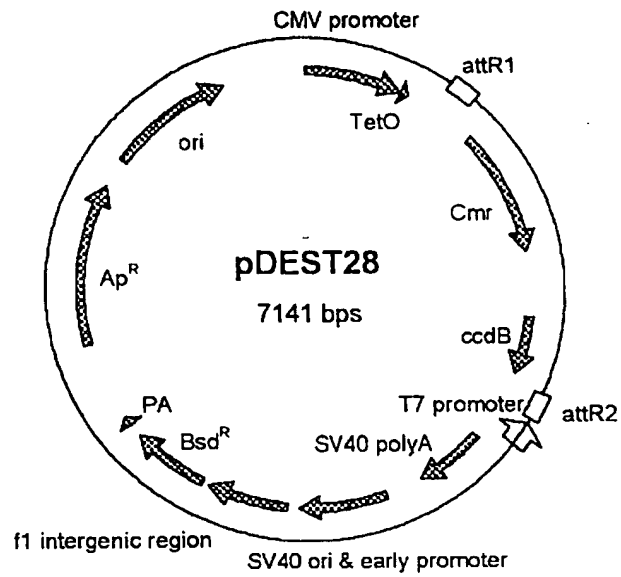


FIGURE 90A

pDEST28 7141 bp

ATGCATGTCGTTACATAAATTACGGTAAATGGCCCGCTGGCTGACCGCCCAACGACCCC
CGCCCATTTGACGTCAATAATGACGTATGTTCCCATAGTAACGCCAATAGGGACTTTCCAT
TGACGTCAATGGGTGGAGTATTTACGGTAAACTGCCCACTTGGCAGTACATCAAGTGTAT
CATATGCCAAGTACGCCCCCTATTGACGTCAATGACGGTAAATGGCCCGCTGGCATTAT
GCCCAGTACATGACCTTATGGGACTTTCTACTTGGCAGTACATCTACGTATTAGTCATC
GCTATTACCATGGTGATGCGGTTTGGCAGTACATCAATGGGCGTGGATAGCGGTTTGAC
TCACGGGGATTTCCAAGTCTCCACCCCATGACGTCAATGGGAGTTTGTGTTGGCACCAA
AATCAACGGGACTTTCCAAAATGTCGTAACAACCTCCGCCCCATTGACGCAAATGGGCGGT
AGGCGTGTACGGTGGGAGGTCTATATAAGCAGAGCTCTCCCTATCAGTGATAGAGATCTC
CCTATCAGTGATAGAGATCGTCGACGAGCTCGTTTAGTGAACCGTCAGATCGCCTGGAGA
CGCCATCCACGCTGTTTGGACCTCCATAGAAGACACCGGGACCGATCCAGCCTCCGGACT
CTAGAGGATCCCTACCGGTGATATCCTCGAGCCCATCAACAAGTTGTACAAAAAAGCTG
AACGAGAAAACGTAAATGATATAAATATCAATATATTAAATTAGATTTTGCATAAAAAAC
AGACTACATAAATCTGTAAACACAAACATATCCAGTCACTATGGCGGCCGATTAGGCAC
CCCAGGCTTTACACTTTATGCTTCCGGCTCGTATAATGTGTGGATTTTGTAGTTAGGATCC
GGCGAGATTTTTCAGGAGCTAAGGAAGCTAAATGGAGAAAAAATCACTGGATATACCAC
CGTTGATATATCCCAATGGCATCGTAAAGAACATTTTGGAGCATTTTCAGTCAGTTGCTCA
ATGTACCTATAACCAGACCGTTTCAGCTGGATATTACGGCCTTTTAAAGACCGTAAAGAA
AAATAAGCACAAGTTTATCCGGCTTTATTCACATTTCTGCCCCCTGATGAATGCTCA
TCCGGAATTCGATATGGCAATGAAAGACGGTGAGCTGGTATATGGGATAGTGTTCACCC
TTGTTACACCGTTTCCATGAGCAAACCTGAAACGTTTTCATCGCTCTGGAGTGAATACCA
CGACGATTTCCGGCAGTTTCTACACATATATTCGCAAGATGTGGCGTGTACGGTGAAAA
CCTGGCCTATTTCCCTAAAGGGTTTATTGAGAATATGTTTTCGTCTCAGCCCAATCCCTG
GGTGAGTTTTCACAGTTTGTATTTAAACGTGGCCCAATATGGACAACCTTCTCGCCCCGT
TTTCACCATGGGCAAATATTATACGCAAGGCGACAAGGTGCTGATGCCGCTGGCGATTCA
GGTTCATCATGCCGTCTGTGATGGCTTCCATGTGGCAGAATGCTTAATGAATTACAACA
GTACTGCGATGAGTGGCAGGGCGGGCGTAAAGATCTGGATCCGGCTTACTAAAAGCCAG
ATAACAGTATGCGTATTTGCGCGCTGATTTTTCGGGTATAAGAATATATACTGATATGTA
TACCCGAAGTATGTCAAAAAGAGGTGTGCTATGAAGCAGCGTATTACAGTGACAGTTGAC
AGCGACAGCTATCAGTTGCTCAAGGCATATATGATGTCAATATCTCCGGTCTGGTGAACA
CAACCATGCAAGTGAAGCCCGTCTGTCTGCGTGGCAGACGCTGGAAAGCGGAAATCAGG
AAGGGATGGCTGAGGTGCGCCCGGTTTATTGAAATGAACGGCTCTTTTGTCTGACGAGAACA
GGGACTGGTGAATGCAAGTTTAAAGTTTACACCTATAAAAGAGAGAGCCGTTATCGTCTG
TTTTGTGGATGTACAGAGTGATATTATTGACACGCCCCGGGCGACGGATGGTGTATCCCCCTG
GCCAGTGCACGTCTGCTGTGATGATAAAGTCTCCCGTGAACCTTACCCGGTGGTGCATATC
GGGGATGAAAGCTGGCGCATGATGACCACCGATATGGCCAGTGTGCCGGTCTCCGTTATC
GGGGAAGAAGTGGCTGATCTCAGCCACCGGAAAATGACATCAAAAACGCCATTAACTG
ATGTTCTGGGGAATATAAATGTGAGGCTCCCTTATACACAGCCAGTCTGCAGGTGACCA
TAGTGACTGGATATGTTGTGTTTACAGTATTATGTAGTCTGTTTTTATGCAAAATCTA
ATTTAATATATTGATATTTATATCATTTTACGTTTCTCGTTACGCTTTCTGTACAAAGT
GGTTGATGGGCGGCCGCTCTAGAGGGCCCAAGCTTACGCGTGCATGCGACGTATAGCTC
TCTCCCTATAGTGAGTCGTATTATAAGCTAGGCACCTGGCCGTCGTTTACAACGTCGTGA
CTGGGAAAACCTGCTAGCTTGGGATCTTTGTGAAGGAACCTTACTTCTGTGGTGTGACATA
ATTGGACAAAACCTACCTACAGAGATTTAAAGCTCTAAGGTAAATATAAAATTTTAAAGTGT
ATAATGTGTTAACTAGCTGCATATGCTTGTCTGTGAGAGTTTGTCTTACTGAGTATGA
TTTATGAAAATATTATACACAGGAGCTAGTGATTCTAATTGTTTGTGTATTTAGATTCA
CAGTCCCAAGGCTCATTTAGGCCCTCAGTCCTCAGAGTCTGTTTATGATCATAATCAG
CCATACCACATTTGTAGAGGTTTTACTTGCTTTAAAAAACCTCCACACCTCCCCCTGAA
CCTGAAACATAAAATGAATGCAATTGTTGTTTAACTTGTATTATGACGCTTATAATGG
TTACAAAATAAAGCAATAGCATCACAAATTTACAAAATAAAGCATTTTTTCACTGCATTC
TAGTTGTGGTTTGTCCAACTCATCAATGTATCTTATCATGTCTGGATCGATCCTGCATT
AATGAATCGGCCAACGCGCGGGGAGAGGCGGTTTGGCTATTGGCTGGCGTAATAGCGAAG
AGGCCGCAACCGATCGCCCTTCCCAACAGTTGCGCAGCCTGAATGGCGAATGGGACGCGC
CCTGTAGCGGCGCATTAAGCGCGGCGGTTGGTGGTTACGCGCAGCGTGACCGCTACAC
TTGCCAGCGCCCTAGCGCCGCTCCTTTCGCTTCTTCCCTTCTTCTCGCCACGTTTCG
CCGGCTTTCCCCGTCAAGCTCTAAATCGGGGCTCCCTTAGGGTTCGATTTAGTGCTT-

Figure 90B

TACGGCACCTCGACCCCAAAAACTTGATTAGGGTGATGGTTACGCTAGTGGGCCATCGC
CCTGATAGACGGTTTTCGCCCTTTGACGTTGGAGTCCACGTTCTTTAATAGTGGACTCT
TGTTCCAACTGGAACAACACTCAACCCATATCTCGGTCTATTCTTTTGATTTATAAGGGA
TTTTGCCGATTTCGGCCTATTGGTTAAAAAATGAGCTGATTAAACAAATATTTAACGCGA
ATTTTAAACAAATATTAACGTTTACAATTTGCGCTGATGCGGTATTTCTCCTTACGCAT
CTGTGCGGTATTTACACCGCATACGCGGATCTGCGCAGCACCATGGCCTGAAATAACCT
CTGAAAGAGGAACCTGGTTAGGTACCTTCTGAGGCGGAAAGAACAGCTGTGGAATGTGT
GTCACTTAGGGTGTGGAAGTCCCCAGGCTCCCCAGCAGGCAGAAGTATGCAAAGCATGC
ATCTCAATTAGTCAGCAACCAGGTGTGGAAGTCCCCAGGCTCCCCAGCAGGCAGAAGTA
TGCAAAGCATGCATCTCAATTAGTCAGCAACCATAGTCCCGCCCTAACTCCGCCCATCC
CGCCCTTAACTCCGCCAGTTCCGCCCATTTCTCCGCCCATGGCTGACTAATTTTTTTTA
TTTATGCAAGGCGGAGGCGCCTCGGCCCTCTGAGCTATTCCAGAAGTAGTGAGGAGGCT
TTTTTGGAGGCTTAGGCTTTTTGCAAAAAGCTTGATTCTTCTGACACAACAGTCTCGAAT
TAAGACCATGGCCAAGCCTTTGTCTCAAGAAGAAATCCACCCTCATTGAAAGAGCAACGGC
TACAATCAACAGCATCCCCATCTCTGAAGACTACAGCGTCGCCAGCGCATCTCTTAG
CGACGGCGCATCTTCACTGGTGTCAATGTATATCATTTTTACTGGGGACCTTGTGCAGA
ACTCGTGTGTCTGGGCACTGTGTGTCTGCGGCAGCTGGCAACCTGACTGTATCTGTGC
GATCGGAAATGAGAACAGGGGCATCTTGAGCCCTGCGGACGTTGCCAGAGTGTCTCT
CGATCTGCATCTCTGGGATCAAGCCATAGTGAAGGACAGTGTGGACAGCGCAGCGCAGT
TGGGATTCGTGAATTGTGTCCTCTGGTTATGTGTGGAGGGCTAAGCACTTCGTGGCGG
AGTTTCGAAATGACCGACCAAGCGAGCGCCCAACCTGCCATCAGATGGCCGCAATAAAATA
CTTTTATTTTTCATTACATCTGTGTGTGGTTTTTTGTGTGAATCGATAGCGATAAGGATC
CGCGTATGGTGCACTCTCAGTACAATCTGCTCTGATGCCGATAGTTAAGCCAGCCCCGA
CACCCGCCAACACCCGCTGACGCGCCCTGACGGGCTTGTCTGTCTCCCGGCATCCGCTTAC
AGACAAGCTGTGACCGTCTCCGGAGCTGCATGTGTGTCAGAGGTTTACCCTCATCACCG
AAACGCGCGAGACGAAAGGGCCTCGTGATACGCTATTTTTATAGGTTAATGTATGATA
ATAATGGTTTTCTTAGACGTGAGGTGGCACTTTTCGGGGAAATGTGCGCGGAACCCCTATT
TGTTTTATTTTCTAAATACATTCAAATATGTATCCGCTCATGAGACAATAACCTGATAA
ATGCTTCAATAATATTGAAAAAGGAGATGAGTATTCAACATTTCCGTGTCCGCCCTT
ATTCCCTTTTTTGCGGCATTTTGCCCTTCCTGTTTTTGCTCACCCAGAAACGCTGGTGAAA
GTAAAGATGCTGAAGATCAGTTGGGTGCACGAGTGGGTTACATCGAACTGGATCTCAAC
AGCGGTAAGATCCTTGAGAGTTTTTCGCCCGAAGAACGTTTTCCAATGATGAGCATTTT
AAAGTTCTGCTATGTGGCGCGGTATTATCCCGTATTGACCGCGGCAAGAGCAACTCGGT
CGCCGCATACACTATTCTCAGAATGACTTGGTTGAGTACTACCACTCACAGAAAAGCAT
ACTGCGGCCAACTTACTTCTGACAACGATCGGAGGACCGAAGGAGCTAACCGCTTTTTTG
CACAAACATGGGGGATCATGTAACCTCGCCTTGATCGTTGGGAACCGGAGCTGAATGAAGCC
ATACCAACGACGAGCGTGACACCAGATGCCGTGATGCAATGGCAACAACGTTGCGCAAA
CTATTAACTGGCGAACTACTTACTCTAGCTTCCCGGCAACAATTAATAGACTGGATGGAG
GCGGATAAAGTTGCAGGACCACTTCTGCGCTCGGCCCTTCCGGCTGGCTGGTTTATTGCT
GATAAATCTGGAGCCGGTGAGCGTGGGTCTCGCGGTATCATTGCAGCACTGGGGCCAGAT
GGTAAGCCCTCCCGTATCGTAGTTATCTACACGACGGGGAGTCAGGCAACTATGGATGAA
CGAAATAGACAGATCGCTGAGATAGGTGCCTCACTGATTAAAGCATTGGTAAGTGTGAGAC
CAAGTTTACTCATATATACTTTAGATTGATTAAAACTTCATTTTTTAATTTAAAGGATC
TAGGTGAAGATCCTTTTTGATAATCTCATGACCAAAATCCCTTAACGTGAGTTTTTCGTTT
CACTGAGCGTCAGACCCCGTAGAAAAGATCAAAGGATCTTCTTGAGATCCTTTTTTTCTG
CGCGTAATCTGCTGCTTGCAAAACAAAAAACACCGCTACCAGCGGTGGTTGTGTGCGG
GATCAAGAGCTACCAACTCTTTTTCCGAAGGTAAGTGGCTTACGAGAGCGCAGATACCA
AATACTGTCCTTCTAGTGTAGCCGTAGTTAGGCCACCACTTCAAGAACTCTGTAGCACCG
CCTACATACCTCGCTCTGTAATCCTGTTACCAAGTGCTGCTGCCAGTGGCGATAAGTCTG
TGTCTTACCGGGTTGGACTCAAGACGATAGTTACCGGATAAGGCGCAGCGTCCGGCTGA
ACGGGGGGTTCGTGCACACAGCCAGCTTGAGCGAAGCAGCTACACCGAACTGAGATAC
CTACAGCGTGAGCATTGAGAAAGCGCCACGCTTCCGAAGGGAGAAAGGCGGACAGGTAT
CCGGTAAGCGGCAGGTCGGAACAGGAGAGCGCACGAGGGAGCTTCCAGGGGAAACCGC
TGGTATCTTTATAGTCTGTGCGGTTTCGCCACCTCTGACTTGAGCGTCGATTTTTGTGA
TGCTCGTCAGGGGGCGGAGCCTATGAAAAACGCCAGCAACGCGGCTTTTTACGGTTC
CTGGCCTTTTGTGCGCTTTTGCTCACATGTTCTTTCTGCGTTATCCCTGATTCTGTG
GATAACCGTATTACCGCCTTTGAGTGAGCTGATACCGCTCGCCGAGCCGAACGACCGAG-

FIGURE 90C

WO 00/52027

204/260

PCT/US00/05432

CGCAGCGAGTCAGTGAGCGAGGAAGCGGAAGAGCGCCCAATACGCAAACCGCCTCTCCCC
GCGCGTTGGCCGATTCATTAATGCAGAGCTTGCAATTGCGCGTTTTTCAATATTATTGA
AGCATTATCAGGGTTATTGTCTCATGAGCGGATACATATTGAATGTATTTAGAAAAAT
AAACAAATAGGGGTTCCGCGCACATTTCCCGAAAAGTGCCACCTGACGTCTAAGAAACC
ATTATTATCATGACATTAACTATAAAAAATAGGCGTAGTACGAGGCCCTTCACTCATTAG
G

FIGURE 901

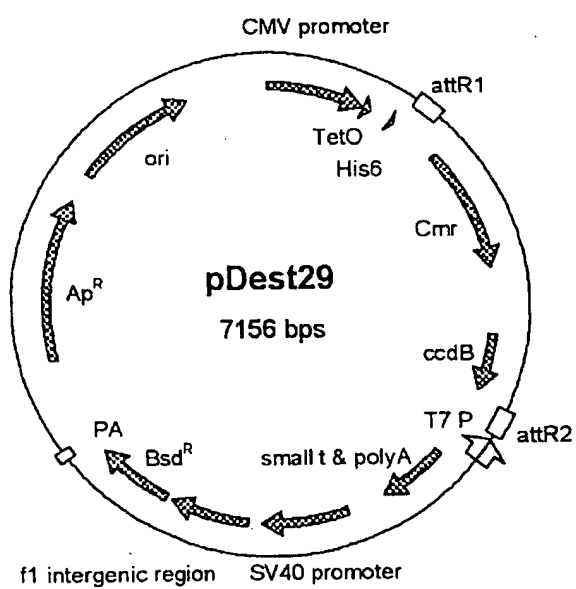


FIGURE 91 A

pDEST29 7156 bp

ATGCATGTCGTTACATAACTTACGGTAAATGGCCCGCCTGGCTGACCGCCCAACGACCCC
CGCCCATTTGACGTCATAATGACGTATGTTCCCATAGTAACGCCAATAGGGACTTCCCAT
TGACGTCAATGGGTGGAGTATTTACGGTAACTGCCCACTTGGCAGTACATCAAGTGTAT
CATATGCCAAGTACGCCCCCTATTGACGTCAATGACGGTAAATGGCCCGCCTGGCATTAT
GCCCAGTACATGACCTTATGGGACTTTCCTACTTGGCAGTACATCTACGTATTAGTCATC
GCTATTACCATGGTGATGCGGTTTTTGGCAGTACATCAATGGGCGTGATAGCGGTTTGAC
TCACGGGGATTTCGAAGTCTCCACCCCATTTGACGTCAATGGGAGTTGTTTTTGGCACCAA
AATCAACGGGACTTTCCAAAATGTCTGAACAACTCCGCCCATTTGACGCAATGGGCGGT
AGGCGGTGACGGTGGGAGGTCTATATAAGCAGAGCTCTCCCTATCAGTGATAGAGATCTC
CCTATCAGTGATAGAGATCGTCGACGAGCTCGTTTAGTGAACCGTCAGATCGCCTGGAGA
CGCCATCCACGCTGTTTTGACCTCCATAGAAGACACCGGACCGATCCAGCCTCCGGACC
ATGGCGTACTACCATCACCATCACCATCACACCGGTGATATCCTCGAGCCCATCAAGT
TTGTACAAAAAGCTGAACGAGAAACGTAAATGATATAAATATCAATATATTAAATTAG
ATTTTGCATAAAAAACAGACTACATAACTGTAAAAACAAACATATCCAGTCACTATGG
CGGCCGCTATTAGCACCCCGAGGCTTTACACTTTATGCTTCCGGCTCGTATAATGTGTGGA
TTTTGAGTTAGGATCCGGCGAGATTTTCAGGAGCTAAGGAAGCTAAATGGAGAAAAAA
TCACTGGATATACCAACGTTGATATATCCCAATGGCATCGTAAAGAACATTTTGAGGCAT
TTCAGTCAGTTGCTCAATGTACCTATAACGACCGTTCAGCTGGATATTACGGCCTTTT
TAAAGACCGTAAAGAAAAAAGCACAAGTTTTATCCGGCCTTTATTACATTCTTGCCC
GCCTGATGAATGCTCATCCGAATTCGGTATGGCAATGAAAGACGGTGAGCTGGTGATAT
GGGATAGTGTTTACCCTTGTACACCGTTTTCCATGAGCAAACTGAAACGTTTTTCATCGC
TCTGGAGTGAATACCACGACGATTTCCGGCAGTTTCTACACATATATTGCAAGATGTGG
CGTGTTACGGTGAAAAACCTGGCCTATTTCCCTAAAGGGTTTTATTGAGAATATGTTTTCG
TCTCAGCCAATCCCTGGGTGAGTTTACCAGTTTTGATTTAAACGTGGCCAATATGGACA
ACTTCTTCGCCCCCGTTTTCCCATGGGCAATATTATACGCAAGGCGACAAGGTGCTGA
TGCCGCTGGCGATTACAGTTTCACTCATGCCGTCTGTGATGGCTTCCATGTCCGGCAGAAATGC
TTAATGAATTACAACAGTACTGCGATGAGTGGCAGGGCGGGCGTAAACCGGTGGATCCG
GCTTACTAAAAGCCAGATAACAGTATGCGTATTTGCGCGCTGATTTTTGCGGTATAAGAA
TATATACTGATATGTATACCCGAAGTATGTCAAAAAGAGGTGTGCTATGAAGCAGCGTAT
TACAGTGACAGTTGACAGCGACAGCTATCAGTTGCTCAAGGCATATATGATGTCAATATC
TCCGGTCTGGTAAGCACAACCATGCAGATGAAGCCCGTCTGCTGCGTGCCGAACCGTGG
AAAGCGGAAAAATCAGGAAGGGATGGCTGAGGTGCGCCCGTTTTATTGAATGAACGGCTCT
TTGCTTGACGAGAACACGGGACTGGTGAATGCAGTTTAAAGTTTACACCTATAAAAGAGA
GAGCCGTTATCGTCTGTTTGTGGATGTACAGAGTGATATTATTGACACGCCCGGGCGACG
GATGGTGATCCCCCTGGCCAGTGACGCTGCTGTGTCAGATAAAGTCTCCCGTGAACCTTA
CCCGTGGTGCATATCGGGGATGAAAGCTGGCGCATGATGACCACCGATATGGCCAGTGT
GCCGCTCTCCGTTATCGGGGAAGAAGTGGCTGATCTCAGCCACCGCGAAAAATGACATCAA
AAACGCCATTAACTGATGTTCTGGGGAATATAAATGTGAGGCTCCGTTATACACAGCCA
GTCTGCAGGTCCACCATAGTGACTGGATATGTTGTGTTTTACAGTATTATGTAGTCTGTT
TTTTATGCAAAATCTAATTTAATATATTGATATTTATATCATTTTACGTTTTCTCGTTTACG
CTTTCTTGTACAAAGTGGTGATGGGCGGCCGCTCTAGAGGGCCCCAAGCTTACGCGTGAT
GCGACGTATAGCTCTCTCCCTATAGTGAGTCGTATTATAAGCTAGGCACTGGCCGTCGT
TTTACAACGTCGTGACTGGGAAAACTGCTAGCTTGGGATCTTTGTGAAGGAACCTTACTT
CTGTGGTGTGACATAATTGGACAACTACCTACAGAGATTTAAAGCTCTAAGGTAAATAT
AAAATTTTAAAGTGATAATGTGTTAACTAGCTGCATATGCTTGTGCTTGTGAGAGTTTT
GCTTACTGAGTATGATTTATGAAAAATATTATACACAGGAGCTAGTGATTCTAATTGTTTTG
TGTATTTTAGATTACAGTCCCAAGGCTCATTTCAGGCCCTCAGTCTCACAGTCTGTT
CATGATCATAATCAGCCATACCATTTGTAGAGGTTTTACTTGCTTTAAAAAACCTCCC
ACACCTCCCCCTGAACCTGAAACATAAAATGAATGCAATTGTTGTTGTTAACTTGTTTAT
TGCAGCTTATAATGGTTACAAATAAAGCAATAGCATCACAAATTTACAAATAAAGCATT
TTTTTCACTGCATTCTAGTTGTGTTTGTCCAACTCATCAATGTATCTTATCATGTCTG
GATCGATCCTGCATTAAATGAATCGGCCAACGCGCGGGGAGAGGCGGTTTTGCGTATTGGCT
GGCGTAATAGCGAAGAGGCCCGCACCGATCGCCCTTCCCAACAGTTGCGCAGCCCTGAATG
GCGAATGGGACGCGCCCTGTAGCGGCGCATTAAGCGCGCGGGGTGTGGTGGTTACGCGCA
GCGTGACCGCTACACTTGCCAGCGCCCTAGCGCCCGCTCCTTTTCGCTTTCTTCCCTTCCT
TTCTCGCCACGTTCCCGGCTTTCCCGTCAAGCTCTAAATCGGGGCTCCCTTTAGGGT-

Figure 91B

TCCGATTAGTGCTTTACGGCACCTCGACCCCAAAAACTTGATTAGGGTGATGGTTAC
GTAGTGGGCCATCGCCCTGATAGACGGTTTTTCGCCCTTTGACGTTGGAGTCCACGTTCT
TTAATAGTGGACTCTTGTTCCAACTGGAACAACACTCAACCCTATCTCGGTCTATTCTT
TTGATTATAAGGGATTTTCCCGATTTCGGCCTATTGGTTAAAAATGAGCTGATTTAAC
AAATATTTAACGGAATTTTAAACAAATATTAACGTTTACAATTTTCGCTGATGCGGTAT
TTTCTCCTTACGCATCTGTGCGGTATTTACACCCGATACGCGGATCTGCGCAGCACCAT
GGCCTGAAATAACCTCTGAAAGAGGAACCTGGTTAGGTACCTTCTGAGGCGGAAGAACC
AGCTGTGGAATGTGTGTCAGTTAGGGTGTGGAAGTCCCCAGGCTCCCCAGCAGGCAGAA
GTATGCAAAGCATGCATCTCAATTAGTCAGCAACCAGGTGTGGAAGTCCCCAGGCTCCC
CAGCAGGCAGAAGTATGCAAAGCATGCATCTCAATTAGTCAGCAACCATAGTCCCGCCCC
TAACTCCGCCCATCCCGCCCCTAACTCCGCCAGTTCCGCCCATTTCTCCGCCCATGGCT
GACTAATTTTTTTTATTTATGTCAGAGGCGAGGCGCCTCGGCCTCTGAGCTATTCCAGA
AGTAGTGAGGAGGCTTTTTTGGAGGCTAGGCTTTTGCAAAAGCTTGATTCTTCTGACA
CAACAGTCTCGAACTTAAGACCATGGCCAGCCTTTGTCTCAAGAAGATCCACCCTCAT
TGAAAGAGCAACGGCTACAATCAACAGCATCCCCATCTCTGAAGACTACAGCGTCGCCAG
CGCAGCTCTCTAGCGACGGCCGCATCTTCACTGGTGTCAATGTATATCATTTTACTGG
GGGACCTTGTGCAGAACTCGTGGTGTGGGCACTGTCTGTCTGCGGCAGCTGGCAACCT
GACTTGTATCGTCGCGATCGGAAATGAGAACAGGGGCATCTTGAGCCCTGCGGACGGTG
CCGACAGGTGCTTCTCGATCTGCATCTCTGGGATCAAGCCATAGTGAAGSACAGTGATGG
ACAGCCGACGGCAGTTGGGATTCTGAATGTCTGCCCTCTGGTTATGTGTGGGAGGGCTA
AGCACTTCTGGCCGAGTTTCAAAATGACCGACCAAGCGACGCCAACCTGCCATCACGAT
GGCCGCAATAAAATATCTTTATTTTCATTACATCTGTGTGTGGTTTTTTGTGTGAATCG
ATAGCGATAAGGATCCGCGTATGGTGCATCTCAGTACAATCTGCTCTGATGCCGCATAG
TTAAGCCAGCCCCGACACCCGCCAACACCCGCTGACGCGCCCTGACGGGCTTGTCTGCTC
CCGCGCATCCGCTTACAGACAAGCTGTGACCGTCTCCGGGAGCTGCATGTGTGTCAGAGTTT
TCACCGTCATCACCGAAACGCGGAGACGAAAGGGCCTCGTGATACGCCTATTTTATAG
GTTAATGTCTATGATAAATAATGGTTTTCTTAGACGTCAGGTGGCACTTTTCCGGGAAATGTG
CGCGGAACCCCTATTTGTTTTATTTTTCTAAATACATTCAAATATGTATCCGCTCATGAGA
CAATAACCCCTGATAAATGCTTCAATAATATTGAAAAAGGAAGATATGAGTATCAACAT
TTCCGTGTGCGCCCTATTCCCTTTTTTGCGGCATTTTGCTTCTGTTTTGTCTACCCA
GAAACGCTGGTGAAGTAAAGATGCTGAAGATCAGTTGGGTGCAGGATGGGTTACATC
GAACCTGGATCTCAACAGCGGTAAAGATCCTTGAGAGTTTTTCGCCCGAAGAAGCTTTTCCA
ATGATGAGCACTTTTAAAGTTCTGCTATGTGGCGCGGTATATCCCGTATTGACGCGGGG
CAAGAGCAACTCGGTGCGCGCATACATATTCTCAGAATGACTTGGTTGAGTACTACCA
GTCACAGAAAAGCATCTTACGGATGGCATGACAGTAAGAGAATTATGCAGTGCTGCCATA
ACCATGAGTGATAACACTGCGGCCAATTACTTCTGACAACGATCGGAGGACCGAAGGAG
CTAACCCTTTTTTGCAACAATGGGGGATCATGTAACCTCGCTTGATCGTTGGGAACCG
GAGCTGAATGAAGCCATACCAACGACGAGCGTGACACCAGATGCCTGTAGCAATGGCA
ACAACGTTGCGCAACTATTAACCTGGCGAACTACTTACTCTAGCTTCCCGGCAACAATTA
ATAGACTGGATGGAGCGGATAAAGTTGAGGACCACTTCTGCGCTCGGCCCTTCCGGCT
GGCTGGTTTTATTGCTGATAAATCTGGAGCCGCTGAGCGTGGGTCTCGCGGTATCATTGCA
GCACTGGGGCCAGATGGTAAGCCCTCCCGTATCGTAGTTATCTACACGACGGGAGTCAG
GCAACTATGGATGAACGAAATAGACAGATCGCTGAGATAGGTGCCTCACTGATTAAAGCAT
TGGTAACCTGTGACCAAGTTTACTCATATATACTTTAGATTGATTTAAAACCTCATTTT
TAATTTAAAAGGATCTAGGTGAAGATCCTTTTTGATAATCTCATGACCAAAATCCCTTAA
CGTGAGTTTTTCTGTTCCACTGAGCGTCAGACCCGCTAGAAAAGATCAAAGGATCTTCTTGA
GATCCTTTTTTCTGCGCGTAATCTGCTGCTTGCAAAACAAAAAACACCGCTACACGCG
GTGGTTTTGTTGCGGATCAAGAGCTACCAACTCTTTTTCCGAAGGTAAGTGGCTTCAAG
AGAGCGCAGATACCAAAATCTGTCTTCTAGTGTAGCCGTAGTTAGGCCACCACTTCAAG
AACTCTGTAGCACCGCTACATACCTCGCTCTGCTAATCTGTACAGTGGCTGCTGCC
AGTGGCGATAAGTCTGTCTTACCGGTTGGACTCAAGACGATAGTTACCGGATAAGGCG
CAGCGGTGCGGCTGAACGGGGGTTGCTGCACACAGCCAGCTTGGAGCGAACGACCTAC
ACCGAACTGAGATACCTACAGCGTGAGCATTGAGAAAGCGCCACGCTTCCGAAGGGAGA
AAGGCGGACAGGTATCCGTAAGCGGCAGGTCGGAACAGGAGAGCGACGAGGAGCTT
CCAGGGGAAAACGCTGCTATCTTTATAGTCTGTGCGGTTTTCGCCACCTCTGACTTGAG
CGTCGATTTTTGTGATGCTCGTCAGGGGGCGGAGCCTATGGA AAAACGCCAGCAACGCG
GCCTTTTTACGTTCTGCGCTTTTGCTGGCCTTTTGCTCACATGTTCTTTCTGCGTTA
TCCCTGATTCTGTGATAACCGTATTACCGCCTTTGAGTGAGCTGATACCGCTCGCCGC-

FIGURE 91C

AGCCGAACGACCGAGCGCAGCGAGTCAGTGAGCGAGGAAGCGGAAGAGCGCCCAATACGC
AAACCGCCTCTCCCCGCGCGTTGGCCGATTCATTAATGCAGAGCTTGCAATTCGCGCGTT
TTTCAATATTATTGAAGCATTATCAGGGTTATTGTCTCATGAGCGGATACATATTTGAA
TGTATTTAGAAAAATAAACAAATAGGGGTTCCGCGCACATTTCCCGAAAAGTGCCACCT
GACGTCTAAGAAACCATTATTATCATGACATTAACTATAAAAATAGGCGTAGTACGAGG
CCCTTTCATCATTAG

FIGURE 91D

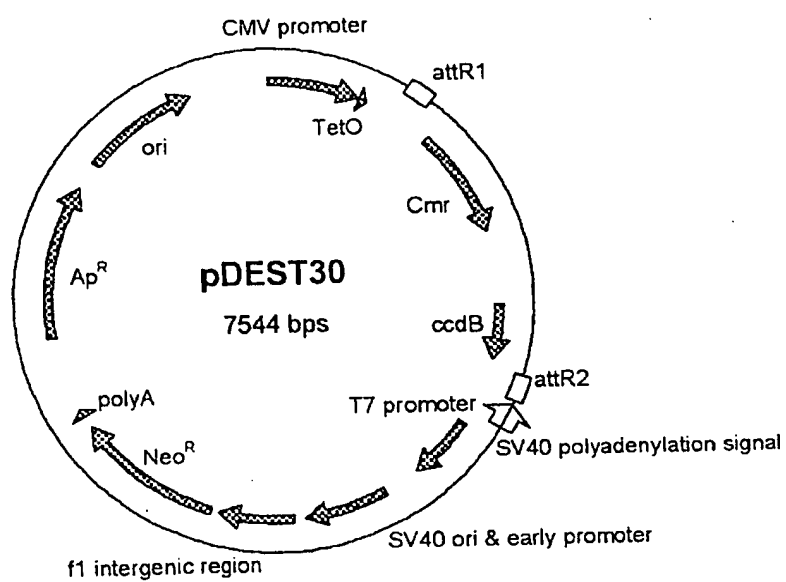


FIGURE 92A

pDEST30 7544 bp

ATGCATGTCGTTACATAAATTACGGTAAATGGCCCGCTGGCTGACCGCCCAACGACCCC
CGCCCATTGACGTCAATAATGACGTATGTTCCCATAGTAACGCCAATAGGGACTTTCCAT
TGACGTCAATGGGTGGAGTATTTACGGTAAACTGCCCACTTGGCAGTACATCAAGTGTAT
CATATGCCAAGTACGCCCCCTATTGACGTCAATGACGGTAAATGGCCCGCTGGCATTAT
GCCCAGTACATGACCTTATGGGACTTTCCTACTTGGCAGTACATCTACGTATTAGTCATC
GCTATTACCATGGTGTATGCGGTTTTGGCAGTACATCAATGGGCGTGGATAGCGGTTTTGAC
TCACGGGGATTTCCAAGTCTCCACCCCATTGACGTCAATGGGAGTTTGTTTTGGCACCAA
AATCAACGGGACTTTCCAAAATGTCTAACAACCTCCGCCCATTGACGCAAATGGGCGGT
AGGCGTGTACGGTGGGAGGTCTATATAAGCAGAGCTCTCCCTATCAGTGATAGAGATCTC
CCTATCAGTGATAGAGATCGTCGACGAGCTCGTTTAGTGAACCGTCAGATCGCCTGGAGA
CGCCATCCACGCTGTTTTGACCTCCATAGAAGACACCGGACCGATCCAGCCTCCGGACT
CTAGAGGATCCCTACCGGTGATATCCTCGAGCCCATCAACAAGTTTGTACAAAAAAGCTG
AACGAGAAACGTAAAATGATATAAATATCAATATATTAAATTAGATTTTGCATAAAAAAC
AGACTACATAAATACTGTAAAAACACAATATCCAGTCACTATGGCGGCCGATTAGGCAC
CCCAGGCTTTTACACTTTATGCTTCCGGCTCGTATAATGTGTGGATTTTGTAGTTAGGATCC
GGCGAGATTTTTCAGGAGCTAAGGAAGCTAAAATGGAGAAAAAATCACTGGATATACCAC
CGTTGTATATATCCCAATGGCATCGTAAAGAACATTTTGGAGCATTTCAGTCAGTTGCTCA
ATGTACCTATAACAGACCGTTTCAGCTGGATATTACGGCCTTTTAAAGACCGTAAAGAA
AAATAAGCACAAAGTTTTATCCGGCCTTTTATCACATTCTTGCCCGCCTGATGAATGCTCA
TCCGGAATTCGGTATGGCAATGAAAGACGGTGAGCTGGTGTATAGGGATAGTGTTCACCC
TTGTTACACCGTTTTTCCATGAGCAAACTGAAACGTTTTCATCGCTCTGGAGTGAATACCA
CGACGATTTCCGGCAGTTTCTACACATATATTTCGAAGATGTGGCGTGTACGGTGAATAA
CCTGGCCTATTTCCCTAAAGGGTTTATTGAGAATATGTTTTTCGTCTCAGCCAATCCCTG
GGTGAGTTTACACAGTTTTGATTTAAACGTGGCCAAATATGGACAACCTTCTCGCCCCGT
TTTACCATGGGCAAAATATTATACGCAAGGCGACAAGGTGCTGATGCCGCTGGCGATTCA
GGTTCATCATGCCGCTGTGTATGGCTTCCATGTCCGCAGAAATGCTTAATGAATTACAACA
GTACTGCGATGAGTGGCAGGGCGGGCGTAAAGATCTGGATCCGCTTACTAAAAGCCAG
ATAACAGTATGCGTATTTGCGCGCTGATTTTTCGGGTATAAGAATATATACTGATATGTA
TACCCGAAGTATGTCAAAAAGAGGTGTGCTATGAAGCAGCGTATTACAGTGACAGTTGAC
AGCGACAGCTATCAGTTGCTCAAGGCATATATGATGTCAATATCTCCGGTCTGGTAAGCA
CAACCATGCAGAAATGAAGCCCGTCTGCGTGGCGAACGCTGGAAAGCGGAAATCAGG
AAGGATGGCTGAGGTGCGCCCGTTTTATTGAAATGAACGGCTCTTTGCTGACGAGAACCA
GGGACTGGTGAAATGCAGTTTAAAGTTTACACCTATAAAGAGAGAGCCGTTATCGTCTG
TTTGTGGATGTACAGAGTGATATTATTGACACGCCCCGGCGACGGATGGTGTATCCCCCTG
GCCAGTGCACGCTGCTGTGTCAGATAAAGTCTCCCGTGAACTTTACCCGGTGGTGATATC
GGGGATGAAAGCTGGCGCATGATGACCACCGATATGGCCAGTGTGCCGGTCTCCGTTATC
GGGGAAGAAGTGGCTGATCTCAGCCACCGCAAAATGACATCAAAAACGCCATTAACTG
ATGTTCTGGGGAATATAAATGTGAGGCTCCCTTATACACAGCCAGTCTGCAGGTCGACCA
TAGTGACTGGATATGTTGTGTTTTACAGTATTATGTAGTCTGTTTTTATGCAAAATCTA
ATTTAATATATTGATATTTATATCATTTTACGTTTCTCGTTCAGCTTTCTGTACAAAGT
GGTTGATGGGCGGCGCTCTAGAGGGCCCAAGCTTACGCGTGCATGCGACGTCATAGCTC
TCTCCCTATAGTGAGTCGTATTATAAGCTAGGCACTGGCCGTCGTTTTTACAACGTCGTGA
CTGGGAAAACCTGCTAGCTTGGGATCTTTGTGAAGGAACCTTACTTCTGTGGTGTGACATA
ATTGGACAAAACCTACAGAGATTAAAGCTCTAAGGTAAATATAAAATTTTTAAGTGT
ATAATGTGTTAAACTAGCTGCATATGCTTGTGCTTGTGAGAGTTTGTCTTACTGAGTATGA
TTTATGAAAATATTATACACAGGAGCTAGTGATTCTAATTGTTTGTGATTTTATAGATTCA
CAGTCCCAAGGCTCATTTAGGCCCCCTCAGTCCTCACAGTCTGTTCATGATCATAATCAG
CCATACCACATTTGTAGAGGTTTTACTTGTCTTAAAAAACCTCCACACCTCCCCCTGAA
CCTGAAACATAAAATGAATGCAATTGTTGTTTAACTGTTTTATTGCAGCTTATAATGG
TTACAAATAAAGCAATAGCATCACAAATTTACAAATAAAGCATTTTTTTCACTGCATT
TAGTTGTGGTTTGTCCAACTCATCAATGTATCTTATCATGTCTGGATCGATCCTGCATT
AATGAATCGGCCAACCGCGGGGAGAGCGGTTTGCATATGGCTGGCGTAATAGCGAAG
AGGCCCGCACCGATCGCCCTTCCCAACAGTTGCGCAGCTGAATGGCGAATGGGACGCGC
CCTGTAGCGGCGATTAAAGCGCGGCGGTGTGGTGGTTACGCGCAGCGTGACCGCTACAC
TTGCCAGCGCCCTAGCGCCGCTCCTTTTCGCTTTCTTCCCTTCTTCCGCAAGTTTCCG
CCGCTTTCCCGTCAAGCTCTAAATCGGGGGCTCCCTTTAGGGTTCCGATTTAGTGCTT-

Figure 928

211/260

TACGGCACCTCGACCCCAAAAACTTGATTAGGGTGATGGTTACGCTAGTGGCCATCGC
CCTGATAGACGGTTTTTCGCCCTTTGACGTTGGAGTCCACGTTCTTTAATAGTGGACTCT
TGTTCCAAACTGGAACAACACTCAACCCTATCTCGGTCTATTCTTTTGATTATAAGGGA
TTTTGCCGATTTCGGCCTATTGGTTAAAAAATGAGCTGATTTAACAAATATTTAACGCGA
ATTTTAACAAATATTAACGTTTACAATTTGCCTGATGCGGTATTTCTCCTTACGCAT
CTGTGCGGTATTTACACCCGCATACGCGGATCTGCGCAGCACCATGGCCTGAAATAACCT
CTGAAAGAGGAACTTGGTTAGGTACCTTCTGAGGCGGAAAGAACCAGCTGTGGAATGTGT
GTCAGTTAGGGTGTGGAAGTCCCCAGGCTCCCCAGCAGGCAGAAGTATGCAAAGCATGC
ATCTCAATTAGTCAGCAACCAGGTGTGGAAGTCCCCAGGCTCCCCAGCAGGCAGAAGTA
TGCAAAGCATGCATCTCAATTAGTCAGCAACCATAGTCCGCCCTTAACCTCCGCCCATCC
CGCCCCCTAATCCGCCCAGTTCCGCCCATTTCTCGCCCCATGGCTGACTAATTTTTTTA
TTTATGACAGGCGGAGGCGCGCTCGGCCCTTGAGCTATTCCAGAAGTAGTGAGGAGGT
TTTTTGGAGGCTTAGGCTTTTGCAAAAAGCTTGATTCTTCTGACACAACAGTCTCGAACT
TAAGGCTAGAGCCACCATGATTGAACAAGATGGATTGCACGCAGGTTCTCCGGCCGCTTG
GGTGGAGAGGCTATTCCGCTATGACTGGGCACAACAGACAATCGGCTGCTCTGATGCCG
CGTGTTCGGCTGTGACGCGAGGGGCGCCGCTTTTGTCAAGACCGACCTGTCCGG
TGCCCTGAATGAACTGCAGGACGAGGCGAGCGCGCTATCGTGGCTGGCCACGACGGGCGT
TCCTTGCGCAGCTGTGCTCGACGTTGTCACTGAAGCGGAAGGACTGGCTGCTATTGGG
CGAAGTGCCGGGCGAGGATCTCTGTGCTCATCTCACCTTGCTGCTGCGGAGAAAGTATCCAT
CATGGCTGATGCAATGCCGCGGCTGCATACGCTTGATCCGCTACCTGCCCATTCGACCA
CCAAGCGAAACATCGCATCGAGCGAGCAGTACTCGGATGGAAGCCGCTCTTGTGCTCA
GGATGATCTGGACGAAGCATCAGGGGCTCGCGCCAGCCGAAGTGTGCGCAGGCTCAA
GGCGGCATGCCGACGGCGAGGATCTCGTCTGACCCATGGCGATGCTGCTTGGCGAA
TATCATGGTGGAAATGGCCGCTTTCTGGATTCTGACGCTGTGGCCGGCTGGGTGTGGC
GGACCGCTATCAGGACATAGCGTTGGCTACCGTGATATTGCTGAAGAGCTTGGCGGCGA
ATGGGCTGACCGCTTCTCGTGTCTTACCGTATCGCGCTCCCGATTGCGAGCGCATCGC
CTTCTATCGCCTTCTTGACGAGTTCTTCTGAGCGGACTCTGGGTTTGAATGACCGAC
CAAGCGACGCCCAACCTGCCATCAGATGGCCGCAATAAAATATCTTTATTTTCAATACA
TCTGTGTGTGGTTTGTGTGAATCGATAGCGATAAGGATCCGCGTATGGTGCCTCT
CAGTACAATCTGCTCTGATGCCGCATAGTTAAGCCAGCCCGACACCCGCCAACCCCGC
TGACCGGCCCTGACGGGCTTGTCTGCTCCCGCATCCGCTTACAGACAAGCTGTGACCGT
CTCCGGGAGCTGCATGTGTGAGGTTTTTACCCTCATCACCGAAACGCGGAGACGAAA
GGGCTCGTATACGCTATTTTATAGGTTAATGTGATGATAAATGTTTCTTAGAC
GTCAGGTGGCACTTTTCGGGGAATGTGCGCGGAACCCCTATTGTTTATTTTCTAAAT
ACATTCAAATATGTATCCGCTCATGAGACAATAACCTGATAAATGCTTCAATAATATG
AAAAAGGAAGAGTATGAGTATTCAACATTTCGCTGTGCGCCTTATTCCTTTTTCGGGC
ATTTTGCCTTCTGTTTTTGTCTCAGCCAGAAACGCTGGTGAAGTAAAGATGCTGAAGA
TCAGTTGGGTGCACGAGTGGGTTACATCGAAGTGGATCTCAACAGCGGTAAGATCCTTGA
GAGTTTTTCGCCCCGAAGAACGTTTTTCCAATGATGAGCACTTTTAAAGTTCTGCTATGTGG
CGCGGTATTATCCGTTATGACGCGGGCAAGAGCAACTCGGTGCGCGCATACACTATTC
TCAGAATGACTTGGTTGAGTACTCACCAGTACAGAAAGCATCTTACGGATGGCATGAC
AGTAAGAGAATTATGCAGTGCTGCCATAACCATGAGTGATAACACTGCGGCCAAGTTACT
CTGACAACGATCGGAGGACCGAAGGAGCTAACCGCTTTTTCACAACATGGGGATCA
TGTAACCTCGCCTTGATCGTTGGGAACCGGAGCTGAATGAAGCCATACCAAACGACGAGCG
TGACACCAGATGCCTGTAGCAATGGCAACAACGTTGCGCAAACTATTAAGTGGCAACT
ACTTACTCTAGCTTCCCGGCAACAATTAATAGACTGGATGGAGGCGGATAAAGTGCAGG
ACCCTTCTGCGCTCGGCCCTTCCGGCTGGCTGGTTTATGCTGATAAATCTGAGCCGG
TGAGCGTGGGTCTCGCGGTATCATTCAGCACTGGGGCCAGATGGTAAGCCCTCCCGTAT
CGTAGTTATCTACAGACGGGGAGTCAGGCAACTATGGATGAACGAAATAGACAGATCGC
TGAGATAGGTGCTCACTGATTAGCATTTGGTAACTGTGACACCAAGTTTACTCATATAT
ACTTTAGATTGATTTAAACTTCTATTTTAAATTTAAAGGATCTAGGTGAAGATCCTTTT
TGATAATCTCATGACCAAAATCCCTTAACGTGAGTTTTTTCGTTCCACTGAGCGTCAGACCC
CGTAGAAAAGATCAAAGGATCTTCTGAGATCCTTTTTTCTGCGCGTAATCTGCTGCTT
GCAACAAAAAACACCGCTACCAGCGGTGGTTTGTGTTGCGGGATCAAGAGCTACCAAC
TCTTTTTCCGAAGGTAAGTGGCTTTCAGCAGAGCGAGATACCAATACTGTCTCTTAGT
GTAGCCGTAGTTAGGCCACCACTTCAAGAACTCTGTAGACCGCTACATACCTCGCTCT
GCTAATCTGTTACAGTGGCTGCTGCCAGTGGCGATAAGTCTGTCTTACCGGGTTGGA
CTCAAGACGATAGTTACCGGATAAGGCGCAGCGTGGGCTGAACGGGGGTTCTGTGCAC

FIGURE 92C

ACAGCCCAGCTTGGAGCGAACGACCTACACCGAACTGAGATACCTACAGCGTGAGCATTG
AGAAAGCGCCACGCTTCCCGAAGGGAGAAAGGCGGACAGGTATCCGGTAAGCGGCAGGGT
CGGAACAGGAGAGCGCACGAGGGAGCTTCCAGGGGGAAACGCCTGGTATCTTTATAGTCC
TGTCGGGTTTCGCCACCTCTGACTTGAGCGTCGATTTTTGTGATGCTCGTCAGGGGGGCG
GAGCCTATGGAAAAACGCCAGCAACGCGGCCTTTTACGGTTCCTGGCCTTTTGCTGGCC
TTTTGCTCACATGTTCTTTCCTGCGTTATCCCCTGATTCTGTGGATAACCGTATTACCGC
CTTTGAGTGAGCTGATACCGCTCGCCGCAGCCGAACGACCGAGCGCAGCGAGTCAGTGAG
CGAGGAAGCGGAAGAGCGCCCAATACGCAAACCGCCTCTCCCGCGCGTTGGCCGATTCA
TTAATGCAGAGCTTGCAATTCGCGCGTTTTTCAATATTATTGAAGCATTATCAGGGTTA
TTGTCTCATGAGCGGATACATATTTGAATGTATTTAGAAAAATAACAAATAGGGGTTCC
GCGCACATTTCCCGAAAAGTGCCACCTGACGTCTAAGAAACCATTATTATCATGACATT
AACCTATAAAAAATAGGCGTAGTACGAGGCCCTTCACTCATTAG

FIGURE 92D

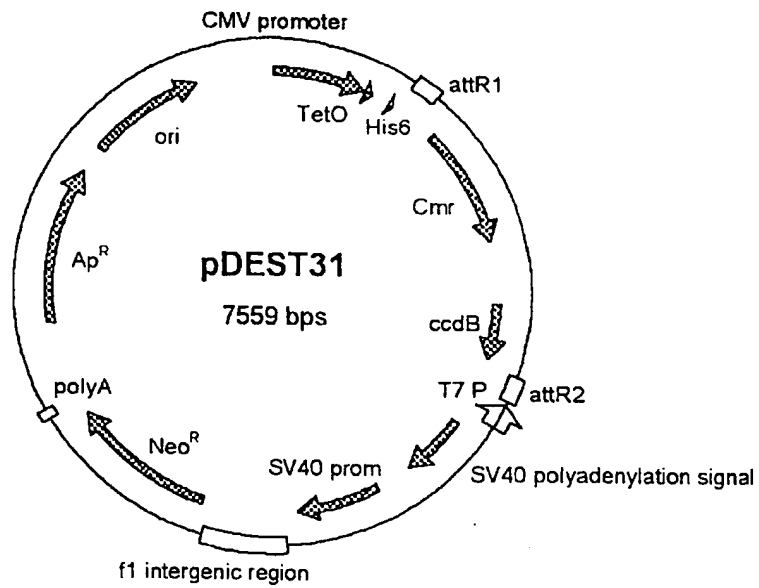


FIGURE 93A

pDEST31 7559 bp

ATGCATGTCGTTACATAACTTACGGTAAATGGCCCGCCTGGCTGACCGCCCAACGACCCC
CGCCCATTTGACGTCAATAATGACGTATGTTCCCATAGTAACGCCAATAGGGACTTTCCAT
TGACGTCAATGGGTGGAGTATTTACGGTAAACTGCCCACTTGGCAGTACATCAAGTGTAT
CATATGCCAAGTACGCCCCCTATTGACGTCAATGACGGTAAATGGCCCGCCTGGCATTAT
GCCAGTACATGACCTTATGGGACTTTCCTACTTGGCAGTACATCTACGTATTAGTCATC
GCTATTACCATGGTGATGCGGTTTTGGCAGTACATCAATGGGCGTGGATAGCGGTTTTGAC
TCACGGGGATTTCCAAGTCTCCACCCATTGACGTCAATGGGAGTTTGTGGTGGACCAA
AATCAACGGGACTTTCCAAAATGTCGTAACAACCTCGCCCATTTGACGCAAAATGGGCGGT
AGGCGTGTACGGTGGGAGGCTCTATATAAGCAGAGCTCTCCCTATCAGTGATAGAGATCTC
CCTATCAGTGATAGAGATCGTCGACGAGCTCGTTTAGTGAACCGTCAGATCGCCTGGAGA
CGCCATCCACGCTGTTTGGACCTCCATAGAAGACACCGGGACCGATCCAGCCTCCGGACC
ATGGCGTACTACCATCACCATCACCATCACACCGGTGATATCCTCGAGCCCATCACAAGT
TTGTACAAAAAGCTGAACGAGAAAACGTAAAATGATATAAAATATCAATATATTAAATTAG
ATTTTGCATAAAAAACAGACTACATAAATCTGTAACACACATATCCAGTCACTATGG
CGGCCGCTTAGGCACCCAGGCTTTACACTTTATGCTTCCGGCTCGTATAATGTGTGGA
TTTTAGTTAGGATCCGGCGAGATTTTCAGGAGCTAAGGAAGCTAAAATGGAGAAAAAAA
TCAGTGGATATACCACCGTTGATATATCCCAATGGCATCGTAAAGAACATTTTGAGGCAT
TTCAGTCAGTTGCTCAATGTACCTATAACAGACCGTTTTCAGCTGGATATTACGGCCCTTT
TAAAGACCGTAAAGAAAAATAAGCAACAAGTTTATCCGGCCTTTATTACATTCTTGCCC
GCCTGATGAATGCTCATCCGGAATTCCGTATGGCAATGAAAGACGGTGAGCTGGTGATAT
GGGATAGTGTTCACCCCTGTTACACCGTTTTCCATGAGCAAACCTGAAACGTTTTCATCGC
TCTGGAGTGAATACCACGACGATTTCCGGCAGTTTCTACACATATATTGCAAGATGTGG
CGTGTTACGGTGAAAACCTGGCCTATTTCCCTAAAGGGTTTATTGAGAATATGTTTTTCG
TCTCAGCCAATCCCTGGGTGAGTTTTCACAGTTTGTATTTAAACGTGGCCAATATGGACA
ACTTCTTCGCCCCCGTTTTACCATGGGCAAAATATTATACGCAAGGCGACAAGGTGCTGA
TGCCGCTGGCGATTTCAGGTTTCATCATGCCGTCTGTGATGGCTTCCATGTCCGCGAATGC
TTAATGAATTACACAGTACTGCGATGAGTGGCAGGGCGGGCGTAAACGCGTGGATCCG
GCTTACTAAAAGCCAGATAACAGTATGCGTATTTGCGCGCTGATTTTGGCGGTATAAGAA
TATATACTGATATGTATACCCGAAGTATGTCAAAAAGAGGTGTGCTATGAAGCAGCGTAT
TACAGTGACAGTTGACAGCGCAGCTATCAGTTGCTCAAGGCATATATGATGTCAATATC
TCCGGTCTGGTAAGCACAACTGCAGATGAAGCCCGTCTGCTGCGTGCCGAACGCTGG
AAAGCGGAAAATCAGGAAGGGATGGCTGAGGTGCGCCGTTTATTGAAATGAACGGCTCT
TTTGCTGACGAGAACAGGGACTGGTGAATGCAGTTTAAAGTTTACACCTATAAAGAGA
GAGCCGTTATCGTCTGTTTGTGGATGTACAGAGTGATATTATGACACGCCCCGGCGACG
GATGGTGATCCCCCTGGCCAGTGACAGTCTGCTGTGATGAAGTCTCCCGTGAACCTTTA
CCCCGTGGTGATATCGGGGATGAAAGCTGGCGCATGATGACCACCGATATGGCCAGTGT
GCCCGTCTCCGTTATCGGGGAAGAAGTGGCTGATCTCAGCCACCGCGAAAATGACATCAA
AAACGCCATTAACTGATGTTCTGGGGAATATAAATGTGAGGCTCCGTTATACACAGCCA
GTCTGCAGGTGACCATAGTGACTGGATATGTTGTGTTTACAGTATTATGTAGTCTGTT
TTTTATGCAAAATCTAATTTAATATATTGATATTTATATCATTTACGTTTCTCGTTCAG
CTTTCTTGTAAGAGTGGTGATGGGCGGCCGCTCTAGAGGGCCCAAGCTTACGCGTGAT
GCGACGTCTAGCTCTCTCCCTATAGTGAGTCGTATTATAAGCTAGGCACTGGCCGTCGT
TTTACAACGTCGTGACTGGGAAAACCTGCTAGCTTGGGATCTTTGTGAAGGAACCTTACTT
CTGTGGTGACATAAATGGACAACTACCTACAGAGATTTAAAGCTCTAAGGTAATAT
AAAAATTTTAAAGTGATATATGTGTTAACTAGCTGCATATGCTTGCTGCTTGAGAGTTT
GCTTACTGAGTATGATTTATGAAAATATTATACACAGGAGCTAGTGATTCTAATTGTTT
TGATTTTATAGATTACAGTCCCAAGGCTCATTTACAGGCCCTCAGTCTCACAGTCTGTT
CATGATCATAATCAGCCATACCACATTTGTAGAGGTTTTACTTGCTTTAAAAACCTCCC
ACACCTCCCCCTGAACCTGAAACATAAAATGAATGCAATTGTTGTTGTTAACTGTTTAT
TGCAGCTTATAATGGTTACAAATAAAGCAATAGCATCACAAATTTACAAATAAAGCATT
TTTTTCACTGCATTCTAGTTGTGGTTTTGTCAAACTCATCAATGTATCTTATCATGTCTG
GATCGATCCTGCATTAATGAATCGGCCAACGCGGGGAGAGGCGGTTTGGCTATTGGCT
GGCGTAATAGCGAAGAGGCGCGCACCGATCGCCCTTCCCAACAGTTGCGCAGCCTGAATG
GCGAATGGGACGCGCCTGTAGCGGCGCATTAGCGCGGCGGTTGGTGGTTACGCGCA
GCGTGACCGCTACACTTCCAGCGCCCTAGCGCCGCTCCTTTCGCTTCTTCCCTTCT
TTCTCGCCACGTTCCGCGGCTTTCCCGCTCAAGCTCTAAATCGGGGGCTCCCTTTAGGGT

FIGURE 93B

TCCGATTTAGTGCTTTACGGCACCTCGACCCCAAAAACTTGATTAGGGTGATGGTTTAC
GTAGTGGGCCATCGCCCTGATAGACGGTTTTTCGCCCTTTGACGTTGGAGTCCACGTTCT
TTAATAGTGGACTCTTGTTCCAACTGGAACAACACTCAACCCATCTCGGTCTATTCTT
TTGATTTATAAGGGATTTTGCCGATTTTCGGCCTATTGGTTAAAAAATGAGCTGATTTAAC
AAATATTTAACGCGAATTTTAACAAAATATTAACGTTTACAATTTGCGCTGATGCGGTAT
TTTCTCCTTACGCATCTGTGCGGTATTTACACCGCATACGCGGATCTGCGCAGCACCAT
GGCCTGAAATAAACCCTGAAAGAGGAACCTGGTTAGGTACCTTCTGAGGCGGAAAGAACC
AGCTGTGGAATGTGTGTGAGTTAGGGTGTGGAAGTCCCCAGGCTCCCCAGCAGGCAGAA
GTATGCAAAGCATGCACTCAATTAGTCAGCAACCAGGTGTGGAAGTCCCCAGGCTCCC
CAGCAGGCAGAAGTATGCAAAGCATGCACTCAATTAGTCAGCAACCATAGTCCCGCCCC
TAACTCCGCCCCATCCCCCCCCCTAACTCCGCCCCAGTTCCGCCCCATTCTCCGCCCCATGGCT
GACTAATTTTATTTATTTATGTCAGAGGCCGAGGCCCTCGGCTCTGAGCTATTCCAGA
AGTAGTGAGGAGGCTTTTTGGAGGCTAGGCTTTTGCAAAAAGCTTGATTCTTCTGACA
CAACAGTCTCGAATTTAAGGCTAGAGCCACCATGATTGAACAAGATGGATTGCACGCAGG
TTCTCCGGCCGCTTGGGTGGAGAGGCTATTGCGCTATGACTGGGCACAACAGACAATCGG
CTGCTCTGATGCGCCGCTGTTCCGGCTGTGAGCGCAGGGGCGCCCGGTTCTTTTTGTCAA
GACCGACCTGTCCGGTGCCCTGAATGAACTGCAGGACGAGGCAGCGCGGCTATCGTGGCT
GGCCACGACGGGCGTTCCTTGCGCAGCTGTGCTCGACGTTGTCACTGAAGCGGGAAGGGA
CTGGCTGCTATTGGGCGAAGTGCCGGGCGAGGATCTCCTGTCACTCACCTTGCTCCTGC
CGAGAAAGTATCCATCATGGCTGATGCAATGCGGCGGCTGCATACGCTTGATCCGGCTAC
TGCCCCATTTCGACCACCAAGCGAAACATCGCATCGAGCGAGCAGTACTCGGATGGAAGC
CGGTCTTTGTCGATCAGGATGATCTGGACGAAGAGCATCAGGGGCTCGCGCCAGCCGAAC
GTTCCGCCAGGCTCAAGGCGCGCATGCCGACGGCGAGGATCTCGTCTGACCCATGGCGA
TGCGCTGCTTGCCGAATATCATGGTGGAAAATGGCCGCTTTCTGGATTTCATCGACTGTGG
CCGGCTGGGTGTGGCGGACCGCTATCAGGACATAGCGTTGGCTACCCGTGATATTGCTGA
AGAGCTTGGCGGCGAATGGGCTGACCGCTTCTCGTGTCTTACGGTATCGCCGCTCCCGA
TTCGCGAGCGCATCGCCTTCTATCGCCTTCTTGACGAGTCTTCTGAGCGGGACTCTGGGG
TTGAAATGACCGACCAAGCGACGCCCAACCTGCCATCAGATGGCCGCAATAAAATATC
TTTTATTTTCAATTACATCTGTGTGTTGTTTTTGTGTGAATCGATAGCGATAAGGATCCG
CGTATGGTGCACTCTCAGTACAATCTGCTCTGATGCCGCATAGTTAAGCCAGCCCCGACA
CCCCCAACACCCGCTGACGCGCCCTGACGGGCTTGTCTGCTCCCGGCATCCGCTTACAG
ACAAGCTGTGACCGTCTCCGGGAGCTGCATGTGTGACAGGTTTTACCGTCTATCACCGAA
ACGCGCGAGACGAAAGGGCCTCGTGATACGCCTATTTTTATAGGTTAATGTCATGATAAT
AATGGTTTCTTAGACGTGAGGTGGCACTTTTCGGGGAAATGTGCGCGGAACCCCTATTG
TTTATTTTCTAAATACATTTCAAAATATGTATCCGCTCATGAGACAATAACCTGATAAAT
GCTTCAATAATATTGAAAAAGGAAGAGTATGAGTATTCAACATTTCCGTGTGCCCCCTAT
TCCCTTTTTTGCGGCATTTTGCCCTTCTGTTTTTGTCTCACCAGAAACGCTGGTGAAAGT
AAAAGATGCTGAAGATCAGTTGGGTGCACGAGTGGGTACATCGAATGGATCTCAACAG
CGGTAAAGATCCTTGAGAGTTTTTCGCCCCGAAGAAGCTTTTCAATGATGAGCACTTTTAA
AGTCTGTCTATGTGGCGCGGTATTATCCCGTATTGACGCGGGCAAGAGCAACTCGGTG
CCGCATACACTATTCTCAGATGACTTGGTTGAGTACTCACCAGTCACAGAAAAGCATCT
TACGGATGGCATGACAGTAAGAGAATTATGCAGTGTGCCATAACCATGAGTGATAACAC
TGCGGCCAACTTACTTCTGACAACGATCGGAGGACCGAAGGAGCTAACCGCTTTTTTGCA
CAACATGGGGGATCATGTAATCGCCTTGATCGTTGGGAACCGGAGCTGAATGAAGCCAT
ACCAAAACGACGAGCGTGACACCACGATGCCGTGTAGCAATGGCAACAACGTTGCGCAAACT
ATTAAGTGGCGAACTACTTACTCTAGCTTCCCGGCAACAATTAATAGACTGGATGGAGGC
GGATAAAGTTGCAGGACCACTTCTGCGCTCGGCCCTTCCGGCTGGCTGGTTATTGCTGA
TAAATCTGGAGCCGGTGAGCGTGGGTCTCGCGGTATCATTGCAGCACTGGGGCCAGATGG
TAAGCCCTCCCGTATCGTAGTTATCTACAGACGGGGAGTCAGGCAACTATGGATGAACG
AAATAGACAGATCGCTGAGATAGGTGCCTCACTGATTAAAGCATTGGTAACTGTGACACCA
AGTTTACTCATATATACTTTAGATTGATTTAAACTTCATTTTTAATTTAAAGGATCTA
GGTGAAGATCCTTTTTTGATAATCTCATGACCAAAATCCCTTAACGTGAGTTTTCGTTCCA
CTGAGCGTCAGACCCCGTAGAAAAGATCAAAGGATCTTCTTGAGATCCTTTTTTCTGCG
CGTAATCTGTGCTTGCAAAACAAAAAACCCGCTACCAGCGGTGGTTGTTTGCCGGA
TCAAGAGCTACCAACTCTTTTTCCGAAGGTAAGTGGCTTCAGCAGAGCGCAGATACCAAA
TACTGTCTTCTAGTGTAGCCGTAGTTAGGCCACCACTTCAAGAACTCTGTAGCACCGCC
TACATACCTCGCTCTGCTAATCCTGTTACCAGTGGCTGTGCGCAGTGGCGATAAGTCTGT
TCTTACCGGGTTGGACTCAAGACGATAGTTACCGGATAAGGCGCAGCGGTGCGGCTGAAC

FIGURE 93C

GGGGGGTTCGTGCACACAGCCCAGCTTGGAGCGAACGACCTACACCGAACTGAGATACCT
ACAGCGTGAGCATTGAGAAAGCGCCACGCTTCCCGAAGGGAGAAAGGCGGACAGGTATCC
GGTAAGCGGCAGGGTCGGAACAGGAGAGCGCACGAGGGAGCTTCCAGGGGGAAACGCCTG
GTATCTTTATAGTCCTGTCGGGTTTCGCCACCTCTGACTTGAGCGTCGATTTTGTGATG
CTCGTCAGGGGGGCGGAGCCTATGGAAAAACGCCAGCAACGCGGCCTTTTACGGTTCCT
GGCCTTTTGCTGGCCTTTTGCTCACATGTTCTTTCCTGCGTTATCCCCTGATTCTGTGGA
TAACCGTATTACCGCCTTTGAGTGAGCTGATACCGCTCGCCGCGAGCCGAACGACCGAGCG
CAGCGAGTCAGTGAGCGAGGAAGCGGAAGAGCGCCCAATACGCAAACCGCCTCTCCCGC
GCGTTGGCCGATTCAATTAATGCAGAGCTTGCAATTCGCGCGTTTTTCAATATTATTGAAG
CATTTATCAGGGTTATTGTCTCATGAGCGGATACATATTTGAATGTATTAGAAAAATAA
ACAAATAGGGGTTCCGCGCACATTTCCCGAAAAGTGCCACCTGACGTCTAAGAAACCAT
TATTATCATGACATTAACTATAAAAATAGGCGTAGTACGAGGCCCTTTCCTCATTAG

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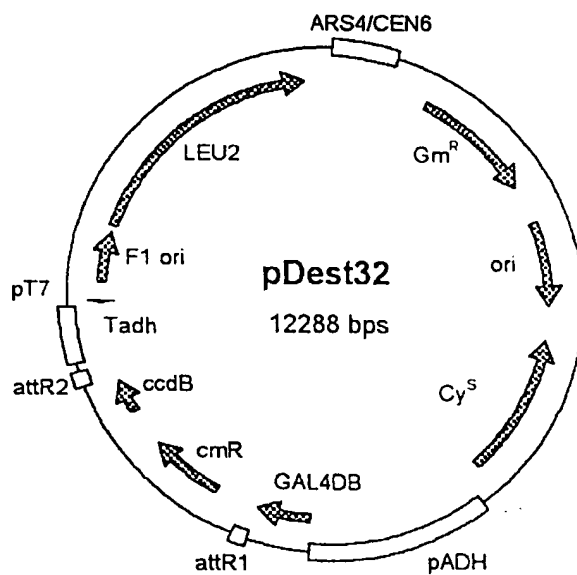


FIGURE 94A

pDEST32 12288 bp

GACGAAAGGGCCTCGTGATACGCCTATTTTTATAGGTTAATGTCATGATAAATGGTTT
CTTAGGACGGATCGCTTGCCTGTAACCTTACACGCGCCTCGTATCTTTAATGATGGAATA
ATTTGGGAATTTACTCTGTGTATTATTTATTTTATGTTTTGTATTGGATTTTAGAAAGT
AAATAAAGAAGGTAGAAGAGTTACGGAATGAAGAAAAAAATAAACAAAGGTTAAAAA
ATTTCAACAAAAAGCGTACTTTACATATATATTATTAGACAAGAAAAGCAGATTAAATA
GATATACATTTCGATTAACGATAAGTAAATGTAAATCACAGGATTTTCGTGTGTGGTCT
TCTACACAGACAAGATGAAACAATTCGGCATTAAATACCTGAGAGCAGGAAGAGCAAGATA
AAAGGTAGTATTTGTGGCGATCCCCCTAGAGTCTTTACATCTTCGGAAAAACAAAACT
ATTTTTCTTTAATTTCTTTTTTACTTTCTATTTTTAAATTTATATATTTATATTAATAA
ATTTAAATTTATAATTATTTTATAGCACGTGATGAAAAGGACCCAGGTGGCACTTTTCGG
GGAAATGTGGCGGGAACCCCTATTTGTTATTTTTCTAAATACATTCAAATATGTATCCG
CTCATGAGACAATAACCTGATAAATGCTTCAATAATCTGCAGTGGCGAGGCGCCGTGTC
TCAAAATCTCTGATGTTACATTGCACAAGATAAAAAATATATCATCATGAACAATAAACT
GTCTGCTTACATAAACAGTAATACAAGGGGTGTTATGAGCCATATTCAACGGGAACGTC
TTGCTGGAGGCCGCGATTAAATTCACATGGATGCTGATTATATGGGTATAAATGGGC
TCGGTAGCCCAACCACTAGAACTATAGCTAGAGTCTTGGGCGAACAAACGATGCTCGCCTT
CCAGAAAACCGAGGATGCGAACCCTTCATCCGGGTGAGCACCACCGGCAAGCGCGCG
ACGGCCGAGGTCTTCCGATCTCCTGAAGCCAGGGCAGATCCGTGCACAGCACCTTGGCGT
AGAAGAACAGCAAGGCCGCAATGCTGACGATGCGTGGAGACCGAAACCTTGGCGTCTGT
TCGCCAGCCAGGACAGAAATGCTCGACTTCGCTGCTGCCCAAGGTGCGCGGTGACGCA
CACCGTGGAAACGGATGAAGGCACGAACCCAGTTGACATAAGCCTGTTCCGTTTCGTAAC
TGTAATGCAAGTAGCGTATGCGCTCAGCCTGCTCCAGAACCTTGACCGAACGCGCG
GTGGTAACGGCGCAGTGGCGGTTTTTCATGGCTTGTTATGAGTGTTTTTGTACAGTCTA
TGCTCGGGCATCCAAGCAGCAAGCGGTTACGCCGTGGGTGATGTTTGTATGTTATGGA
GCAGCAACGATGTTACGCAGCAGCAACGATGTTACGCAGCAGGGCAGTCCGCCATAAACA
AAGTTAGGTGGTCAAGTATGGGCATCATTCCGACATGTAGGCTCGGCCCTGACCAAGTC
AAATCCATGCGGGCTGCTCTTGATCTTTTCGGTCTGAGTTCCGAGACGTAGCCACCTAC
TCCCAACATCAGCCGGACTCCGATTACCTCGGGAACCTGCTCCGTAGTAAGACATTATC
CGCCTTGCTGCCTTCGACCAAGAAGCGGTTGTTGGCGCTCTCGCGCTTACGTTCTGCC
AGGTTTGAGCAGCGCGTAGTGAGATCTATATCTATGATCTCGCAGTCTCCGCGAGCAC
CGGAGGCAGGGCATTGCCACCGCGCTCATCAATCTCCTCAAGCATGAGGCCAAGCGGCTT
GGTGCTTATGTGATCTACGTGCAAGCAGATTACGGTGACGATCCCGCAGTGGCTCTCTAT
ACAAGTTGGGCATACGGGAAGAAGTGATGCACTTTGATATCGACCCCAAGTACCGCCACC
TAACAATTGTTCAAGCCGAGATCGGCTTCCCGGCTAATAGGTTGTATTGATGTTGGAC
GAGTCGGAATCGCAGACCGATACCGAGATCTTGCCATCTTATGGAACCTGCCTCGGTGAGT
TTTCTCCTTCATTACAGAAACGGCTTTTCAAAAATATGGTATTGATAATCCTGATATGA
ATAAATTCAGTTTTCATTTGATGCTCGATGAGTTTTTCTAATCAGAATTGGTTAATTGGT
TGTAACACTGGCAGAGCATTACGCTGACTTGACGGGACGGCGNCATGACCAAAATCCCTT
AACGTGAGTTTTCGTTCCACTGAGCGTCAGACCCCGTAGAAAAGATCAAAGGATCTTCTT
GAGATCCTTTTTTCTGCGCGTAATCTGCTGCTTGCAAACAAAAAACCCACCGCTACCAG
CGGTGTTTTGTTTCCGGATCAAGAGCTACCAACTCTTTTCCGAAGGTAACCTGGCTTCA
GCAGAGCGCAGATACCAAACTGTCCTTCTAGTGTAGCCGTAGTTAGGCCACCACTTCA
AGAACTCTGTAGCACCGCTACATACCTCGCTCTGCTAATCCTGTTACCAGTGGCTGCTG
CCAGTGGCGATAAGTCTGTCTTACCGGGTTGGACTCAAGACGATAGTTACCGGATAAAG
CGCAGCGGTGCGGCTGAACGGGGGTTTCTGTCACACAGCCAGCTTGGAGCGAACGACCT
ACACCGAACTGAGATACCTACAGCGTGAGCATTGAGAAAGCGCCACGCTTCCCGAAGGGA
GAAAGGCGGACAGGTATCCGGTAAGCGGCAGGGTCGGAACAGGAGAGCGCAGAGGGAGC
TTCCAGGGGGGAACGCTGGTATCTTTATAGTCTGTGCGGTTTTCGCCACCTCTGACTTG
AGCGTGCATTTTTGTGATGCTCGTCAGGGGGGCGAGCCTATGGAAAAACGCCAGCAACG
CGGCCTTTTACGGTTCTTGGCCTTTTGCTGGCCTTTTGCTCACATGTTCTTCTCTGCGT
TATCCCTGATTCTGTGGATAACCGTATTACCGCCTTTGAGTGAGCTGATACCGCTCGCC
GCAGCCGAACGACCGAGCGCAGCGAGTCACTGAGCGAGGAAGCGGAAGAGCGCCCAATAC
GCAACCGCCTCTCCCGCGCTTGGCCGATTCAATTAATGTCAGCTGGCACGACAGGTTTC
CCGACTGGAAGCGGCGAGTGAAGCGCAACGCAATTAATGTGAGTTACCTCACTCATTAGG
CACCCAGGCTTACACTTTATGCTTCCGGCTCTATGTTGTGTGGAATTGTGAGCGGAT
AACAAATTCACACAGGAACAGCTATGACCATGATTACGCCAAGCTCGGAATTAACCTC-

FIGURE 94B

ACTAAAGGGAACAAAGCTGGTACCGATCCCGAGCTTTGCAAATTAAGCCTTCGAGCGT
CCCAAAACCTTCTCAAGCAAGGTTTTCAGTATAATGTTACATGCGTACACGCGTCTGTAC
AGAAAAAAGAAAAATTTGAAATATAAATAACGTTCTTAATACTAACATAACTATAAAA
AAATAAATAGGGACCTAGACTTCAGGTTGTCTAACTCCTTCCTTTTCGGTTAGAGCGGAT
GTGGGGGGAGGGCGTGAATGTAAGCGTGACATAAATAATTACATGATATCGACAAAGGAA
AAGGGGCTGTCTTACTCACAGGCTTTTTTCAAGTAGGTAATTAAGTCGTTTCTGTCTTTT
TCCTTCTTCAACCCACCAAGGCCATCTTGGTACTTTTTTTTTTTTTTTTTTTTTTTTTT
TT
TTTTTTTTTATAGAAAATAATACAGAAGTAGATGTTGAATTAGATTAACTGAAGATATAT
AATTTATTGGAAAATACATAGAGCTTTTGTGATGCGCTTAAGCGATCAATTCAACAAC
ACCACCAGCAGCTCTGATTTTTCTTCAGCCAACCTTGGAGACGAATCTAGCTTTGACGAT
AACTGGAAACATTGGAAATCTACCTTACCCAAGATCTTACCGTAACCGGCTGCCAAAGT
GTCAATAACTGGAGCAGTTTCTTAGAAGCAGATTTCAGTATTGGTCTCTCTGTCTTCTC
TGGGTCAATGTCCACAATTTGTCCAGTTCAAGACTGGCTTCCAGAAATGAGCTTGTGTG
CTTGTGGAAGTATCTCATACCAACCTTACCGAAATAAACCCTGGATGGTATTATCCATGTT
AATTCGTGGTGTGTTGACCACCGGCCATACCTCTACCACCGGGGTGCTTTCTGTGCTT
ACCGATACGACCTTTACCGGCTGAGACGTGACCTCTGTGCTTTCTAGTCTTAGTGAATCT
GGAAGGCATTCTTGATTAGTTGGATGATGTTCTGGGATTAAATGCAAAAATCACTTAAAG
AAGGAAAATCAACGGAGAAAAGCAAAACGCCATCTTAAATATACGGGATACAGATGAAAGGG
TTTGAACCTATCTGAAAATAGCATTAAACAAGCGAAAACCTGCGAGGAAAATTTGTTGC
GTCTCTCGGGGCTATTACGCGCCAGAGGAAAATAGGAAAATAACAGGGCATTAGAAA
ATAATTTTGATTTTGGTAATGTGTGGGCTTGGTGTACAGATGTTACATTGGTTACAGTA
CTCTTGTTTTTGCTGTGTTTTTCGATGAATCTCCAAAATGGTTGTTAGCACATGGAAGAG
TCACCGATGCTAAGTTATCTCTATGTAAGCTACGTGGCGTGACTTTTGATGAAGCCGCAC
AAGAGATACAGGATTGGCAACTGCAAAATAGAATCTGGGGATCCCCCTCGAGATCCGGGA
TCGAAGAAATGATGGTAAATGAAATAGGAAATCAAGGAGCATGAAGGCAAAAGACAATA
TAAGGGTCGAACGAAAATAAAGTGAAAAGTGTGATATGATGATTTGGCTTTGCGGCG
CCGAAAAACGAGTTTACGCAATTGCACAATCATGCTGACTCTGTGGCGGACCCGCGCTC
TTGCGGGCCCGGCGATAACGCTGGGCGTGAGGCTGTGCCCGGCGAGTTTTTGTGCCCTG
CATTTTCCAAGGTTTACCCTGCGCTAAGGGCGCGAGATTGGAGAAGCAATAAGAATGCCGG
TTGGGGTTGCGATGATGACGACCAGCAACTGGTGTATTATTAAAGTTGCCGAAAGAA
CCTGAGTGCAATTTGCAACATGAGTATACTAGAAGAATGAGCCAAGACTTGCAGACGCGA
GTTTGGCGGTGGTGCGAACAAATAGAGCGACCATGACCTTGAAGGTGAGACGCGCATAAAC
GCTAGAGTACTTTGAAGAGGAAACAGCAATAGGGTTGCTACCAGTATAAATAGACAGGTA
CATACAACACTGGAAATGGTTGTCTGTTTGTAGTACGCTTTCAATTCATTGGGTGTGCAC
TTTATTATGTTACAATATGGAAGGGAACCTTTACACTTCTCTATGCACATATATTAATTA
AAGTCCAATGCTAGTAGAGAAGGGGGTAACCCCTCCGCGCTCTTTCCGATTTTTTT
CTAAACCGTGGAAATTTTCGGATATCCTTTTGTGTTTTCCGGGTGTACAATATGGACTT
CTCTTTTCTGGCAACCAACCCATACATCGGGATTCTTATAATACCTTCGTTGGTCTCCC
TAACATGTAGGTGGCGGAGGGGAGATATACAATAGAACAGATACCAGACAAGACATAATG
GGCTAAACAAGACTACACCAATTACACTGCCTCATTGATGGTGGTACATAACGAACATAAT
ACTGTAGCCCTAGACTTGATAGCCATCATCATATCGAAGTTTCACTACCTTTTTCATT
TGCCATCTATTGAAGTAATAATAGGCGCATGCAACTTCTTTCTTTTCTTTTCTTTCTC
TCTCCCCCGTTGTTGTCTCACCATATCCGCAATGACAAAAAAATGATGGAAGACACTAA
AGGAAAAAATTAACGACAAAGACAGCACCAACAGATGTCGTTGTTCCAGAGCTGATGAGG
GGTATCTTCAACACACGAAACTTTTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCT
AGCAACGGTATACGGCCTTCTTCCAGTTACTTGAATTTGAAATAAAAAAAGTTTGCCGC
TTTGCTATCAAGTATAAATAGACCTGCAATTATTAATCTTTGTTTCTCGTCAATGTTT
TCGTTCCCTTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCT
AATCAACTCCAAGCTTGAAGCAAGCCTCTGAAAGATGAAGCTACTGTCTTCTATCGAAC
AAGCATGCGATATTTGCCGACTTAAAAAGCTCAAGTGCTCCAAAGAAAACCGAAGTGCG
CCAAGTGTCTGAAGAACAACCTGGGAGTGTGCTACTCTCCCAAAACCAAAAGGTCTCCGC
TGACTAGGGCACATCTGACAGAAGTGGAATCAAGGCTAGAAAGACTGGAACAGCTATTTT
TACTGATTTTCTCGAGAAGACCTTGACATGATTTGAAATGGATTCTTTACAGGATA
TAAAGCATTTGTTAACAGGATTATTTGTACAAGATAATGTGAATAAAGATGCCGTCACAG
ATAGATTGGCTTCAGTGGAGACTGATATGCCTCTAACATTGAGACAGCATAGAATAAGTG
CGACATCATCATCGGAAGAGTAGTAACAAAGGTCAAAGACAGTTGACTGTATCGTCGA
GGTCCAATCAAACAAGTTTGTACAAAAAGCTGAACGAGAAACGTAATGATATAAATA-

Figure 94C

TCAATATATTAAATTAGATTTTGCATAAAAAACAGACTACATAATACTGTAAAAACACAAC
ATATCCAGTCACTATGGCGGCCGCTAAGTTGGCAGCATCACCCGACGCACTTTGCGCCGA
ATAAATACCTGTGACGGAAGATCACTTCGCAGAATAAATAAATCCTGGTGTCCCTGTTGA
TACCGGGAAGCCCTGGGCCAACTTTTGGCGAAAAATGAGACGTTGATCGGCACGTAAGAGG
TTCCAACCTTTCACCATAATGAAATAAGATCACTACCGGGCGTATTTTGTGATTATCGAG
ATTTTCAGGAGCTAAGGAAGCTAAAAATGGAGAAAAAATCACTGGATATACCACCGTTGA
TATATCCCAATGGCATCGTAAAGAACATTTTGAGGCATTTCACTGAGTTGCTCAATGTAC
CTATAACAGACCGCTTCAGCTGGATATTACGGCCTTTTAAAGACCGTAAAGAAAAATAA
GCACAAGTTTTATCCGGCCTTTATTACATTCTTGCCCGCCTGATGAATGCTCATCCGGA
ATTCGGTATGGCAATGAAAGACGGTGAGCTGGTATATGGGATAGTGTTCACCTTGTGA
CACCCTTTTCCATGAGCAAACTGAAACGTTTTCATCGCTCTGGAGTGAATACCACGACGA
TTTCCGGCAGTTTCTACACATATATTTCGAAGATGTGGCGTGTACGGTGAAGAACCTGGC
CTATTTCCTAAAGGGTTTATTGAGAATATGTTTTCTGCTCAGCCAATCCCTGGGTGAG
TTTACCAGTTTTGATTTAAACGTGGCCAATATGGACAACCTTCTTCGCCCCCGTTTTAC
CATGGGCAAAATATTATACGCAAGGCGACAAGGTGCTGATGCCGCTGGCGATTCAAGTTCA
TCATGCCGTCTGTGATGGCTTCCATGTCCGCAGAAATGCTTAATGAATTACAACAGTACTG
CGATGAGTGGCAGGGCGGGCGTAATCTAGAGGATCCGGCTTACTAAAAGCCAGATAACA
GTATGCGTATTTGCGCGCTGATTTTTGCGGTATAAGAATATATACTGATATGTATACCCG
AAGTATGTCAAAAAGAGGTGTGCTATGAAGCAGCGTATTACAGTGACAGTTGACAGCGAC
AGCTATCAGTTGCTCAAGGCATATATGATGTCAATATCTCCGGTCTGGTAAGCAACAACCA
TGCAGAAATGAAGCCGCTCGTCTGCGTCCGAACGCTGGAAGCGGAAAATCAGGAAGGGA
TGGCTGAGGTGCGCCCGTTTATTGAAATGAACGCTCTTTGCTGACGAGAACAGGGACT
GGTGAATGCAAGTTTAAAGTTTACACCTATAAAAGAGAGACCGTTATCGTCTGTTGTG
GATGTACAGAGTGATATTATGACACGCCCGGGCGACGGATGGTATCCCCCTGGCCAGT
GCACGTCTGCTGTGATGATAAAGTCTCCCGTGAACCTTACCCGGTGGTGCATATCGGGGAT
GAAAGCTGGCGCATGATGACCACCGATATGGCCAGTGTGCCGGTCTCCGTTATCGGGGAA
GAAGTGGCTGATCTCAGCCACCGCGAAAATGACATCAAAAACGCCATTAACTGATGTTT
TGGGGAATATAAATGTGAGGCTCCCTTATACACAGCCAGTCTGCAAGTTCGACCATAGTGA
CTGGATATGTTGTGTTTTACAGTATTATGTAGTCTGTTTTTATGCAAAATCTAATTTAA
TATATTGATATTTATATCATTTTACGTTTCTCGTTTCAGCTTTCTTGTACAAAGTGGTTG
ATGGCCGCTAAGTAAGTAAGACGCTCGAGCTCTAAGTAAGTAACGGCCGCCACCGCGTGG
AGCTTTGGACTTCTTCGCCAGAGGTTTGGTCAAGTCTCAATCAAGGTTGTGCGCTGTG
TACCTTGCCAGAAATTTACGAAAAGATGGAAGGGTCAAATCGTTGGTAGATACGTTGT
TGACACTTCTAAATAAGCGAATTTCTTATGATTTATGATTTTTATTATTAATAAGTTAT
AAAAAAAATAAGTGATACAAATTTTAAAGTGACTCTTAGGTTTAAACGAAAATTTCTT
GTTCTTGAGTAACCTTTCTGTAGGTGAGTTGCTTTCTCAGGTATAGCATGAGGTGCG
TCTTATTGACCACACCTCTACCGGCATGCCGAGCAATGCCTGCAAAATCGCTCCCCATTT
CACCAATTGTAGATATGCTAACTCCAGCAATGAGTTGATGAATCTCGGTGTGTATTTA
TGTCTCAGAGGACAATACCTGTGTAAATCGTTCTTCCACACGGATCCCAATTGCGCCCTA
TAGTGAGTCGTATTACAATTCAGTGGCCGTCGTTTACAACGTCGTGACTGGGAAAACCC
TGGCGTTACCCAACCTAATCGCCTTGACGACATCCCCCTTTCGCCAGCTGGCGTAATAG
CGAAGAGGGCCCGACCGATCGCCCTTCCCAACAGTTGCGCAGCCTGAATGGCGAATGGAC
GCGCCCTGTAGCGGCGCATTAAGCGCGGCGGGTGTGGTGGTTACGCGCAGCGTGACCGCT
ACACTTGCCAGCGCCCTAGCGCCCGCTCCTTTGCTTTCTTCCCTTCTTCTCGCCACG
TTGCGCCGGCTTTCCCGCTCAAGCTCTAAATCGGGGGCTCCCTTTAGGGTTCCGATTTAGT
GCTTTACGGCACCTCGACCCCAAAAACTTGATTAGGGTGATGTTTACGTAAGTGGGCCA
TCGCCCTGATAGACGGTTTTTTCGCCCTTGACGTTGGAGTCCACGTTCTTTAATAGTGA
CTCTTGTTCCAACTGGAACAACACTCAACCTATCTCGGTCTATCTTTTGATTATATAA
GGGATTTTGGCGATTTGCGCCTATTGGTTAAAAAATGAGCTGATTAAACAAAAATTTAAC
GCGAATTTTAAACAAATATTAAAGTTTACAAATTTCTGATGCGGTATTTTCTCCTTACGC
ATCTGTGCGGTATTTACACCGCATATCGACCGGTCGAGGAGAACTTCTAGTATATCCAC
ATACCTAATATTATTGCCCTTATTAAAAATGGAATCGGAACAATTACATCAAAATCCACAT
TCTCTTCAAAATCAATTGTCTGTACTTCTTGTTCATGTGTGTTCAAAAACGTTATATT
TATAGGATAATTATACTCTATTTCTCAACAAGTAATTGGTTGTTTGGCCGAGCGGTCTAA
GGCGCCTGATTCAAGAAATATCTTGACCGCAGTTAACTGTGGGAATACTCAGGTATCGTA
AGATGCAAGAGTTGCAATCTTTAGCAACCATTATTTTTTCTTCAACATAACGAGAACA
CACAGGGGCGCTATCGCACAGAATCAAAATTCGATGACTGGAAATTTTGTAAATTTAG
AGGTGCGCCTGACGCATATACCTTTTTCAACTGAAAAATTTGGGAGAAAAAGGAAGGTGAG-

FIGURE 94D

AGGCCGGAACCGGCTTTTCATATAGAATAGAGAAGCGTTTCATGACTAAATGCTTGCATCA
CAATACTTGAAGTTGACAATATTATTAAAGGACCTATTGTTTTTCCAATAGGTGGTTAG
CAATCGTCTTACTTTCTAATTTTCTTACCTTTTACATTTTCAGCAATATATATATATATT
TCAAGGATATACCATTTCTAATGTCTGCCCCCTATGTCTGCCCCCTAAGAAGATCGTCGTTTT
GCCAGGTGACCACGTTGGTCAAGAAATCACAGCCGAAGCCATTAAGGTTCTTAAAGCTAT
TTCTGATGTTCTGTTCCAATGTCAAGTTCGATTTTCGAAAAATCATTTAATTGGTGGTGTCTGC
TATCGATGTACAGGTGTCCCACTTCCAGATGAGGCGCTGGAAGCCTCCAAGAAGGTTGA
TGCCGTTTTGTTAGGTGCTGTGGGTGGTCTTAAATGGGGTACCGGTAGTGTTAGACCTGA
ACAAGGTTTTACTAAAAATCCGTAAAGAACTTCAATTGTACGCCAACTTAAGACCATGTAA
CTTTGTCATCCGACTCTCTTTTAGACTTATCTCCAATCAAGCCACAATTTGCTAAAGGTAC
TGACTTCGTTGTTGTCTCAGAGAATTAGTGGGAGGTATTTACTTTGGTAAGAGAAAGGAAGA
CGATGGTGTATGGTGTCTGCTTGGGATAGTGAACAATACACCGTTCCAGAAGTGCAAAGAAT
CACAAGAATGGCCGCTTTTCATGGCCCTACAACATGAGCCACCATTGCCTATTTGGTCCCTT
GGATAAAGCTAATGTTTTGGCCCTCTTCAAGATTATGGAGAAAAACTGTGGAGGAAACCAT
CAAGAACGAATTCCTTACATTGAAGGTTCAACATCAATTGATTGATTCTGCCGCCATGAT
CCTAGTTAAGAACCCCAACCCACCTAAATGGTATTATAATCACCAGCAACATGTTTGGTGA
TATCATCTCCGATGAAGCCTCCGTTATCCAGGTTCTTGGGTTTGTGCCATCTGCGTCT
CTTGGCCCTCTTGGCCAGACAAGAACACCGCATTTGGTTTGTACGAACCATGCCACGGTTC
TGCTCCAGATTTGCCAAAGAATAAGGTTGACCCCTATCGCCACTATCTTGTCTGCTGCAAT
GATGTTGAAATTGTCTATTGAACCTTGCCTGAAGAAGGTAAGGCCATTGAAGATGCAGTTAA
AAAGGTTTTGGATGCGAGGTATCAGAACTGGTGAATTTAGGTGGTTCACACAGTACCACCGA
AGTCGGTGATGCTGTGCGCCGAAGAAGTTAAGAAATCCTTGCTTAAAAAGATTCTCTTTT
TTTATGATATTTGTACATAAACTTTATAAATGAAATTCATAATAGAAACGACACGAAATT
ACAAATGGAATATGTTTCATAGGGTAGACGAACTATATACGCAATCTACATACATTTAT
CAAGAAGGAGAAAAAGGAGGATAGTAAAGGAATACAGGTAAGCAAATTGATACTAATGGC
TCAACGTGATAAGGAAAAAGAATTGCACTTTAACATTAATATTGACAAGGAGGAGGGCAC
CACACAAAAAGTTAGGTGTAAACAGAAAATCATGAACTACGATTCTTAATTTGATATTGG
AGGATTTTCTTAAAAAATAAATAACAACAATAAAAAACACTCAATGACCTGACCAT
TTGATGGAGTTTAAGTCAATACCTTCTTGAACCATTTCCATAATGGTGAAAGTTCCCTC
AAGAATTTTACTCTGTCTCAGAAACGGCCTTACGACGTAGTCTGATATGGTGCACCTCTCAGTA
CAATCTGCTCTGATGCCGCATAGTTAAGCCAGCCCCGACCCGCCAACACCGCTGACG
CGCCCTGACGGGCTTGTCTGCTCCCGCATCCGCTTACAGACAAGCTGTGACCGTCTCCG
GGAGCTGCATGTCTCAGAGGTTTTACCGTCATCACCGAACGCGCGA

FIGURE 94E

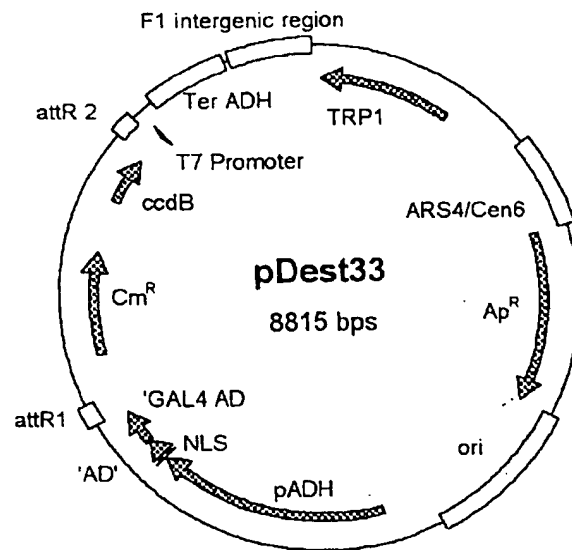


FIGURE 95A

pDEST33 8815 bp

GCCTTACGCATCTGTGCGGTATTTACACCGCAGGCAAGTGCACAAACAATACTTAAATA
AATACTACTCAGTAATAACCTATTTCTTAGCATTTTTGACGAAATTTGCTATTTTGTAG
AGTCTTTTACACCAATTTGTCTCCACACCTCCGCTTACATCAACACCAATAACGCCATTTA
ATCTAAGCGCATCACCAACATTTTCTGGCGTCAGTCCACCAGCTAACATAAAATGTAAGC
TTTCGGGGCTCTCTTGCCCTTCCAACCCAGTCAGAAATCGAGTTCCAATCCAAAAGTTCAC
CTGTCCACCTGCTTCTGAATCAAACAAGGAATAAACGAATGAGGTTTCTGTGAAGCTG
CACTGAGTAGTATGTTGCAGTCTTTTGAAATACGAGTCTTTAATAACTGGCAAACCGA
GGAATCTTGTGATTCTTGCCACGACTCATCTCCATGCAGTTGGACGATATCAATGCCGT
AATCATTGACCAGAGCCAAAACATCCTCCTTAGGTTGATTACGAAACACGCCAACCAAGT
ATTTCCGGAGTGCTGAACTATTTTATATGCTTTTACAAGACTTGAAATTTTCTTGCAA
TAACCGGGTCAATTGTTCTCTTTCTATTGGGCACACATATAATACCAGCAAGTCAGCAT
CGGAATCTAGAGCACATTCTGCGGCTCTGTGCTCTGCAAGCCGCAAACTTTACCAATG
GACCAGAACTACCTGTGAAATTAATAACAGACATACTCCAAGCTGCCTTTGTGTGCTTAA
TCAGTATACTCAGTGCTCAATAGTCACCAATGCCCTCCCTCTTGCCCTCTCCTTTTC
TTTTTTCGACCGAATTAATTTCTTAATCGGCAAAAAAGAAAGCTCCGGATCAAGATTGT
ACGTAAGGTGACAAGCTATTTTCAATAAAGAATATCTTCCACTACTGCCATCTGGCGTC
ATAACTGCAAAGTACACATATATTACGATGCTGTCTATTAAATGCTTCTATATTATATA
TATAGTAATGTCGTTTATGTTGCACTCTCAGTACAATCTGCTCTGATGCCGCAATAGTTAA
GCCAGCCCCGACACCCGCCAACACCCGCTGACGCGCCCTGACGGGCTTGTCTGCTCCCGG
CATCCGCTTACAGACAAGCTGTGACCGTCTCCGGGAGCTGCATGTGTGAGAGGTTTTCAC
CGTCATCACGAAACGCGCGAGACGAAAGGGCCTCGTGATACGCCTATTTTATAGGTTA
ATGTCATGATAATAATGGTTTCTTAGGACGGATCGCTTGCTGTAACCTTACACGCGCCTC
GTATCTTTTAATGATGGAATAATTTGGGAATTTACTCTGTGTTTATTTATTTTATGTTT
TGTATTTGGATTTTAGAAAGTAAATAAAGAAGGTAGAAGACTTACGGAATGAAGAAAAA
AAATAAACAAAGGTTAAAAAATTTCAACAAAAAGCGTACTTTACATATATATTATTAG
ACAAGAAAAGCAGATTAAATAGATATACATTGATTAACGATAAGTAAATGTAAATCA
CAGGATTTTCTGTGTGGTCTTCTACACAGACAAGATGAAACAATTCGGCATTAATACCT
GAGAGCAGGAAGAGCAAGATAAAAGGTAGTATTTGTTGGCGATCCCCCTAGAGTCTTTTA
CATCTTCGGAACAAAACTATTTTCTTTAATTTCTTTTACTTTCTATTTTAA
TTTATATATTTATATTAATAAATTTAAATTATAATTTATTTTATAGCAGTGATGAAAAG
GACCCAGGTGGCACTTTTCGGGGAAATGTGCGCGGAACCCCTATTTGTTATTTTCTAA
ATACATTCAAATATGTATCCGCTCATGAGACAATAACCCGTGATAATGCTTCAATAATAT
TGAAAAGGAAGAGTATGAGTATTAACATTTCCGTGTGCGCCTTATTCCTTTTGTGCG
GCATTTTGCCTTCTGTGTTTGTCTCACCCAGAAACGCTGGTGAAGTAAAGATGCTGAA
GATCAGTTGGGTGCACGAGTGGGTACATCGAAGTGGATCTCAACAGCGGTAAGATCCTT
GAGAGTTTTCGCCCCGAAGAACGTTTTTCAATGATGAGCACTTTAAAGTTCTGTATGT
GGCGCGGTATTTATCCCGTATTGACGCGGGCAAGAGCAACTCGGTGCGCGCATACACTAT
TCTCAGAAATGACTTGGTTGAGTACTCACAGTCACAGAAAAGCATCTTACGGATGGCATG
ACAGTAAGAGAATTATGCAAGTGTGCCATAACCATGAGTGATAACACTGCGGCCAACTTA
CTTCTGACAAACGATCGGAGGACCGAAGGAGCTAACCGCTTTTTCACAACATGGGGGAT
CATGTAACCTCGCCTTGATCGTTGGGAACCGGAGCTGAATGAAGCCATACCAAACGACGAG
CGTGACACCACGATGCTGTAGCAATGGCAACAACGTTGCGCAAACTATTAAGTGGCGAA
CTACTTACTCTAGCTTCCCGGCAACAATTAATAGACTGGATGGAGGCGGATAAAGTTGCA
GGACCACTTCTGCGCTCGGCCCTTCCGGCTGGCTGGTTTATTGCTGATAAATCTGGAGCC
GGTGAGCGTGGGTCTCGCGGTATCATTTGCAGCACTGGGGCCAGATGGTAAGCCCTCCCGT
ATCGTAGTTATCTACACGACGGGCAGTCAGGCAACTATGGATGAACGAAATAGACAGATC
GCTGAGATAGGTGCTCACTGATTAAGCATTGGTAACGTGTCAGACCAAGTTTACTCATAT
ATACTTTAGATTGATTTAAACCTTCAATTTTAAATTTAAAGGATCTAGGTGAAGATCCTT
TTTGATAATCTCATGACCAAAATCCCTTAACGTGAGTTTTCGTTCCACTGAGCGTCAGAC
CCCGTAGAAAAGATCAAAGGATCTTCTTGAGATCCTTTTCTGCGCGTAATCTGCTGC
TTGCAAAACAAAAAACCCGCTACCAGCGGTGGTTTGTGCGGATCAAGAGCTACCA
ACTCTTTTCCGAAGGTAAGTGGCTTCAGCAGAGCGCAGATACCAAATACTGTCTCTTCTA
GTGTAGCCGTAGTTAGGCCACCACTTCAAGAACTCTGTAGCACCGCCTACATACCTCGCT
CTGCTAATCCTGTTACCAAGTGGCTGCTGCCAGTGGCGATAAGTCGTGCTTACCGGGTTG
GACTCAAGACGATAGTTACCGGATAAGGCGCAGCGGTGCGGCTGAACGGGGGTTCTGTG
ACACAGCCCAGCTTGGAGCGAACGACCTACACCGAACTGAGATACCTACAGCGTGAGCAT-

FIGURE 95B

TGAGAAAGCGCCACGCTTCCCAGGGAGAAAGGCGGACAGGTATCCGGTAAGCGGCAGG
GTCGGAACAGGAGAGCGCACGAGGGAGCTTCCAGGGGGAAACGCTGGTATCTTTATAGT
CCTGTGCGGGTTTCGCCACCTCTGACTTGAGCGTCGATTTTGTGATGCTCGTCAGGGGGG
CCGAGCCTATGGAACACGCCAGCAACGCGGCTTTTTACGGTTCTTGGCCTTTTGCTGG
CCTTTTGCTCAGATGTTCTTCTGCGTTATCCCCTGATTCTGTGGATAACCGTATTACC
GCCTTTGAGTGAGCTGATACCGCTCGCCGAGCCGAACGACCGAGCGCAGCGAGTCAGTG
AGCGAGGAAGCGGAAGAGCGCCCAATACGCAACCGCCTCTCCCCGCGCGTTGGCCGATT
CATTAATGCAGCTGGCACGACAGGTTTCCCAGCTGGAAGCGGGCAGTGAGCGCAACGCA
ATTAATGTGAGTTACCTCACTCATTAGGCACCCAGGCTTTACACTTTATGCTCCGGCT
CCTATGTTGTGTGGAATTGTGAGCGGATAACAATTTACACAGGAAACAGCTATGACCAT
GATTACGCCAAGCTCGGAATTAACCTCACTAAAGGGAACAAAAGCTGGGTACCGGGCCC
CCCCTCGAGATCCGGGATCGAAGAAATGATGGTAAATGAAATAGGAAATCAAGGAGCATG
AAGGCAAAAGACAAATATAAGGGTCGAACGAAAAATAAAGTGAAAGTGTGATATGATG
TATTTGGCTTTGCGCGCGCGAAAAACAGTTTACGCAATTGCACAATCATGCTGACTCT
GTGGCGGACCCGCGCTCTTGC CGGCGCGGATAACGCTGGGCGTGAGGCTGTGCCGCGC
GGAGTTTTTTGCGCCTGCATTTTCCAAGGTTTACCCTGCGCTAAGGGGCGAGATTGGAGA
AGCAATAAGAATGCCGTTGGGGTTGCGATGATGACGACCACGACAACCTGGTGTCAATTAT
TTAAGTTGCCGAAAGAACCTGAGTGCATTTGCAACATGAGTATACTAGAAGAATGAGCCA
AGACTTGGCGAGACGCGAGTTTGC CGGTGGTGCGAACAATAGAGCGACCATGACCTTGAAG
GTGAGACGCGCATAACCGCTAGAGTACTTTGAAGAGGAAACAGCAATAGGGTTGCTACCA
GTATAAATAGACAGGTACATACAACTGGAATGGTGTCTGTTTGTGATACGCTTTCAA
TTCATTTGGGTGTGCATTTATTATGTTTACAATATGGAAGGGAACCTTTACACTTCTCCTA
TGCACATATATTAATTAAGTCCAATGCTAGTAGAGAAGGGGGTAACACCCCTCCGCGC
TCCTTTCCGATTTTTTTCTAAACCGTGGAATATTTCCGATATCCTTTTGTGTTTCCGGG
TGTACAATATGGACTTCTCTTTCTGGCAACCAACCATACATCGGGATTCCATATAAT
ACCTTCGTTGGTCTCCCTAACATGTAGGTGGCGGAGGGGAGATATACAATAGAACAGATA
CCAGACAAGACATAATGGGCTAAACAAGACTACACCAATTACATGCGCTCATTGATGGTG
GTACATAACGAACTAATACTGTAGCCCTAGACTTGATAGCCATCATCATATCGAAGTTTC
ACTACCTTTTCCATTTGCCATCTATTGAAGTAATAATAGGCGCATGCAACTTCTTTTC
TTTTTTTTCTTTCTCTCTCCCCGTTGTTGTCTCACCATATCCGCAATGACAAAAAAA
ATGATGGAAGACACTAAAGGAAAAAATTAACGACAAAGACAGCACCAACAGATGTCGTTG
TTCCAGAGCTGATGAGGGGTATCTTCGAACACACGAAACTTTTTCTTCTTCTTCAAG
CACACTACTCTCTAATGAGCAACGGTATACGGCCTTCTTCCAGTTACTTGAATTTGAAA
TAAAAAAGTTTTCGCGCTTTGCTATCAAGTATAAATAGACCTGCAATTATTAATCTTTTG
TTTCTCGTCAATGTTCTCGTTCCCTTTCTTCTTGTCTTTTCTGCACAATATTTCA
AGCTATACCAAGCATACAATCAACTCCAAGCTTATGCCCAAGAAGAAGCGGAAGGTCTCG
AGCGGCGCCAATTTTAATCAAAGTGGGAATATTGCTGATAGCTCATTGTCTTCACTTTC
ACTAACAGTAGCAACGGTCCGAACCTCATAACAACCTCAAACAAATTCTCAAGCGCTTTC
CAACCAATTGCCCTCCTCTAACGTTTATGATAACTTCATGAATAATGAAATCACGGCTAGT
AAAATTGATGATGGTAATAATTCAAACCACTGTACCTGGTTGGACGGACCAAACTGCG
TATAACGCGTTTGGGAATCACTACAGGGATGTTTAATACCACTACAATGGATGATGTATAT
AACTATCTATTGATGATGAAGATACCCACCAAAACCAAAAAAAGAGGGTGGGTGGAAT
CAAAACAAGTTTGTACAAAAAGCTGAACGAGAAACGTAAAATGATATAAATATCAATATA
TTAAATTAGATTTTGCATAAAAAACAGACTACATAAATACTGTAAAAACACAACATATCCAG
TCACTATGGCGGCGCTAAGTTGGCAGCATCACCCGACGCACTTTGCGCGGAATAAATAC
CTGTGACGGAAGATCACTTCGAGAAATAAATAAATCCTGGTGTCCCTGTTGATACCGGGA
AGCCCTGGGCCAATTTTGGCGAAATGAGACGTTGATCGGCACGTAAGAGGTTCCAAC
TTCAACATAATGAAATAAGATCACTACCGGGCGTATTTTTGAGTTATCGAGATTTTCAG
GAGCTAAGGAAGCTAAAATGGAGAAAAAATCACTGGATATACCACGTTGATATATCCC
AATGGCATCGTAAAGAACATTTTGGGCATTTCACTAGTTGCTCAATGTACCTATAAACC
AGACCGTTCACTGATGATATACCGGCTTTTTAAAGACCGTAAAGAAAAATAAGCACAACT
TTTATCCGCGCTTTATTACATTTCTGCGCGCTGATGAATGCTCATCCGGAATTCGTA
TGGCAATGAAAGACGGTGAGCTGGTATGAGGATAGTGTTCACCTTGTACACCGTTT
TCCATGAGCAAACTGAAACGTTTTTCATCGCTCTGGAGTGAATACCACGACGATTTCCGGC
AGTTTCTACACATATATTGCAAGATGTGGCGTGTACGGTGAAAACTGGCCTATTTCC
CTAAAGGGTTTATTGAGAATATGTTTTCTGCTCAGCCCAATCCCTGGGTGAGTTTACCA
GTTTTGATTTAAACGTGGCAATATGGACAACCTTCTCGCCCCGTTTTTACCATGGGCA
AATATTATACGCAAGGCGACAAGGTGCTGATGCCGCTGGCGATTACAGTTTATCATGCGG-

FIGURE 95C

TCTGTGATGGCTTCCATGTCCGCAGAATGCTTAATGAATTACAACAGTACTGCGATGAGT
GGCAGGGCGGGCGTAATCTAGAGGATCCGGCTTACTAAAAGCCAGATAACAGTATGCGT
ATTTGCGCGCTGATTTTTGCGGTATAAGAATATATACTGATATGTATACCCGAAGTATGT
CAAAAAGAGGTGTGCTATGAAGCAGCGTATTACAGTGACAGTTGACAGCGACAGCTATCA
GTTGCTCAAGGCATATATGATGTCAATATCTCCGGTCTGGTAAGCACAACCATGCAGAAT
GAAGCCCGTCTGCTGCGTGCCGAACGCTGGAAAGCGGAAAATCAGGAAGGGATGGCTGAG
GTCGCCCCGGTTTATTGAAATGAACGGCTCTTTGCTGACGAGAACAGGGACTGGTGAAAT
GCAGTTTAAGGTTTACACCTATAAAAGAGAGAGCCGTTATCGTCTGTTTGTGGATGTACA
GAGTGATATTATTGACACGCCCGGGCGACGGATGGTGATCCCCCTGGCCAGTGCACGCTCT
GCTGTGAGATAAAGTCTCCCGTGAACCTTACC CGGTGGTGATATCGGGGATGAAAGCTG
GCGCATGATGACCACCGATATGGCCAGTGTGCCGCTCTCCGTTATCGGGGAAGAAGTGGC
TGATCTCAGCCACCGCGAAAATGACATCAAAAACGCCATTAACTGATGTTCTGGGGAAT
ATAAATGTCAGGCTCCGTTATACACAGCCAGTCTGCAGGTCGACCATAGTGAAGTGGATAT
GTTGTGTTTTTACAGTATTATGTAGTCTGTTTTTATGCAAAATCTAATTTAATATATTGA
TATTTATATCATTTACGTTTCTCGTTCAGCTTCTTGTACAAAGTGGTTTGATGGCCGC
TAAGTAAGTAAGACGTCGAGCTCCCTATAGTGAGTCGTATTACACTGGCCGTCGTTTTAC
AACGTCGTGACTGGGAAAACACCGGTGAGCTCTAAGTAAGTAACGGCCGCCACCGCGGTG
GAGCTTTGGACTTCTTCGCCAGAGGTTTGGTCAAGTCTCCAATCAAGGTTGTCCGCTTGT
CTACCTTGCCAGAAATTTACGAAAAGATGGAAAAGGGTCAAATCGTTGGTAGATACGTTG
TTGACACTTCTAAATAAGCGAATTTCTTATGATTTATGATTTTTATTATTAAATAAGTTA
TAAAAAAAATAAGTGTATACAAATTTTAAAGTGACTCTTAGGTTTTTAAACGAAAATTCT
TGTTCTTGAGTAACTCTTTCCTGTAGGTGAGTTGCTTTCTCAGGTATAGCATGAGGTG
CTCTTATTGACCACACCTCTACCGGCATGCCGAGCAAATGCCTGCAAATCGCTCCCCATT
TCACCCAATTGTAGATATGCTAACTCCAGCAATGAGTTGATGAATCTCGGTGTGTATTTT
ATGTCCTCAGAGGACAATACCTGTTGTAATCGTTCTTCCACACGGATCCGCATCAGGCGA
AATTGTAAACGTTAATATTTTGTAAATTCGCGTTAAATATTTGTTAAATCAGCTCATT
TTTTAACCAATAGGCCGAAATCGGCAAAATCCCTTATAAATCAAAAGAATAGACCGAGAT
AGGGTTGAGTGTTGTTCCAGTTTGAACAAGAGTCCACTATTAAAGAACGTGGACTCCAA
CGTCAAAGGGCGAAAAACCGTCTATCAGGGCGATGGCCCACTACGTGAACCATCACCTTA
ATCAAGTTTTTTGGGGTCGAGGTGCCGTAAAGCACTAAATCGGAACCTTAAAGGGAGCCC
CCGATTTAGAGCTTGACGGGGAAAGCCGGCGAACGTGGCGAGAAAGGAAGGGAAGAAAGC
GAAAGGAGCGGGCGCTAGGGCGCTGGCAAGTGTAGCGGTACGCTGCGCGTAACCAACAC
ACCCGCCGCGCTTAATGCGCCGCTACAGGGCGCGTCCCATTCGCCATTCACTGCA

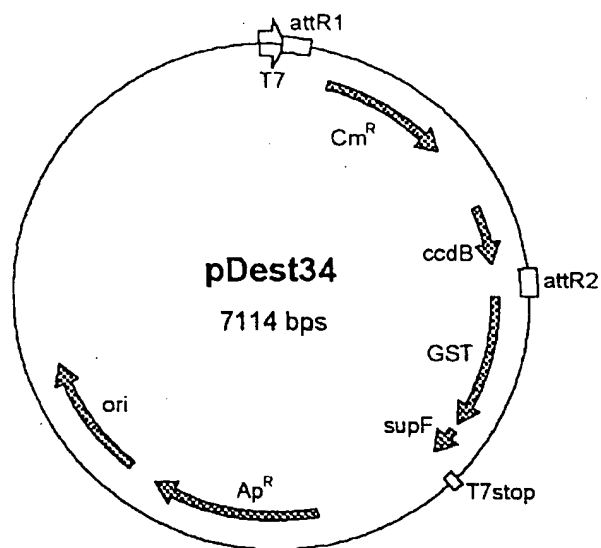


FIGURE 96A

pDEST34 7114 bp

<u>Location (Base Nos.)</u>	<u>Gene Encoded</u>
195..71	attR1
304..963	CmR
1305..1610	ccdB
1651..1775	attR2
1780..2472	GST
2675..2720	T7stop
3334..4194	ampR
4343..4982	ori

ATCGAGATCTCGATCCCGCGAAATTAATACGACTCACTATAGGGAGACCACAACGGTTTC
 CCTCTAGATCACAAAGTTTGTACAAAAAGCTGAACGAGAAAACGTAAAAATGATATAAATAT
 CAATATATTAATTTAGATTTTGCATAAAAAACAGACTACATAACTGTAAAACACAACA
 TATCCAGTCACTATGGCGGCCGATTAGGCACCCAGGCTTTACACTTTATGCTTCCGGC
 TCGTATAATGTGTGGATTTTGTAGTTAGGATCCGCGAGATTTTCAGGAGCTAAGGAAGCT
 AAAATGGAGAAAAAATCACTGGATATACCACCGTTGATATATCCCAATGGCATCGTAAA
 GAACATTTTGAGGCATTTCACTAGTTCATCAATGTACCTATAACCAGACCGTTTCACTG
 GATATTACGGCCTTTTAAAGACCGTAAGAAAAATAAGCACAAAGTTTTATCCGGCCTTT
 ATTCACATTTCTTGCCCGCTGATGAATGCTCATCCGGAATTCGATATGGCAATGAAAGAC
 GGTGAGCTGGTGATATGGGATAGTGTTCACCCTTGTACACCGTTTCCATGAGCAAAC
 GAAACGTTTTCATCGCTCTGGAGTGAATACCACGACGATTTCCGGCAGTTTCTACACATA
 TATTCGCAAGATGTGGCGTGTTCGGGTGAAAACCTGGCCTATTCCCTAAAGGGTTTATT
 GAGAATATGTTTTCTGCTCAGCCAATCCCTGGGTGAGTTTACCAGTTTGTATTTAAAC
 GTGGCCAATATGGACAACCTTCTCGCCCCGTTTTACCATGGGCAAAATATTATACGCAA
 GGCGACAAGGTGCTGATGCCGCTGCGGATTCAAGTTTATCATGCGCTCTGTGATGGCTTC
 CATGTCCGGCAGAAATGCTTAATGAATTACAACAGTACTGCGATGAGTGGCAGGGCGGGCG
 TAAACGCGTGGATCCGGCTTACTAAAAGCCAGATAACAGTATGCGTATTGCGCGCTGAT
 TTTTGGCGGTATAAGAATATATACTGATATGTATACCCGAAGTATGTCAAAAAGAGGTGT
 CTATGAAGCAGCGTATTACAGTGACAGTTGACAGCGACAGCTATCAGTTGCTCAAGGCAT
 ATATGATGTCAATATCTCCGCTCTGGTAAGCACAACCATGACAGATGAAGCCCGTCTGCT
 GCGTGCCGAACGCTGGAAAGCGGAAATCAGGAAGGGATGGCTGAGGTGCGCCCGTTTAT
 TGAATGAACGCTCTTTTGTCTGACGAGAACAGGGACTGGTGAAATGCAGTTTAAAGTTT
 ACACCTATAAAAGAGAGAGCCGTTATCGTCTGTTTGTGGATGTACAGAGTGATATTATG
 ACACGCCCCGGGCGACGGATGGTGATCCCCCTGGCCAGTGCACGCTGCTGTGATGATAAG
 TCTCCCGTGAACCTTACCCTGGTGATATCGGGGATGAAAGCTGGCGCATGATGACCA
 CCGATATGGCCAGTGTGCCGCTCTCCGTTATCGGGGAAGAAGTGGCTGATCTCAGCCACC
 GCGAAAATGACATCAAAAACGCCATTAACTGATGTTCTGGGGAATATAAATGTCAGGCT
 CCCTTATACACAGCCAGTCTGCAGGTGACCATAGTGAAGTGGATATGTTGTGTTTTACAG
 TATTATGTAGTCTGTTTTTTATGCAAAATCTAATTTAATATATTGATATTTATATCAATT
 TACGTTTCTCGTTCAGCTTTCTTGTACAAAGTGGTGATTATGTCCCTTATACTAGGTTAT
 TGGAAAATTAAGGCGCTTGTGCAACCCACTCGACTTCTTTTGAATATCTTGAAGAAAAA
 TATGAAGAGCATTGTATGAGCGCGATGAAGGTGATAAATGGCGAAACAAAAAGTTTGAA
 TTGGGTTTGGAGTTTCCCAATCTTCTTATTATATTGATGGTGATGTTAAATTAACACAG
 TCTATGGCCATCATACGTTATATAGCTGACAAGCACAACATGTTGGGTGGTGTGCCAAAA
 GAGCGTGCAGAGATTTCAATGCTTGAAGGAGCGGTTTTGGATATTAGATACGGTGTTCG
 AGAATTGCATATAGTAAAGACTTTGAAACTCTCAAAGTTGATTTTCTTAGCAAGCTACCT
 GAAATGCTGAAAATGTTTGAAGATCGTTTATGTATATAAATCATATTTAAATGGTGATCAT
 GTAACCCATCCTGACTTCATGTTGTATGACGCTCTTGATGTTGTTTTATACATGGACCCA
 ATGTGCCCTGGATGCGTTCCCAAAATTAGTTTGTGTTTTAAAAAACGATTGAAGCTATCCCA
 CAAATTGATAAGTACTTGAATCCAGCAAGTATATAGCATGGCCTTTGCGAGGCTGGCAA
 GCCACGTTTGGTGGTGGCGACCATCTCCAAAATCGGATCTGGTTCGCGCTCCATGGGGA
 TCCGGCTGCTAACAAAGCCCGAAAGGAAGCTGAGTTGGCTGCTGCCACCGCTGAGCGCTT
 CCCGATAAGGGAGCAGGCCAGTAAAGCATTACCCGTGGTGGGTTCCCGAGCGGCCAAA
 GGGAGCAGACTCTAAATCTGCCGTCATCGACTTCGAAGGTTTGAATCCTTCCCCCACCAC
 CATCACTTTCAAAAGTGAATTCGCTGAGCAATACTAGCATAACCCCTTGGGGCCTCTAA-

FIGURE 96B

ACGGGTCTTGAGGGGTTTTTGTCTGAAAGGAGGAACATATCCGGATATCCACAGGACGG
GTGTGGTTCGCCATGATCGCGTAGTCGATAGTGGCTCCAAGTAGCGAAGCGAGCAGGACTG
GGCGCGCGCCAAAGCGGTCCGACAGTGTCTCCGAGAACGGGTGCGCATAGAAATTGCATCA
ACGCATATAGCGCTAGCAGCACGCCATAGTGACTGGCGATGCTGTCCGAATGGACGATAT
CCCCGAAGAGGCCCGGCAGTACCGGCATAACCAAGCCTATGCCTACAGCATCCAGGGTGA
CGGTGCCGAGGATGACGATGAGCGCATTGTTAGATTTTCATACACGGTGCTGACTGCGTT
AGCAATTTAACTGTGATAAACTACCGCATTAAGCTTATCGATGATAAGCTGTCAAACAT
GAGAATTTCTGAAGACGAAAGGGCCTCGTGATACGCCCTATTTTATAGGTTAATGTCATG
ATAATAATGGTTTTCTTAGACGTACAGGTGGCACTTTTCGGGGAAATGTGCGCGGAACCCCT
ATTTGTTTATTTTTCTAAATACATTCAAATATGTATCCGCTCATGAGACAATAACCCCTGA
TAAATGCTTCAATAATATTGAAAAAGGAGAGTATGAGTATTCAACATTTCCGTGTCGCC
CTTATTCCTTTTTTTCGGGCATTTTGCCTTCTGTTTTTTGTCTACCCAGAAACGCTGGTG
AAAGTAAAGATGCTGAAGATCAGTTGGGTGCACGAGTGGGTTACATCGAACTGGATCTC
AACAGCGGTAAGATCCTTGAGAGTTTTCGCCCCGAGAAACGTTTTCCAATGATGAGCACT
TTTAAAGTTCTGCTATGTGGCGGTATTATCCCGTGTGACGCCGGGCAAGAGCAACTC
GTTCCGCCCATACACTATTCTCAGAATGACTGGTTGAGTACTCACCAGTCACAGAAAAAG
CATCTTACGGATGGCATGACAGTAAGAGAATTATGAGTGCTGCCATAACCATGAGTGAT
AACACTGCGGCCAACTTACTTCTGACAACGATCGGAGGACCGAAGGAGCTAACCGCTTTT
TTGCACAACATGGGGGATCATGTAACCTCGCTTGATCGTTGGGAACCGGAGCTGAATGAA
GCCATACCAAACGACGAGCGTGACACACGATGCCTGCAGCAATGGCAACAACGTTGCGC
AAACTATTAACTGGCGAACTACTTACTCTAGCTTCCCGGCAACAATTAATAGACTGGATG
GAGCGGGATAAAGTTGCAGGACCACTTCTGCGCTCGGCCCTTCCGGCTGGCTGGTTTTATT
GCTGATAAATCTGGAGCCGGTGAGCGTGGGTCTCGCGGTATCATTGACGCACTGGGGCCA
GATGGTAAGCCCTCCCGTATCGTAGTTATCTACACGACGGGGAGTCAGGCAACTATGGAT
GAACGAATAGACAGATCGCTGAGATAGGTGCCTCACTGATTAAAGCATTGGTAACGTCA
GACCAAGTTTACTCATATATACTTTAGATTGATTTAAACTTCATTTTTAATTTAAAGG
ATCTAGGTGAAGATCCTTTTTGATAATCTCATGACCAAAATCCCTTAACGTGAGTTTTCG
TTCCACTGAGCGTCAGACCCCGTAGAAAAGATCAAAGGATCTTCTTGAGATCCTTTTTTT
TTCCGCGTAATCTGCTGCTTGCAAAACAAAAAACCCAGCTACACGCGGTGGTTTTGTTG
CCGGATCAAGAGCTACCAACTCTTTTCCGAAGGTAACGCTTCCAGCAGAGCGCAGATA
CCAAATACTGTCTTCTAGTGAGCGTAGTTAGGCCACCACTTCAAGAACTCTGTAGCA
CCGCTACATACCTCGCTCTGCTAATCTGTTACAGTGGCTGCTGCCAGTGGCGATAAG
TCGTGTCTTACCGGGTTGGACTCAAGACGATAGTTACCGGATAAGGCGCAGCGGTGGGC
TGAACGGGGGGTTTCGTGCACACAGCCAGCTTGGAGCGAACGACCTACACCGAACTGAGA
TACCTACAGCGTGAGCTATGAGAAAGCGCCACGCTTCCGAAGGGAGAAAGGCGGACAGG
TATCCGGTAAGCGGCAGGGTCCGAACAGGAGAGCGCACGAGGGAGCTTCCAGGGGAAAC
GCCTGGTATCTTTATAGTCCTGTGCGGTTTTCGCCACCTCTGACTTGAGCGTCGATTTTTG
TGATGCTCGTCAGGGGGCGGAGCCTATGAAAAACGCCAGCAACCGCGCCTTTTTACGG
TTCTGGCCTTTTGTGGCCTTTTGTCTACATGTTCTTTCTGCGTTATCCCTGATTCT
GTGGATAACCGTATTACCGCCTTTGAGTGAGCTGATACGCTCGCCGAGCCGAACGACC
GAGCGCAGCGAGTCAGTGAGCGAGGAAGCGGAAGAGCGCTGATGCGGTATTTCTCCTT
ACGCATCTGTGCGGTATTTACACCGCATATATGGTGCACTCTCAGTACAATCTGCTCTG
ATGCCGCATAGTTAAGCCAGTATACACTCCGCTATCGCTACGTGACTGGGTCTATGGCTGC
GCCCGACACCCGCCAACACCCGCTGACGCGCCCTGACGGGCTTGTCTGCTCCCGGCATC
CGCTTACAGACAAGCTGTGACCGTCTCCGGGAGCTGCATGTGTGAGAGGTTTTACCGTC
ATCACCGAAACGCGGAGGACGCTGCGGTAAAGCTCATCAGCGTGGTCTGAGCGGATTC
ACAGATGTCTGCCGTGTTTCATCCGCGTCCAGCTCGTTGAGTTTCTCCAGAAGCGTTAATGT
CTGGCTTCTGATAAAGCGGGCCATGTTAAGGGCGGTTTTTCTGTTTGGTCACTGATGC
CTCCGTGTAAGGGGGATTCTGTTTCATGGGGTAATGATACCGATGAAACGAGAGAGGAT
GCTCACGATACGGGTACTGATGATGAACATGCCCGGTTACTGGAACGTTGTGAGGGTAA
ACAACTGGCGGTATGGATGCGGCGGGACAGAGAAAAATCACTCAGGGTCAATGCCAGCG
CTTCGTTAATACAGATGTAGGTGTTCCACAGGGTAGCCAGCAGCATCTGCGATGCAGAT
CCGGAACATAATGTGACAGGGCGCTGACTTCCGCGTTTCCAGACTTTACGAACACGGAA
ACCGAAGACCATTCATGTTGTTGCTCAGGTGCGCAGACGTTTTGACAGCAGCAGTCGCTTCA
CGTTCTGCTCGCGTATCGGTGATTCACTTCTGCTAACCAAGTAAGGCAACCCCGCCAGCCTAG
CCGGGTCTCAACGACAGGAGCAGCATCATGCGCACCCGTGGCCAGGACCCCAACGCTGCC
CGAGATGCGCCCGTGCCTGCTGAGATGGCGGACGCGATGGATATGTTCTGCCAAGG
GTTGGTTTTGCGCATTCACAGTTCTCCGCAAGAAATGATTGGCTCCAATCTTGGAGTGGT-

FIGURE 96C

GAATCCGTTAGCGAGGTGCCGCCGGCTTCCATTTCAGGTGAGGTGGCCCCGGCTCCATGCA
CCGCGACGCAACGCGGGGAGGCAGACAAGGTATAGGGCGCGCCTACAATCCATGCCAAC
CCGTTCCATGTGCTCGCCGAGGCGGCATAAATCGCCGTGACGATCAGCGGTCCAGTGATC
GAAGTTAGGCTGGTAAGAGCCGCGAGCGATCCTTGAAGCTGTCCCTGATGGTCGTCTATCT
ACCTGCCTGGACAGCATGGCCTGCAACGCGGGCATCCCGATGCCGCCGGAAGCGAGAAGA
ATCATAATGGGAAGGCCATCCAGCCTCGCGTCGCGAACGCCAGCAAGACGTAGCCCAGC
GCGTCGGCCGCCATGCCGGCGATAATGGCCTGCTTCTCGCCGAAACGTTTGGTGGCGGGA
CCAGTGACGAAGGCTTGAGCGAGGGCGTGCAAGATTCCGAATACCGCAAGCGACAGGCCG
ATCATCGTCGCGCTCCAGCGAAAGCGGTCTCTCGCCGAAATGACCCAGAGCGCTGCCGGC
ACCTGTCTTACGAGTTGCATGATAAAGAAGACAGTCATAAGTGCGGCGACGATAGTCATG
CCCCGCGCCCAACGGAAGGAGCTGACTGGGTTGAAGGCTCTCAAGGGCATCGGTGATCG
ACGCTCTCCCTTATGCGACTCCTGCATTAGGAAGCAGCCCAGTAGTAGGTTGAGGCCGTT
GAGCACCGCCGCGCAAGGAATGGTGATGCAAGGAGATGGCGCCCAACAGTCCCCCGGC
CACGGGGCCTGCCACCATACCACGCCGAAACAAGCGCTCATGAGCCCGAAGTGGCGAGC
CCGATCTTCCCATCGGTGATGTCGGCGATATAGGCGCCAGCAACCGCACCTGTGGCGCC
GGTGATGCCGGCCACGATGCGTCCGGCGTAGAGG

FIGURE 9(a)

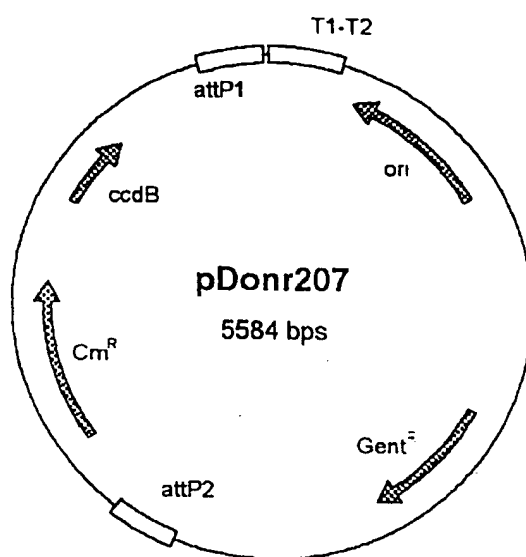


FIGURE 97A

pDONR207 5584 bp

GCGAGAGTAGGGAAGTCCAGGCATCAAATAAAACGAAAGGCTCAGTCGGAAGACTGGGC
CTTTTCGTTTTATCTGTTGTTGTTCGGTGAACGCTCTCCTGAGTAGGACAAATCCGCCGGG
AGCGGATTGAAAGTTGTGAAGCAACGCCCGAGGGTGGCGGGCAGGACGCCCGCCATA
AACTGCCAGGCATCAAATAAGCAGAAGGCCATCCTGACGGATGGCCTTTTTGCGTTTCT
ACAAACTCTTCTGGCTAGCGGTAATACGGTTATCCACAGAATCAGGGGATAACGCAGGA
AAGAACATGTGAGCAAAAGGCCAGCAAAAGGCCAGGAACCGTAAAAAGGCCGCTGTCTG
GCGTTTTTCCATAGGCTCCGCCCTGACGAGCATCAGAAAAATCGACGCTCAAGTCAG
AGGTGGCGAAACCCGACAGGACTATAAAGATACAGGCGTTTCCCCCTGGAAGCTCCCTC
GTGCGCTCTCCTGTTCCGACCTGCCGCTTACCGGATACCTGTCCGCTTTCTCCCTTCG
GGAAGCGTGGCGCTTTCTCATAGCTCAGCTGTAGGTATCTCAGTTCCGTTAGGTCGTT
CGCTCCAAGCTGGGCTGTGTGCAGAACCCCCCTTCAGCCCGACCGCTGCGCCTTATCC
GGTAACATATCGTCTTGAGTCCAACCCGGTAAGACACGACTTATCGCCACTGGCAGCAGCC
ACTGGTAACAGGATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAGAGTTCTTGAAGTGG
TGGCCTAACTACGGCTACACTAGAAGGACAGTATTTGGTATCTGCGCTCTGCTGAAGCCA
GTTACCTTCGGAAGAGAGTTGGTAGCTCTTGATCCGGCAAAACAAACCCGCTGGTAGC
GGTGGTTTTTTGTTTGCAAGCAGCAGATTACGCGCAGAAAAAGGATCTCAAGAAGAT
CCTTTGATCTTTTACGGGGTGTGACGCTCAGTGAACGAAAACTCAGTTAAGGGATT
TTGGTCATGAGCTTGCGCCGTCCCGTCAAGTCAGCGTAATGCTCTGCCAGTGTTACAACC
AATTAACCAATTCTGATTAGAAAACTCTCGAGCATCAAATGAACTGCAATTTATTCAT
TATCAGGATTATCAATACCATATTTTGAAGAAAGCGTTTCTGTAATGAAGGAGAAAT
CACCGAGGCAGTTCCATAGGATGGCAAGATCCTGGTATCGGTCTGCGATTCCGACTCGTC
CAACATCAATACAACCTATTAGTAGCCAACCACTAGAACTATAGCTAGAGTCTGGGCGA
ACAAACGATGCTCGCCTTCCAGAAAAACCGAGGATGCGAACCACTTCATCCGGGGTCAGCA
CCACCGGCAAGCGCCGCGACGGCCGAGGTCTTCCGATCTCTGAAGCCAGGGCAGATCCG
TGCACAGCACCTTGGCCTAGAAGAACAGCAAGGCCGCAATGCTGACGATGCGTGGAGA
CCGAAACCTTGGCCTCGTTCGCCAGCCAGGACAGAAATGCCTCGACTTCGCTGCTGCCA
AGGTTGCCGGGTGACGCACACCGTGGAAACGGATGAAGGCACGAACCCAGTTGACATAAG
CCTGTTCCGGTTCGTAAACTGTAATGCAAGTAGCGTATGCGCTCAGCCTGCTCCAGAA
CCTTGACCGAAGCGAGCGGTGTTAACGGCGCAGTGGCGGTTTTTCATGGCTTGTATGACT
GTTTTTTGTACAGTCTATGCTCGGCTCGGGCATCAAGCAGCAAGCGGTTACCGCGTGGGTC
GATGTTTGTATGTTATGGAGCAGCAACGATGTTACGCAGCAGCAACGATGTTACGCAGCAG
GGCAGTCGCCCTAAACAAAGTTAGGTGGCTCAAGTATGGGCATCATTCCGACATGTAGG
CTCGGCCCTGACCAAGTCAAATCCATGCGGGCTGCTCTTGATCTTTTCGGTCTGAGTTT
GGAGACGTAGCCACCTACTCCCAACATCAGCCGACTCCGATTACCTCGGGAACCTTGCTC
CGTAGTAAGACATTATCGCGCTTGCTGCTTCGACCAAGAAGCGGTTGTTGGCGCTCTC
GCGGCTTACGTTCTGCCAGGTTTGAGCAGCCGCGTAGTGAGATCTATATCTATGATCTC
GCAGTCTCCGGCGAGCACCGGAGGCAGGGCATTGCCACCGCGCTCATCAATCTCCTCAAG
CATGAGGCCAACGCGCTTGGTGCTTATGTGATCTACGTGCAAGCAGATTACGGTGACGAT
CCCCGAGTGGCTCTCTATACAAAGTTGGGCATACGGGAAGAAGTGATGCACTTTGATATC
GACCCAAAGTACCGCCACCTAACAATTGTTCAAGCCGAGATCGGCTTCCCGGCTAATTT
CCCCTCGTCAAAAAAAGGTTATCAAGTGAGAAATCACCATGAGTGACGACTGAATCCGG
TGAGAAATGGCAAAAGTTTATGCAATTTCTTCCAGACTTGTTCACAGGCCAGCCATTACG
CTCGTCAATAAATCACTCGCATCAACCAAAACCGTTATTCAATTCGTGATTGCGCTGAGC
GAGACGAAATACCGGATCGCTGTTAAAAGGACAATTACAAACAGGAATCGAATGCAACCG
GCGCAGGAACACTGCCAGCGCATCAACAATATTTTCACTGAATCAGGATATTTCTCTAA
TACCTGGAATGCTGTTTTTCCGGGATCGCAGTGGTGAGTAACCATGCATCATCAGGAGT
ACGGATAAAATGCTTGATGGTGGGAAGAGGCATAAATCCGTCAGCCAGTTTAGTCTGAC
CATCTCATCTGTAAACATCATTTGGCAACGCTACCTTTGCCATGTTTCAGAAACAACCTCTGG
CGCATCGGGCTTCCCATACAAGCGATAGATTGTGCGACCTGATTGCCCGACATTATCGCG
AGCCCATTTATACCCATATAAATCAGCATCCATGTTGGAATTTAATCGCGGCTCGACGT
TTCCCGTTGAATATGGCTCATACACCCCTGTATTACTGTTTATGTAAGCAGACAGTTT
TATTGTTTCATGATGATATATTTTATCTGTGCAATGTAACATCAGAGATTTTGAGACAC
GGGCCAGAGCTGCAGCTGGATGGCAAAATATGATTTTATTTGACTGATAGTGACCTGTT
CGTTGCAACAAATTGATAAGCAATGCTTTCTTATAATGCCAATTTGTACAAGAAAGCTG
AACGAGAAACGTAAATGATATAAATATCAATATATTAATTAGATTTCATATAAAAAAC
AGACTACATAATACTGTAAAAACAAATATCCAGTCACTATGAATCAACTACTTAGATG-

FIGURE 97B

GTATTAGTGACCTGTAGTCGACTAAGTTGGCAGCATCACCCGACGCACCTTTGCGCCGAAT
AAATACCTGTGACGGGAAGATCACTTCGCAGAATAAAATAAATCCTGGTGTCCCTGTTGATA
CCGGGAAGCCCTGGGCCAACTTTGGCGAAAATGAGACGTTGATCGGCACGTAAGAGGTTTC
CAACTTTCACCATAATGAAATAAGATCACTACCGGGCGTATTTTTTGTAGTTATCGAGATT
TTCAGGAGCTAAGGAAGCTAAAAATGGAGAAAAAAATCACTGGATATACCACCGTTGATAT
ATCCCAATGGCATCGTAAAGAACATTTTGTAGGCATTTTCAGTCAGTTGCTCAATGTACCTA
TAACCAGACCGTTTCAGCTGGATATTACGGCCTTTTAAAGACCGTAAAGAAAAATAAGCA
CAAGTTTTTATCCGGCCTTTATTCACATTTCTGCCCCGCTGATGAATGCTCATCCGGAATT
CCGTATGGCAATGAAAGACGGTGAGCTGGTGATATGGGATAGTGTTCACCCTTGTTACAC
CGTTTTCCATGAGCAAACGTGAAACGTTTTTCATCGCTCTGGAGTGAATACCACGACGATTT
CCGGCAGTTTTCTACACATATATTGCAAGATGTGGCGTGTACGGTGAAAACCTGGCCCTA
TTTCCCTAAAGGTTTTATTGAGAATATGTTTTTCGTCTCAGCCAATCCCTGGGTGAGTTT
CACCAGTTTTTGATTTAAACGTGGCCAATATGGACAACTTCTTCGCCCCCGTTTTTCACCAT
GGGCAAAATATTATACGCAAGGCGACAAGGTGCTGATGCCGCTGGCGATTTCAGGTTTCATCA
TGCCGCTCTGTGATGGCTTCCATGTTCGGCAGAATGCTTAATGAATTACAACAGTACTGCCGA
TGAGTGGCAGGGCGGGCGTAATCGCGTGGATCCGGCTTACTAAAAGCCAGATAACAGTA
TGCGTATTTGCGCGCTGATTTTTCGGGTATAAGAATATATACTGATATGTATACCCGAAG
TATGTCAAAAAGAGGTGTGCTATGAAGCAGCGTATTACAGTGACAGTTGACAGCGACAGC
TATCAGTTGCTCAAGGCATATATGATGTCAATATCTCCGGTCTGGTAAAGCACAACCATGC
AGAATGAAGCCCCGCTGCTGCGTGCCGAACGCTGGAAAGCGGAAAAATCAGGAAGGGATGG
CTGAGGTGCGCCCGGTTTTATTGAAATGAACGGCTCTTTTGTGACGAGAACAGGGACTGGT
GAAATGCAGTTTTAAGGTTTTACACCTATAAAAGAGAGAGCCGTTATCGTCTGTTTGTGGAT
GTACAGAGTGATATATTGACACGCCCCGGCGACGGATGGTGATCCCCCTGGCCAGTGCA
CGTCTGCTGTGATAGATAAAGTCTCCCGTGAACCTTTACCCGGTGGTGATATCGGGGATGAA
AGCTGGCGCATGATGACCACCGATATGGCCAGTGTGCCGGTCTCCGTTATCGGGGAAGAA
GTGGCTGATCTCAGCCACCGGAAAATGACATCAAAAACGCCATTAACCTGATGTTCTGG
GGAATATAAATGTAGGCTCCCTTATACACAGCCAGTCTGCAAGTTCGATACAGTAGAAAT
TACAGAACTTTTATCACGTTTTAGTAAGTATAGAGGCTGAAAATCCAGATGAAGCCGAACG
ACTTGTAAGAGAAAAAGTATAAGAGTTGTGAAATTTGTTCTTGATGCAGATGATTTTCAGGA
CTATGACACTAGCGTATATGAATAGGTAGATGTTTTTATTTTGTACACAAAAAAGAGGC
TCGCACCTCTTTTTCTTATTTCTTTTTATGATTTAATACGGCATTGAGGACAATAGCGAG
TAGGCTGGATACGACGATTCCGTTTGAGAAGAACATTTGGAAGGCTGTCGGTTCGACTAAG
TTGGCAGCATCACCGAAGAACATTTGGAAGGCTGTCGGTTCGACTACAGGTCACTAATAC
CATCTAAGTAGTTGATTCATAGTGACTGGATATGTTGTGTTTTACAGTATTATGTAGTCT
GTTTTTTATGCAAAATCTAATTTAATATATTGATATTTATATCATTTTTACGTTTTCTCGTT
CAGCTTTTTTTGTACAAAGTTGGCATTATAAAAAAGCATTGCTCATCAATTTGTTGCAACG
AACAGGTCACTATCAGTCAAAATAAAATCATTATTTGGGGCCCCGAGATCCATGCTAGCGT
TAAC

FIGURE 97C

pMAB85

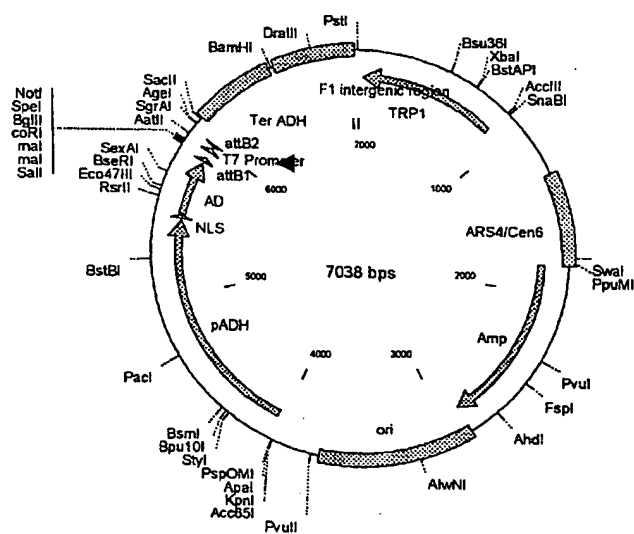


FIGURE 98A

pMAB85 7038 bp

GCCTTACGCATCTGTGCGGTATTTACACCGCAGGCAAGTGCACAAACAATACTTTAAATA
AATACTACTCAGTAATAACCTATTTCTTAGCATTTTGTACGAAATTTGCTATTTTGTAG
AGTCTTTTACACCATTGTCTCCACACCTCCGCTTACATCAACACCAATAACGCCATTTA
ATCTAAGCGCATCACCAACATTTCTGGCGTCAGTCCACCAGCTAACATAAAATGTAAGC
TTTCGGGGCTCTCTTGCCTTCCAACCCAGTCAGAAATCGAGTTCCAATCCAAAAGTTCAC
CTGTCCCACCTGCTTCTGAATCAACAAGGGAATAAACGAATGAGGTTTCTGTGAAGCTG
CACTAGTAGTATGTTGCAGTCTTTGGAAATACGAGTCTTTTAATAACTGGCAAAACCGA
GGAACTCTTGGTATTCTTGCCACGACTCATCTCCATGCAGTTGGACGATATCAATGCCGT
AATCATTGACCAGAGCCAAAACATCCTCCTTAGGTTGATTACGAAACACGCCAACCAAGT
ATTTCCGAGTGCCTGAATATTTTATATGCTTTTACAAGACTTGAAATTTTCTTGCAA
TAACCGGTCATTGTTCTCTTTCTATTGGGCACACATATAATACCCAGCAAGTCAGCAT
CGGAATCTAGAGCACATTCTGCGGCTCTGTGCTCTGCAAGCCGCAAACTTTACCAATG
GACCAGAACTACCTGTGAATTAATAACAGACATACTCCAAGCTGCCTTTGTGTGCTTAA
TCACGTATACTCAGTGTCTCAATAGTCACCAATGCCCTCCCTCTGGCCCTCTCCTTTTC
TTTTTTCGACCGAATTAATTCTTAATCGGCAAAAAAGAAAGCTCCGGATCAAGATTGT
ACGTAAGGTGACAAGCTATTTTTCAATAAAGAATATCTTCCACTACTGCCATCTGGCGTC
ATAAGTGCAAAGTACACATATATTACGATGCTGTCTATTAAATGCTTCTATATTATATA
TATAGTAATGTGTTTTATGGTGCACTCTCAGTACAATCTGCTCTGATGCCGATAGTTAA
GCCAGCCCCGACACCCGCCAACCCGCTGACGCGCCCTGACGGGCTTGTCTGCTCCCGG
CATCCGCTTACAGACAAGCTGTGACCGTCTCCGGGAGCTGCATGTGTGAGAGGTTTTAC
CGTCATCACGAAACGCGGAGACGAAAGGGCTCTGATACGCCTATTTTATAGTTA
ATGTCATGATAATAATGGTTTCTTAGGACGGATCGCTTGCCCTGTAACCTACACGCGCCTC
GTATCTTTTAAATGATGGAATAATTTGGGAATTTACTCTGTGTTTATTTATTTTATGTTT
TGTTTTGGATTTTAGAAAGTAAATAAAGAAGGTAGAAGAGTTACGGAATGAAGAAAAA
AAATAAACAAAGGTTTTAAAAAATTTCAACAAAAAGCGTACTTTACATATATATTTATTAG
ACAAGAAAAGCAGATTAAATAGATATACATTGATTAACGATAAGTAAATGTAAATCA
CAGGATTTTCGTGTGTGGTCTTCTACACAGACAAGATGAAACAATTCGGCATTAAATACCT
GAGAGCAGGAAGAGCAAGATAAAGGTAGTATTTGTTGGCGATCCCCCTAGAGTCTTTTA
CATCTTCGAAAAACAAAACTATTTTTCTTTAATTTCTTTTTTACTTTCTATTTTTAA
TTTTATATATTTATATTAATAAATTTAAATTATAATTATTTTATAGCACGTGATGAAAAG
GACCCAGGTGGCACTTTTCGGGAAATGTGCGGGAACCCCTATTTGTTTATTTTCTAA
ATACATTTCAAATATGTATCCGCTCATGAGACAATAACCCGATAAATGCTTCAATAATAT
TGAAAAAGGAAGAGTATGAGTATTCAACATTTCCGTGTGCGCCCTTATTCCTTTTTCGCG
GCATTTTGCCTTCTGTGTTTTGCTCACCAGAACGCTGGTGAAAGTAAAGATGCTGAA
GATCAGTTGGGTGCACGAGTGGGTACATCGAAGTGGATCTCAACAGCGGTAAGATCCTT
GAGAGTTTTCGCCCCGAAGAACGTTTTCCAATGATGAGCACTTTTAAAGTTCTGTATGT
GGCGCGGTATTTATCCCGTATTGACGCGGGCAAGAGCAACTCGGTGCGCGCATACACTAT
TCTCAGAATGACTTGGTTGAGTACTCACCAGTCACAGAAAAGCATCTTACGGATGGCATG
ACAGTAAGAGAATTATGCAGTGTGCCATAACCATGAGTGATAACACTGCGGCCAATCTTA
CTTCTGACAACGATCGGAGGACCGAAGGAGCTAACCGCTTTTTTCAACATGGGGGAT
CATGTAACCTCGCCTTGATCGTTGGGAACCGGAGCTGAATGAAGCCATACCAACGACGAG
CGTGACACCACGATGCCTGTAGCAATGGCAACAACGTTGCGCAAACTATTAACCTGGCGAA
CTACTTACTCTAGCTTCCCGGCAACAATTAATAGACTGGATGGAGGCGGATAAAGTTGCA
GGACCCTTCTGCGCTCGGCCCTTCCGGCTGGCTGGTTTATTGCTGATAAATCTGGAGCC
GGTGAGCGTGGGTCTCGCGGTATCATTGCAGCACTGGGGCCAGATGGTAAGCCCTCCCGT
ATCGTAGTTATCTACACGACGGGAGTCAGGCAACTATGGATGAACGAAATAGACAGATC
GCTGAGATAGGTGCCCTCACTGATTAAAGCATTGGTAAGTGTGAGACCAAGTTTACTCATAT
ATACTTTAGATTGATTAAAACTTCATTTTAAATTTAAAGGATCTAGGTGAAGATCCTT
TTTGATAATCTCATGACCAAAATCCCTTAACGTGAGTTTTCGTTCCACTGAGCGTCAGAC
CCCGTAGAAAAGATCAAAGGATCTTCTTGAGATCCTTTTTTCTGCGCGTAATCTGTGTC
TTGCAACAAAAAACCCGCTACCAGCGGTGGTTTGTGTTGCGGATCAAGAGCTACCA
ACTCTTTTCCGAAGGTAAGTGGCTTCAGCAGAGCGCAGATACCAATACTGTCTTTCTA
GTGTAGCCGTAGTTAGGCCACCACTTCAAGAACTCTGTAGCACCGCTACATACCTCGCT
CTGCTAATCCTGTTACCAAGTGGCTGCTGCCAGTGGCGATAAGTGTGCTTACCGGGTTG
GACTCAAGACGATAGTTACCGGATAAGGCGCAGCGGTGCGGCTGAACGGGGGGTTCTGTC-

Figure 98B

ACACAGCCCAGCTTGGAGCGAACGACCTACACCGAACTGAGATACCTACAGCGTGAGCAT
TGAGAAAGCGCCACGCTTCCCGAAGGGAGAAAGGCGGACAGGTATCCGGTAAGCGGCAGG
GTCGGAACAGGAGAGCGCACGAGGGAGCTTCCAGGGGGGAACGCCTGGTATCTTTATAGT
CCTGTGCGGGTTTCGCCACCTCTGACTTGAGCGTCGATTTTGTGATGCTCGTCAGGGGGG
CCGAGCCTATGGAAAAACGCCAGCAACGCGGCCCTTTTACGGTTCTGGCCTTTTGTCTGG
CCTTTTGCTCACATGTTCTTTCTGCGTTATCCCTGATTCTGTGGATAACCGTATTACC
GCCTTTGAGTGAGCTGATACCGCTCGCCGACGCGAACGACCGAGCGCAGCGAGTCAGTG
AGCGAGGAAGCGGAAGAGCGCCCAATACGCAAAACCGCCTCTCCCGCGCGTTGGCCGATT
CATTAAATGCAGCTGGCACGACAGGTTTCCCGACTGGAAAGCGGGCAGTGAGCGCAACGCA
ATTAATGTGAGTTACCTCACTCATTAGGCACCCAGGCTTTACACTTTATGCTTCCGGCT
CCTATGTTGTGTGAATTGTGAGCGGATAACAATTTACACAGGAAACAGCTATGACCATT
GATTACGCCAAGCTCGGAATTAACCTCACTAAAGGGAACAAAAGCTGGGTACCGGGCCC
CCCCTCGAGATCCGGGATCGAAGAAATGATGGTAAATGAAATAGGAAATCAAGGAGCATG
AAGGCAAAAGACAAATATAAGGGTCGAACGAAAAATAAAGTGAAAGTGTGTATATGATG
TATTTGGCTTTGCGGCGCCGAAAAACGAGTTTACGCAATTGCACAATCATGCTGACTCT
GTGGCGGACCCGCGCTCTTGCCGCGCCGCGGATAACGCTGGGCGTGAGGCTGTGCGCCGGC
GGAGTTTTTTGCGCCTGCATTTTCCAAGGTTTACCTGCGCTAAGGGGCGAGATTGGAGA
AGCAATAAGAATGCCGTTGGGGTTGGGATGATGACGACCACGCAACTGGTGTCTATTAT
TTAAGTTCCGGAAGAACCTGAGTGCAATTGCAACATGAGTATACTAGAAGAATGAGCCA
AGACTTGGGAGACGCGAGTTTGGCGGTGGTGCAACAATAGAGCGACCATGACCTTGAAG
GTGAGACCGGCATAACCGCTAGAGTACTTTGAAGAGGAAACAGCAATAGGGTTGTACCA
GTATAAATAGACAGGTACATACAACTGGAATGGTGTCTGTTTGTGAGTACGCTTTCAA
TTCAATTTGGGTGTGCACTTTATTTATGTTTACAATATGGAAGGGAACCTTTACACTTCTCCTA
TGCACATATATTAATTAAGTCCAATGCTAGTAGAGAAGGGGGTAACACCCCTCCGCGC
TCTTTTCCGATTTTTTTCTAAACCGTGGAATATTTGCGATATCCTTTTGTGTTTCCGGG
TGTAACAATATGGACTTCTCTTTTCTGGCAACCAACCCATACATCGGGATTCTATAAT
ACCTTCGTTGGTCTCCCTAACATGATAGGTGGCGGAGGGGAGATATACAATAGAACAGATA
CCAGACAAGACATAATGGGCTAAACAAGACTACACCAATTACACTGCCTCATTGATGGTG
GTACATAACGAACCTAATACTGTAGCCCTAGACTTGATAGCCATCATCATATCGAAGTTT
ACTACCCTTTTTCCATTTGCCATCTATTGAAGTAATAATAGGCGCATGCAACTTCTTTTC
TTTTTTTTTCTTTCTCTCTCCCCGTTGTTGTCTCACCATATCCGCAATGACAAAAAAA
ATGATGGAAGACACTAAAGGAAAAAATTAACGACAAAGACAGCACCAACAGATGTCTGTG
TTCCAGAGCTGATGAGGGGTATCTTGAACACACGAAACTTTTTCTTCTTCAATTCACG
CACACTACTCTAATGAGCAACGGTATACGGCCTTCTTCCAGTTACTTGAATTTGAAA
TAAAAAAGTTTGGCGCTTGTCTATCAAGTATAAATAGACCTGCAATATTAAATCTTTTG
TTTTCTTCGTCATTGTTCTCGTTCCCTTTCTTCTTCTTCTTTTCTGCAATATTTTCA
AGCTATACCAAGCATACAATCAACTCCAAGCTTATGCCCAAGAAGAAGCGGAAGGTCTCG
AGCGGCGCCAATTTAATCAAAAGTGGGAATATTGCTGATAGCTCATTGTCTTCACTTTC
ACTAACAGTAGCAACGGTCCGAACCTCATAACAACCTCAAACAAATTCTCAAGCGCTTTCA
CAACCAATTGCCCTCCTCTAACGTTTCATGATACTTCATGAATAATGAAATCACGGCTAGT
AAAATTGATGATGGTAATAATTCAAACCACTGTCACTGGTTGGACGGACCAAACTGCG
TATAACGCGTTTGAATCACTACAGGGATGTTTAAATACCACTACAATGGATGATGATAT
AACTATCTATTTCGATGATGAAGATACCCACCAACCAAAAAAGAGGGTGGGTGATC
ACAAGTTGTACAAAAAGCAGGCTTGTGACCCCGGGAATTGAGATCTACTAGTGCGGC
CGCACGCGTACCCAGCTTTCTGTACAAAGTGGTGACGTCGAGCTCCCTATAGTGAGTCG
TATTACACTGGCCGTCTGTTTACAACGTCGTGACTGGGAAAAACACCGGTGAGCTCTAAGT
AAGTAACGGCCGCCACCGCGGTGGAGCTTTGACTTCTTCGCCAGAGGTTTGGTCAAGTC
TCCAATCAAGGTTGTGCGCTTGTCTACCTTGGCAGAAATTTACGAAAAGATGGAAGGG
TCAAATCGTTGGTAGATACGTTGTTGACACTTCTAAATAAGCGAATTCTTATGATTTAT
GATTTTTATTATTAAATAAGTTATAAAAAAATAAGTGATACAAATTTAAAGTGAATC
TTAGGTTTTAAACGAAATTTCTGTTCTTGAGTAACTCTTCTGTAGGTGAGGTGCT
TTCTCAGGTATAGCATGAGGTGCTCTTATTGACCACACCTTACCGCATGCCGAGCAA
ATGCTTGCAAATCGCTCCCCATTTACCCCAATTGTAGATATGCTAACTCCAGCAATGAGT
TGATGAATCTCGGTGTGATTTTATGTCTCAGAGGACAATACCTGTTGTAATCGTTCTT
CCACACGGATCCGCATCAGGCGAAATTGTAAACGTTAATTTTTGTTAAATTCGCGTTA
AATATTTGTTAAATCAGCTCATTTTTTAACCAATAGGCCGAAATCGGCAAAATCCCTTAT
AAATCAAAAGAATAGACCGAGATAGGGTTGAGTGTGTTCCAGTTTGAACAAGAGTCCA
CTATTAAAGAACGTGGACTCCAACGTCAAAGGGCGAAAAACCGTCTATCAGGGCGATGGC-

FIGURE 98C

CCACTACGTGAACCATCACCCCTAATCAAGTTTTTTGGGGTCGAGGTGCCGTAAAGCACTA
AATCGGAACCCCTAAAGGGAGCCCCGATTTAGAGCTTGACGGGGAAAGCCGGCGAACGTG
GCGAGAAAGGAAGGGAAGAAAGCGAAAGGAGCGGGCGCTAGGGCGCTGGCAAGTGTAGCG
GTCACGCTGCGCGTAACCACACCCGCGCGCTTAATGCGCCGCTACAGGGCGCGTCC
CATTCGCCATTCACTGCA

FIGURE 98D

237/240

pMAB86

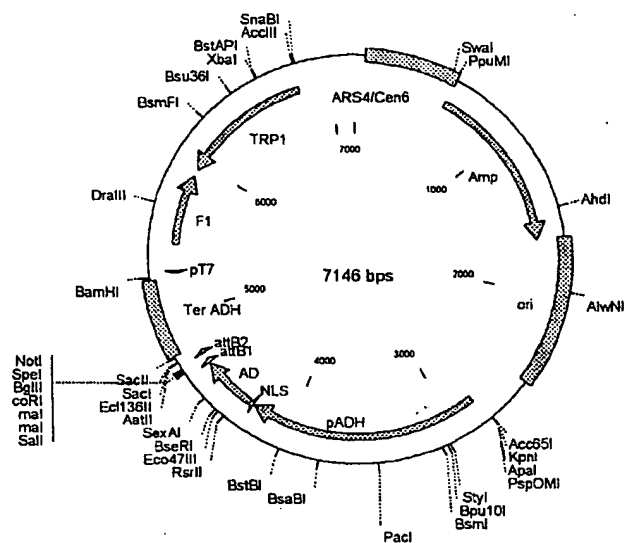


FIGURE 99A

pMAB86 7146 bp

GACGAAAGGGCCTCGTGATACGCCTATTTTATAGGTTAATGTCATGATAATAATGGTTT
CTTAGGACGGATCGCTTGCTGTAACTTACACGCGCTCGTATCTTTTAAATGATGGAATA
ATTTGGGAATTTACTCTGTGTTTATTTATTTTATGTTTGTATTTGGATTTTAGAAAAGT
AAATAAAGAAGGTAGAAGAGTTACGGAATGAAGAAAAAATAAACAAAGGTTTAAAAA
ATTTCAACAAAAGCGTACTTTACATATATATTTATTAGACAAGAAAAGCAGATTAAATA
GATATACATTCGATTAAACGATAAGTAAATGTAAATCACAGGATTTTCGTGTGTGGTCT
TCTACACAGACAAGATGAAACAATTCGGCATTAACTCTGAGAGCAGGAAGAGCAAGATA
AAAGGTAGTATTTGTTGGCGATCCCCCTAGAGTCTTTACATCTTCGGAAAACAAAACT
ATTTTTTCTTAAATTTCTTTTTTACTTTTCTATTTTAAATTTATATATTTATATTAAAA
ATTTAAATTATAATTATTTTATAGCACGTGATGAAAAGGACCCAGGTGGCAGTTTTCGG
GGAAATGTGCGCGGAACCCCTATTGTTTATTTTCTAAATACATTCAAATATGTATCCG
CTCATGAGACAATAACCTGATAAATGCTTCAATAATTGAAAAAGGAAGATGAGT
ATTCAACATTTCCGTGTGCGCCCTTATTCCTTTTTCGCGGCATTTGCGCTCCTGTTTT
GCTCACCCAGAAAACGCTGGTGAAAGTAAAGATGCTGAAGATCAGTTGGGTGCACGAGTG
GGTTACATCGAACTGGATCTCAACAGCGGTAAAGATCCTTGAGAGTTTTCGCCCCGAAGAA
CGTTTCCAAATGATGAGCACTTTTAAAGTTCTGCTATGTGGCGCGGTATTATCCCGTATT
GACGCGGGGCAAGAGCAACTCGGTGCGCGCATACACTATTCTCAGAATGACTTGGTTGAG
TACTCACCAGTCACAGAAAAGCATCTTACGATGGCATGACAGTAAGAGAATTATGCACT
GCTGCCATAACCATGAGTGATAACACTGCGGCCAACTTACTTCTGACAACGATCGGAGGA
CCGAAGGAGCTAACCGCTTTTTTTCACAACATGGGGGATCATGTAACCTCGCCTTGATCGT
TGGAACCGGAGCTGAATGAAGCCATACCAACGACGAGCGTGACACCAGATGCCTGTA
GCAATGGCAACAACGTTGCGCAAACTATTAACTGGCGAACTACTTACTTAGCTTCCCG
CAACAATTAATAGACTGGATGGAGGCGGATAAAGTTGCAGGACCACTTCTGCGCTCGGCC
CTTCCGGCTGGCTGGTTTATTGCTGATAAATCTGGAGCCGGTGAGCGTGGGTCTCGCGGT
ATCATTGAGCACTGGGCGCAGATGGTAAGCCCTCCCGTATCGTAGTTATCTACACGACG
GGCAGTCAGGCAACTATGGATGAACGAAATAGACAGATCGCTGAGATAGGTGCCCTCACTG
ATTAAGCATTGGTAACTGTGAGACCAAGTTTACTCATATATACCTTTAGATTGATTTAAAA
CTTCATTTTTTAAATTTAAAGGATCTAGGTGAAGATCCTTTTGTATAATCTCATGACCAAA
ATCCTTTAACGTGAGTTTTCGTTTCCACTGAGCGTCAGACCCCGTAGAAAAGATCAAAGGA
TCTTCTTGAGATCCTTTTTTCTGCGCGTAATCTGCTGCTTGCAAAACAAAAAACACCG
CTACCAGCGGTGGTTTGTGTCGGGATCAAGAGCTACCAACTCTTTTCCGAAGGTAAC
GGCTTCAGCAGAGCGCAGATACCAATACTGTCTTCTAGTGTAGCCGTAGTTAGGCCAC
CACTTCAAGAACTCTGTAGCACCGCCTACATACCTCGCTCTGCTAATCCTGTTACCACTG
GCTGCTGCCAGTGGCGATAAGTCTGTCTTACCGGGTTGGACTCAAGACGATAGTTACCG
GATAAGGCGCAGCGGTGCGGCTGAACGGGGGTTCTGTCACACAGCCAGCTTGGAGCGA
ACGACCTACACCGAACTGAGATACCTACAGCGTGAGCATTGAGAAAGCGCCACGCTTCCC
GAAGGGAGAAAGGCGGACAGGTATCCGTAAGCGGCAGGCTCGGAACAGGAGAGCGCACG
AGGGAGCTTCCAGGGGGGAACGCCTGGTATCTTTATAGTCCTGTGCGGTTTCGCCACCTC
TGACTTGAGCGTCGATTTTGTGATGCTCGTCAGGGGGGCCGAGCCTATGGA AAAACGCC
AGCAACGCGGCCTTTTTACGGTTCTTGGCCTTTTGCTGGCCTTTTGCTCACATGTTCTTT
CCTGCGTTATCCCTGATTCTGTGGATAACCGTATTACCGCCTTTGAGTGAGCTGATACC
GCTCGCCGAGCCGAACGACCGAGCGCAGCGAGTCAGTGAGCGAGGAAGCGGAAGAGCGC
CCAATACGCAAAACCGCCTCTCCCGCGCGTTGGCCGATTCAATTAATGACGCTGGCACGAC
AGGTTTCCCGACTGGAAAGCGGGCAGTGAGCGCAACGCAATTAATGTGAGTTACCTCACT
CATTAGGCACCCAGGCTTTACACTTTATGCTTCCGGCTCCTATGTTGTGTGGAATTGTG
AGCGGATAACAAATTTACACAGGAAACAGCTATGACCATGATTACGCCAAGCTCGGAATT
AACCCTCACTAAAGGGGAACAAAAGCTGGGTACCGGGCCCCCTCGAGATCCGGGATCGA
AGAAATGATGGTAAATGAAATAGGAAATCAAGGAGCATGAAGGCAAAAGACAAATATAAG
GGTCGAACGAAAAATAAAGTGAAAAGTGTGATATGATGATTTTGGCTTTGCGGCGCCGA
AAAAACGAGTTTACGCAATTGCACAATCATGCTGACTCTGTGGCGGACCCGCGCTCTTGC
CGGCCCGGCGATAACGCTGGGCGTGAGGCTGTGCCCGCGGAGTTTTCGCGCTGCAATT
TTCCAAGGTTTACCCTGCGCTAAGGGGCGAGATTGGAGAAGCAATAAGAAATGCCGGTTGG
GGTTGCGATGATGACGACCACGACAACCTGGTGTCAATTATTTAAGTTGCCGAAAGAACCTG
AGTGCAATTTGCAACATGAGTATACTAGAAGAAATGAGCCAAGACTTGCAGACCGGAGTTT
GCCGTTGGTGCAACAATAGAGCGACCATGACCTTGAAGGTGAGACGCGCATAACCGCTA-

FIGURE 99B

GAGTACTTTGAAGAGGAAACAGCAATAGGGTTGCTACCACTATAAATAGACAGGTACATA
CAACACTGGAAATGGTTGTCTGTTTGTAGTACGCTTCAATTCATTGGGGTGTCACTTTA
TTATGTTACAATATGGAAGGGAACCTTACACTTCTCCTATGCACATATATTAATTAAGT
CCAATGCTAGTAGAGAAGGGGGTAAACCCCTCCGCGCTCTTTTCCGATTTTTTCTAA
ACCGTGGAAATATTCGGATATCCTTTTGTGTTTCCGGGTGTACAATATGGACTTCTCT
TTTCTGGCAACCAACCCATACATCGGGATTCTATAATACCTTCGTTGGTCTCCCTAAC
ATGTAGGTGGCGGAGGGGAGATATACAATAGAACAGATACCAGACAAGACATAATGGGCT
AAACAAGACTACACCAATTACACTGCCTCATTTGATGGTGGTACATAACGAACATAACTG
TAGCCCTAGACTTGATAGCCATCATCATATCGAAGTTTCACTACCCTTTTTTCATTGGCC
ATCTATTGAAGTAATAATAGGCGCATGCAACTTCTTTCTTTTTTTTTCTTTCTCTCTC
CCCCGTTGTTGTCTCACCATATCCGCAATGACAAAAAATGATGGAAGACACTAAAGGA
AAAAATTAACGACAAAGACAGCACCAACAGATGTCGTTGTTCCAGAGCTGATGAGGGTA
TCTTCGAACACACGAACTTTTTCTTCTTCTTCACTTACGACACTACTCTCTAATGAGCA
ACGGTATACGGCTTCTTCCAGTTACTTGAATTTGAAATAAAAAAGTTTGGCCGCTTTG
CTATCAAGTATAAATAGACCTGCAATTATTAATCTTTTGTTCCTCGTCATTGTTCTCGT
TCCCTTCTTCTTCTTCTTTCTTTTCTGCACAATATTTCAAGCTATACCAAGCATACAATC
AATCCAAGCTTATGCCCCAAGAAGCGGAAGGTCTCGAGCGGCGCAATTTTAATCAA
AGTGGGAATATTGCTGATAGCTCATTGTCCTTCACTTCACTAACAGTAGCAACGGTCCG
AACCTGATAACAACTCAAAACAAATTCTCAAGCGCTTTCACAACCAATTGCCTCTCTAAC
GTTTCATGATAACTTCATGAATAATGAAATCACGGCTAGTAAAATTGATGATGGTAATAAT
TCAAAACCACTGTCACTGGTTGGACGGACCAACTGCGTATAACGGCTTTGGAATCACT
ACAGGGATGTTTAAATACCACTACAATGGATGATGTATATACTATCTATTGATGAGAA
GATACCCCAACCAACCAAAAAAAGAGGGTGGGTGATCACAAGTTTGTACAAAAAGCA
GGCTTGTGACCCCGGGAATTGAGATCTACTAGTGGCGGCGCAGCGTACCCAGCTTTCT
TGTACAAAGTGGTGACGTCGAGCTCTAAGTAAGTAACGGCCGCCACCGGGTGGAGCTTT
GGACTTCTTCCGCGAGGTTTGGTCAAGTCTCAATCAAGGTTGTCGGCTTGTCTACCTT
GCCAGAAATTTACGAAAAGATGGAAGGGTCAAATCGTTGGTAGATACGTTGTTGACAC
TTCTAAATAAGCGAATTTCTTATGATTTATGATTTTATTATTAAATAAGTTATAAAAA
AATAAGTGTATACAAATTTTAAAGTGACTCTTAGGTTTAAACGAAAAATTTCTTCTT
GAGTAATCTTTCTGTAGGTGAGGTTGCTTTCTCAGGTATAGCATGAGGTGCTCTTAT
TGACCACACCTCTACCGGCATGCGGAGCAATGCCTGCAATCGCTCCCATTTACCCCA
ATTGATAGATATGCTAACTCCAGCAATGAGTTGATGAATCTCGGTGTGATTTTATGCTCT
CAGAGGACAATACCTGTTGTAATCGTTCTTCCACACGGATCCCAATTCGCCCTATAGTGA
GTCGTATTACAATTCACTGGCCGTCGTTTACAACGTCGTCGAGTGGGAAACCCCTGGCGT
TACCCAACTTAATCGCTTGCAGCACATCCCCCTTTCGCCAGCTGGCGTAATAGCGAAGA
GGCCCGCACCGATCGCCCTTCCCAACAGTTGCGCAGCTGAATGGCGAATGGACGGCCC
TGTAGCGGCGCATTAAAGCGCGCGGGGTGTGGTGGTTACGCGCAGCGTGACCGCTACACTT
GCCAGCGCCCTAGCGCCGCTCCTTTCCGCTTCTTCCCTTCTTCTCGCCACGTTCCGCC
GGCTTTCCCGTCAAGCTCTAAATCGGGGGCTCCCTTAGGGTTCCGATTAGTGCTTTA
CGGCACCTCGACCCCAAAAACTTGATTAGGGTGATGGTTACAGTAGTGGGCCATCGCCC
TGATAGACGGTTTTTCCGCCCTTTGACGTTGGAGTCCAGTTCTTTAATAGTGGACTCTTG
TTCCAACTGGAACAACACTCAACCTATCTCGGTCTATTCTTTGATTTATAAGGGATT
TTGCCGATTTCCGCCCTATTGGTTAAAAAATGAGCTGATTTAACAAAAATTTAACCGGAAT
TTAACAAAAATATTACGTTTACAATTTCTTGATGCGGTATTTTCTCCTTACGCATCTGT
GCGGTATTTACACCGCAGGCAAGTGCACAAACAATACTTAAATAAATACTACTCAGTAA
TAACCTATTTCTTAGCATTTTGTACGAAATTTGCTATTTGTTAGAGTCTTTTACCCAT
TTGTCTCCACACCTCCGCTTACATCAACACCAATAACGCCATTTAATCTAAGCGCATCAC
CAACATTTCTGGCGTCAGTCCACCAGCTAACATAAAATGTAAGCTTTCGGGGCTCTCTT
GCCTTCCAACCCAGTCAGAAATCGAGTTCCAATCCAAAAGTTCACTGTCCACCTGCTT
CTGAATCAAACAAGGGAATAACGAATGAGGTTTCTGTGAAGCTGCACTGAGTAGTATGT
TGCAGTCTTTTGGAAATACGAGTCTTTAATAACTGGCAAACCGAGGAACCTTGGTATT
CTTGCCCAAGACTCATCTCCATGCAAGTTGGACGATATCAATGCCGTAATCATTGACAGAG
CCAAAACATCTCCTTAGGTTGATTACGAACACGCCAACCAAGTATTTCCGAGTGCCTG
AACTATTTTATATGCTTTTACAAGACTTGAATTTTCTTGCAATAACCGGGTCAATTG
TTCTCTTTCTATTGGGCACACATATAATACCCAGCAAGTCAGCATCGGAATCTAGAGCAC
ATTCTGCGGCTCTGTGCTCTGCAAGCCGCAAACTTTACCAATGGACAGAACTACCTG
TGAATTAATAACAGACATACTCCAAGCTGCCTTTGTGTGCTTAATCACGTATACTCAG
TGCTCAATAGTACCAATGCCCTCCCTCTTGGCCCTCTCTTTTCTTTTTCGACCGAAT-

FIGURE 99C

TAATTCCTTAATCGGCAAAAAAGAAAAGCTCCGGATCAAGATTGTACGTAAGGTGACAAG
CTATTTTTCAATAAAGAATATCTTCCACTACTGCCATCTGGCGTCATAACTGCAAAGTAC
ACATATATTACGATGCTGTCTATTAAATGCTTCTATATTATATATATAGTAATGTCGTT
TATGGTGCACTCTCAGTACAATCTGCTCTGATGCCGCATAGTTAAGCCAGCCCCGACACC
CGCCAAACACCCGCTGACGCGCCCTGACGGGCTTGTCTGCTCCCGGCATCCGCTTACAGAC
AAGCTGTGACCGTCTCCGGGAGCTGCATGTGTCAGAGGTTTTCACCGTCATCACCAGAAAC
GCGCGA

FIGURE 99D

INDICATIONS RELATING TO A DEPOSITED MICROORGANISM
(PCT Rule 13bis)

REC'D

A. The indications made below relate to the microorganism referred to in the description on page <u>54</u> , line <u>8</u> .	
B. IDENTIFICATION OF DEPOSIT Further deposits are identified on an additional sheet <input checked="" type="checkbox"/>	
Name of depositary institution Agricultural Research Culture Collection (NRRL) International Depository Authority	
Address of depositary institution (including postal code and country) 1815 N. University Street Peoria, Illinois 61604 United States of America	
Date of deposit February 27, 1999	Accession Number NRRL B-30103
C. ADDITIONAL INDICATIONS (leave blank if not applicable) This information is continued on an additional sheet <input type="checkbox"/>	
Escherichia coli DB3.1(pEZC15101) In respect of those designations in which a European Patent is sought a sample of the deposited microorganism will be made available until the publication of the mention of the grant of the European patent or until the date on which the application has been refused or withdrawn or is deemed to be withdrawn, only by the issue of such a sample to an expert nominated by the person requesting the sample (Rule 28(4) EPC).	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)	
E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)	
The indications listed below will be submitted to the international Bureau later (specify the general nature of the indications, e.g., "Accession Number of Deposit")	

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INDICATIONS RELATING TO A DEPOSITED MICROORGANISM
(PCT Rule 13bis)

A. The indications made below relate to the microorganism referred to in the description on page <u>55</u> , line <u>16</u> .	
B. IDENTIFICATION OF DEPOSIT Further deposits are identified on an additional sheet <input checked="" type="checkbox"/>	
Name of depositary institution Agricultural Research Culture Collection (NRRL) International Depository Authority	
Address of depositary institution (including postal code and country) 1815 N. University Street Peoria, Illinois 61604 United States of America	
Date of deposit February 27, 1999	Accession Number NRRL B-30100
C. ADDITIONAL INDICATIONS (leave blank if not applicable) This information is continued on an additional sheet <input type="checkbox"/>	
Escherichia coli DB3.1(pENTR-1A) In respect of those designations in which a European Patent is sought a sample of the deposited microorganism will be made available until the publication of the mention of the grant of the European patent or until the date on which the application has been refused or withdrawn or is deemed to be withdrawn, only by the issue of such a sample to an expert nominated by the person requesting the sample (Rule 28(4) EPC).	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)	
E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)	
The indications listed below will be submitted to the international Bureau later (specify the general nature of the indications, e.g., "Accession Number of Deposit")	

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INDICATIONS RELATING TO A DEPOSITED MICROORGANISM
(PCT Rule 13bis)

A. The indications made below relate to the microorganism referred to in the description on page <u>55</u> , line <u>16</u> .	
B. IDENTIFICATION OF DEPOSIT Further deposits are identified on an additional sheet <input checked="" type="checkbox"/>	
Name of depositary institution Agricultural Research Culture Collection (NRRL) International Depository Authority	
Address of depositary institution (including postal code and country) 1815 N. University Street Peoria, Illinois 61604 United States of America	
Date of deposit February 27, 1999	Accession Number NRRL B-30102
C. ADDITIONAL INDICATIONS (leave blank if not applicable) This information is continued on an additional sheet <input type="checkbox"/>	
Escherichia coli DB3.1(pENTR-3C) In respect of those designations in which a European Patent is sought a sample of the deposited microorganism will be made available until the publication of the mention of the grant of the European patent or until the date on which the application has been refused or withdrawn or is deemed to be withdrawn, only by the issue of such a sample to an expert nominated by the person requesting the sample (Rule 28(4) EPC).	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)	
E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)	
The indications listed below will be submitted to the international Bureau later (specify the general nature of the indications, e.g., "Accession Number of Deposit")	

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INDICATIONS RELATING TO A DEPOSITED MICROORGANISM
(PCT Rule 13bis)

A. The indications made below relate to the microorganism referred to in the description on page <u>55</u> , line <u>16</u> .	
B. IDENTIFICATION OF DEPOSIT Further deposits are identified on an additional sheet <input checked="" type="checkbox"/>	
Name of depositary institution Agricultural Research Culture Collection (NRRL) International Depository Authority	
Address of depositary institution (including postal code and country) 1815 N. University Street Peoria, Illinois 61604 United States of America	
Date of deposit February 27, 1999	Accession Number NRRL B-30101
C. ADDITIONAL INDICATIONS (leave blank if not applicable) This information is continued on an additional sheet <input type="checkbox"/>	
Escherichia coli DB3.1(pENTR-2B) In respect of those designations in which a European Patent is sought a sample of the deposited microorganism will be made available until the publication of the mention of the grant of the European patent or until the date on which the application has been refused or withdrawn or is deemed to be withdrawn, only by the issue of such a sample to an expert nominated by the person requesting the sample (Rule 28(4) EPC).	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (If the indications are not for all designated States)	
E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)	
The indications listed below will be submitted to the international Bureau later (specify the general nature of the indications, e.g., "Accession Number of Deposit")	

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INDICATIONS RELATING TO A DEPOSITED MICROORGANISM
(PCT Rule 13bis)

A. The indications made below relate to the microorganism referred to in the description on page <u>20-21</u> WIPO PCT	
B. IDENTIFICATION OF DEPOSIT Further deposits are identified on an additional sheet <input checked="" type="checkbox"/>	
Name of depositary institution Agricultural Research Culture Collection (NRRL) International Depository Authority	
Address of depositary institution (including postal code and country) 1815 N. University Street Peoria, Illinois 61604 United States of America	
Date of deposit February 27, 1999	Accession Number NRRL B-30108
C. ADDITIONAL INDICATIONS (leave blank if not applicable) This information is continued on an additional sheet <input type="checkbox"/>	
Escherichia coli DB10B(pCMVSPORT6) In respect of those designations in which a European Patent is sought a sample of the deposited microorganism will be made available until the publication of the mention of the grant of the European patent or until the date on which the application has been refused or withdrawn or is deemed to be withdrawn, only by the issue of such a sample to an expert nominated by the person requesting the sample (Rule 28(4) EPC).	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)	
E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)	
The indications listed below will be submitted to the international Bureau later (specify the general nature of the indications, e.g., "Accession Number of Deposit")	

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INDICATIONS RELATING TO A DEPOSITED MICROORGANISM
(PCT Rule 13bis)

A. The indications made below relate to the microorganism referred to in the description on page <u>54</u> , line <u>9</u>	
B. IDENTIFICATION OF DEPOSIT Further deposits are identified on an additional sheet <input checked="" type="checkbox"/>	
Name of depositary institution Agricultural Research Culture Collection (NRRL) International Depository Authority	
Address of depositary institution (including postal code and country) 1815 N. University Street Peoria, Illinois 61604 United States of America	
Date of deposit February 27, 1999	Accession Number NRRL B-30105
C. ADDITIONAL INDICATIONS (leave blank if not applicable) This information is continued on an additional sheet <input type="checkbox"/>	
Escherichia coli DB3.1(pEZC15103) In respect of those designations in which a European Patent is sought a sample of the deposited microorganism will be made available until the publication of the mention of the grant of the European patent or until the date on which the application has been refused or withdrawn or is deemed to be withdrawn, only by the issue of such a sample to an expert nominated by the person requesting the sample (Rule 28(4) EPC).	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)	
E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)	
The indications listed below will be submitted to the international Bureau later (specify the general nature of the indications, e.g., "Accession Number of Deposit")	

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INDICATIONS RELATING TO A DEPOSITED MICROORGANISM
(PCT Rule 13bis)

A. The indications made below relate to the microorganism referred to in the description on page <u>54</u> , line <u>9</u> .	
B. IDENTIFICATION OF DEPOSIT Further deposits are identified on an additional sheet <input checked="" type="checkbox"/>	
Name of depositary institution Agricultural Research Culture Collection (NRRL) International Depository Authority	
Address of depositary institution (including postal code and country) 1815 N. University Street Peoria, Illinois 61604 United States of America	
Date of deposit February 27, 1999	Accession Number NRRL B-30104
C. ADDITIONAL INDICATIONS (leave blank if not applicable) This information is continued on an additional sheet <input type="checkbox"/>	
Escherichia coli DB3.1(pEZC15102) In respect of those designations in which a European Patent is sought a sample of the deposited microorganism will be made available until the publication of the mention of the grant of the European patent or until the date on which the application has been refused or withdrawn or is deemed to be withdrawn, only by the issue of such a sample to an expert nominated by the person requesting the sample (Rule 28(4) EPC).	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)	
E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)	
The indications listed below will be submitted to the international Bureau later (specify the general nature of the indications, e.g., "Accession Number of Deposit")	

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INDICATIONS RELATING TO A DEPOSITED MICROORGANISM
(PCT Rule 13bis)

A. The indications made below relate to the microorganism referred to in the description on page <u>52</u> , line <u>31</u> .	
B. IDENTIFICATION OF DEPOSIT Further deposits are identified on an additional sheet <input checked="" type="checkbox"/>	
Name of depositary institution Agricultural Research Culture Collection (NRRL) International Depository Authority	
Address of depositary institution (including postal code and country) 1815 N. University Street Peoria, Illinois 61604 United States of America	
Date of deposit February 27, 1999	Accession Number NRRL B-30099
C. ADDITIONAL INDICATIONS (leave blank if not applicable) This information is continued on an additional sheet <input type="checkbox"/>	
Escherichia coli DB3.1(pAHPKan) or Escherichia coli DB3.1(pAttPKan) In respect of those designations in which a European Patent is sought a sample of the deposited microorganism will be made available until the publication of the mention of the grant of the European patent or until the date on which the application has been refused or withdrawn or is deemed to be withdrawn, only by the issue of such a sample to an expert nominated by the person requesting the sample (Rule 28(4) EPC).	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)	
E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)	
The indications listed below will be submitted to the international Bureau later (specify the general nature of the indications, e.g., "Accession Number of Deposit")	

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Escherichia coli DB3.1(pENTR-3C)

ICELAND

The applicant hereby requests that, until the application has been laid open to public inspection (by the Icelandic Patent Office), or has been finally decided upon by the Icelandic Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art.

NETHERLANDS

The applicant hereby requests that until the date of a grant of a Netherlands patent or until the date on which the application is refused or withdrawn or lapsed, the microorganism shall be made available as provided in Rule 31F(1) of the Patent Rules only by the issue of a sample to an expert. The request to this effect must be furnished by the applicant with the Netherlands Industrial Property Office before the date on which the application is made available to the public under Section 22C or Section 25 of the Patents Act of the Kingdom of the Netherlands, whichever of the two dates occurs earlier.

NORWAY

The applicant hereby requests that, until the application has been laid open to public inspection (by the Norwegian Patent Office), or has been finally decided upon by the Norwegian Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the Norwegian Patent Office not later than at the time when the application is made available to the public under Sections 22 and 33(3) of the Norwegian Patents Act. If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the Norwegian Patent office or any person approved by the applicant in the individual case.

SINGAPORE

The applicant hereby requests that the furnishing of a sample of a microorganism shall only be made available to an expert. The request to this effect must be filed by the applicant with the International Bureau before the completion of the technical preparations for international publication of the application.

Escherichia coli DB3.1(pENTR-3C)

SWEDEN

The applicant hereby requests that, until the application has been laid open to public inspection (by the Swedish Patent Office), or has been finally decided upon by the Swedish Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the International Bureau before the expiration of 16 months from the priority date (preferably on the Form PCT/RO/134 reproduced in annex Z of Volume I of the PCT Applicant's Guide). If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the Swedish Patent office or any person approved by the applicant in the individual case.

UNITED KINGDOM

The applicant hereby requests that the furnishing of a sample of a microorganism shall only be made available to an expert. The request to this effect must be filed by the applicant with the International Bureau before the completion of the technical preparations for international publication of the application.

Escherichia coli DB3.1(pENTR-2B)

AUSTRALIA

The applicant hereby gives notice that the furnishing of a sample of a microorganism shall only be effected prior to the grant of a patent, or prior to the lapsing, refusal or withdrawal of the application, to a person who is a skilled addressee without an interest in the invention (Regulation 3.25(3) of the Australian Patents Regulations).

CANADA

The applicant hereby requests that, until either a Canadian patent has been issued on the basis of the application or the application has been refused, or is abandoned and no longer subject to reinstatement, or is withdrawn, the furnishing of a sample of deposited biological material referred to in the application only be effected to an independent expert nominated by the Commissioner of Patents.

DENMARK

The applicant hereby requests that, until the application has been laid open to public inspection (by the Danish Patent Office), or has been finally decided upon by the Danish Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the Danish Patent Office not later than at the time when the application is made available to the public under Sections 22 and 33(3) of the Danish Patents Act. If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the Danish Patent office or any person approved by the applicant in the individual case.

FINLAND

The applicant hereby requests that, until the application has been laid open to public inspection (by the National Board of Patents and Registration), or has been finally decided upon by the National Board of Patents and Registration without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the International Bureau before the expiration of 16 months from the priority date (preferably on the Form PCT/RO/134 reproduced in annex Z of Volume I of the PCT Applicant's Guide). If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the National Board of Patents and Registration or any person approved by the applicant in the individual case.

Escherichia coli DB3.1(pENTR-2B)

ICELAND

The applicant hereby requests that, until the application has been laid open to public inspection (by the Icelandic Patent Office), or has been finally decided upon by the Icelandic Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art.

NETHERLANDS

The applicant hereby requests that until the date of a grant of a Netherlands patent or until the date on which the application is refused or withdrawn or lapsed, the microorganism shall be made available as provided in Rule 31F(1) of the Patent Rules only by the issue of a sample to an expert. The request to this effect must be furnished by the applicant with the Netherlands Industrial Property Office before the date on which the application is made available to the public under Section 22C or Section 25 of the Patents Act of the Kingdom of the Netherlands, whichever of the two dates occurs earlier.

NORWAY

The applicant hereby requests that, until the application has been laid open to public inspection (by the Norwegian Patent Office), or has been finally decided upon by the Norwegian Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the Norwegian Patent Office not later than at the time when the application is made available to the public under Sections 22 and 33(3) of the Norwegian Patents Act. If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the Norwegian Patent office or any person approved by the applicant in the individual case.

SINGAPORE

The applicant hereby requests that the furnishing of a sample of a microorganism shall only be made available to an expert. The request to this effect must be filed by the applicant with the International Bureau before the completion of the technical preparations for international publication of the application.

Escherichia coli DB3.1(pENTR-2B)**SWEDEN**

The applicant hereby requests that, until the application has been laid open to public inspection (by the Swedish Patent Office), or has been finally decided upon by the Swedish Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the International Bureau before the expiration of 16 months from the priority date (preferably on the Form PCT/RO/134 reproduced in annex Z of Volume I of the PCT Applicant's Guide). If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the Swedish Patent office or any person approved by the applicant in the individual case.

UNITED KINGDOM

The applicant hereby requests that the furnishing of a sample of a microorganism shall only be made available to an expert. The request to this effect must be filed by the applicant with the International Bureau before the completion of the technical preparations for international publication of the application.

Escherichia coli DB3.1(pENTR-1A)

AUSTRALIA

The applicant hereby gives notice that the furnishing of a sample of a microorganism shall only be effected prior to the grant of a patent, or prior to the lapsing, refusal or withdrawal of the application, to a person who is a skilled addressee without an interest in the invention (Regulation 3.25(3) of the Australian Patents Regulations).

CANADA

The applicant hereby requests that, until either a Canadian patent has been issued on the basis of the application or the application has been refused, or is abandoned and no longer subject to reinstatement, or is withdrawn, the furnishing of a sample of deposited biological material referred to in the application only be effected to an independent expert nominated by the Commissioner of Patents.

DENMARK

The applicant hereby requests that, until the application has been laid open to public inspection (by the Danish Patent Office), or has been finally decided upon by the Danish Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the Danish Patent Office not later than at the time when the application is made available to the public under Sections 22 and 33(3) of the Danish Patents Act. If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the Danish Patent office or any person approved by the applicant in the individual case.

FINLAND

The applicant hereby requests that, until the application has been laid open to public inspection (by the National Board of Patents and Registration), or has been finally decided upon by the National Board of Patents and Registration without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the International Bureau before the expiration of 16 months from the priority date (preferably on the Form PCT/RO/134 reproduced in annex Z of Volume I of the PCT Applicant's Guide). If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the National Board of Patents and Registration or any person approved by the applicant in the individual case.

Escherichia coli DB3.1(pENTR-1A)

ICELAND

The applicant hereby requests that, until the application has been laid open to public inspection (by the Icelandic Patent Office), or has been finally decided upon by the Icelandic Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art.

NETHERLANDS

The applicant hereby requests that until the date of a grant of a Netherlands patent or until the date on which the application is refused or withdrawn or lapsed, the microorganism shall be made available as provided in Rule 31F(1) of the Patent Rules only by the issue of a sample to an expert. The request to this effect must be furnished by the applicant with the Netherlands Industrial Property Office before the date on which the application is made available to the public under Section 22C or Section 25 of the Patents Act of the Kingdom of the Netherlands, whichever of the two dates occurs earlier.

NORWAY

The applicant hereby requests that, until the application has been laid open to public inspection (by the Norwegian Patent Office), or has been finally decided upon by the Norwegian Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the Norwegian Patent Office not later than at the time when the application is made available to the public under Sections 22 and 33(3) of the Norwegian Patents Act. If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the Norwegian Patent office or any person approved by the applicant in the individual case.

SINGAPORE

The applicant hereby requests that the furnishing of a sample of a microorganism shall only be made available to an expert. The request to this effect must be filed by the applicant with the International Bureau before the completion of the technical preparations for international publication of the application.

Escherichia coli DB3.1(pENTR-1A)

SWEDEN

The applicant hereby requests that, until the application has been laid open to public inspection (by the Swedish Patent Office), or has been finally decided upon by the Swedish Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the International Bureau before the expiration of 16 months from the priority date (preferably on the Form PCT/RO/134 reproduced in annex Z of Volume I of the PCT Applicant's Guide). If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the Swedish Patent office or any person approved by the applicant in the individual case.

UNITED KINGDOM

The applicant hereby requests that the furnishing of a sample of a microorganism shall only be made available to an expert. The request to this effect must be filed by the applicant with the International Bureau before the completion of the technical preparations for international publication of the application.

Escherichia coli DB10B(pCMVSPORT6)

AUSTRALIA

The applicant hereby gives notice that the furnishing of a sample of a microorganism shall only be effected prior to the grant of a patent, or prior to the lapsing, refusal or withdrawal of the application, to a person who is a skilled addressee without an interest in the invention (Regulation 3.25(3) of the Australian Patents Regulations).

CANADA

The applicant hereby requests that, until either a Canadian patent has been issued on the basis of the application or the application has been refused, or is abandoned and no longer subject to reinstatement, or is withdrawn, the furnishing of a sample of deposited biological material referred to in the application only be effected to an independent expert nominated by the Commissioner of Patents.

DENMARK

The applicant hereby requests that, until the application has been laid open to public inspection (by the Danish Patent Office), or has been finally decided upon by the Danish Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the Danish Patent Office not later than at the time when the application is made available to the public under Sections 22 and 33(3) of the Danish Patents Act. If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the Danish Patent office or any person approved by the applicant in the individual case.

FINLAND

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Escherichia coli DB3.1(*pAHPKan*) or *Escherichia coli* DB3.1(*pAttPKan*)

AUSTRALIA

The applicant hereby gives notice that the furnishing of a sample of a microorganism shall only be effected prior to the grant of a patent, or prior to the lapsing, refusal or withdrawal of the application, to a person who is a skilled addressee without an interest in the invention (Regulation 3.25(3) of the Australian Patents Regulations).

CANADA

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DENMARK

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FINLAND

The applicant hereby requests that, until the application has been laid open to public inspection (by the National Board of Patents and Registration), or has been finally decided upon by the National Board of Patents and Registration without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the International Bureau before the expiration of 16 months from the priority date (preferably on the Form PCT/RO/134 reproduced in annex Z of Volume I of the PCT Applicant's Guide). If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the National Board of Patents and Registration or any person approved by the applicant in the individual case.

Escherichia coli DB3.1(pAHPKan) or *Escherichia coli* DB3.1(pAttPKan)

ICELAND

The applicant hereby requests that, until the application has been laid open to public inspection (by the Icelandic Patent Office), or has been finally decided upon by the Icelandic Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art.

NETHERLANDS

The applicant hereby requests that until the date of a grant of a Netherlands patent or until the date on which the application is refused or withdrawn or lapsed, the microorganism shall be made available as provided in Rule 31F(1) of the Patent Rules only by the issue of a sample to an expert. The request to this effect must be furnished by the applicant with the Netherlands Industrial Property Office before the date on which the application is made available to the public under Section 22C or Section 25 of the Patents Act of the Kingdom of the Netherlands, whichever of the two dates occurs earlier.

NORWAY

The applicant hereby requests that, until the application has been laid open to public inspection (by the Norwegian Patent Office), or has been finally decided upon by the Norwegian Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the Norwegian Patent Office not later than at the time when the application is made available to the public under Sections 22 and 33(3) of the Norwegian Patents Act. If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the Norwegian Patent office or any person approved by the applicant in the individual case.

SINGAPORE

The applicant hereby requests that the furnishing of a sample of a microorganism shall only be made available to an expert. The request to this effect must be filed by the applicant with the International Bureau before the completion of the technical preparations for international publication of the application.

Escherichia coli DB3.1(pAHPKan) or Escherichia coli DB3.1(pAttPKa λ)

SWEDEN

The applicant hereby requests that, until the application has been laid open to public inspection (by the Swedish Patent Office), or has been finally decided upon by the Swedish Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the International Bureau before the expiration of 16 months from the priority date (preferably on the Form PCT/RO/134 reproduced in annex Z of Volume I of the PCT Applicant's Guide). If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the Swedish Patent office or any person approved by the applicant in the individual case.

UNITED KINGDOM

The applicant hereby requests that the furnishing of a sample of a microorganism shall only be made available to an expert. The request to this effect must be filed by the applicant with the International Bureau before the completion of the technical preparations for international publication of the application.

Escherichia coli DB10B(pCMVSPORT6)

ICELAND

The applicant hereby requests that, until the application has been laid open to public inspection (by the Icelandic Patent Office), or has been finally decided upon by the Icelandic Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art.

NETHERLANDS

The applicant hereby requests that until the date of a grant of a Netherlands patent or until the date on which the application is refused or withdrawn or lapsed, the microorganism shall be made available as provided in Rule 31F(1) of the Patent Rules only by the issue of a sample to an expert. The request to this effect must be furnished by the applicant with the Netherlands Industrial Property Office before the date on which the application is made available to the public under Section 22C or Section 25 of the Patents Act of the Kingdom of the Netherlands, whichever of the two dates occurs earlier.

NORWAY

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SINGAPORE

The applicant hereby requests that the furnishing of a sample of a microorganism shall only be made available to an expert. The request to this effect must be filed by the applicant with the International Bureau before the completion of the technical preparations for international publication of the application.

Escherichia coli DB10B(pCMVSPORT6)

SWEDEN

The applicant hereby requests that, until the application has been laid open to public inspection (by the Swedish Patent Office), or has been finally decided upon by the Swedish Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the International Bureau before the expiration of 16 months from the priority date (preferably on the Form PCT/RO/134 reproduced in annex Z of Volume I of the PCT Applicant's Guide). If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the Swedish Patent office or any person approved by the applicant in the individual case.

UNITED KINGDOM

The applicant hereby requests that the furnishing of a sample of a microorganism shall only be made available to an expert. The request to this effect must be filed by the applicant with the International Bureau before the completion of the technical preparations for international publication of the application.

Escherichia coli DB3.1(pEZC15103)

AUSTRALIA

The applicant hereby gives notice that the furnishing of a sample of a microorganism shall only be effected prior to the grant of a patent, or prior to the lapsing, refusal or withdrawal of the application, to a person who is a skilled addressee without an interest in the invention (Regulation 3.25(3) of the Australian Patents Regulations).

CANADA

The applicant hereby requests that, until either a Canadian patent has been issued on the basis of the application or the application has been refused, or is abandoned and no longer subject to reinstatement, or is withdrawn, the furnishing of a sample of deposited biological material referred to in the application only be effected to an independent expert nominated by the Commissioner of Patents.

DENMARK

The applicant hereby requests that, until the application has been laid open to public inspection (by the Danish Patent Office), or has been finally decided upon by the Danish Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the Danish Patent Office not later than at the time when the application is made available to the public under Sections 22 and 33(3) of the Danish Patents Act. If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the Danish Patent office or any person approved by the applicant in the individual case.

FINLAND

The applicant hereby requests that, until the application has been laid open to public inspection (by the National Board of Patents and Registration), or has been finally decided upon by the National Board of Patents and Registration without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the International Bureau before the expiration of 16 months from the priority date (preferably on the Form PCT/RO/134 reproduced in annex Z of Volume I of the PCT Applicant's Guide). If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the National Board of Patents and Registration or any person approved by the applicant in the individual case.

Escherichia coli DB3.1(pEZC15103)

ICELAND

The applicant hereby requests that, until the application has been laid open to public inspection (by the Icelandic Patent Office), or has been finally decided upon by the Icelandic Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art.

NETHERLANDS

The applicant hereby requests that until the date of a grant of a Netherlands patent or until the date on which the application is refused or withdrawn or lapsed, the microorganism shall be made available as provided in Rule 31F(1) of the Patent Rules only by the issue of a sample to an expert. The request to this effect must be furnished by the applicant with the Netherlands Industrial Property Office before the date on which the application is made available to the public under Section 22C or Section 25 of the Patents Act of the Kingdom of the Netherlands, whichever of the two dates occurs earlier.

NORWAY

The applicant hereby requests that, until the application has been laid open to public inspection (by the Norwegian Patent Office), or has been finally decided upon by the Norwegian Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the Norwegian Patent Office not later than at the time when the application is made available to the public under Sections 22 and 33(3) of the Norwegian Patents Act. If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the Norwegian Patent office or any person approved by the applicant in the individual case.

SINGAPORE

The applicant hereby requests that the furnishing of a sample of a microorganism shall only be made available to an expert. The request to this effect must be filed by the applicant with the International Bureau before the completion of the technical preparations for international publication of the application.

Escherichia coli DB3.1(pEZC15103)

SWEDEN

The applicant hereby requests that, until the application has been laid open to public inspection (by the Swedish Patent Office), or has been finally decided upon by the Swedish Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the International Bureau before the expiration of 16 months from the priority date (preferably on the Form PCT/RO/134 reproduced in annex Z of Volume I of the PCT Applicant's Guide). If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the Swedish Patent office or any person approved by the applicant in the individual case.

UNITED KINGDOM

The applicant hereby requests that the furnishing of a sample of a microorganism shall only be made available to an expert. The request to this effect must be filed by the applicant with the International Bureau before the completion of the technical preparations for international publication of the application.

Escherichia coli DB3.1(pEZC15102)

AUSTRALIA

The applicant hereby gives notice that the furnishing of a sample of a microorganism shall only be effected prior to the grant of a patent, or prior to the lapsing, refusal or withdrawal of the application, to a person who is a skilled addressee without an interest in the invention (Regulation 3.25(3) of the Australian Patents Regulations).

CANADA

The applicant hereby requests that, until either a Canadian patent has been issued on the basis of the application or the application has been refused, or is abandoned and no longer subject to reinstatement, or is withdrawn, the furnishing of a sample of deposited biological material referred to in the application only be effected to an independent expert nominated by the Commissioner of Patents.

DENMARK

The applicant hereby requests that, until the application has been laid open to public inspection (by the Danish Patent Office), or has been finally decided upon by the Danish Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the Danish Patent Office not later than at the time when the application is made available to the public under Sections 22 and 33(3) of the Danish Patents Act. If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the Danish Patent office or any person approved by the applicant in the individual case.

FINLAND

The applicant hereby requests that, until the application has been laid open to public inspection (by the National Board of Patents and Registration), or has been finally decided upon by the National Board of Patents and Registration without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the International Bureau before the expiration of 16 months from the priority date (preferably on the Form PCT/RO/134 reproduced in annex Z of Volume I of the PCT Applicant's Guide). If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the National Board of Patents and Registration or any person approved by the applicant in the individual case.

Escherichia coli DB3.1(pEZC15102)

ICELAND

The applicant hereby requests that, until the application has been laid open to public inspection (by the Icelandic Patent Office), or has been finally decided upon by the Icelandic Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art.

NETHERLANDS

The applicant hereby requests that until the date of a grant of a Netherlands patent or until the date on which the application is refused or withdrawn or lapsed, the microorganism shall be made available as provided in Rule 31F(1) of the Patent Rules only by the issue of a sample to an expert. The request to this effect must be furnished by the applicant with the Netherlands Industrial Property Office before the date on which the application is made available to the public under Section 22C or Section 25 of the Patents Act of the Kingdom of the Netherlands, whichever of the two dates occurs earlier.

NORWAY

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SINGAPORE

The applicant hereby requests that the furnishing of a sample of a microorganism shall only be made available to an expert. The request to this effect must be filed by the applicant with the International Bureau before the completion of the technical preparations for international publication of the application.

Escherichia coli* DB3.1(pEZC15102)*SWEDEN**

The applicant hereby requests that, until the application has been laid open to public inspection (by the Swedish Patent Office), or has been finally decided upon by the Swedish Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the International Bureau before the expiration of 16 months from the priority date (preferably on the Form PCT/RO/134 reproduced in annex Z of Volume I of the PCT Applicant's Guide). If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the Swedish Patent office or any person approved by the applicant in the individual case.

UNITED KINGDOM

The applicant hereby requests that the furnishing of a sample of a microorganism shall only be made available to an expert. The request to this effect must be filed by the applicant with the International Bureau before the completion of the technical preparations for international publication of the application.

Escherichia coli DB3.1(pEZC15101)**AUSTRALIA**

The applicant hereby gives notice that the furnishing of a sample of a microorganism shall only be effected prior to the grant of a patent, or prior to the lapsing, refusal or withdrawal of the application, to a person who is a skilled addressee without an interest in the invention (Regulation 3.25(3) of the Australian Patents Regulations).

CANADA

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Escherichia coli DB3.1(pEZC15101)

ICELAND

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NORWAY

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SINGAPORE

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Escherichia coli DB3.1(pEZC15101)

SWEDEN

The applicant hereby requests that, until the application has been laid open to public inspection (by the Swedish Patent Office), or has been finally decided upon by the Swedish Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the International Bureau before the expiration of 16 months from the priority date (preferably on the Form PCT/RO/134 reproduced in annex Z of Volume I of the PCT Applicant's Guide). If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the Swedish Patent office or any person approved by the applicant in the individual case.

UNITED KINGDOM

The applicant hereby requests that the furnishing of a sample of a microorganism shall only be made available to an expert. The request to this effect must be filed by the applicant with the International Bureau before the completion of the technical preparations for international publication of the application.

Escherichia coli DB3.1(pENTR-3C)**AUSTRALIA**

The applicant hereby gives notice that the furnishing of a sample of a microorganism shall only be effected prior to the grant of a patent, or prior to the lapsing, refusal or withdrawal of the application, to a person who is a skilled addressee without an interest in the invention (Regulation 3.25(3) of the Australian Patents Regulations).

CANADA

The applicant hereby requests that, until either a Canadian patent has been issued on the basis of the application or the application has been refused, or is abandoned and no longer subject to reinstatement, or is withdrawn, the furnishing of a sample of deposited biological material referred to in the application only be effected to an independent expert nominated by the Commissioner of Patents.

DENMARK

The applicant hereby requests that, until the application has been laid open to public inspection (by the Danish Patent Office), or has been finally decided upon by the Danish Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the Danish Patent Office not later than at the time when the application is made available to the public under Sections 22 and 33(3) of the Danish Patents Act. If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the Danish Patent office or any person approved by the applicant in the individual case.

FINLAND

The applicant hereby requests that, until the application has been laid open to public inspection (by the National Board of Patents and Registration), or has been finally decided upon by the National Board of Patents and Registration without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the International Bureau before the expiration of 16 months from the priority date (preferably on the Form PCT/RO/134 reproduced in annex Z of Volume I of the PCT Applicant's Guide). If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the National Board of Patents and Registration or any person approved by the applicant in the individual case.

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US00/05432

A. CLASSIFICATION OF SUBJECT MATTER IPC(7) : Please See Extra Sheet. US CL : 435/91.2, 252.3, 320.1; 530/350; 536/ 23.1, 24.1 According to International Patent Classification (IPC) or to both national classification and IPC		
B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) U.S. : 435/91.2, 252.3, 320.1; 530/350; 536/ 23.1, 24.1 Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched NONE Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) Please See Extra Sheet.		
C. DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X,P --- Y,P	US 5,888,732 A (HARTLEY et al.) 30 March 1999, see entire document.	1-21, 25-30 36-38 ----- 22-24, 31-35
X - Y	HASAN et al. Escherichia coli genome targeting, I. Cre-lox-mediated in vitro generation of ori- plasmids and their in vivo chromosomal integration and retrieval. Gene. 1994, Vol. 150, pages 51-56, see entire document.	1-5, 10, 11, 19-21 ----- 15-18, 22-38
X - Y	KATZ et al. Site-specific recombination in Escherichia coli between the att sites of plasmid pSE211 from Saccharopolyspora erythraea. Mol. Gen. Genet. 1991, Vol. 227, pages 155-159, see entire document.	1-11, 19-21 ----- 15-18, 22-38
<input checked="" type="checkbox"/> Further documents are listed in the continuation of Box C. <input type="checkbox"/> See patent family annex.		
* Special categories of cited documents: *A* document defining the general state of the art which is not considered to be of particular relevance *E* earlier document published on or after the international filing date *L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) *O* document referring to an oral disclosure, use, exhibition or other means *P* document published prior to the international filing date but later than the priority date claimed *T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention *X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone *Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art *A* document member of the same patent family		
Date of the actual completion of the international search 08 MAY 2000		Date of mailing of the international search report 23 MAY 2000
Name and mailing address of the ISA/US Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231 Facsimile No. (703) 305-3230		Authorized officer <i>Ortalia Lawrence</i> IREM YUCEL Telephone No. (703) 308-0196

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US00/05432

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X - Y	ASTUMIAN et al. Site-specific recombination between cloned attP and attB sites from the Haemophilus influenzae bacteriophage HP1 propagated in recombination deficient Escherichia coli. J of Bacteriology. March 1989, Vol. 171, No. 3, pages 1747-1750, see entire document.	1-11, 19-21 ----- 15-18, 22-38

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US00/05432

A. CLASSIFICATION OF SUBJECT MATTER:

IPC (7):

C07H 21/04; C07K 1/00, 14/00; C12N 1/21, 15/00, 15/09, 15/63, 15/70; C12P 19/34

B. FIELDS SEARCHED

Electronic data bases consulted (Name of data base and where practicable terms used):

WEST, STN (CAPLUS); DIALOG (MEDLINE, BIOSIS, SCISEARCH, PASCAL)

Terms: att (B?, P?, R?, L?), MCS, POLYLINKER, PLASMID, VECTOR, LOCALIZATION, SIGNAL, TRANSCRIPTION, TERMIN?, TRANSLATION?, ORI, REPLICON, GST, HEXHIST?, THIOREDOX?, CLEAVAGE, SITE?, SPECIF?, DIRECT?, RECOMBIN?, CLON?, INSERT?